

MDPI

Revieu

Role of Neuroglobin in the Neuroprotective Actions of Estradiol and Estrogenic Compounds

George E. Barreto ¹, Andrew J. McGovern ¹ and Luis M. Garcia-Segura ²,*

- Department of Biological Sciences, University of Limerick, V94 T9PX Limerick, Ireland; George.Barreto@ul.ie (G.E.B.); Andrew.McGovern@ul.ie (A.J.M.)
- ² Instituto Cajal, Consejo Superior de Investigaciones Científicas (C.S.I.C.), 28002 Madrid, Spain
- * Correspondence: lmgs@cajal.csic.es; Tel.: +34-915-854-729

Abstract: Estradiol exerts neuroprotective actions that are mediated by the regulation of a variety of signaling pathways and homeostatic molecules. Among these is neuroglobin, which is upregulated by estradiol and translocated to the mitochondria to sustain neuronal and glial cell adaptation to injury. In this paper, we will discuss the role of neuroglobin in the neuroprotective mechanisms elicited by estradiol acting on neurons, astrocytes and microglia. We will also consider the role of neuroglobin in the neuroprotective actions of clinically relevant synthetic steroids, such as tibolone. Finally, the possible contribution of the estrogenic regulation of neuroglobin to the generation of sex differences in brain pathology and the potential application of neuroglobin as therapy against neurological diseases will be examined.

Keywords: astrocytes; estrogen receptors; microglia; mitochondria; neuron; sex differences; tibolone



Citation: Barreto, G.E.; McGovern, A.J.; Garcia-Segura, L.M. Role of Neuroglobin in the Neuroprotective Actions of Estradiol and Estrogenic Compounds. *Cells* **2021**, *10*, 1907. https://doi.org/10.3390/ cells10081907

Academic Editors: Roberta Misasi, Maria Marino and Margherita Ruoppolo

Received: 5 July 2021 Accepted: 26 July 2021 Published: 27 July 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

The central nervous system (CNS) is a target for the ovarian hormone estradiol, which not only regulates the activity of the neuronal circuits controlling reproductive physiology, sexual behavior and energy homeostasis, but also of those participating in the processing of tactile, nociceptive, visual and auditory information, motor coordination, emotions, verbal skills, cognition, learning and memory. Estradiol also activates protective homoeostatic responses in neurons, astrocytes, microglia, oligodendrocytes and endothelial cells to maintain the function of all of these circuits under pathological conditions.

After CNS injury, the hormone reduces edema, oxidative stress, neuroinflammation, altered ionic balance, apoptosis, loss of cerebral blood flow and other tissue alterations that cause secondary neuronal death and contribute to expanding the damage over other CNS regions. In parallel, estradiol activates reparative mechanisms through the regulation of autophagy and the promotion of synaptic plasticity, postnatal neurogenesis and myelin repair [1]. In consequence, estradiol is neuroprotective in animal models of depression, stroke, traumatic CNS injury, multiple sclerosis, amyotrophic lateral sclerosis, Parkinson's disease, diabetic and hypertensive encephalopathies, Alzheimer's disease and other illnesses of the nervous system [1–6].

Both under physiological conditions and after CNS injury, estradiol regulates neuronal and glial cell function by a coordinated activation of extranuclear and nuclear signaling events [1]. Nuclear-initiated estrogen signaling is mediated by classical estrogen receptors (ERs), α (ER α) and β (ER β), which are transcription factors expressed by neurons, astrocytes and microglia [7,8] and are widely distributed in numerous brain regions, including those that are frequently involved in neurodegenerative diseases, such as the hippocampus, cerebral cortex and the striatum [7,8]. These receptors can be coexpressed in the same individual brain cells [7] and, after being activated by the hormone, form homo or heterodimers, recruit coactivators, corepressors and other components of the transcriptional machinery and regulate transcription by binding to estrogen response elements in the

Cells 2021, 10, 1907 2 of 12

promoters of target genes, or by interacting with other transcription factors in the cell nucleus [9].

