269

in

# **Iron-Induced** Fibrin in Cardiovascular Disease

Boguslaw Lipinski<sup>1</sup> and Etheresia Pretorius<sup>2,\*</sup>

<sup>1</sup>Joslin Diabetes Center, Harvard Medical School, Boston, MA 02215, USA

<sup>2</sup>Department of Physiology, Faculty of Health Sciences, University of Pretoria, Arcadia 0007, South Africa

**Abstract:** Accumulating evidence within the last two decades indicates the association between cardiovascular disease (CVD) and chronic inflammatory state. Under normal conditions fibrin clots are gradually degraded by the fibrinolytic enzyme system, so no permanent insoluble deposits remain in the circulation. However, fibrinolytic therapy in coronary and cerebral thrombosis is ineffective unless it is installed within 3-5 hours of the onset. We have shown that trivalent iron (FeIII) initiates a hydroxyl radical-catalyzed conversion of fibrinogen into a fibrin-like polymer (parafibrin) that is remarkably resistant to the proteolytic dissolution and thus promotes its intravascular deposition. Here we suggest that the persistent presence of proteolysis-resistant fibrin clots causes chronic inflammation. We study the effects of certain amphiphilic substances on the iron- and thrombin-induced fibrinogen polymerization visualized using scanning electron microscopy. We argue that the culprit is an excessive accumulation of free iron in blood, known to be associated with CVD. The only way to prevent iron overload is by supplementation with iron chelating agents. However, administration of free radical scavengers as effective protection against persistent presence of fibrin-like deposits should also be investigated to contribute to the prevention of cardiovascular and other degenerative diseases.

Keywords: Cardiovascular disease, fibrinogen, free radicals, inflammation, iron, parafibrin, thrombosis.

# **INTRODUCTION**

Numerous epidemiological and large prospective studies have shown that hypercoagulability [1] and the increased blood level of fibrinogen (FBG) are important risk factors for cardiovascular disease [2-4]. It is generally agreed that the higher content of fibrinogen in plasma the greater the chance of thrombus formation, therefore its level should be maintained as low as possible. It should be remembered, however, that critical factor in thrombosis is not the absolute amount of FBG, but the fate of its thrombin-generated product, fibrin. Under physiological conditions fibrin clots are gradually, albeit completely, removed from the site of vessel wall injury by the powerful fibrinolytic system of blood [5].

However, if for some reason the fibrinolytic system is inefficient, persisting thrombi will obstruct the flow of blood with all its pathological consequences. The best example of such a situation is the use of a thrombin-like enzyme of viper venom (Ancrod) for the prevention of thrombosis [6]. Thus, despite a complete conversion of all circulating FBG to fibrin, no persistent thrombi are produced due to the solid phase activation of fibrinolysis by fibrin [7]. Disseminated intravascular thrombosis occur only when fibrinolysis is inhibited by eg. *Aprotinin* [8].

The presence of fibrin-like material in atherosclerotic plaques was first observed over 150 years ago by Karl

Received: April 21, 2013 Revised: May 22, 2013 Accepted: May 24, 2013

Rokitansky [8] and later confirmed by other researchers [1, 9-11]. In 1995 Elspeth Smith documented the existence of

fibrinogen-related antigens (FRA) the insoluble fraction

obtained from atherosclerotic intima [12]. According to this

author FRA is present at the surface as well as deep inside

the plaques thus providing scaffolding for migration and

proliferation of smooth muscle cells. It should be

emphasized that this type of insoluble FGB deposits are

morphologically different from fibrin present in thrombi

formed as a result of plaque raptures [13]. This fact obscures

identification of fibrin-like material present inside the

arterial wall, and may perhaps explain why the attention of

researches were drawn away and directed to a more popular

atherosclerotic plaques was also attempted to be explained

by the loss of negative charge of the arterial wall of subjects

with CVD. It was demonstrated that the amount of acid

mucopolysaccharides (AMPS) extractable from the arterial

intima was significantly lower in CVD patients as compared

to young and healthy subjects [15]. According to this idea

AMPS would form soluble complexes with blood fibrin

monomers thus preventing their anchoring and deposition on

the endothelial cells. However, this mechanism would

require a chronic activation of intravascular blood

presence of insoluble FBG deposits

concept of the role of cholesterol [14].

coagulation that has still to be proven.

