



# Draft Genome Sequence of *Candida auris* Strain LOM, a Human Clinical Isolate from Greater Metropolitan Houston, Texas

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**ABSTRACT** *Candida auris* is an emerging pathogen of considerable public health importance. We present the draft genome sequence of a strain recently cultured from the urine of a patient hospitalized in the greater Houston metropolitan region. Two combined Oxford Nanopore sequencing runs provided sufficient data to rapidly generate a draft genome.

*Candida auris* recently has emerged worldwide and caused outbreaks in health care facilities (1, 2). An isolate of *C. auris* was cultured from the urine of a patient from the Houston metropolitan region on 5% sheep blood tryptic soy agar and identified by Brucker matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) analysis using the research use only (RUO) v4.1.80 database. The patient had not recently traveled outside the United States and had transferred from a long-term-care facility. This work was approved by our institutional review board (IRB1010-0199).

The organism was classified as present on admission. Given the potential infection control and public health importance of this isolate, we rapidly characterized its genome using two Oxford Nanopore GridION runs using FLO-MIN106 flow cells and the Guppy v2.0.10 base caller. DNA was extracted from overnight growth on solid agar using ballistic lysis with FastPrep matrix B (first run) or Y (second run) and the Qiagen DNA blood and tissue kit. The first GridION run used the rapid barcoding kit (catalog number SQK-RBK004), yielding 1.29 million reads and 3.48 gigabases (Gb) of sequence with an average read length of 2.7 kb. The second run used the ligation sequencing kit (catalog number SQK-LSK109) with a long-read wash, yielding 2.98 million reads and 13.72 Gb of sequence with an average read length of 4.6 kb. The reads from both runs were combined and filtered using Filtlong v0.1.1 with a 5-kb-read cutoff and 100-fold coverage (<https://github.com/rrwick/Filtlong>). Unicycler v0.4.3 was used to assemble the genome with miniasm and pilon polishing (3). The assembly had 9 contigs (total length, 12,293,266 bp). The largest contig was 4,297,164 bp, and the  $N_{50}$  value was 2,304,466 bp. The average GC content was 45.19%.

We compared our assembly with 7 reference genome assemblies in GenBank (strains B11221, RCPF-1821, 6684, B8441, B11220, B11243, and VPCI 479/P/13) using progressiveMauve v2.4.0 and discovered that our 7 longest contigs correspond to the 7 chromosomes present in other *C. auris* strains (4). Phylogenetic analysis using MUMmer v4.0 (with the show-snps setting) showed that *C. auris* strain LOM is most closely related to strain B11221, a strain from clade III (1,177 single-nucleotide polymorphisms [SNPs] distant) (5, 6). Prephix and phrecon (<https://github.com/codinghedgehog/>) were used to generate an alignment from the MUMmer SNPs to create a neighbor-joining tree using FastTree2 (Fig. 1) (7). The LOM genome has the *erg11* gene with an F126L amino acid replacement that is common to clade III strains (6).

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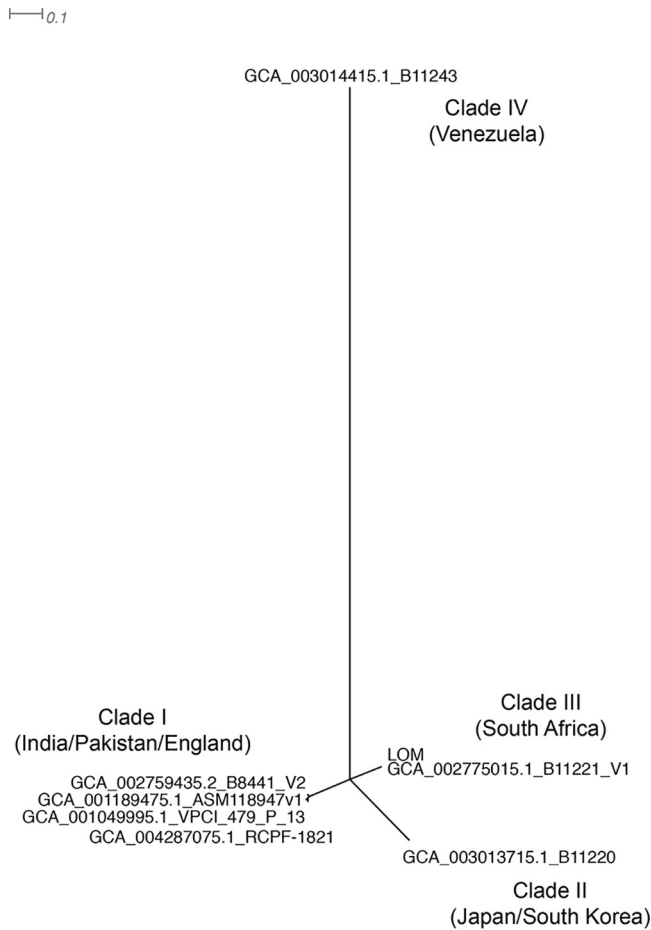
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**FIG 1** Neighbor-joining radial phylogenetic tree showing the relationship of *C. auris* strain LOM to the 7 reference strain genomes, B11221 (GenBank accession number [GCA\\_002775015](#)), RCPF-1821 ([GCA\\_004287075](#)), 6684 ([GCA\\_001189475](#)), B8441 ([GCA\\_002759435](#)), B11220 ([GCA\\_003013715](#)), B11243 ([GCA\\_003014415](#)), and VPCI 479/P/13 ([GCA\\_001049995](#)). The four clades and their respective geographic associations are indicated.

