



# Draft Genome Sequences of *Pseudomonas* sp. Strains MWU 12-2029, MWU 12-3088, and MWU 12-3091, Isolated from Wild and Cultivated Massachusetts Cranberry Bogs

 Akhila Manthena,<sup>a</sup>  Arin Pittman,<sup>a</sup>  Scott Soby<sup>b</sup>

<sup>a</sup>Arizona College of Osteopathic Medicine, Midwestern University, Glendale, Arizona, USA

<sup>b</sup>Biomedical Sciences, College of Graduate Studies, and College of Veterinary Medicine, Midwestern University, Glendale, Arizona, USA

**ABSTRACT** *Pseudomonas* sp. strains MWU12-2029, MWU12-3088, and MWU12-3091 were isolated from wild and cultivated cranberry bog soils in southeastern Massachusetts. The three isolates are closely related to *Pseudomonas kribbensis*, a not validly published member of the *P. fluorescens* group, and contain three putative insecticidal protein genes, including the toxin complex A gene (*tcaC*).

*Pseudomonas* spp. comprise a considerable component of bacteria isolated from cultivated and wild cranberry bogs during a multiyear culture-dependent survey conducted at the Cape Cod National Seashore (MWU12-2029) (42.070624N, 70.210548W) and the University of Massachusetts State Bog (MWU12-3088 and MWU12-3091) (41.766767N, 70.66842W) (1–8). Although presumably an important constituent of these wetland microbiomes, little is known about how these organisms affect the dynamics of the bog ecosystem, including their effects on insects. As an initial foray into understanding the cranberry bog soil microbiome, 5 cm by 5 cm soil samples were taken from cultivated and wild bogs in July 2012 for isolating and characterizing bacterial populations. Approximately 1 g from soil cores was vortexed in sterile water, and the rinsate was plated onto King's medium B (KMB) agar supplemented with 50  $\mu\text{g mL}^{-1}$  each of cycloheximide and ampicillin and incubated at 26°C for 48 h. Fluorescent colonies were purified three times on KMB and stored at –80°C in 34% glycerol. Isolates from frozen storage were recovered on KMB agar, and populations were inoculated into KMB broth cultures grown overnight for genomic DNA (gDNA) isolation. All kits described below were used according to the manufacturers' instructions. gDNA was extracted with a DNeasy blood and tissue kit (Qiagen, USA), and Kapa Biosystems Hyperplus library preparation kits (catalog number KK8514; Roche, USA) were used to generate Illumina-compatible genomic DNA libraries: DNA was enzymatically sheared to ~500 bp, end repaired, and A tailed; Illumina-compatible adapters with unique indexes (catalog number 00989130v2; IDT, Coralville, IA) were then ligated to each sample; and adapter-ligated molecules were cleaned using Kapa pure beads (catalog number KK8002) and amplified with Kapa Hifi enzyme (catalog number KK2502). Library fragment sizes were determined on an Agilent TapeStation system and quantified by quantitative PCR (qPCR) (Kapa library quantification kit, catalog number KK4835) on a QuantStudio 5 system (Thermo Fisher, USA). Samples were multiplex pooled and sequenced on an Illumina MiSeq platform in a 2-by-250 flow cell. The software was set to default settings except as indicated below. Raw reads were assembled with Unicycler v0.4.8 (9) and polished with Pilon v1.23 (10) within the PATRIC v3.6.12 comprehensive genome analysis pipeline, except for the trim setting, which was set to "true" (11). The comprehensive analysis pipeline includes quality control and trimming by QAST v5.0.2 (12) and Trim Galore v0.4.0 (13) and annotation by RASTtk v1.073 (14), supplemented with antiSMASH v6.0 (15) for the recognition of secondary metabolite gene clusters. Using the Type (Strain) Genome Server (TYGS), isolates were placed with high confidence within the genus *Pseudomonas* (16). All three isolates were most closely related to *Pseudomonas kribbensis* 46-2<sup>T</sup> (GenBank accession number [CP029608](https://doi.org/10.1128/mra.00889-22))

**Editor** Catherine Putonti, Loyola University Chicago

**Copyright** © 2022 Manthena et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Scott Soby, [ssobyx@midwestern.edu](mailto:ssobyx@midwestern.edu).