THE ROLE OF IRON

© 2013 Bentham Science Publishers

The

<sup>\*</sup>Address correspondence to this author at the Department of Physiology, Faculty of Health Sciences, University of Pretoria, Private Bag x323, ARCADIA, 0007, South Africa; Tel: +27 12 319 2907; E-mail: resia.pretorius@up.ac.za

phenomenon is compatible with the observed thrombolytic resistance in patients with coronary and/or cerebral thrombosis [17].

Another important fact is that there is a relationship between body iron overload and pathogenesis of numerous degenerative diseases, including atherosclerosis [18-23]. Particularly relevant is the extensive review by D.B.Kell on the role of free blood iron in various pathological conditions. [24]. In addition, links have been found to exist between iron body stores, cardiovascular risk factors and hypercoagulability [25, 26]. Moreover, in experimental models the infusion of trivalent iron salts was shown to cause diffused thrombosis [27]. It is a common belief that free blood iron, via the Fenton-like reaction, is responsible for so-called oxidative stress that, in turn, leads to atherosclerosis and related cardiovascular diseases [28]. Yet, despite this attractive, albeit simplistic, concept no effectiveness of antioxidant therapy has been demonstrated [29]. As a result numerous natural products (specifically polyphenols) are not being clinically tried because they had been labeled as antioxidants. This highly controversial and, in fact, damaging notion was dealt with in a recent article, which emphasized the importance of polyphenolic substances as iron chelating and free radical scavenging agents that may be neither oxidants nor antioxidants [30].

## **Iron-Induced Conversion of Fibrinogen to Parafibrin**

We have recently documented that trivalent iron ion (FeIII) generates in aqueous solutions powerful hydroxyl radicals that subsequently modify fibrinogen molecules converting them to insoluble fibrin-like polymer [31]. It should be emphasized that such a polymer is not only resistant to fibrinolytic dissolution, but also to proteolytic digestion, i.e. with chymotrypsin, that normally degrades fibrin(ogen) into smaller polypeptide fragments. Protein chemistry teaches us that undesirable molecular interactions in blood proteins are prevented by holding their hydrophobic groups inside the interior of protein tridimensional structures stabilized by intra-molecular disulfide bonds. Once these bonds are broken the polypeptide chains become unfolded with the exposure of hydrophobic domains which form intermolecular bonds resulting in the formation of large aggregates. It is of great importance to note that such aggregates cannot be degraded by the proteolytic enzymes as is the case with human prion proteins [32] and bacterial hydrophobins [33]. Consequently, it is often very difficult to identify insoluble fibrin deposits in pathologically affected organs in various chronic diseases, because no antigen can be released into the liquid phase of the extracted tissues. In concordance with this, the presence of insoluble fibrin(ogen) deposits can only be demonstrated by a direct immunochemical staining of the tissue sections.

## Parafibrin as an Inflammation Inducer

The resistance of fibrin clots to enzymatic degradation can now be explained by our finding of the alternative ironinduced mechanism of blood coagulation (Fig. 1). According to this concept free iron of blood (Fe III) generates hydroxyl radicals, which in turn convert circulating FBG into an insoluble fibrin-like material ( or parafibrin) without the action of thrombin [31]. It should be strongly emphasized that this pseudo or parafibrin is one of very few proteins, such as prions [32] and bacterial hydrophobins [33], that are totally resistant to enzymatic proteolysis. As a consequence such a dense fibrin polymer acquires the features of a foreign body and attracts macrophages resulting in a permanent state of inflammation known to be associated with atherosclerosis [34-37]. Also it is of interest to note the reports on the relationship between inflammation and blood coagulation. Moreover there are numerous experimental and clinical studies that indicate the relationship between inflammation, iron overload and cardiovascular diseases [20, 38-40].

