

RESEARCH ARTICLE

Open Access

Epigenetic regulation of matrix metalloproteinase expression in ameloblastoma

Lucyana Conceição Farias¹, Carolina Cavaliéri Gomes², Marcela Carolina Rodrigues¹, Wagner Henriques de Castro¹, Júlio César Tanos Lacerda³, Efigênia Ferreira e Ferreira⁴ and Ricardo Santiago Gomez^{1*}

Abstract

Background: An ameloblastoma is a benign odontogenic neoplasm with aggressive behaviour and high recurrence rates. The increased expression of matrix metalloproteinases (MMPs) has been reported in ameloblastomas. In the present study, we hypothesised that epigenetic alterations may regulate *MMP* expression in ameloblastomas.

Methods: We investigated the methylation status of the genes *MMP-2* and *MMP-9* in addition to mRNA transcription and protein expression in ameloblastomas. Methylation analysis was performed by both methylation-specific polymerase chain reaction (MSP-PCR) and restriction enzyme digestion to evaluate the methylation profile of *MMP-2* and *MMP-9* in 12 ameloblastoma samples and 12 healthy gingiva fragments, which were included as controls. Furthermore, we investigated the transcription levels of the genes by quantitative reverse-transcription PCR (qRT-PCR). Zymography was performed to verify protein expression in ameloblastomas.

Results: The ameloblastomas showed a high frequency of unmethylated *MMP-2* and *MMP-9*, whereas the healthy gingival samples presented a sharp prevalence of methylated MMPs. Higher expression levels of *MMP-9* were found in ameloblastomas compared to healthy gingiva. However, no significant differences in the *MMP-2* mRNA expression between groups was found. All ameloblastomas showed positive expression of *MMP-2* and *MMP-9* proteins.

Conclusions: Our findings suggest that expression of *MMP-9* is increased in ameloblastomas and is possibly modulated by unmethylation of the gene.

Keywords: Ameloblastoma, Odontogenic tumours, Matrix metalloproteinases, *MMP-2*, *MMP-9*, Methylation, Epigenetic

Background

An ameloblastoma is a benign odontogenic tumour that exhibits a high recurrence risk, aggressive behaviour and local invasiveness [1]. Histologically, an ameloblastoma consists of epithelial strands or islands of ameloblastic epithelium. The peripheral cells are columnar, while the cells lying more centrally are fusiform to polyhedral and are loosely connected to each other [1].

Different studies have demonstrated genetic alterations in odontogenic tumours [2-4], but few studies have

analysed epigenetic events in these tumours [5-7]. Methylation is an epigenetic alteration that plays an important role in controlling gene activity, embryonic development, and genomic imprinting. It has been associated with gene silencing by transcriptional inactivation [8]. DNA methylation or hypomethylation of the *p16*, *p21* and *LINE-1* genes was reported in ameloblastomas by our group and others [5,9,10], but the significance of this data remains to be determined.

Matrix metalloproteinases (MMPs) are zinc-dependent enzymes that are important in extracellular matrix remodelling and are associated with tumour growth and invasion through collagen matrix degradation [11]. The invasive characteristic of ameloblastomas has been associated with the expression of genes related to bone turnover and

* Correspondence: rsgomez@ufmg.br

¹Department of Oral Surgery and Pathology, School of Dentistry, Universidade Federal de Minas Gerais, Av. Antonio Carlos, 6627, CEP 31270 901 Belo Horizonte, Minas Gerais, Brazil

Full list of author information is available at the end of the article

extracellular matrix remodelling; these include *BMP*, *RANKL* and its receptor, *MMP* and *TIMP* [12-17]. As MMPs may be regulated by DNA methylation in malignant neoplasms [18,19], such phenomenon might be important in ameloblastoma pathogenesis and should be investigated. Therefore, the purpose of this study was to investigate the association between *MMP-2* and *MMP-9* methylation and their mRNA transcription and protein expression in ameloblastomas.

