

Increase in Multidrug Resistant *Neisseria gonorrhoeae* FC428-Like Isolates Harboring the Mosaic *penA* 60.001 Gene, in Nanjing, China (2017-2020)

Yuanyuan Zhao¹, Wenjing Le¹, Caroline A Genco², Peter A Rice³, Xiaohong Su¹

¹Sexually Transmitted Disease Clinic, Institute of Dermatology, Chinese Academy of Medical Sciences and Peking Union Medical College, Nanjing, People's Republic of China; ²Department of Immunology, Tufts University School of Medicine, Boston, MA, USA; ³Division of Infectious Diseases and Immunology, Department of Medicine, University of Massachusetts Chan Medical School, Worcester, MA, USA

Correspondence: Xiaohong Su, Sexually Transmitted Disease Clinic, Institute of Dermatology, Chinese Academy of Medical Sciences and Peking Union Medical College, No. 12, Jiangwangmiao Road, Nanjing, 210042, Jiangsu Province, People's Republic of China, Email suxh@ncstdlc.org

Background: Since the first Chinese report of the ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone in 2016, additional FC428-like, *penA* 60.001 isolates have been identified in China.

Objective: To document the rise in *penA* 60.001 isolates in Nanjing, China, and characterize their molecular and epidemiological features.

Methods: *N. gonorrhoeae* minimum inhibitory concentrations (MICs, mg/L) for ceftriaxone, cefixime, penicillin, tetracycline, ciprofloxacin, azithromycin, spectinomycin, gentamicin and zoliflodacin were determined by agar dilution. MICs for ertapenem were measured by E-test. *N. gonorrhoeae* antimicrobial sequence typing (NG-STAR) of seven loci (*penA*, *mtrR*, *porB*, *ponA*, *gyrA*, *parC* and *23S rRNA*) was analyzed together with *N. gonorrhoeae* multiantigen sequence typing (NG-MAST) and multilocus sequence typing (MLST). Phylogenetic analysis was also performed using whole genomic sequencing (WGS).

Results: Fourteen FC428-related *penA* 60.001 *N. gonorrhoeae* infections were identified out of 677 infections from 2017 to 2020, in Nanjing, representing an incremental yearly rise in the percentage of the city's *N. gonorrhoeae* isolates that were FC428-related. Seven FC428-related *N. gonorrhoeae* infections were acquired in Nanjing, proper; four others in eastern Chinese cities and three from unknown locations. All FC428-related isolates were resistant to ceftriaxone, cefixime, ciprofloxacin, tetracycline and penicillin but susceptible to spectinomycin, gentamicin, ertapenem and zoliflodacin; three strains were resistant to azithromycin. *penA* 60.001 isolates displayed closely related MLST types and NG-STAR types but relatively distant NG-MAST types. WGS showed a phylogenetic analysis that intermingled with other international isolates.

Conclusion: *penA* 60.001 *N. gonorrhoeae* isolates emerged in Nanjing, China, beginning in 2017, and have continued to rise.

Keywords: *Neisseria gonorrhoeae*, *penA* 60.001, ceftriaxone-resistance, FC428, WGS phylogenetic analysis

Introduction

Gonorrhea is the second most common bacterial sexually transmitted infection (Chlamydia is first), with a worldwide estimated incidence of new gonococcal infections in 2020 of 82.4 million cases.¹ If untreated, gonorrhea in women can lead to severe complications, such as endometritis and salpingitis (pelvic inflammatory disease [PID]) that may result in substantial morbidity including infertility and ectopic pregnancy; in men epididymitis can also result in infertility.² Furthermore, *Neisseria gonorrhoeae* has developed antimicrobial resistance to almost all antibiotic classes used for treatment that now requires intensive surveillance of gonococcal isolates for antibiotic susceptibility.³

Presently, ceftriaxone, an extended-spectrum cephalosporin (ESC) represents first-line treatment for gonorrhea. However, individual strains of *N. gonorrhoeae* with high-level resistance to ESCs have emerged sporadically outside of China; in Japan (eg, *N. gonorrhoeae* strains H041, GU140106 and most recently FC428),⁴⁻⁶ Australia (strain A8806),⁷ France (strain F89),⁸ Italy (strain G2891).⁹ Notably, the FC428 clone remains prominent; first described in Japan in

2015;⁶ since then, FC428-related sub-clones or FC428-like isolates (with a close phylogenomic relationship to FC428) have been identified in Australia (strains A7846 and A7536),¹⁰ Canada (47,707),¹¹ Denmark (GK124),¹² France (F90)¹³ and Ireland (IR72),¹⁴ thereby confirming international spread of FC428-related sub-clones that possess ESC resistance. A common feature of FC428 and FC428-like strains is possession of an ESC-resistant genetic determinant; the mosaic *penA* 60.001 gene.^{6,10–14} At present, the surveillance of FC428/FC428-like isolates is a focus in tracking emergence and spread of resistant *N. gonorrhoeae* strains. Notably, infections with FC428/FC428-related sub-clones have been associated with travel to the Asia-Pacific region, now including its identification in several distinct regions in China.^{15–21} Here, we report 14 cases of *N. gonorrhoeae* that possessed the FC428-associated *penA* 60.001 gene, isolated from 2017 to 2020 in Nanjing, China. We compared their molecular types, also including whole genome sequences (WGSs), with other FC428-like isolates and identified their contribution, overall, to the reported emergence of FC428-like *N. gonorrhoeae* isolates in China.

Materials and Methods

Isolation of *Neisseria gonorrhoeae*

Gonococcal isolates were collected from male subjects with symptomatic urethritis (urethral discharge and/or dysuria) and their female sex partners who were patients attending the STD Clinic at the Institute of Dermatology, Chinese Academy of Medical Sciences, Nanjing, China. Urethral or cervical exudates were collected using cotton swabs and plated immediately onto Thayer-Martin medium (Zhuhai DL Biotech, China) and incubated in candle jars at 36°C for 24 to 48 h. Colonial morphology, Gram's stain, and oxidase testing were used to identify *N. gonorrhoeae*, which was then sub-cultured onto GC medium (chocolate agar base) (Difco, Detroit, MI) supplemented with 1% Isovitalex (Oxoid, USA). Gonococcal colonies were suspended in tryptone-based soy broth and stored frozen (−80°C).