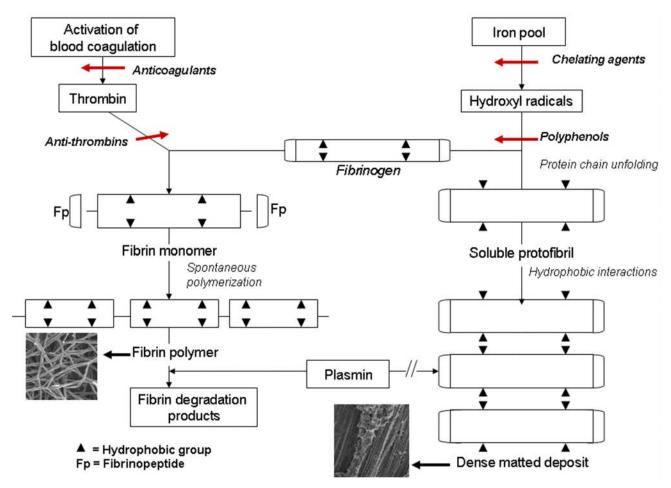
#### Protective Mechanisms Against Iron-Induced Pathology

As shown in Fig. (1) the intravascular formation of modified fibrin can be inhibited at two stages. First, and perhaps the most important, is the inactivation of free iron, usually achievable by the administration of a variety of iron chelators [24, 41-47], as well as by other means of the reduction of body iron stores [48]. If this fails, the next step is elimination of hydroxyl radicals by means of a number of natural and/or synthetic scavengers. The hydroxyl radical scavenging reaction occurs by virtue of aromatic hydroxylation, as exemplified by the reaction with salicylic acid known to prevent inflammation and its consequences [47]. This reaction is believed to be responsible for the beneficial health effects of polyphenolic substances present in fruits and vegetables of the so-called Mediterranean diet [49]. Small molecular weight phenolic compounds such as chlorogenic acid, ferulic and coumaric acids, consumed with certain food products, become even more effective hydroxyl radical scavengers due to their enhanced absorption from the alimentary track. The larger molecules of polyphenols have to be first metabolized by the intestinal flora (probiotics) in order to achieve their in vivo health beneficial effect [50].

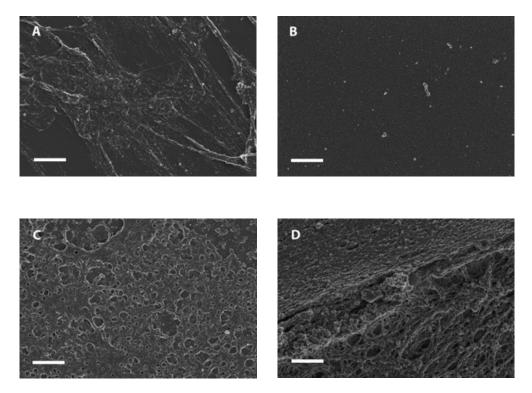
In concert with this observation it is the fact that the altered fibrin structure argued to be associated with cardiovascular disease (CVD) can be normalized by the pretreatment with hydrophilic substances e.g. high-density lipoprotein (HDL) [51] and/or human serum albumin known to be decreased in CVD [52-55]. The protective effect of such substances is documented here using SEM method for HDL (Fig. 2C), and for a non-ionic detergent Tween 20 (Fig. 2A and B). These results stand in contrast with the potentiation of iron-induced dense parafibrin formation exerted by low density lipoprotein (Fig. 2D). It is also possible that the health beneficial effect of human serum albumin (HSA), a highly hydrophilic protein, is due to the restoration of normal fibrin strands generated with thrombin (Fig. 2E and F). Therefore, it can be concluded that it is not just iron homeostasis, but the blood content of hydrophilic and polyphenolic agents that is important in the prevention of atherosclerosis.