Methods

Patients and tissue samples

Twelve fresh ameloblastoma specimens were collected during surgical care in the Department of Oral Surgery and Pathology, Universidade Federal de Minas Gerais, Brazil. These samples comprised eleven solid-multicystic follicular ameloblastomas and one unicystic case. Diagnoses were confirmed by histopathologic analysis based on the World Health Organization classification of histological typing of odontogenic tumours [1]. Other clinical data are shown in Table 1. Twelve fragments of healthy gingival samples with no clinical evidence of inflammation were collected during third molar extractions and used as controls. The samples were obtained following informed consent and with the approval of the *Universidade Federal de Minas Gerais* Ethics Committee (reference number 266/11).

DNA isolation and methylation analysis of *MMP-2* and *MMP-9*

Genomic DNA was isolated from the tissue samples using a Qiagen DNeasy Tissue Kit (Qiagen Inc., Valencia, CA, USA) according to the manufacturer's instructions. *Meth-Primer* software [20] was used to search CpG islands and sparse CG dinucleotides. Distinct methods are suggested to analyse methylation profiles according to the presence

Table 1 Distribution of subjects according to gender, age and anatomic site

Parameters	Ameloblastoma (n = 12)	Healthy gingiva (n = 12)
Gender		
Male	05 (41.7%)	07 (58.3%)
Female	07 (58.3%)	05 (41.7%)
Age (years)		
Variation	8-51	19-28
Mean ± SD	31.0 ± 14.6	25.4 ± 5.3
Anatomic Sites		
Mandible		
anterior region	01 (8.3%)	0
posterior region	11 (91.7%)	4 (33.3%)
Maxilla	0	8 (66.7%)

of CpG islands or sparse CG dinucleotides located in the promoter region or in exons near to that region [21].

To assess the *MMP-2* gene CpG island methylation, genomic DNA was modified by sodium bisulfite as described previously [6] and subsequently amplified with primer sets designed to specifically recognise methylated (F 5'-GCG GTTATACGTATCGAGTTAGC-3' and R 5'-ACTCTTTA TCCGTTTTAAAAACGAC-3'; 205 bp) and unmethylated DNA (F 5'-GGTGGTTATATGTATTGAGTTAGTGA-3' and R 5'-ACTCTTTATCCATTTAAAAACAAC-3' 206 bp). Bisulfite-treated unmethylated DNA from (peripheral blood mononuclear cells) cells was used as a positive control for unmethylated amplification of the *MMP-2* gene. Methylation-induced DNA of same cells by the MSssI methylase enzyme (New England Biolabs, Beverly, USA) was used as positive control for methylated amplification.

The methylation-sensitive restriction enzymes HhaI and AclI (New England BioLabs, Beverly, MA, USA) were used to assess the methylation of CG dinucleotides in the *MMP-9* promoter, including the CG sites located at positions -35, -185, -223, -233, as described previously [21]. Restriction enzymes cleave DNA at unmethylated CG sites, but they are unable to cut methylated cytosines. Analysis using a bioinformatics web site (<http://www.restrictionmapper.org>) showed that the HhaI enzyme cleaves the restriction site at position -35 and that the other sites are cleaved by AclI. The CG dinucleotides analysed in this study are located close to the transcription start of the *MMP-9* gene. Two hundred nanograms of genomic DNA was digested separately with each of the restriction enzymes HhaI and AclI according to manufacturer's protocol to cleave the specific regions containing CG sites (New England BioLabs, Beverly, MA, USA). Digestion was followed by PCR amplification (primers: F 5'-GCTTCATCCCCCTCCCTCC-3'; R 5'-AG CACCAGGACCAGGGGC-3'; 369 bp). PCR products were subjected to electrophoresis in 6.5% polyacrylamide gels. While methylated cytosine produces a band equivalent to that of control methylated DNA of placenta tissue, the cleavage by restriction enzyme at unmethylated CpG induces DNA strand breaks, and thus no band is detected. In each PCR reaction, undigested DNA of each sample was also carried out as controls. Undigested and digested PCR products were electrophoresed in parallel. Human unmethylated DNA (Qiagen Inc., Valencia, CA, USA), which is sensitive to action of the enzyme, was also used as unmethylated positive control.