Antimicrobial Susceptibility Testing

Mean inhibitory concentrations (MICs) for penicillin, tetracycline, ciprofloxacin, azithromycin, cefixime, ceftriaxone, spectinomycin, gentamicin and zoliflodacin were determined by agar dilution, according to the Clinical and Laboratory Standards Institute (CLSI) guidelines. For ertapenem, the E-test method was used.^{22,23} Using CLSI²⁴ and the European Committee on Antimicrobial Susceptibility Testing (EUCAST)²⁵ criteria for antimicrobial resistance (AMR), the following MIC breakpoints were used to ascertain resistance: ≥ 0.25 mg/L, ceftriaxone and cefixime; ≥ 1 mg/L, azithromycin and ciprofloxacin; ≥ 128 mg/L, spectinomycin and ≥ 2 mg/L, penicillin and tetracycline. Breakpoints for gentamicin, ertapenem and zoliflodacin have not been established formally by CLSI but we used MICs ≥ 32 mg/L for gentamicin and ≥ 0.5 mg/L for zoliflodacin, which have been used previously.^{26,27} Using *Haemophilus influenzae* criteria defined by CLSI, an MIC ≤ 0.5 mg/L for ertapenem was considered susceptible.²⁸ β -lactamase production was determined by paper acidometric testing.²⁹

Molecular Epidemiologic Typing

We employed molecular epidemiologic typing: NG-MAST (<http://www.ng-mast.net>); MLST (<http://pubmlst.org/neisseria/>) and NG-STAR (<https://ngstar.canada.ca>), which were analyzed as described previously,^{30–32} for NG-MAST; *porB* and *tbpB*, for MLST; *abcZ*, *adk*, *aroE*, *fumC*, *gdh*, *pdh* and *pgm* and for NG-STAR; *penA*, *mtrR*, *porB*, *ponA*, *gyrA*, *parC* and *23S rRNA*.

Whole Genome Sequencing (WGS) and Phylogenetic Analysis

Genomic DNA was extracted using QIAextractor DX Kits (QIAGEN, Hilden, Germany). Whole genomes of 13/14 strains were successfully sequenced using the HiSeq (Illumina, San Diego, CA, USA) and PacBio RSII SMRT (Pacific Biosciences, Menlo Park, CA, USA) platforms at Genewiz Inc. (Suzhou, China). The complete genomic sequences of the thirteen isolates were uploaded to NCBI (BioProject ID: PRJNA553852, PRJNA553854 and PRJNA916595). Phylogenetic analysis was performed as previously described.¹⁶ Sequences were aligned with the FC428 genome (BioProject ID: PRJDB5915) and single nucleotide polymorphisms (SNPs) identified using MUMmer software. Repetitive regions and SNPs in putative recombinogenic regions were removed using SyRI software. All available

whole-genome sequences or short-read sequences of FC428-related clones were similarly analyzed. Iqtree was used to analyze the phylogenetic relationship (maximum-likelihood method, bootstrap 1000) and to visualize the phylogenetic tree.

Statistical Analyses

SPSS 23 was used to perform statistical analyses. Comparisons of the rates of isolation of FC428-like isolates over time were performed using a logistic regression test of trend; $P \leq 0.05$ (2-sided) was considered significant.

Results

Between 2017 and 2020, we identified 14 *N. gonorrhoeae* FC428-like isolates with high-level resistance to ceftriaxone and cefixime in Nanjing, which represented an incremental yearly rise in the percentage of the city's *N. gonorrhoeae* isolates that were FC428-like: 0.44% in 2017 (1/229); 0.49% (1/204) in 2018; 4.73% in 2019 (7/148) and 5.21% in 2020 (5/96), ($\chi^2=12.467$, $P < 0.001$) (Figure 1). Characteristics of subjects infected with FC428-like *N. gonorrhoeae* are shown in Table 1. Thirteen isolates were from men and one from a woman (sexual contact of a man with gonococcal infection with positive nucleic amplification test result for *N. gonorrhoeae*); none had traveled outside of China. All subjects had uncomplicated gonococcal infection and reported 1 or 2 sex partners in the recent month; partners were from Nanjing, Shanghai, Suzhou, Taizhou and Yancheng, indicating the presence of ESC-resistant FC428-like *N. gonorrhoeae* in (certain) eastern Chinese cities. Ten men provided sexual histories: five indicated coitus with regular partners only, five with non-regular partners. 9/14 took antibiotics (levofloxacin or cephalosporin) before attending the clinic. Microscopic examination of gram stained urethral (or cervical) smears revealed >10 polymorphonuclear neutrophils (PMNs)/per high-power field and gram-negative intracellular diplococci in all 14. All subjects were treated with spectinomycin: two that were coinfecting with *Chlamydia trachomatis* were also treated with doxycycline (0.1g, twice a day) for ten days. Twelve

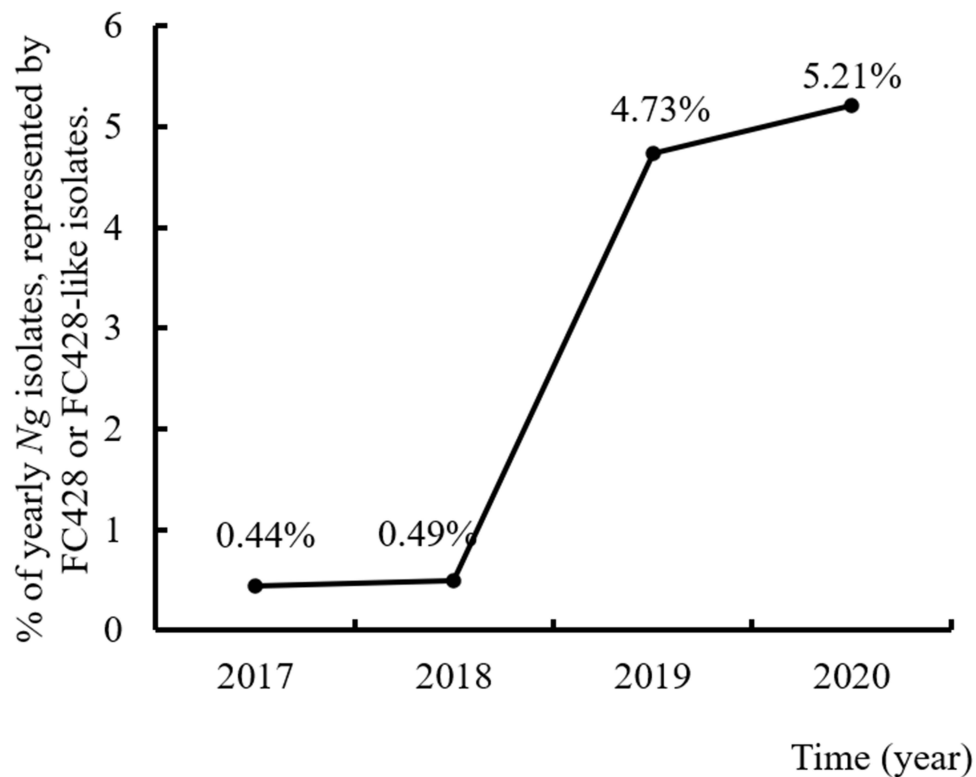


Figure 1 % of yearly *N. gonorrhoeae* (Ng) isolates, represented by FC428 or FC428-like isolates (2017–20). Rate (%) of FC428 or FC428-like isolates increased yearly across the 4 years, taken as a whole ($P < 0.001$; $\chi^2=12.467$; Test of Trend) (between 2017 and 2020); linear association of the rates of FC428 isolates and year examined by logistic regression.