Extranuclear-initiated estrogen signaling is mediated by classical ERs localized at extranuclear sites, such as at lipid rafts of the plasma membrane [10] or at the mitochondria [11], or by activating other types of ERs, such as G protein-coupled estrogen receptor-1 or Gq-coupled membrane estrogen receptor at the plasma membrane or the endoplasmic reticulum [12,13]. Extranuclear estrogen signaling results in the modification of the activity of ion channels, kinases, phosphatases and the signaling pathways of membrane-associated receptors, such as insulin-like growth factor-1 (IGF-1), Wnt and Notch signaling [1,12–14]. All these signaling pathways modulated by estradiol converge in the regulation of transcriptional events that are in turn coordinated with the transcriptional regulation activated by nuclear estrogen signaling [15]. Under conditions of brain injury, the activation of these nuclear and extranuclear initiated signaling mechanisms mediate the neuroprotective actions of estradiol.

The neuroprotective actions of estradiol are in part exerted through the transcriptional upregulation of neurotrophic factors and other protective molecules [1,14]. One such molecule is neuroglobin (Ngb) [16], a globin protein family member that is expressed by neurons and glial cells in the mammalian central and peripheral nervous system [17,18] and exerts neuroprotection [19,20]. Here we will review the protective actions of neuroglobin over glial and neuronal cells, focusing especially on the neuroglobin-enhancing actions of hormonal compounds as a therapeutic strategy to alleviate brain damage.

2. Neuroglobin Neuroprotective Actions

In neural cells, neuroglobin interacts with proteins involved in signal transduction [16] and regulates the activity of different signaling pathways, including PI3K [21], AMP-activated protein kinase (AMPK) [22] and Wnt signaling [23]. In addition, neuroglobin regulates mitochondrial dynamics [19], mitochondrial respiratory function and ATP production [24,25] and regulates cell metabolism [22]. However, the physiological consequences of all these effects of neuroglobin in the mammalian brain have been only partially explored. Nevertheless, it is known that neuroglobin promotes neurogenesis, possibly by increasing PSA-NCAM, Tuj1 and involving Wnt signaling [23,25,26] and neuritogenesis [27,28].

In aquatic mammals, such as whales and seals, neuroglobin is involved in the physiological adaptation to hypoxia when submerged [29], suggesting that in terrestrial mammals, neuroglobin may participate in the adaptation to brain hypoxia under pathological conditions. In agreement with this possibility, it was shown that neuroglobin protects neural cells against hypoxia-ischemia [24,30,31], oxidative stress [32] and oxygen/glucose deprivation [33] and reduces infarct lesion after experimental stroke [34–36]. Moreover, the fact that vascular endothelial growth factor (VEGF) stimulates neuroglobin through VEGFR2/Flk1 [37] may also suggest the involvement of neuroglobin in neovascularization and angiogenesis, two essential repairing cellular mechanisms that are severely disturbed in cerebral ischemia.

The investigation of the possible protective actions of neuroglobin and its molecular mechanisms has been addressed using a variety of cellular and animal models [38]. These studies have revealed that the neuroprotective actions of neuroglobin are not limited to hypoxic-ischemic insults. For instance, overexpression of neuroglobin in transgenic mice using adenoviral vectors reduces lesion size [39] and improves sensorimotor behavior [40] after traumatic brain injury (TBI) and promotes locomotor recovery after spinal cord injury [41]. In addition, neuroglobin overexpression protects neuronal cells from NMDA [42] and β -amyloid toxicity [42–44] and decreases behavioral deficits in mice with Alzheimer's disease [40]. On the other hand, the expression of neuroglobin declines in the brain in Huntington's disease mouse model and in Alzheimer's disease patients in parallel to the increase in pathological alterations [45,46].

Cells **2021**, *10*, 1907 3 of 12

Mitochondria have emerged as a central component in the neuroprotective actions of neuroglobin [19,20]. Indeed, neuroglobin is physically localized in neuronal mitochondria [47] and its overexpression in primary neurons exposed to hypoxia results in the preservation of mitochondrial membrane potential and respiration, the enhancement of ATP production and reduced oxidative stress [48]. In addition, neuroglobin interacts with several mitochondrial proteins involved in the regulation of the intrinsic pathway of apoptosis, such as cytochrome-c₁, voltage-dependent anion channel 1 (VDAC) or huntingtin (HTT) [16,49–51] and protects neuronal cells by inhibiting this cell death pathway [52].