RNA extraction and Quantitative Real-time PCR (qRT-PCR) of *MMP-2* and *MMP-9*

Total RNA was extracted from tissue samples using Trizol reagent according to the manufacturer's protocol (Invitrogen, Carlsbad, CA, USA). RNA integrity was analysed by

1% agarose gel electrophoresis. Reverse transcription of 1 µg of RNA to cDNA was performed using SuperScript III First-Strand (Invitrogen, Carlsbad, CA, USA) following the manufacturer's instructions. Primer sequences were designed using the *PrimerExpress* software (v.3, Applied Biosystems) as follows: *MMP-2* (F: 5'-AGCTCCCGG AAAAGATTGATG-3'; R: 5'-CAGGGTGCTGGCTG AGTAGAT-3'; 101 bp) and *MMP-9* (F: 5'-GAGGTT CGACGTGAAGGCGCAGAT-3'; R: 5'-CATAGGTCACG TAGCCCACT TGGT3'; 200 bp). All reactions were run in duplicate in a StepOne Real time PCR System using the SYBR-green fluorescence quantification system (Applied Biosystems, Warrington, UK). The comparative C_t method was used [22]. Expression levels of the *MMP-2* and *MMP-9* genes relative to a calibrator sample (placenta tissue) were obtained by normalisation to endogenous β -actin.

Gelatin zymography

Ameloblastoma protein was extracted and subjected to electrophoresis under nonreducing conditions on SDS-polyacrylamide gels copolymerised with 1 mg/ml gelatin (Sigma Chemical Co, St Louis, MO, USA) as previously described [23]. After electrophoresis, the gels were washed in 2.5% Triton-X 100 and incubated for at least 18 h at 37°C in incubation buffer (50 mM Tris-HCl, pH 7.5, containing 5 mM CaCl₂, 100 mM NaCl, 0.01% Triton X-100). Zymographic gels were stained in 0.2% Coomassie Brilliant Blue R-250 and de-stained. The gels were scanned to analyse the bands representative of MMPs, according to molecular weight. Analysis of protein expression in healthy gingiva was not performed due to the scarcity of tissue samples.

Statistical analysis

Mann-Whitney tests were used to compare the relative quantification of *MMP-2* and *MMP-9* between groups. Chi-squared or Fisher's exact were used when appropriate. The analyses were carried out using SPSS 17.0

software, and probability values <0.05 were considered statistically significant.

Results

MMP-2 and *MMP-9* methylation statuses are shown in Table 2 and represented in Figure 1. While all healthy gingival samples showed *MMP-2* methylation, approximately half of ameloblastomas were unmethylated. Similarly, an increased frequency of unmethylated *MMP-9* of specific CG sites digested by HhaI was identified in the ameloblastomas. Almost all of the ameloblastoma samples showed an unmethylated profile for *MMP-9*. No difference was found in the methylation of CG sites digested by AciI among the groups studied.

The qRT-PCR results are summarised in Figures 2a and 2b. Higher expression levels of *MMP-9* were found in ameloblastomas compared to healthy gingiva. However, significant differences in the *MMP-2* mRNA expression levels were not found.

When we investigated the influence of the methylation status of both genes on their transcription, no association was found between *MMP-2* transcription and its methylation in ameloblastomas ($p = 0.319$). Almost all of the tumour samples showed an unmethylated *MMP-9* pattern in conjunction with increased mRNA levels. As most of the ameloblastomas were unmethylated at the *MMP-9* gene, considering all of the restriction sites, it was not possible to statistically compare the transcription of the gene in the cases with or without methylated sequences.

All of the ameloblastoma samples showed expression of *MMP-2* and *MMP-9* proteins, as verified by zymography (Figure 3). However, pro-*MMP-2* and pro-*MMP-9* forms were not identified in ameloblastomas.

Discussion

The underlying molecular pathways associated with the pathogenesis of ameloblastomas are not well established yet. Previous investigations have assessed the molecular

Table 2 Methylation status of *MMP-2* and *MMP-9* genes in ameloblastoma and healthy gingiva

Parameters	Ameloblastoma (n = 12)	Healthy gingiva (n = 12)	<i>p</i> -value*
MMP-2 methylation			
Methylated	05 (41.7%)	12 (100.0%)	0.002*
Unmethylated	07 (58.3%)	0 (0.0%)	
MMP-9 methylation/restriction site digested by HhaI			
Methylated	0 (0.0%)	06 (50.0%)	0.006*
Unmethylated	12 (100.0%)	06 (50.0%)	
MMP-9 methylation/restriction site digested by AciI			
Methylated	01 (8.3%)	0 (0.0%)	0.317
Unmethylated	11 (91.7%)	12 (100.0%)	

* Fisher's exact test ($p < 0.05$).