The authors declare no conflict of interest.

**Received** 25 August 2022

**Accepted** 2 October 2022

**Published** 17 October 2022

**TABLE 1** Genomic data summary

| Isolate    | BioSample<br>accession no.   | GenBank<br>accession no.         | SRA<br>accession no.        | Genome<br>size (bp) | No. of<br>contigs | $N_{50}$<br>(bp) | G+C<br>content (%) | Mean read<br>length (bp) | No. of<br>reads | Coverage<br>(x) | dDDH <sub>46-2</sub> /ANIb<br>with <i>P. kribbensis</i><br>46-2 <sup>T</sup> (%) |
|------------|------------------------------|----------------------------------|-----------------------------|---------------------|-------------------|------------------|--------------------|--------------------------|-----------------|-----------------|--|
| MWU12-2029 | <a href="#">SAMN26814158</a> | <a href="#">JALJDY0000000000</a> | <a href="#">SRR18644892</a> | 6,318,446           | 38                | 583,019          | 60.68              | 233.75                   | 3,403,188       | 126             | 65.1/95.48   |
| MWU12-3088 | <a href="#">SAMN26803934</a> | <a href="#">JALJEA0000000000</a> | <a href="#">SRR18645887</a> | 6,305,590           | 37                | 392,904          | 60.67              | 229.72                   | 3,774,982       | 138             | 65.6/95.65   |
| MWU12-3091 | <a href="#">SAMN26896969</a> | <a href="#">JALJET0000000000</a> | <a href="#">SRR18508983</a> | 6,305,411           | 38                | 392,833          | 60.67              | 224.57                   | 3,190,686       | 114             | 65.6/95.64   |

but fell below the 70% digital DNA-DNA hybridization (dDDH<sub>ca</sub>) (TYGS v342) (16–18) or 95 to 96% average nucleotide identity by BLAST analysis (ANIb) (JSpeciesWS v3.9.5) (19, 20) cutoff to be included in that species (Table 1).

MWU12-2029, MWU12-3088, and MWU12-3091 all contain presumptive genes for insecticidal toxin complex A (*tcaC*) (21) and two additional putative insecticidal toxins that are widespread among members of the *Pseudomonas fluorescens* group.

**Data availability.** The whole-genome shotgun sequencing project has been deposited in the DDBJ/EMBL/GenBank database under BioProject accession number [PRJNA691338](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA691338) with the accession numbers in Table 1. The versions described in this paper are GenBank accession numbers [JALJDY000000000.1](https://www.ncbi.nlm.nih.gov/nuclot/JALJDY000000000.1) (MWU12-2029), [JALJEA000000000.1](https://www.ncbi.nlm.nih.gov/nuclot/JALJEA000000000.1) (MWU12-3088), and [JALJET000000000.1](https://www.ncbi.nlm.nih.gov/nuclot/JALJET000000000.1) (MWU12-3091). RASTtk annotations are available under an open license at Zenodo (see <https://zenodo.org/record/6413129#.YwaiEUfMKUK>, <https://zenodo.org/record/6413119#.Ywah6kfMKUK>, and <https://zenodo.org/record/6413231#.YwainOfMKUK>).

## ACKNOWLEDGMENTS

This research was supported by the Office of Research and Sponsored Programs, the College of Graduate Studies, and the Biomedical Sciences Program, Midwestern University.

We appreciate and acknowledge Alisha Harrison for the isolation of genomic DNA. Library construction and Illumina sequencing were performed at the Arizona State University Genomics Core Facility.

This work fulfills a course requirement for Special Topics in Bacterial Genomics for A.M. and A.P.