Table 1 Characteristics of 14 Subjects with Uncomplicated Gonococcal Infection, Nanjing, China

Characteristics	1	2	3	4	5	6	7
Subject							
Date	Sept, 2017	July,2018	May, 2019	June, 2019	June, 2019	Sept, 2019	Nov, 2019
Gender	Male	Male	Male	Male	Male	Male	Male
Age	24	34	59	19	23	16	47
City of origin	Nanjing	Nanjing	Nanjing	Nanjing	Nanjing	Nanjing	Nanjing
Foreign travel	No	No	No	No	No	No	No
History of STIs	No	No	No	NGU	CT, PPLO infection	No	HPV, PPLO infection
Administered antibiotics prior to the clinic visit	LVFX	LVFX	LVFX	No	LVFX Cephalosporin	Cephalosporin	Yes, type undetermined
Microscopic exam							
Gram's stain of exudate	>10 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs
Microbiology							
NG isolate	NJ 1711654	NJ 189125	NJ 195417	NJ 196610	NJ 197542	NJ 1911400	NJ 1913940
MG (PCR)	NA	–	–	+	–	–	–
CT (PCR)	–	–	–	–	+	–	–
Treatment	Spe	Spe	Spe	Spe	Spe	Spe	Spe
Follow-up	Resolution of symptoms and signs	Resolution of symptoms and signs	Resolution of symptoms and signs	Urethral discomfort No sign	Urethral discomfort No sign	Resolution of symptoms and signs	Resolution of symptoms and signs
Sex-contact(s)							
No. of Contacts (recent month)	1	1	1	1	1	2	2
City/ies where contact occurred	Unknown	Shanghai	Nanjing	Unknown	Taizhou	Nanjing	1. Nanjing 2. Suzhou
Nature of the sex contact	Unknown	Unknown	Sex-worker	Regular sex-partner	Non-Regular sex-partner	Non-Regular sex-partner	1. Wife 2. Non-Regular sex-partner
Sex-contact treated	Unknown	Unknown	Unknown	Yes	Unknown	Unknown	1. Yes 2. Unknown

Characteristics	8	9	10	11	12	13	14
Subject							
Date	Nov, 2019	Dec, 2109	Jun, 2020	July, 2020	Oct, 2020	Oct, 2020	Nov, 2020
Gender	Male	Male	Male	Female	Male	Male	Male
Age	20	42	20	26	20	21	20
City of origin	Nanjing	Nanjing	Nanjing	Nanjing	Nanjing	Unknown	Nanjing
Foreign travel	No	No	No	No	No	No	No
History of STIs	No	No	No	No	No	No	NG infection
Administered antibiotics prior to the clinic visit	Yes, type undetermined	Cefdinir	No	No	No	LVFX	No
Microscopic exam							
Gram's stain of exudate	>10 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>20 PMN /hpf + GNIDs	>10 PMN /hpf + GNIDs
Microbiology							
NG isolate	NJ 1914215	NJ 1914646	NJ 203279	NJ 204705	NJ 208430	NJ 208756	NJ 209469
MG (PCR)	-	-	-	-	NA	-	-
CT (PCR)	-	-	+	-	NA	-	-
Treatment	Spe	Spe	Spe	Spe	Spe	Spe	Spe
Follow-up	Resolution of symptoms and signs	Resolution of symptoms and signs	Resolution of symptoms and signs	Resolution of symptoms and signs TOC (-)	Resolution of symptoms and signs TOC (-)	Resolution of symptoms and signs	Resolution of symptoms and signs
Sex-contact(s)							
No. of Contacts (recent month)	1	1	1	1	1	2	1
City/ies where contact occurred	Nanjing	Yancheng	Nanjing	Nanjing	Nanjing	Unknown	Nanjing
Nature of the sex contact	Regular Sex-partner	Non-Regular sex-partner	Regular Sex-partner	Regular Sex-partner	Unknown	Unknown	Regular Sex-partner
Sex-contact treated	Yes	Yes	No	Yes	Unknown	Unknown	No

Abbreviations: STIs, Sexually transmitted infections; NGU; non-gonococcal urethritis; MG, Mycoplasma genitalium; PPLo, mycoplasma; CT, Chlamydia trachomatis; HPV, human papilloma virus; LVFX, levofloxacin; NA, not available; TOC, test of cure; PMN, polymorphonuclear neutrophils; hpf, high power field; GNID, Gram-negative Intracellular Diplococci; Spe, spectinomycin.

had total resolution of symptoms and signs; 2 had residual urethral symptoms but no discharge. Two subjects had tests-of-cure; both of which were negative (Table 1).

Antimicrobial susceptibility testing of the 14 isolates showed similar resistance profiles (Table 2); all were resistant to ceftriaxone (MICs ≥ 0.5 mg/L), cefixime (MICs ≥ 2 mg/L), ciprofloxacin (MICs ≥ 16 mg/L), tetracycline (MICs ≥ 1 mg/L) and penicillin (MICs ≥ 2 mg/L) but were susceptible to spectinomycin, gentamicin, ertapenem and zoliflodacin. Three strains (NJ1914215, NJ203279 and NJ209649) were resistant to azithromycin (MIC, 8 mg/L); the remaining isolates were sensitive (0.25 mg/L).

Sequence typing (ST) analysis showed that the 14 isolates displayed closely related MLST and NG-STAR types but relatively distant NG-MAST types (Table 3). Three strains (NJ189125, NJ195417 and NJ1911400) belonged to ST3435, identical to FC428; the NG-MAST of NJ1711654 (ST17114) differed in *porB* by a single base pair (*tbpB* was identical). However, the remaining 10 isolates belonged to 6 different MG-MAST types and displayed relatively large differences in the number of SNPs in *porB* and *tbpB* compared to FC428: ST19387 (22 SNPs in *porB* [1 isolate]); ST19389 (23 SNPs in *porB* [1 isolate]); ST19489 (24 SNPs in *porB* [2 isolates]); ST20048 (25 SNPs in *porB* and 53 SNPs in *tbpB* [3 isolates]); ST20051 (16 SNPs in *porB* and 81 SNPs in *tbpB* [1 isolate]); ST22091 (29 SNPs in *porB* [1 isolate]) and ST21683 (25 SNPs in *porB* and 81 SNPs in *tbpB* [1 isolate]). Nonetheless, the 14 isolates belonged to only 3 MLST types: 10 were MLST1903, identical to FC428; three were MLST 7363 (1 SNP in *abcZ* and 1 SNP in *fumC*) and one was MLST 11710 (7 SNPs in *aroE* and 1 SNP in *fumC*). NG-STAR types were identified as: ST 233 (5 isolates); ST 2238 (3 isolates); ST 1961 and ST 1143 (2 isolates each) and ST 3523 and ST 2239 (1 isolate each). A few SNPs were identified in *porB* (ST 1143, ST 2239 and ST 3523), *parC* (ST 2238), *23S rRNA* (ST 2238) and *gyrA* (ST 1961) alleles. NG-STAR typing identified mosaic *penA* 60.001 in all 14 strains. Notably, 3 strains (NJ1914215, NJ203279 and NJ209649) possessed a C2611T mutation in *23S rRNA* and were resistant to azithromycin.