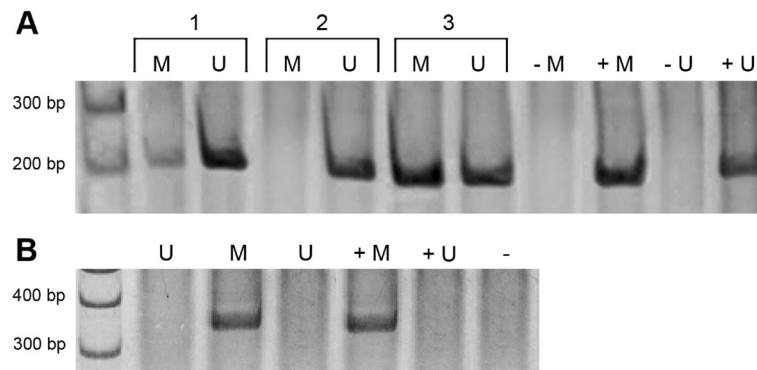


Figure 1 Representative figure of the methylation analysis. **A:** Methylation status of *MMP-2* in ameloblastoma. M: PCR products when amplified by methylated primers (205 bp); U: PCR products when amplified by unmethylated primers (206 bp); +M: positive control for methylated reaction; +U: positive control for unmethylated reaction. -M and -U: negative controls without DNA. Lines 1 to 3 represent DNA from ameloblastoma samples. **B:** Methylation status of *MMP-9* in ameloblastoma. DNA samples were digested by the *AclI* restriction enzyme followed by PCR, flanking the restriction sites. Absent band indicates unmethylated profile (U) due to DNA cleavage by the restriction enzyme. Presence of the PCR band represents methylated profile (M) of the *MMP-9* gene. +M: methylated positive control; +U: unmethylated positive control; -: negative control without DNA.

and genetic alterations related mainly to apoptosis, allelic loss of tumour suppressor genes, deregulation of the Sonic Hedgehog signalling pathway, and the clonality of these tumours [3,24,25].

Matrix metalloproteinases are involved in the degradation of collagen, as well as bone matrix, and have been shown to play a key role in the local invasiveness of ameloblastoma cells [15,26]. Overexpression of *MMP-2* and *MMP-9* was associated with the infiltrative behaviour of ameloblastomas, as well as that of several malignant neoplasms [17,27]. The suppression of *MMP-2* activity was able to inhibit the invasiveness of ameloblastoma cells *in vitro* [14,15]. Furthermore, it was suggested that increased expression of *MMP-9* may be involved in the proliferation and invasive behaviour of ameloblastomas [26].

Some papers, including studies from our research group, have demonstrated epigenetic alterations in

odontogenic tumours [5,6,9,10,28]. In the present study, we hypothesised that methylation may regulate the expression of *MMP-2* and *MMP-9* in ameloblastomas. We also investigated the association between methylation and the transcription levels of these genes. As most of the ameloblastoma samples were of the solid follicular type, we were not able to analyse possible associations between each clinical or histological type and the molecular data.

MMPs play an important role in collagen matrix remodelling in physiologic and pathologic processes, such as those found in basal membranes, dental follicle tissue and tumour metastasis [27,29]. Although *MMP-2* is related to ameloblastoma pathogenesis, it seems to be constitutively expressed in physiologic tissues and many cell types and to exhibit characteristics of a housekeeping gene [30-32]. Perhaps this could explain the similar