## REFERENCES

- Ebadzadsahrai G, Soby S. 2020. Draft genome sequence of *Pseudomonas* sp. strain MWU12-2323, isolated from a wild cranberry bog in Truro, Massachusetts. *Microbiol Resour Announc* 9(2):e01387-19. <https://doi.org/10.1128/MRA.01387-19>.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequences of *Pseudomonas* MWU13-2625 and MWU12-2115, isolated from a wild cranberry bog at the Cape Cod National Seashore. *Microbiol Resour Announc* 7(12):e00992-18. <https://doi.org/10.1128/MRA.00992-18>.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequence of *Pseudomonas* sp. strain MWU12-2534b, isolated from a wild cranberry bog in Truro, Massachusetts. *Microbiol Resour Announc* 7(13):e01005-18. <https://doi.org/10.1128/MRA.01005-18>.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequence of *Pseudomonas* sp. strain MWU13-2860, isolated from a wild cranberry bog in Truro, Massachusetts. *Microbiol Resour Announc* 7(13):e01007-18. <https://doi.org/10.1128/MRA.01007-18>.
- Yaeger J, Soby S. 2022. Draft genome sequence of *Pseudomonas* sp. strain MWU15-20650, isolated from a wild cranberry bog in Provincetown, MA. *Microbiol Resour Announc* 11(8):e00545-22. <https://doi.org/10.1128/mra.00545-22>.
- Anasi A, Soby S. 2022. Draft genome sequences of *Pseudomonas* sp. strains MWU12-2037 and MWU12-2345, isolated from peat and sandy bog soils in the Cape Cod National Seashore, Massachusetts. *Microbiol Resour Announc* 11(8):e00536-22. <https://doi.org/10.1128/mra.00536-22>.
- Sholl T, Soby S. 2022. Draft genome sequence of *Pseudomonas* sp. strain MWU15-20650, isolated from wild cranberry fruit in the Cape Cod National Seashore. *Microbiol Resour Announc* 11(8):e00547-22. <https://doi.org/10.1128/mra.00547-22>.
- Koszewski MH, Motevalli S, Soby SD. 2021. Draft genome sequences of *Pseudomonas* spp. isolated from berry surfaces in commercial cranberry bogs in Massachusetts, USA. *Microbiol Resour Announc* 10(27):e00204-21. <https://doi.org/10.1128/MRA.00204-21>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial bioinformatics database and analysis resource center. *Nucleic Acids Res* 45:D535–D542. <https://doi.org/10.1093/nar/gkw1017>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Krueger F. 2014. Trim Galore: a wrapper tool around Cutadapt and FastQC to consistently apply quality and adapter trimming to FastQ files, with some extra functionality for MspI-digested RRBS-type (reduced representation bisulfite [sic]-Seq) libraries. [https://www.bioinformatics.babraham.ac.uk/projects/trim\\_galore/](https://www.bioinformatics.babraham.ac.uk/projects/trim_galore/).
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Blin K, Shaw S, Kloosterman AM, Charlop-Powers Z, van Wezel GP, Medema MH, Weber T. 2021. antiSMASH 6.0: improving cluster detection and comparison capabilities. *Nucleic Acids Res* 49:W29–W35. <https://doi.org/10.1093/nar/gkab335>.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.
- Wayne LG, Good RC, Krichevsky MI, Blacklock Z, David HL, Dawson D, Gross W, Hawkins J, Vincent Levy-Frebault V, McManus C, Portaels F, Rusch-Gerdes S, Schroder KH, Silcox VA, Tsukamura M, Van Den Breen L, Yakrus MA. 1991. Report of the Ad Hoc Committee on Reconciliation of Approaches to Bacterial Systematics. *Int J Syst Bacteriol* 41:463–472. <https://doi.org/10.1099/00207713-41-4-463>.
- Auch AF, von Jan M, Klenk H-P, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome

- sequence comparison. *Stand Genomic Sci* 2:117–134. <https://doi.org/10.4056/sigs.531120>.
19. Thompson CC, Chimetto L, Edwards RA, Swings J, Stackebrandt E, Thompson FL. 2013. Microbial genomic taxonomy. *BMC Genomics* 14:913. <https://doi.org/10.1186/1471-2164-14-913>.
  20. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
  21. Blackburn M, Golubeva E, Bowen D, ffrench-Constant R. 1998. A novel insecticidal toxin from *Photobacterium luminescens*: histopathological effects of toxin complex A (Tca) on the midgut of *Manduca sexta*. *Appl Environ Microbiol* 64:3036–3041. <https://doi.org/10.1128/AEM.64.8.3036-3041.1998>.