Table 2 Antimicrobial Susceptibilities of Mosaic *penA* 60.001 Multidrug Resistant *N. Gonorrhoeae* Strains Isolated in Nanjing, China

MIC (mg/L)	NJ1711654	NJ189125	NJ195417	NJ196610	NJ197542	NJ1911400	NJ1913940	NJ1914215
Ceftriaxone	1	1	1	1	1	1	0.5	1
Cefixime	2	≥ 4	≥ 4	≥ 4	≥ 4	2	2	2
Spectinomycin	16	32	32	32	32	32	16	16
Tetracycline	4	4	2	2	2	2	1	1
Ciprofloxacin	≥ 16	≥ 16	≥ 32	≥ 32	≥ 32	≥ 32	≥ 32	16
Azithromycin	0.25	0.25	0.5	0.5	0.25	0.25	0.25	8
Penicillin	4	4	4	8	4	4	4	2
Gentamicin	≤ 4	8	8	8	8	8	8	≤ 4
AZD	0.03	0.06	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125
PPNG	-	-	-	-	-	-	-	-
TRNG	-	-	-	-	-	-	-	-
MIC (mg/L)	NJ1914646	NJ203279	NJ204705	NJ208430	NJ208756	NJ209649	FC428	
Ceftriaxone	0.5	1	1	0.5	1	1	0.5	
Cefixime	2	2	2	2	2	2	1	
Spectinomycin	32	32	16	32	32	16	8	
Tetracycline	1	2	≥ 32	1	2	1	NA	
Ciprofloxacin	16	16	16	≥ 32	≥ 32	16	>32	
Azithromycin	0.25	8	0.25	0.5	0.5	8	0.25	
Penicillin	2	2	4	≥ 16	4	2	NA	
Gentamicin	8	8	8	8	8	8	NA	
AZD	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125	≤ 0.125	NA	
PPNG	-	-	-	+	-	-	NA	
TRNG	-	-	+	-	-	-	NA	

Table 3 Molecular Characteristics of Mosaic penA 60.001 Multidrug Resistant *N. Gonorrhoeae* Strains Isolated in Nanjing, China

Characteristics	NJ1711654	NJ189125	NJ195417	NJ196610	NJ197542	NJ1911400	NJ1913940	NJ1914215
NG-MAST	17114	3435	3435	19,387	19,389	3435	19,489	20,048
<i>porB</i>	2183	1053	1053	11,301	11,303	1053	3531	206
<i>tbpB</i>	21	21	21	21	21	21	21	3021
MLST	1903	1903	1903	1903	1903	1903	1903	7363
<i>abcZ</i>	126	126	126	126	126	126	126	59
<i>adk</i>	39	39	39	39	39	39	39	39
<i>aroE</i>	67	67	67	67	67	67	67	67
<i>fumC</i>	157	157	157	157	157	157	157	78
<i>gdh</i>	148	148	148	148	148	148	148	148
<i>pdhC</i>	153	153	153	153	153	153	153	153
<i>pgm</i>	65	65	65	65	65	65	65	65
NG-STAR	233	233	233	1961	1961	233	1143	2238
<i>penA</i>	60.001	60.001	60.001	60.001	60.001	60.001	60.001	60.001
<i>mtrR</i>	1	1	1	1	1	1	1	1
<i>porB</i>	8	8	8	8	8	8	12	8
<i>ponA</i>	1	1	1	1	1	1	1	1
<i>gyrA</i>	7	7	7	101	101	7	7	7
<i>parC</i>	3	3	3	3	3	3	3	53
23S rRNA	100	100	100	100	100	100	100	2
Characteristics	NJ1914646	NJ203279	NJ204705	NJ208430	NJ208756	NJ209649	FC428	
NG-MAST	20051	20,048	21,683	22,091	19,489	20,048	3435	
<i>porB</i>	6485	206	206	12,096	3531	206	1053	
<i>tbpB</i>	110	3021	3131	21	21	3021	21	
MLST	1903	7363	11,710	1903	1903	7363	1903	
<i>abcZ</i>	126	59	126	126	126	59	126	
<i>adk</i>	39	39	39	39	39	39	39	
<i>aroE</i>	67	67	170	67	67	67	67	
<i>fumC</i>	157	78	78	157	157	78	157	
<i>gdh</i>	148	148	148	148	148	148	148	
<i>pdhC</i>	153	153	153	153	153	153	153	
<i>pgm</i>	65	65	65	65	65	65	65	
NG-STAR	2239	2238	233	3523	1143	2238	233	
<i>penA</i>	60.001	60.001	60.001	60.001	60.001	60.001	60.001	
<i>mtrR</i>	1	1	1	1	1	1	1	
<i>porB</i>	4	8	8	48	12	8	8	
<i>ponA</i>	1	1	1	1	1	1	1	
<i>gyrA</i>	7	7	7	7	7	7	7	
<i>parC</i>	3	53	3	3	3	53	3	
23S rRNA	100	2	100	100	100	2	100	

Whole genomic sequencing (WGS) and resultant phylogenetic analysis demonstrated that 13 FC428-related clones were derived from five clades (A-E in Figure 2 [sequencing of NJ203279, the 14th isolate, was unsuccessful]). NJ1711654, NJ189125, NJ195417 were in clade A (similar to BJ16148 isolated in Beijing)¹⁵ and possessed 100–200 SNPs compared to FC428. NJ196610, NJ197542 and NJ1911400 belonged to a newly identified clade in our collection (clade B) and possessed 283–438 SNPs compared to FC428. NJ208430, NJ208756 and NJ1913940, also a new clade in our collection (clade C), possessed 297–566 SNPs compared to FC428. NJ1914215, NJ206649, NJ1914646 (clade D) represented isolates similar to seven additional isolates in the phylogenetic tree from Hangzhou.¹⁹ Genomic sequences of NJ20204705 and FC428 differed by >1000 SNPs; NJ20204705 was more closely related to G97687 and G7944, which