The neuroprotective mechanisms of neuroglobin also include actions outside the mitochondria. For instance, oxidative stress promotes the recruitment of neuroglobin into lipid raft microdomains of the plasma membrane. There, neuroglobin interacts with flotillin-1 and α -subunits of heterotrimeric G proteins, preventing the decrease in cAMP levels induced by oxidative stress and promoting neuronal viability [32]. Neuroglobin also preserves the activity of Na⁺/K⁺ ATPases in the membrane, as it has been detected in the hippocampus in a model of transient global cerebral ischemia [53]. Na⁺/K⁺ ATPases are essential to maintain intracellular ion homeostasis and membrane excitability. Indeed, the preservation of its activity by neuroglobin after transient global cerebral ischemia is associated with reduced damage to CA1 hippocampal neurons [53]. Another localization of neuroglobin are the neuronal growth cones [54], structures involved in axonal growth and axonal regeneration. Indeed, neuroglobin has been shown to promote the regeneration of CNS axons after traumatic injury or ischemia [54,55].

The preservation of the activity of Na^+/K^+ ATPases by neuroglobin is associated with an increased membranous level of Na^+/K^+ ATPases $\beta 1$ subunit in astrocytes [53], suggesting a role of these glial cells in the mechanisms of neuroprotection. This is in agreement with the fact that neuroglobin expression is enhanced in astrocytes and microglia after brain injury [53,56–59]. Furthermore, it has been shown that exogenous intravitreal injection of neuroglobin reduces the activation of microglia, the expression of inflammatory cytokines and the apoptosis of retinal ganglion cells in hypoxic retina [60]. Therefore, the control of neuroinflammation by astrocytes and microglia may participate in the neuroprotective response mediated by neuroglobin.

Astrocytes not only mediate neuroglobin neuroprotection, but they are also protected themselves by neuroglobin. Thus, neuroglobin increases the viability of astrocytes exposed to glucose deprivation or oxidative stress [61–63]. To better understand the importance of neuroglobin for cell survival, neuroglobin-depleted astrocytes lose mitochondrial membrane potential ($\Delta \Psi m$), alongside increases in superoxide (O^-_2) and hydrogen peroxide levels (H_2O_2), while attenuating the levels of two antioxidant enzymes: catalase and superoxide dismutase 2 (SOD2, mitochondrial) [64,65]. This may be indicative of loss of mitochondria integrity and function, thereby promoting cell death. In agreement with this, when knocking down neuroglobin, pAKT/AKT is downregulated in an in vitro scratch and metabolic injury model, therefore worsening cell viability [65]. Given the essential trophic and metabolic support provided by astrocytes to neurons, it should be expected that the protection exerted by neuroglobin on astrocytes should also contribute to sustain neuronal viability. In this regard, it is noticeable that neuroglobin is transported in exosomes from astrocytes to neurons [66], a mechanism that may help to coordinate the protective responses in both cell types.

Despite the benefits of neuroglobin over neuronal and glial cells post-injury, its clinical use is hampered because it is unable to cross membranes, or even penetrate through the blood–brain barrier (BBB), due to its physical properties and hydrophilic exterior. To circumvent this problem, transactivator of transcription protein (TAT) has been used to deliver neuroglobin to the brain. Indeed, rabbits treated with neuroglobin coupled with TAT transduction domain (TAT-Ngb) had improved neuronal outcome in terms of a significant reduction in apoptotic mechanisms in a model of subarachnoid hemorrhage [67]. Furthermore, TAT-Ngb fusion proteins preserved mitochondrial function upon oxidative damage by inducing a significant rise in superoxide dismutase, heme-oxygenase 1 (HO-1)