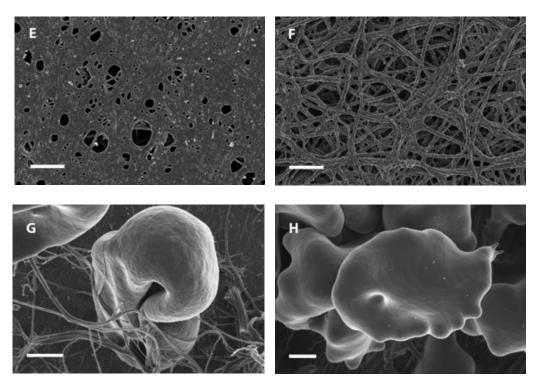
## THE ROLE OF RED BLOOD CELLS

Another pathologic process leading to atherosclerosis is the impaired blood flow caused by the intravascular aggregation of red blood cells (RBC). Although its mechanism is not completely understood, it is well known that the elevated erythrocyte sedimentation rate (ESR) is associated with inflammation and CVD. (56-59) We have



**Fig. (1).** Conversion of plasma fibrinogen into insoluble polymers catalyzed by thrombin (left panel), and the formation of dense fibrin deposits generated with iron (right panel). By contrast to the enzymatically formed fibrin susceptible to fibrinolysis, the iron-induced fibrin polymer is remarkably resistant to proteolytic degradation. Reprinted with permission from Pol Arch Med Wewn. Vol.122, p.120 (Fig.6), 2012.<sup>31</sup>





**Fig. (2).** Effects of certain amphiphilic substances on the iron- and thrombin-induced fibrinogen (FGB) polymerization visualized USING scanning electron microscopy, **A.** Control: Purified fibrinogen (PF) and ferric chloride (FC); **B.** PF + Tween 20 + FC; **C.** PF + high-density lipoprotein + FC; **D.** PF + low-density lipoprotein + FC; **E.** Low-albumin plasma (LAP)+ thrombin; **F.** LAP + purified human albumin + thrombin; **G.** Whole blood of a stroke patient; **H.** Normal whole blood + FC. Scale = 1  $\mu$ m.

shown for the first time that the abnormal RBC morphology induced by iron ions added to normal blood is strikingly similar to that observed in blood of stroke patients [60] as well as of subjects with diabetes mellitus [61]. These changes are shown here in Fig. (2G) (Stroke) and H (healthy blood with added ferric iron). It is argued that the close association between RBC and the modified fibrinogen molecules can be caused by the interaction between hydrophobic epitopes on the cell membranes and those of the soluble fibrin protofibrils generated with iron (Fig. 1). This mechanism may explain significant reduction of blood flow [62] and increased blood viscosity in patients with thrombotic arterial disease [63]. The relationship between inflammation and elevated ESR was emphasized years ago by Zacharski and Kyle [64]. Finally, it should be noted that RBC aggregation and sedimentation were originally thought to be caused by blood soluble fibrin monomers by virtue of their interaction with hydrophobic epitopes on RBC membranes [65]. However, this concept was abandoned in view of the absence of any evidence of the link between chronic activation of intravascular blood coagulation and ESR. The concept of iron-induced parafibrin formation offers more plausible mechanism of the relationship between hemorheologic disturbances and inflammation.

# CONCLUSION

In conclusion, we postulate in this paper that the excess of blood free iron is responsible for the non-enzymatic generation of insoluble fibrin-like material (parafibrin) that, when deposited on the arterial wall, initiates inflammatory reactions. This pathological process, very different from the classical activation of blood coagulation, can be prevented by substances that chelate iron, scavenge hydroxyl radicals, and inhibit hydrophobic interactions in proteins. However, in view of the fact that, so far, there is no known agent or a biological process that can degrade parafibrin, an interdisciplinary research approach is needed to find an effective method for the elimination from the human body this unique inducer of chronic inflammation.