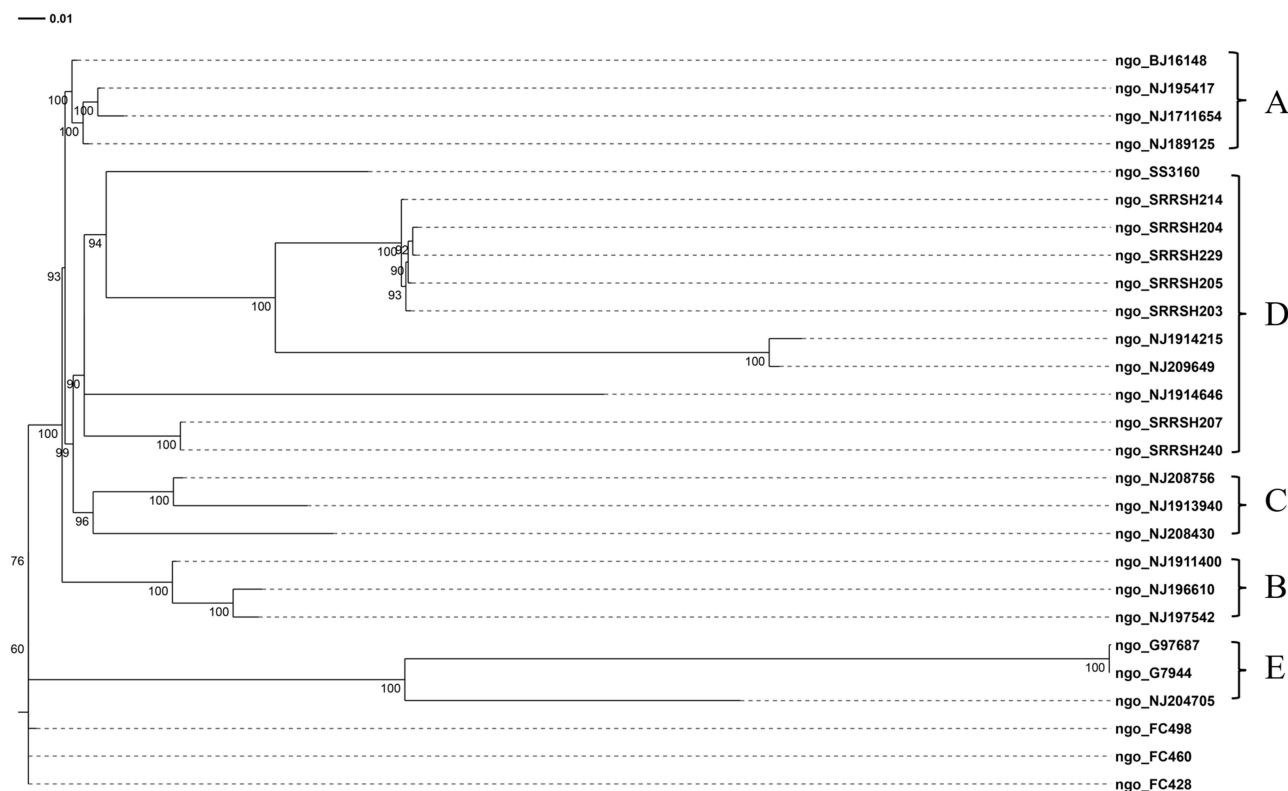


Figure 2 Maximum Likelihood (ML) phylogeny of 13 FC428-like isolates in this study (termed NJ) compared with FC428-like strains from other Chinese and international sites: FC428 is placed as the phylogenetic tree root. 13 FC428-related clones were derived from five clades (A–E). The scale bar indicates the average number of SNPs per site. Whole genome sequences (WGSs) of Nanjing (NJ) isolates were determined in this study. Other WGSs were obtained from public databases.

are English isolates.³³ However, NJ20204705 was susceptible to azithromycin (MIC 0.25ug/mL) while G97687 and G7944 were highly resistant to azithromycin (and ESCs).

Discussion

Sustained spread of *N. gonorrhoeae* FC428 and FC428-like isolates has been demonstrated by their continued emergence, suggesting a possible fitness advantage in humans. However, experiments with *N. gonorrhoeae* FC428 (in vitro and in vivo in mouse models) have shown ambiguous changes in fitness,³⁴ distinct from ceftriaxone-resistant gonococcal strains H041 and F89, for example, that harbor mosaic *penA* alleles and where fitness is reduced,³⁵ possibly explaining why these strains have not been transmitted widely.³⁵ Resistance of *N. gonorrhoeae* FC428 to ESCs, first reported in Japan in 2015,⁶ and identified subsequently in Europe,^{10,12–14,36,37} North America,^{11,38} Southeast Asia (Vietnam,³⁹ Singapore⁴⁰ but not Thailand^{41–43}), is caused by the mosaic *penA*-60.001 allele. In addition to the 14 isolates reported here, thirty-five FC428-like isolates have also been reported from 7 eastern Chinese cities (Figure 3),^{15–19,21,44,45} surpassing the 31 isolates that have been reported from other parts of the world.^{10–14,36–40,46} We report that FC428/FC428-like gonococcal isolates identified in Nanjing increased markedly from 2017 to 2020, exceeding the numbers of isolates reported from any other Chinese city (Figure 3).

In nearly six years, from 2015 to 2021, nine cases of ceftriaxone-resistant *N. gonorrhoeae* were reported from the United Kingdom (UK); all were associated international travel, including two that possessed documented whole genomic similarity to *N. gonorrhoeae* strain FC428, also exhibiting multilocus sequence type ST-1903; *N. gonorrhoeae* multi-antigen sequence type ST-1614 (*porB* 1053, *tbpB* 33) and antimicrobial resistance sequence type ST233.³⁶ In just 6 months, between December 2021 and June 2022, 9 mosaic *penA* 60.001 *N. gonorrhoeae* isolates were obtained from patients presenting to sexual health service across the UK.⁴⁷ MLST types for the 9 isolates were ST16406 (1 isolate) and ST8123 (8 isolates), differing from ST1903, the most prevalent Nanjing MLST type (11 of 14 isolates). ST8123 was the