Cells 2021, 10, 1907 4 of 12

and nuclear factor erythroid 2-related factor 2 (Nrf2) [68]. However, some challenges have been mentioned in the attempts made to facilitate neuroglobin delivery to the brain. For example, TAT-Ngb-FITC has been shown to penetrate cortical neurons in vitro, but a caveat of its therapeutic efficacy was the absence of fluorescent signals after 48 h [69]. This is a major downside when considering times post-injury, a critical period in neurological diseases as traumatic brain injury. Furthermore, SH-SY5Y (dopaminergic neurons) pretreated with TAT-Ngb (400 nM-1 μM) 2 h prior to oxygen glucose deprivation (OGD) for 18 or 36 h had no improvements on cell viability [70]. On the contrary, TAT-Ngb rescued differentiated PC12 cells from OGD and inhibited mitochondrial apoptosis, possibly by modulating the Jak stat family [71]. In accordance with this in vitro study, administration of TAT-Ngb to mice 2 h prior to middle cerebral artery occlusion (MCAO) reduced infarct volume at 24 h reperfusion and decreased neuronal death by 72 h after MCAO [35]. Although promising, most studies using neuroglobin delivery approaches have been performed on brain ischemia in vivo and in vitro models, demonstrating the urgent need to implement these approaches for other brain diseases. More recently, the use of sodium hyaluronate to facilitate the delivery of neuroglobin following MCAO and 24 h reperfusion has been explored [72]. Although this nanocarrier successfully delivered neuroglobin to the brain, the potential biological implication of the rise of its cytosolic levels was not fully explored.

3. Interaction of Estradiol and Neuroglobin Neuroprotective Actions

The neuroprotective actions of estradiol and neuroglobin share several common mechanisms. In part, this is due to the fact that the neuroglobin gene is upregulated by estradiol in neurons [73] and glial cells [57]. The regulation of neuroglobin expression by estradiol is exerted through membrane-initiated estrogen signaling, involving ER β activation of p38/MAPK [73], and also by direct nuclear-initiated transcriptional regulation of the neuroglobin gene by ER α and ER β , which probably act by binding to other transcription factors, such as Sp1, activator protein 1 (AP-1) or NF- κ B, given the absence of canonical estrogen response elements the neuroglobin promoter [73–76].

It is possible that the mechanisms of neuroglobin regulation by estradiol may differ between different cell types and depending on the physiological conditions. For instance, in astrocytes, the regulation of neuroglobin expression is modified by inflammatory signals. Thus, under basal conditions the induction of neuroglobin expression by estradiol in astrocytes is mainly regulated through ER β . However, a synergic action of ER α and ER β , which are coexpressed in astrocytes, is necessary to maintain the neuroglobin expression upregulated when NF- κ B is activated by lipopolysaccharide (LPS) in these cells [57]. Another factor involved in the estrogenic regulation of neuroglobin is huntingtin (HTT), because in the mouse hippocampus and in murine striatal neurons the polyQ mutation of HTT causes the disease and impairs the upregulation of neuroglobin by estradiol [51].

In addition to increasing neuroglobin expression, estradiol promotes the accumulation of neuroglobin in the mitochondria in neuronal cells [50]. HTT is also necessary for this hormonal effect [51]. Indeed, estradiol promotes the expression of HTT and the formation of an HTT-neuroglobin complex and its translocation to the mitochondria [51]. In the mitochondria, HTT [77] and neuroglobin [49] interact with proteins involved in the control of apoptosis, such as VDAC. Through VDAC, neuroglobin inhibits the opening of mitochondrial permeability transition pore (mPTP) and the subsequent cytochrome c release [33]. Interestingly, estradiol also promotes the association of neuroglobin with cytochrome c [50]. This effect of estradiol is enhanced under conditions of oxidative stress and is one of the mechanisms activated by the hormone to prevent apoptosis. Thus, in neuroblastoma cells exposed to H_2O_2 , the hormone further increases the mitochondrial association of neuroglobin with cytochrome c, preventing its release from the mitochondria to the cytosol and, therefore, the initiation of the apoptotic cascade [50].