This work further demonstrates the usefulness of real-time long-read whole-genome sequencing to rapidly provide relevant information concerning emerging pathogens of significant concern (8, 9). The availability of this high-quality draft genome sequence will serve as a useful resource if additional isolates are recovered in the greater Houston metropolitan area. The sequence data also will assist translational research efforts designed to more fully understand the molecular mechanisms underlying host interactions in an emerging pathogen that has substantial detrimental public health potential.

**Data availability.** The BioProject accession number for *C. auris* strain LOM is [PRJNA540998](#), the GridION runs are in the SRA (numbers [SRR9017243](#) and [SRR9017244](#)), and the draft genome is in GenBank (accession number [SZYF00000000](#)). The 7 reference strains and their GenBank genome assembly accession numbers are as follows: B11221 ([GCA\\_002775015](#)), RCPF-1821 ([GCA\\_004287075](#)), 6684 ([GCA\\_001189475](#)), B8441 ([GCA\\_002759435](#)), B11220 ([GCA\\_003013715](#)), B11243 ([GCA\\_003014415](#)), and VPCI 479/P/13 ([GCA\\_001049995](#)).

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## REFERENCES

1. Spivak ES, Hanson KE. 2018. *Candida auris*: an emerging fungal pathogen. *J Clin Microbiol* 56:e01588-17. <https://doi.org/10.1128/JCM.01588-17>.
2. Jeffery-Smith A, Taori SK, Schelenz S, Jeffery K, Johnson EM, Borman A, Manuel R, Brown CS. 2017. *Candida auris*: a review of the literature. *Clin Microbiol Rev* 31:e00029-17. <https://doi.org/10.1128/CMR.00029-17>.
3. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
4. Darling AE, Tritt A, Eisen JA, Facciotti MT. 2011. Mauve assembly metrics. *Bioinformatics* 27:2756–2757. <https://doi.org/10.1093/bioinformatics/btr451>.
5. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12. <https://doi.org/10.1186/gb-2004-5-2-r12>.
6. Muñoz JF, Gade L, Chow NA, Loparev VN, Juieng P, Berkow EL, Farrer RA, Litvintseva AP, Cuomo CA. 2018. Genomic insights into multidrug-resistance, mating and virulence in *Candida auris* and related emerging species. *Nat Commun* 9:5346. <https://doi.org/10.1038/s41467-018-07779-6>.
7. Price MN, Dehal PS, Arkin AP. 2010. FastTree 2—approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. <https://doi.org/10.1371/journal.pone.0009490>.
8. Long SW, Kachroo P, Musser JM, Olsen RJ. 2017. Whole-genome sequencing of a human clinical isolate of emm28 *Streptococcus pyogenes* causing necrotizing fasciitis acquired contemporaneously with Hurricane Harvey. *Genome Announc* 5:e01269-17. <https://doi.org/10.1128/genomeA.01269-17>.
9. Quick J, Loman NJ, Duraffour S, Simpson JT, Severi E, Cowley L, Bore JA, Koundouno R, Dudas G, Mikhail A, Ouedraogo N, Afrough B, Bah A, Baum JH, Becker-Ziaja B, Boettcher JP, Cabeza-Cabrero M, Camino-Sanchez A, Carter LL, Doerrbecker J, Enkirch T, Dorival IGG, Hetzelt N, Hinzmann J, Holm T, Kafetzopoulou LE, Koropogui M, Kosgey A, Kuisma E, Logue CH, Mazzarelli A, Meisel S, Mertens M, Michel J, Ngabo D, Nitzsche K, Pallash E, Patrono LV, Portmann J, Repits JG, Rickett NY, Sachse A, Singethan K, Vitoriano I, Yemanaberhan RL, Zekeng EG, Trina R, Bello A, Sall AA, Faye O, et al. 2016. Real-time, portable genome sequencing for Ebola surveillance. *Nature* 530:228–232. <https://doi.org/10.1038/nature16996>.