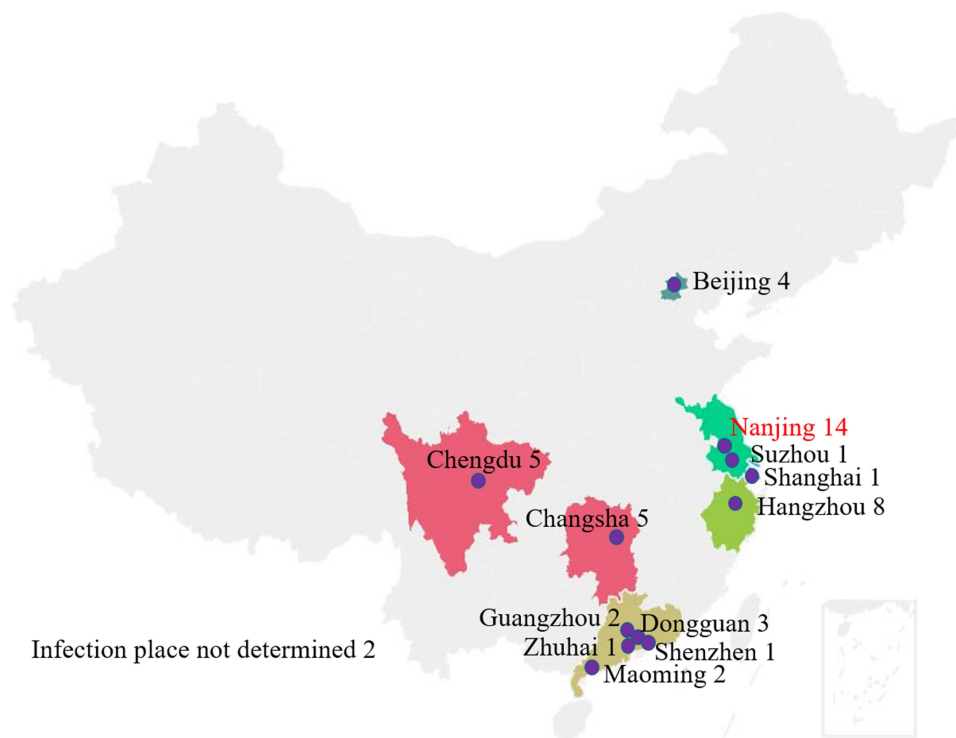


Figure 3 Distribution of FC428-like isolates reported from Chinese cities.

most common ST type among isolates collected in Shenzhen, China, between 2014 and 2018.⁴⁸ The rapid emergence of mosaic *penA* 60.001 *N. gonorrhoeae* in two separate areas of the world suggests that mosaic *penA* 60.001 strains are spreading rapidly throughout the world.

Effective antimicrobial treatment of gonorrhea is essential to interrupt transmission of FC428/FC428-like isolates. Although differences in antimicrobial susceptibilities of the 14 isolates in our study were evident, MICs of ceftriaxone for the 14 isolates were similar to those for FC428.²⁰ Three isolates showed a moderate level of resistance to azithromycin, which has also been reported in FC428-like isolates from several Chinese cities including Hangzhou, Changsha and Shenzhen,^{18,19,21} indicating that FC428-related sub-clones have evolved further and spread in China.

FC428-like genotypes (including NG-MAST, MLST and NG-STAR) have evolved to possess additional gene mutations during their spread. The 14 strains displayed relatively distant NG-MAST types; only one other FC428-like isolate, ST3435, has been reported to harbor an NG-MAST type that differed from the FC428 “signature” NG-MAST type. SNPs in *porB* resulted in different NG-MAST types. MLST1903, which is identical to FC428, was the predominant MLST type;¹⁹ however, three isolates were typed as MLST 7363 (identical to SC18-68 isolated in Chengdu, China¹⁷) and one was typed as MLST 11710 (not yet reported). NG-STAR types were similar to ST 233 (same as the FC428 type). NG-STAR ST1961, ST 2238, ST 2239 and ST 3523 were all new types. NG-STAR ST1143 was identical to FC428-like isolates from Changsha.¹⁸

Phylogenetic analysis showed that 13/14 of the isolates reported in this study were closely related to other FC428-like isolates (Figure 2); these subdivided into 5 clades, which were linked to other clades from Beijing,¹⁵ Hangzhou¹⁹ and England.³³

Conclusion

In summary, we describe fourteen recent ceftriaxone-resistant FC428-related *penA* 60.001 *N. gonorrhoeae* infections in Nanjing, China, which represents a significant rise in this infection locally. Other Chinese cities have seen similar cases; however, reporting from nearby SE Asian countries has been sporadic or absent. In the West, isolated infections and even outbreaks have been caused by this organism. The rapid emergence of FC428-related *penA* 60.001 *N. gonorrhoeae* infections in separate areas of the world suggests that this infection is rapidly gaining a foothold worldwide.

Ethics Approval and Informed Consent

Our study complies with the Declaration of Helsinki. The Ethics Committee of the Institute of Dermatology, Chinese Academy of Medical Sciences and Peking Union Medical College (approval number: 2016-KY 022), and the Institutional Review Board (IRB) at Tufts University School of Medicine (approval number: 12219) approved the study; all enrolled participants provided written informed consent. Participants themselves and a parent or legal guardian of participants under 18 years of age provided written informed consents.

Data Sharing Statement

The data of complete genomic sequences of the thirteen isolates were available from NCBI (BioProject ID: PRJNA553852, PRJNA553854 and PRJNA916595).

Funding

This work was supported by grants from the Chinese Academy of Medical Sciences Initiative for Innovative Medicine (2016-I2M-3-021) and the US National Institutes of Health (AI116969).

Disclosure

No potential conflict of interest was reported by the authors.

