






Complete Genome Sequences of Four *Campylobacter jejuni* Strains Isolated from Retail Chicken Meat and Broiler Feces

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ABSTRACT *Campylobacter jejuni* is the leading pathogen that causes foodborne infections. Here, we report the complete genome sequences of four *C. jejuni* strains isolated from retail chicken meat and broiler feces samples. Genes encoding type VI secretion and antibiotic resistance were detected among these isolates.

Campylobacteriosis, caused by *Campylobacter jejuni*, is one of the major foodborne infections that cause human gastroenteritis, and poultry is considered the major reservoir host (1–6). Therefore, to understand the variation in molecular characteristics of *C. jejuni* poultry isolates, whole-genome sequencing was performed to determine detailed information about the virulence and antimicrobial resistance genes (6–8).

Four *C. jejuni* strains isolated and identified in our previous study (9) were further utilized. Each *C. jejuni* strain was cultured in Bolton broth (Thermo Fisher Scientific, USA) and incubated under microaerophilic conditions for 48 h at 42°C; genomic DNA was then extracted using the GeneJET genomic DNA purification kit (Thermo Fisher Scientific). For long-read sequencing, genomic DNA was fragmented using a g-TUBE device (Covaris, USA) and the fragments size selected to a mean of 8 to 12 kb using AMPure XP beads (Beckman Coulter, USA) for library preparation. A multiplexing library pool was prepared and barcoded using a genomic DNA ligation kit (SQK-LSK109) and the native barcoding genomic DNA kit (SQK-NBD104), respectively; the library was sequenced on an R9.4 MinION flow cell using the Nanopore GridION sequencer (Oxford Nanopore Technologies, Oxford, UK) for 48 h. For short-read sequencing, Illumina TruSeq DNA PCR-free (insert size, 350 bp) library preparation and sequencing using the paired-end 150-bp (PE150) method was performed by Novogene on the HiSeq X Ten sequencer (Illumina, USA). The raw Nanopore reads were assembled into contigs using Canu v1.9 (10). The first 1,000 bp of each contig was mapped to the entire contig using blastn from the BLAST+ v2.9.0 tool suite (11); then, the overlapping regions were removed using SAMtools v1.9 (12). The start of each circular contig that had an overlapping region was moved upstream of the *dnaA* gene, when present, or upstream of the gene closest to the middle of the contig using Circlator v1.5.5 (13). The junction site of circular contigs was verified using PCR amplification and Sanger sequencing (14). The Illumina and Nanopore reads were mapped to the assembled contigs using BWA v0.7.17-r1188 (15) and minimap2 v2.17 (16), respectively. Any contig without supporting Illumina data was removed. The remaining contigs were polished using Pilon v1.23 (17). The genome sequences were annotated using Prokka v1.13 (18).

The genome sizes and number of protein-coding DNA sequences of the isolates range from 1.63 Mb to 1.84 Mb and 1,421 to 1,552, respectively, and two isolates

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TABLE 1 Summary of genome assembly and genotype characteristics of four *C. jejuni* isolates

| Strain | Source | GenBank accession no. | Size (bp) | Illumina | | Nanopore | | GC content (%) | No. of CDSs ^a | ARG(s) ^b | Virulence genes | |
|--------------------------------------|---------------------|-----------------------|-----------|--------------|---------------------------|--------------|---------------------------|----------------|--------------------------|---------------------|--|---|
| | | | | No. of reads | Avg per-base coverage (×) | No. of reads | Avg per-base coverage (×) | | | | | |
| <i>C. jejuni</i> MS2005 ^c | Retail chicken meat | CP084080 | 1,757,242 | 3,476,551 | 571 | 164,000 | 736 | 10,111 | 30.5 | 1,541 | <i>aph(3')-IIIa</i> , <i>sat-4</i> , <i>cmeR</i> | <i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssI</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i> |
| <i>C. jejuni</i> MS2005 ^d | Retail chicken meat | CP084081 | 5,208 | | 6,746 | | 405 | | 28.5 | 6 | | |
| <i>C. jejuni</i> MS2058 ^c | Retail chicken meat | CP084082 | 1,758,823 | 5,455,985 | 890 | 104,000 | 472 | 10,256 | 30.5 | 1,552 | <i>aph(3')-IIIa</i> , <i>sat-4</i> , <i>cmeR</i> | <i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssI</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i> |
| <i>C. jejuni</i> MS2058 ^d | Retail chicken meat | CP084083 | 40,825 | | 1,086 | | 231 | | 28.5 | 41 | <i>tet(O)</i> | |
| <i>C. jejuni</i> MS2074 ^c | Retail chicken meat | CP084084 | 1,629,343 | 5,376,179 | 986 | 144,000 | 691 | 9,909 | 30.7 | 1,491 | <i>bla</i> _{OXA-61} , <i>cmeR</i> | |
| <i>C. jejuni</i> MS2167 ^c | Broiler feces | CP084085 | 1,711,301 | 5,762,419 | 1,007 | 220,000 | 951 | 9,302 | 30.5 | 1,491 | <i>cmeR</i> | <i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssI</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i> |

^a CDSs, protein-coding DNA sequences.

^b ARG(s), antimicrobial resistance gene(s).

^c Chromosome.

^d Plasmid.

(MS2005 and MS2058) contain one plasmid each. The respective fold coverages for Illumina and Nanopore ranged from 541 to 864 and 440 to 851, and the N_{50} values ranged from 9,302 to 10,256 bp, respectively. *In silico* prediction of antimicrobial resistance genes using CARD v3.1.0 (19) identified *aph(3')-IIIa*, *sat-4*, and *bla*_{OXA-61} genes in the chromosomes, whereas plasmid pMS2058 contains the *tet(O)* gene. Using SecReT6 (20), the presence of type VI secretion genes was predicted in the chromosomes of three *C. jejuni* strains MS2005, MS2058, and MS2167 (Table 1). The genome information obtained in this study can be further utilized for the identification of potential vaccine candidates and functional analysis of type VI secretion genes.

Data availability. The genome sequences and raw data are available at NCBI GenBank under the BioProject accession number [PRJNA655459](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA655459). The specific parameters and code used for sequencing can be found at https://github.com/IGBB/campylobacter_jejuni.

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