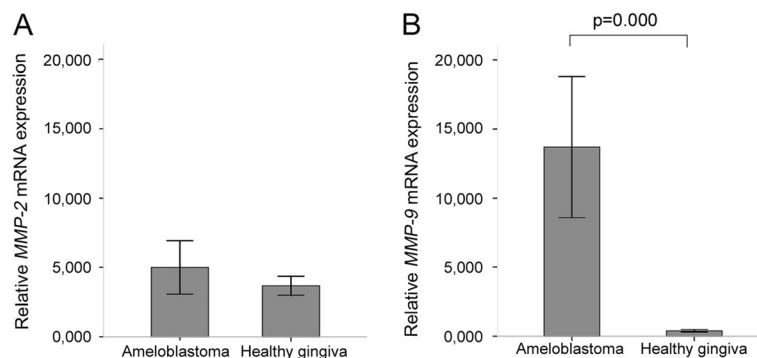
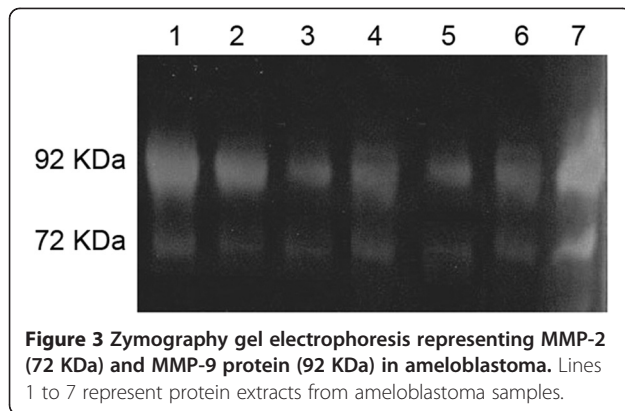


Figure 2 qRT-PCR results of *MMP-2* (A) and *MMP-9* (B) mRNA transcription in ameloblastoma and healthy gingiva. In both analyses, gene expression is shown as the mean \pm SE. All reactions were normalised to β -actin and are relative to a calibrator sample. * Mann-Whitney test ($p < 0.05$).



expression levels of *MMP-2* mRNA in ameloblastomas and healthy gingiva. Furthermore, our data suggest that *MMP-2* expression in ameloblastomas may not be modulated by methylation because the methylation profile for this gene did not correlate with *MMP-2* transcript levels in this odontogenic tumour.

The ameloblastomas presented an unmethylated profile of *MMP-2* and *MMP-9* genes compared to gingiva. Furthermore, along with unmethylated *MMP-9*, this tumour showed increased transcription of *MMP-9* when compared to the control group. The important role of methylation in epigenetic silencing is well established, particularly through regulatory mechanisms of transcription. Accordingly, our data suggest that an unmethylated profile of the *MMP-9* gene in ameloblastomas may be a permissive event allowing the binding of transcription factors to DNA, thus favouring *MMP-9* gene transcription.

All of the ameloblastomas showed *MMP-9* protein expression and were mostly unmethylated for *MMP-9*, so it was not possible to assess if the transcription of the gene was correlated with its methylation status. However, our study suggests that the increased transcription of *MMP-9* in ameloblastomas could possibly be influenced by unmethylation of the gene. The evident protein expression, identified by zymography, provides additional evidence supporting the possible gene regulation by unmethylated *MMP-9*. It is interesting to note that hypomethylation of the *MMP-2* and *MMP-9* genes increases gene expression and contributes to cancer cell invasiveness and tumorigenesis in malignant neoplasms, such as prostate cancer and lymphoma [18,33].

Conclusion

In conclusion, our study provides new insights into the epigenetic regulation of MMPs in ameloblastomas and points to the hypomethylation of *MMP-9* as a possible mechanism involved in the increased transcription of the gene in this tumour. However, functional studies are needed to better explain the role the methylation of

matrix metalloproteinases plays in the pathogenesis of ameloblastoma.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RSG, CCG, and LCF participated in the study design and drafted the manuscript. LCF and MCS performed the experiments. EFF performed the statistical analysis. WHC and JCTL collected the ameloblastoma samples and clinical information. RSG and CCG reviewed all of the histological diagnoses. All of the authors read and approved the final manuscript.

Acknowledgements

The authors gratefully acknowledge grants from Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), and MS/SCITIE/Decit, Brazil. Professor RS Gomez is a research fellow of CNPq.