References

1. WHO. Gonorrhoea: latest antimicrobial global surveillance results and guidance for vaccine development published; 2021. Available from: <https://www.who.int/news/item/22-11-2021-gonorrhoea-antimicrobial-resistance-results-and-guidance-vaccine-development>. Accessed May 12, 2023.
2. Costa-Lourenço A, Barros Dos Santos KT, Moreira BM, Fracalanza SEL, Bonelli RR. Antimicrobial resistance in *Neisseria gonorrhoeae*: history, molecular mechanisms and epidemiological aspects of an emerging global threat. *Braz J Microbiol*. 2017;48(4):617–628. doi:10.1016/j.bjm.2017.06.001
3. Unemo M, Shafer WM. Antimicrobial resistance in *Neisseria gonorrhoeae* in the 21st century: past, evolution, and future. *Clin Microbiol Rev*. 2014;27(3):587–613. doi:10.1128/cmr.00010-14
4. Ohnishi M, Golparian D, Shimuta K, et al. Is *Neisseria gonorrhoeae* initiating a future era of untreatable gonorrhea?: detailed characterization of the first strain with high-level resistance to ceftriaxone. *Antimicrob Agents Chemother*. 2011;55(7):3538–3545. doi:10.1128/aac.00325-11
5. Lindberg R, Fredlund H, Nicholas R, Unemo M. *Neisseria gonorrhoeae* isolates with reduced susceptibility to cefixime and ceftriaxone: association with genetic polymorphisms in *penA*, *mtrR*, *porB1b*, and *ponA*. *Antimicrob Agents Chemother*. 2007;51(6):2117–2122. doi:10.1128/aac.01604-06
6. Nakayama S, Shimuta K, Furubayashi K, Kawahata T, Unemo M, Ohnishi M. New Ceftriaxone- and Multidrug-Resistant *Neisseria gonorrhoeae* Strain with a Novel Mosaic *penA* Gene Isolated in Japan. *Antimicrob Agents Chemother*. 2016;60(7):4339–4341. doi:10.1128/aac.00504-16
7. Lahra MM, Ryder N, Whaley DM. A new multidrug-resistant strain of *Neisseria gonorrhoeae* in Australia. *N Engl J Med*. 2014;371(19):1850–1851. doi:10.1056/NEJMc1408109
8. Unemo M, Golparian D, Nicholas R, Ohnishi M, Gallay A, Sednaoui P. High-level cefixime- and ceftriaxone-resistant *Neisseria gonorrhoeae* in France: novel *penA* mosaic allele in a successful international clone causes treatment failure. *Antimicrob Agents Chemother*. 2012;56(3):1273–1280. doi:10.1128/aac.05760-11
9. Anselmo A, Ciammaruconi A, Carannante A, et al. Draft genome sequence of *Neisseria gonorrhoeae* sequence type 1407, a multidrug-resistant clinical isolate. *Genome Announc*. 2015;3(4):e00903–00915. doi:10.1128/genomeA.00903-15
10. Lahra MM, Martin I, Demczuk W, et al. Cooperative recognition of internationally disseminated ceftriaxone-resistant *Neisseria gonorrhoeae* strain. *Emerg Infect Dis*. 2018;24(4):735–740. doi:10.3201/eid2404.171873
11. Lefebvre B, Martin I, Demczuk W, et al. Ceftriaxone-Resistant *Neisseria gonorrhoeae*, Canada, 2017. *Emerg Infect Dis*. 2018;24(2):381–383. doi:10.3201/eid2402.171756
12. Terkelsen D, Tolstrup J, Johnsen CH, et al. Multidrug-resistant *Neisseria gonorrhoeae* infection with ceftriaxone resistance and intermediate resistance to azithromycin, Denmark, 2017. *Euro Surveill*. 2017;22(42):17–00659. doi:10.2807/1560-7917.Es.2017.22.42.17-00659
13. Poncin T, Fouere S, Braille A, et al. Multidrug-resistant *Neisseria gonorrhoeae* failing treatment with ceftriaxone and doxycycline in France, November 2017. *Euro Surveill*. 2018;23(21):1800264. doi:10.2807/1560-7917.Es.2018.23.21.1800264
14. Golparian D, Rose L, Lynam A, et al. Multidrug-resistant *Neisseria gonorrhoeae* isolate, belonging to the internationally spreading Japanese FC428 clone, with ceftriaxone resistance and intermediate resistance to azithromycin, Ireland, August 2018. *Euro Surveill*. 2018;23(47):1800617. doi:10.2807/1560-7917.Es.2018.23.47.1800617
15. Chen SC, Han Y, Yuan LF, Zhu XY, Yin YP. Identification of internationally disseminated ceftriaxone-resistant *Neisseria gonorrhoeae* strain FC428, China. *Emerg Infect Dis*. 2019;25(7):1427–1429. doi:10.3201/eid2507.190172
16. Chen SC, Yuan LF, Zhu XY, van der Veen S, Yin YP. Sustained transmission of the ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone in China. *J Antimicrob Chemother*. 2020;75(9):2499–2502. doi:10.1093/jac/dkaa196
17. Wang H, Wang Y, Yong G, et al. Emergence and genomic characterization of the ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone in Chengdu, China. *J Antimicrob Chemother*. 2020;75(9):2495–2498. doi:10.1093/jac/dkaa123
18. Yuan Q, Li Y, Xiu L, et al. Identification of multidrug-resistant *Neisseria gonorrhoeae* isolates with combined resistance to both ceftriaxone and azithromycin, China, 2017–2018. *Emerg Microbes Infect*. 2019;8(1):1546–1549. doi:10.1080/22221751.2019.1681242