## ETHICAL APPROVAL DISCLOSURE

Ethical approval was granted at the University of Pretoria (Human Ethics Committee: Faculty OF Health Sciences) under the name of E Pretorius (corresponding author). All human blood samples obtained were analyzed at the University of Pretoria and all participants filled in informed consent forms.

## **CONFLICT OF INTEREST**

The authors confirm that this article content has no conflicts of interest.

## **ACKNOWLEDGEMENTS**

The authors would like to thank Natasha Vermeulen and Janette Bester for technical assistance with sample preparation.

#### REFERENCES

- Chan MY, Andreotti F, Becker RC. Hypercoagulable states in cardiovascular disease. Circulation. 2008;118(22):2286-97. Epub 2008/11/26.
- [2] Kannel WB, Wolf PA, Castelli WP, D'Agostino RB. Fibrinogen and risk of cardiovascular disease. The Framingham Study. JAMA. 1987;258(9):1183-6. Epub 1987/09/04.

- [3] Levenson J, Giral P, Razavian M, Gariepy J, Simon A. Fibrinogen and silent atherosclerosis in subjects with cardiovascular risk factors. Arterioscler Thromb Vasc Biol. 1995;15(9):1263-8. Epub 1995/09/01.
- [4] Kaptoge S, Di Angelantonio E, Pennells L, Wood AM, White IR, Gao P, et al. C-reactive protein, fibrinogen, and cardiovascular disease prediction. N Engl J Med. 2012;367(14):1310-20. Epub 2012/10/05.
- [5] Astrup T. The biological significance of fibrinolysis. Lancet. 1956;271(6942):565-8. Epub 1956/09/15.
- [6] Liu S, Marder VJ, Levy DE, Wang SJ, Yang F, Paganini-Hill A, et al. Ancrod and fibrin formation: perspectives on mechanisms of action. Stroke. 2011;42(11):3277-80. Epub 2011/08/27.
- [7] Lipinski B, Nowak A, Gurewich V. The organ distribution of 125Ifibrin in the generalized Shwartzman reaction and its relation to leucocytes. Br J Haematol. 1974;28(2):221-31. Epub 1974/10/01.
- [8] Rokitansky K. A Manual of Pathological Anatomy 1852;Vol.IV. London: Sydenham Society.
- [9] Duguid JB. Thrombosis as a factor in the pathogenesis of coronary atherosclerosis. J Pathol Bacteriol. 1946;58:207-12. Epub 1946/04/01.
- [10] Kadish JL. Fibrin and atherogenesis--a hypothesis. Atherosclerosis. 1979;33(4):409-13. Epub 1979/08/01.
- [11] Bini A, Kudryk BJ. Fibrinogen and fibrin in the arterial wall. Thromb Res. 1994;75(3):337-41. Epub 1994/08/01.
- [12] Smith EB. Fibrinogen, fibrin and the arterial wall. Eur Heart J. 1995;16 Suppl A:11-4; discussion 4-5. Epub 1995/03/01.
- [13] Ogata J, Yutani C, Kaneko T, Kuriyama Y, Sawada T. Rupture of atheromatous plaque as a cause of thrombotic occlusion of the internal carotid artery. Stroke. 1987;18(6):1175-6. Epub 1987/11/01.
- [14] Bennett PC, Silverman SH, Gill PS, Lip GY. Peripheral arterial disease and Virchow's triad. Thromb Haemost. 2009;101(6):1032-40. Epub 2009/06/06.
- [15] Lipinski B, Lewicki Z, Zajdel M, Miks B, Hagel E. Saline extractable acid mucopolysaccharides of human aorta in relation to age, sex and atherosclerosis. Mater Med Pol. 1972;4(1):19-22. Epub 1972/01/01.
- [16] Undas A, Szuldrzynski K, Stepien E, Zalewski J, Godlewski J, Tracz W, et al. Reduced clot permeability and susceptibility to lysis in patients with acute coronary syndrome: effects of inflammation and oxidative stress. Atherosclerosis. 2008;196(2):551-7. Epub 2007/07/21.
- [17] Lipinski B. Modification of fibrin structure as a possible cause of thrombolytic resistance. J Thromb Thrombolysis. 2010;29(3):296-8. Epub 2009/06/25.
- [18] Ahluwalia N, Genoux A, Ferrieres J, Perret B, Carayol M, Drouet L, et al. Iron status is associated with carotid atherosclerotic plaques in middle-aged adults. J Nutr. 2010;140(4):812-6. Epub 2010/02/26.
- [19] Brewer GJ. Iron and copper toxicity in diseases of aging, particularly atherosclerosis and Alzheimer's disease. Exp Biol Med (Maywood). 2007;232(2):323-35. Epub 2007/01/30.
- [20] Depalma RG, Hayes VW, Chow BK, Shamayeva G, May PE, Zacharski LR. Ferritin levels, inflammatory biomarkers, and mortality in peripheral arterial disease: a substudy of the Iron (Fe) and Atherosclerosis Study (FeAST) Trial. J Vasc Surg. 2010;51(6):1498-503. Epub 2010/03/23.
- [21] Hahalis G, Kalogeropoulos A, Terzis G, Tselepis AD, Kourakli A, Mylona P, et al. Premature atherosclerosis in non-transfusiondependent beta-thalassemia intermedia. Cardiology. 2011;118(3):159-63. Epub 2011/06/08.
- [22] Kiechl S, Willeit J, Egger G, Poewe W, Oberhollenzer F. Body iron stores and the risk of carotid atherosclerosis: prospective results from the Bruneck study. Circulation. 1997;96(10):3300-7. Epub 1997/12/13.
- [23] Merono T, Rosso LG, Sorroche P, Boero L, Arbelbide J, Brites F. High risk of cardiovascular disease in iron overload patients. Eur J Clin Invest. 2011;41(5):479-86. Epub 2010/12/07.
- [24] Kell DB. Towards a unifying, systems biology understanding of large-scale cellular death and destruction caused by poorly liganded iron: Parkinson's, Huntington's, Alzheimer's, prions, bactericides, chemical toxicology and others as examples. Arch Toxicol. 2010;84(11):825-89. Epub 2010/10/23.
- [25] Franchini M, Targher G, Montagnana M, Lippi G. Iron and thrombosis. Ann Hematol. 2008;87(3):167-73. Epub 2007/12/11.