Author details

¹Department of Oral Surgery and Pathology, School of Dentistry, Universidade Federal de Minas Gerais, Av. Antonio Carlos, 6627, CEP 31270 901 Belo Horizonte, Minas Gerais, Brazil. ²Department of Pathology, Biological Sciences Institute, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ³Department of Oral Surgery and Diagnosis, Odilon Behrens Hospital, Belo Horizonte, Brazil. ⁴Department of Social and Preventive Dentistry, School of Dentistry, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil.

Received: 12 April 2012 Accepted: 28 July 2012

Published: 6 August 2012

References

1. Barnes L, Eveson JW, Reichart PA, Sidransky D: *World Health Organization classification of tumors: pathology and genetics of tumours of the head and neck*. Lyon: IARC; 2005:296–300.
2. Barreto DC, Gomez RS, Bale AE, Bosen WL, De Marco L: **PTCH gene mutations in odontogenic keratocysts**. *J Dent Res* 2000, **79**:1418–1422.
3. Nodit L, Barnes L, Childers E, Finkelstein S, Swalsky P, Hunt J: **Allelic loss of tumor suppressor genes in ameloblastic tumors**. *Mod Pathol* 2004, **17**:1062–1067.
4. Kumamoto H: **Molecular pathology of odontogenic tumors**. *J Oral Pathol Med* 2006, **35**:65–74.
5. Kitkumthorn N, Mutirangura A: **LINE-1 methylation difference between ameloblastoma and keratocystic odontogenic tumor**. *Oral Dis* 2010, **16**:286–291.
6. Moreira PR, Guimarães MM, Guimarães AL, Diniz MG, Gomes CC, Brito JA, Gomez RS: **Methylation of P16, P21, P27, RB1 and P53 genes in odontogenic keratocysts**. *J Oral Pathol Med* 2009, **38**:99–103.
7. Pan S, Dong Q, Sun LS, Li TJ: **Mechanisms of inactivation of PTCH1 gene in nevoid basal cell carcinoma syndrome: modification of the two-hit hypothesis**. *Clin Cancer Res* 2010, **16**:442–450.
8. Esteller M: **CpG island hypermethylation and tumor suppressor genes: a booming present, a brighter future**. *Oncogene* 2002, **21**:5427–5440.
9. Zhong M, Yan Y, Wang J, Liu J, Li L, Gong YB: **DNA methylation of human telomerase reverse transcriptase promoter region in human ameloblastoma**. *Zhonghua Kou Qiang Yi Xue Za Zhi* 2007, **42**:304–307.
10. Moreira PR, Guimarães MM, Gomes CC, Diniz MG, Brito JA, de Castro WH, Gomez RS: **Methylation frequencies of cell-cycle associated genes in epithelial odontogenic tumours**. *Arch Oral Biol* 2009, **54**:893–897.
11. Hadler-Olsen E, Fadnes B, Sylte I, Uhlin-Hansen L, Winberg JO: **Regulation of matrix metalloproteinase activity in health and disease**. *FEBS J* 2011, **278**:28–45.
12. Sandra F, Hendarmin L, Kukita T, Nakao Y, Nakamura N, Nakamura S: **Ameloblastoma induces osteoclastogenesis: a possible role of ameloblastoma in expanding in the bone**. *Oral Oncol* 2005, **41**:637–644.
13. Kumamoto H, Ooya K: **Expression of bone morphogenetic proteins and their associated molecules in ameloblastomas and adenomatoid odontogenic tumors**. *Oral Dis* 2006, **12**:163–170.