19. Yan J, Chen Y, Yang F, et al. High percentage of the ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone among isolates from a single hospital in Hangzhou, China. *J Antimicrob Chemother.* 2021;76(4):936–939. doi:10.1093/jac/dkaa526
20. Lin X, Chen W, Xie Q, et al. Dissemination and genome analysis of high-level ceftriaxone-resistant penA 60.001 *Neisseria gonorrhoeae* strains from the Guangdong Gonococcal antibiotics susceptibility Programme (GD-GASP), 2016–2019. *Emerg Microbes Infect.* 2022;11(1):344–350. doi:10.1080/22221751.2021.2011618
21. Zhang L, Zhang C, Zeng Y, et al. Emergence and characterization of a ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone evolving moderate-level resistance to azithromycin in Shenzhen, China. *Infect Drug Resist.* 2021;14:4271–4276. doi:10.2147/idr.S336212
22. Unemo M, Golparian D, Linnios A, et al. In vitro activity of ertapenem versus ceftriaxone against *Neisseria gonorrhoeae* isolates with highly diverse ceftriaxone MIC values and effects of ceftriaxone resistance determinants: ertapenem for treatment of gonorrhoea? *Antimicrob Agents Chemother.* 2012;56(7):3603–3609. doi:10.1128/aac.00326-12
23. Li X, Le W, Lou X, Genco CA, Rice PA, Su X. In vitro activity of ertapenem against *Neisseria gonorrhoeae* clinical isolates with decreased susceptibility or resistance to extended-spectrum Cephalosporins in Nanjing, China (2013 to 2019). *Antimicrob Agents Chemother.* 2022;66(5):e0010922. doi:10.1128/aac.00109-22
24. Clinical and Laboratory Standards Institute. *Performance Standards for Antimicrobial Susceptibility Testing*. 28th ed. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.
25. Testing TECoAS. Breakpoint tables for interpretation of MICs and zone diameters. Version 50; 2015. Available from: <http://www.eucast.org>. Accessed May 12, 2023.
26. Mann LM, Kirkcaldy RD, Papp JR, Torrone EA. Susceptibility of *Neisseria gonorrhoeae* to Gentamicin-Gonococcal Isolate Surveillance Project, 2015–2016. *Sex Transm Dis.* 2018;45(2):96–98. doi:10.1097/olq.0000000000000693
27. Taylor SN, Marrazzo J, Batteiger BE, et al. Single-Dose Zoliflodacin (ETX0914) for treatment of urogenital gonorrhoea. *N Engl J Med.* 2018;379(19):1835–1845. doi:10.1056/NEJMoa1706988
28. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing: twenty-third informational supplement. CLSI document M100-S23. *Clinical Lab Standards Institute.* 2013;33:100–102.
29. Su X, Jiang F, Qimuge D. Surveillance of antimicrobial susceptibilities in *Neisseria gonorrhoeae* in Nanjing, China, 1999–2006. *Sex Transm Dis.* 2007;34(12):995–999. doi:10.1097/OLQ.0b013e3180ca8f24
30. Martin IM, Ison CA, Aanensen DM, Fenton KA, Spratt BG. Rapid sequence-based identification of gonococcal transmission clusters in a large metropolitan area. *J Infect Dis.* 2004;189(8):1497–1505. doi:10.1086/383047
31. Jolley KA, Maiden MC. BIGSdb: scalable analysis of bacterial genome variation at the population level. *BMC Bioinform.* 2010;11:595. doi:10.1186/1471-2105-11-595
32. Demczuk W, Sidhu S, Unemo M, et al. *Neisseria gonorrhoeae* sequence typing for antimicrobial resistance, a novel antimicrobial resistance multilocus typing scheme for tracking global dissemination of *N. gonorrhoeae* strains. *J Clin Microbiol.* 2017;55(5):1454–1468. doi:10.1128/jcm.00100-17
33. Eyre DW, Sanderson ND, Lord E, et al. Gonorrhoea treatment failure caused by a *Neisseria gonorrhoeae* strain with combined ceftriaxone and high-level azithromycin resistance, England, February 2018. *Euro Surveill.* 2018;23(27):1800323. doi:10.2807/1560-7917.Es.2018.23.27.1800323
34. Zhou K, Chen SC, Yang F, van der Veen S, Yin YP. Impact of the gonococcal FC428 penA allele 60.001 on ceftriaxone resistance and biological fitness. *Emerg Microbes Infect.* 2020;9(1):1219–1229. doi:10.1080/22221751.2020.1773325
35. Vincent LR, Kerr SR, Tan Y, et al. In vivo-selected compensatory mutations restore the fitness cost of mosaic penA alleles that confer ceftriaxone resistance in *Neisseria gonorrhoeae*. *mBio.* 2018;9(2):e01905–01917. doi:10.1128/mBio.01905-17
36. Eyre DW, Town K, Street T, et al. Detection in the United Kingdom of the *Neisseria gonorrhoeae* FC428 clone, with ceftriaxone resistance and intermediate resistance to azithromycin, October to December 2018. *Euro Surveill.* 2019;24(10):1900147. doi:10.2807/1560-7917.Es.2019.24.10.1900147
37. Poncin T, Merimeche M, Braille A, et al. Two cases of multidrug-resistant *Neisseria gonorrhoeae* related to travel in south-Eastern Asia, France, June 2019. *Euro Surveill.* 2019;24(36):1900528. doi:10.2807/1560-7917.Es.2019.24.36.1900528
38. Berenger BM, Demczuk W, Gratrix J, Pabbaraju K, Smyczek P, Martin I. Genetic characterization and enhanced surveillance of ceftriaxone-resistant *Neisseria gonorrhoeae* strain, Alberta, Canada, 2018. *Emerg Infect Dis.* 2019;25(9):1660–1667. doi:10.3201/eid2509.190407
39. Trinh TM, Nguyen TT, Le TV, et al. *Neisseria gonorrhoeae* FC428 Subclone, Vietnam, 2019–2020. *Emerg Infect Dis.* 2022;28(2):432–435. doi:10.3201/eid2802.211788
40. Chio MTW, Goh SS, Tan AL, Koh TH, Abdul Rahman NB. First case of ceftriaxone-resistant multidrug-resistant *Neisseria gonorrhoeae* in Singapore. *Antimicrob Agents Chemother.* 2019;63(5):e02624–02618. doi:10.1128/aac.02624-18
41. Nokchan N, Wongsurawat T, Jenjaroenpun P, Nitayanon P, Tribuddharat C. Whole-genome sequence analysis of high-level penicillin-resistant strains and antimicrobial susceptibility of *Neisseria gonorrhoeae* clinical isolates from Thailand. *PLoS One.* 2022;17(7):e0271657. doi:10.1371/journal.pone.0271657
42. Golparian D, Kittiyaowamarn R, Paopang P, et al. Genomic surveillance and antimicrobial resistance in *Neisseria gonorrhoeae* isolates in Bangkok, Thailand in 2018. *J Antimicrob Chemother.* 2022;77(8):2171–2182. doi:10.1093/jac/dkac158
43. Kueakulpattana N, Wannigama DL, Luk-In S, et al. Multidrug-resistant *Neisseria gonorrhoeae* infection in heterosexual men with reduced susceptibility to ceftriaxone, first report in Thailand. *Sci Rep.* 2021;11(1):21659. doi:10.1038/s41598-021-00675-y
44. Lin X, Qin X, Wu X, et al. Markedly increasing antibiotic resistance and dual treatment of *Neisseria gonorrhoeae* isolates in Guangdong, China, from 2013 to 2020. *Antimicrob Agents Chemother.* 2022;66(4):e0229421. doi:10.1128/aac.02294-21
45. Yang F, Zhang H, Chen Y, et al. Detection and analysis of two cases of the internationally spreading ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone in China. *J Antimicrob Chemother.* 2019;74(12):3635–3636. doi:10.1093/jac/dkz384
46. Lee K, Nakayama SI, Osawa K, et al. Clonal expansion and spread of the ceftriaxone-resistant *Neisseria gonorrhoeae* strain FC428, identified in Japan in 2015, and closely related isolates. *J Antimicrob Chemother.* 2019;74(7):1812–1819. doi:10.1093/jac/dkz129
47. Day M, Pitt R, Mody N, et al. Detection of 10 cases of ceftriaxone-resistant *Neisseria gonorrhoeae* in the United Kingdom, December 2021 to June 2022. *Euro Surveill.* 2022;27(46):2200803. doi:10.2807/1560-7917.Es.2022.27.46.2200803
48. Li Y, Li Y, Xiu L, et al. Typing of *Neisseria Gonorrhoeae* isolates in Shenzhen, China from 2014–2018 reveals the shift of genotypes associated with antimicrobial resistance. *Antimicrob Agents Chemother.* 2021;65(5):e02311–02320. doi:10.1128/aac.02311-20

Infection and Drug Resistance

Dovepress

Publish your work in this journal

Infection and Drug Resistance is an international, peer-reviewed open-access journal that focuses on the optimal treatment of infection (bacterial, fungal and viral) and the development and institution of preventive strategies to minimize the development and spread of resistance. The journal is specifically concerned with the epidemiology of antibiotic resistance and the mechanisms of resistance development and diffusion in both hospitals and the community. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/infection-and-drug-resistance-journal>