- [26] Salonen JT, Nyyssonen K, Korpela H, Tuomilehto J, Seppanen R, Salonen R. High stored iron levels are associated with excess risk of myocardial infarction in eastern Finnish men. Circulation. 1992;86(3):803-11. Epub 1992/09/01.
- [27] Tian J, Hu S, Sun Y, Ban X, Yu H, Dong N, et al. A novel model of atherosclerosis in rabbits using injury to arterial walls induced by ferric chloride as evaluated by optical coherence tomography as well as intravascular ultrasound and histology. J Biomed Biotechnol. 2012;2012:121867. Epub 2012/06/06.
- [28] Griendling KK, FitzGerald GA. Oxidative stress and cardiovascular injury: Part I: basic mechanisms and in vivo monitoring of ROS. Circulation. 2003;108(16):1912-6. Epub 2003/10/22.
- [29] Steinhubl SR. Why have antioxidants failed in clinical trials? Am J Cardiol. 2008;101(10A):14D-9D. Epub 2008/06/24.
- [30] Lipinski B. Hydroxyl radical and its scavengers in health and disease. Oxid Med Cell Longev. 2011;2011:809696. Epub 2011/09/10.
- [31] Lipinski B, Pretorius E. Novel pathway of ironinduced blood coagulation: implications for diabetes mellitus and its complications. Pol Arch Med Wewn. 2012;122(3):115-22. Epub 2012/03/31.
- [32] Das D, Luo X, Singh A, Gu Y, Ghosh S, Mukhopadhyay CK, et al. Paradoxical role of prion protein aggregates in redox-iron induced toxicity. PLoS ONE. 2010;5(7):e11420. Epub 2010/07/14.
- [33] Kwan AH, Winefield RD, Sunde M, Matthews JM, Haverkamp RG, Templeton MD, et al. Structural basis for rodlet assembly in fungal hydrophobins. Proc Natl Acad Sci U S A. 2006;103(10):3621-6. Epub 2006/03/16.
- [34] Kleemann R, Verschuren L, Morrison M, Zadelaar S, van Erk MJ, Wielinga PY, et al. Anti-inflammatory, anti-proliferative and antiatherosclerotic effects of quercetin in human in vitro and in vivo models. Atherosclerosis. 2011;218(1):44-52. Epub 2011/05/24.
- [35] Libby P. Inflammation in atherosclerosis. Arterioscler Thromb Vasc Biol. 2012;32(9):2045-51. Epub 2012/08/17.
- [36] Ross R. Atherosclerosis--an inflammatory disease. N Engl J Med. 1999;340(2):115-26. Epub 1999/01/14.
- [37] Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med. 2005;352(16):1685-95. Epub 2005/04/22.
- [38] Hu FB. The iron-heart hypothesis: search for the ironclad evidence. JAMA. 2007;297(6):639-41. Epub 2007/02/15.
- [39] Menke A, Fernandez-Real JM, Muntner P, Guallar E. The association of biomarkers of iron status with peripheral arterial disease in US adults. BMC Cardiovasc Disord. 2009;9:34. Epub 2009/08/05.
- [40] Lele S, Shah S, McCullough PA, Rajapurkar M. Serum catalytic iron as a novel biomarker of vascular injury in acute coronary syndromes. EuroIntervention. 2009;5(3):336-42. Epub 2009/09/09.
- [41] Duffy SJ, Biegelsen ES, Holbrook M, Russell JD, Gokce N, Keaney JF, Jr., et al. Iron chelation improves endothelial function in patients with coronary artery disease. Circulation. 2001;103(23):2799-804. Epub 2001/06/13.
- [42] Ferrara DE, Taylor WR. Iron chelation and vascular function: in search of the mechanisms. Arterioscler Thromb Vasc Biol. 2005;25(11):2235-7. Epub 2005/11/01.
- [43] Kwiatkowski JL. Oral iron chelators. Hematol Oncol Clin North Am. 2010;24(1):229-48. Epub 2010/02/02.
- [44] Matthews AJ, Vercellotti GM, Menchaca HJ, Bloch PH, Michalek VN, Marker PH, et al. Iron and atherosclerosis: inhibition by the iron chelator deferiprone (L1). J Surg Res. 1997;73(1):35-40. Epub 1998/02/14.
- [45] Zhang WJ, Wei H, Frei B. The iron chelator, desferrioxamine, reduces inflammation and atherosclerotic lesion development in experimental mice. Exp Biol Med (Maywood). 2010;235(5):633-41. Epub 2010/05/14.
- [46] Yuan XM, Li W. The iron hypothesis of atherosclerosis and its clinical impact. Ann Med. 2003;35(8):578-91. Epub 2004/01/08.
- [47] Mehta SR. Aspirin for prevention and treatment of cardiovascular disease. Ann Intern Med. 2009;150(6):414-6. Epub 2009/03/19.
- [48] Zacharski LR, Shamayeva G, Chow BK. Effect of controlled reduction of body iron stores on clinical outcomes in peripheral arterial disease. Am Heart J. 2011;162(5):949-57 e1. Epub 2011/11/19.
- [49] Nadtochiy SM, Redman EK. Mediterranean diet and cardioprotection: the role of nitrite, polyunsaturated fatty acids, and polyphenols. Nutrition. 2011;27(7-8):733-44. Epub 2011/04/02.