14. Wang A, Zhang B, Huang H, Zhang L, Zeng D, Tao Q, Wang J, Pan C: **Suppression of local invasion of ameloblastoma by inhibition of matrix metalloproteinase-2 *in vitro*.** *BMC Cancer* 2008, **8**:182.
15. Zhang B, Zhang J, Huang HZ, Chen WL, Tao Q, Zeng DL, Zhang LT, Xu JH: **Inhibition of ameloblastoma invasion *in vitro* and *in vivo* by inhibitor of metalloproteinase-2 activity.** *J Oral Pathol Med* 2009, **38**:731–736.
16. Zhang B, Zhang J, Huang HZ, Xu ZY, Xie HL: **Expression and role of metalloproteinase-2 and endogenous tissue regulator in ameloblastoma.** *J Oral Pathol Med* 2010, **39**:219–222.
17. Qian Y, Huang HZ: **The role of RANKL and MMP-9 in the bone resorption caused by ameloblastoma.** *J Oral Pathol Med* 2010, **39**:592–598.
18. Shukeir N, Pakneshan P, Chen G, Szyf M, Rabbani SA: **Alteration of the methylation status of tumor-promoting genes decreases prostate cancer cell invasiveness and tumorigenesis *in vitro* and *in vivo*.** *Cancer Res* 2006, **66**:9202–9210.
19. Chernov AV, Sounni NE, Remacle AG, Strongin AY: **Epigenetic control of the invasion-promoting MT1-MMP/MMP-2/TIMP-2 axis in cancer cells.** *J Biol Chem* 2009, **284**:12727–12734.
20. Li LC, Dahiya R: **MethPrimer: designing primers for methylation PCRs.** *Bioinformatics* 2002, **18**:1427–1431.
21. Roach HI, Yamada N, Cheung KS, Tilley S, Clarke NM, Oreffo RO, Kokubun S, Bronner F: **Association between the abnormal expression of matrix-degrading enzymes by human osteoarthritic chondrocytes and demethylation of specific CpG sites in the promoter regions.** *Arthritis Rheum* 2005, **52**:3110–3124.
22. Livak KJ, Schmittgen TD: **Analysis of relative gene expression data using real-time quantitative PCR and the 2^{-ΔΔCT} method.** *Methods* 2001, **25**:402–408.
23. Snoek-van Beurden PA, Von den Hoff JW: **Zymographic techniques for the analysis of matrix metalloproteinases and their inhibitors.** *Biotechniques* 2005, **38**:73–83.
24. Gomes CC, Duarte AP, Diniz MG, Gomez R: **Current concepts of ameloblastoma pathogenesis.** *J Oral Pathol Med* 2010, **39**:585–591.
25. Farias LC, Gomes CC, Brito JA, Galvão CF, Diniz MG, de Castro WH, Bernardes VD, De Marco LA, Gomez RS: **Loss of heterozygosity of the PTCH gene in ameloblastoma.** *Hum Pathol* 2012, **43**:1229–1233.
26. Kumamoto H, Yamauchi K, Yoshida M, Ooya K: **Immunohistochemical detection of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) in ameloblastomas.** *J Oral Pathol Med* 2003, **32**:114–120.
27. Zhou CX, Gao Y, Johnson NW, Gao J: **Immunoexpression of matrix metalloproteinase-2 and matrix metalloproteinase-9 in the metastasis of squamous cell carcinoma of the human tongue.** *Aust Dent J* 2010, **55**:385–389.
28. Moreira PR, Cardoso FP, Brito JA, Batista AC, Gomes CC, Gomez RS: **Hypomethylation of tumor suppressor genes in odontogenic myxoma.** *Braz Dent J* 2011, **22**:422–427.
29. Omar NF, Gomes JR, Neves JD, Salmon CR, Novaes PD: **MT1-MMP expression in the odontogenic region of rat incisors undergoing interrupted eruption.** *J Mol Histol* 2011, **42**:505–511.
30. Huhtala P, Chow LT, Tryggvason K: **Structure of the human type IV collagenase gene.** *J Biol Chem* 1990, **265**:11077–11082.
31. Crowther M, Goodall S, Jones JL, Bell PR, Thompson MM: **Localization of matrix metalloproteinase-2 within the aneurysmal and normal aortic wall.** *Br J Surg* 2000, **87**:1391–1400.
32. Goodall S, Crowther M, Hemingway DM, Bell PR, Thompson MM: **Ubiquitous elevation of matrix metalloproteinase-2 expression in the vasculature of patients with abdominal aneurysms.** *Circulation* 2001, **104**:304–309.
33. Chicoine E, Estève PO, Robledo O, Van Themsche C, Potworowski EF, St-Pierre Y: **Evidence for the role of promoter methylation in the regulation of MMP-9 gene expression.** *Biochem Biophys Res Commun* 2002, **297**:765–772.

doi:10.1186/1472-6890-12-11

Cite this article as: Farias et al.: Epigenetic regulation of matrix metalloproteinase expression in ameloblastoma. *BMC Clinical Pathology* 2012 12:11.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