- [50] Kahle K, Huemmer W, Kempf M, Scheppach W, Erk T, Richling E. Polyphenols are intensively metabolized in the human gastrointestinal tract after apple juice consumption. J Agric Food Chem. 2007;55(26):10605-14. Epub 2007/12/01.
- [51] Zabczyk M, Hondo L, Krzek M, Undas A. High-density cholesterol and apolipoprotein AI as modifiers of plasma fibrin clot properties in apparently healthy individuals. Blood Coagul Fibrinolysis. 2012. Epub 2012/10/06.
- [52] Alvarez-Perez FJ, Castelo-Branco M, Alvarez-Sabin J. Albumin level and stroke. Potential association between lower albumin level and cardioembolic aetiology. Int J Neurosci. 2011;121(1):25-32. Epub 2010/10/20.
- [53] Folsom AR, Lutsey PL, Heckbert SR, Cushman M. Serum albumin and risk of venous thromboembolism. Thromb Haemost. 2010;104(1):100-4. Epub 2010/04/15.
- Hostmark AT, Tomten SE. Serum albumin and self-reported [54] prevalence of stroke: a population-based, cross-sectional study. Eur J Cardiovasc Prev Rehabil. 2006;13(1):87-90. Epub 2006/02/02.
- [55] Kim KJ, Yang WS, Kim SB, Lee SK, Park JS. Fibrinogen and fibrinolytic activity in CAPD patients with atherosclerosis and its correlation with serum albumin. Perit Dial Int. 1997;17(2):157-61. Epub 1997/03/01.
- Andresdottir MB, Sigfusson N, Sigvaldason H, Gudnason V. [56] Erythrocyte sedimentation rate, an independent predictor of coronary heart disease in men and women: The Reykjavik Study. Am J Epidemiol. 2003;158(9):844-51. Epub 2003/10/31.
- [57] Erikssen G, Liestol K, Bjornholt JV, Stormorken H, Thaulow E, Erikssen J. Erythrocyte sedimentation rate: a possible marker of

Lipinski and Pretorius

atherosclerosis and a strong predictor of coronary heart disease mortality. Eur Heart J. 2000;21(19):1614-20. Epub 2000/09/16.

- [58] Nagy E, Eaton JW, Jeney V, Soares MP, Varga Z, Galajda Z, et al. Red cells, hemoglobin, heme, iron, and atherogenesis. Arterioscler Thromb Vasc Biol. 2010;30(7):1347-53. Epub 2010/04/10.
- [59] Natali A, L'Abbate A, Ferrannini E. Erythrocyte sedimentation rate, coronary atherosclerosis, and cardiac mortality. Eur Heart J. 2003;24(7):639-48. Epub 2003/03/27.
- Lipinski B, Pretorius E, Oberholzer HM, van der Spuy WJ. [60] Interaction of fibrin with red blood cells: the role of iron. Ultrastruct Pathol. 2012;36(2):79-84. Epub 2012/04/05.
- [61] Pretorius E, Lipinski B. Iron alters red blood cell morphology. Blood. 2013;121(1):9.
- [62] Arbel Y, Banai S, Benhorin J, Finkelstein A, Herz I, Halkin A, et al. Erythrocyte aggregation as a cause of slow flow in patients of acute coronary syndromes. Int J Cardiol. 2012;154(3):322-7. Epub 2011/07/26.
- [63] Cairneross D, Collins GM, Kostalas G, Ludbrook J. Blood viscosity and erythrocyte sedimentation rate in patients with thrombotic arterial disorders. Med J Aust. 1969;1(26):1348-52. Epub 1969/06/28.
- [64] Zacharski LR, Kyle RA. Significance of extreme elevation of erythrocyte sedimentation rate. JAMA. 1967;202(4):264-6. Epub 1967/10/23.
- Lipinski B, Worowski K, Mysliwiec M, Farbiszewski R. [65] Erythrocyte sedimentation and soluble fibrin monomer complexes. Thromb Diath Haemorrh. 1969;21(2):196-202. Epub 1969/04/30.