Neuroglobin may also be involved in the neuroprotective actions of estradiol mediated by glial cells. Indeed, part of the neuroprotective actions of estradiol are mediated by the

Cells 2021, 10, 1907 5 of 12

control of reactive gliosis and the reduction of the inflammatory response of astrocytes and microglia [78,79].

Neuroinflammatory response requires complex homeostatic regulation, because from one side neuroinflammation is an acute protective response but it may enhance neuronal damage when it is decontrolled as, for instance, under chronic neurodegenerative conditions. The regulation of neuroglobin expression in astrocytes is a good example of this complex homeostatic regulation. Thus, from one side inflammatory signaling through NF- κ B, and estradiol signaling through ER β , promote neuroglobin expression in astrocytes, probably as a protective response [57]. However, in parallel, estradiol inhibits NF- κ B activation and the inflammatory response of astrocytes by a mechanism involving ER α , and, in doing so, the hormone also reduces NF- κ B-mediated induction of neuroglobin [57]. These opposite actions of estradiol may be interpreted as a mechanism to maintain the level of the inflammatory response under an adequate control.

4. Potential Role of Neuroglobin in the Generation of Sex Differences in Brain Pathology

The estrogenic actions in the nervous system contribute to sex differences in the incidence, prevalence, age of onset or symptomatology that are observed in numerous neurodegenerative and affective disorders [8], including pathological conditions in which neuroglobin has been reported to exert neuroprotective actions in animal models. It is plausible that a sex-specific regulation of neuroglobin expression in neurons and glial cells by circulating estradiol, or by estradiol locally synthesized by brain aromatase enzyme [80], may contribute to the generation of these differences. However, this possibility has not received enough attention in the experimental designs, which usually utilize only one animal sex. Nevertheless, it has been reported that female mice have higher basal levels of neuroglobin in the hippocampus [51] and the striatum [46] than males. However, it is still unknown if these differences offer a better protection of the female hippocampus and striatum against pathological insults.

Sex differences in the expression of neuroglobin after brain injury have also been scarcely explored, although a significant increase in neuroglobin expression has been observed in the striatum of transgenic male mice affected by Huntington's disease, but not in female animals affected by the pathology [46]. In contrast, neuroglobin mRNA levels increase in the cerebral cortex of female rats, but not in the male cortex, after ischemic stroke [81]. Since there is a significant negative correlation between the expression of neuroglobin and the volume of cortical infarct [81], sex differences in neuroglobin expression after brain injury may be potentially involved in the decreased brain infarct volume observed in young female animals in the acute phase of ischemic stroke [82].

Sex differences have also been observed in the cellular localization of neuroglobin in a mouse model of a penetrating traumatic brain injury. In this model, neuroglobin expression in the injured cerebral cortex was restricted to the tissue located in proximity to the wound and was detected in neurons, reactive astrocytes and microglia [57,59]. The area of neuroglobin expression near the wound on day 7 post-injury was similar in 2-month-old males and females. However, the cellular distribution of neuroglobin was different, with males presenting a higher colocalization of neuroglobin with the microglia marker ionized calcium binding adaptor molecule 1 (Iba1) than females [59]. A similar outcome was also observed in 5-month-old male animals under a controlled cortical impact model showing an upregulation of neuroglobin in the ipsilateral hemisphere by day 7 after injury [40]. Interestingly, a positive correlation seems to exist between the severity of the damage and the degree of neuroglobin expression in males [83]. However, the pattern of neuroglobin expression over post-TBI periods might depend on the time of assessment. Thus, a thorough evaluation of spatiotemporal neuroglobin levels is utterly necessary for a precise time course of its evolution during acute and chronic TBI periods.

The consequences of these differences in neuroglobin cellular distribution for the functional response of the cerebral cortex to injury are still unknown. Although males show a higher neuronal survival in the lesion border of the cerebral cortex than females [59], this

Cells 2021, 10, 1907 6 of 12

could not be ascribed to the differences in neuroglobin localization in microglia. However, this question merits further investigation, considering that neuroglobin expression in microglia is observed after different forms of cellular injury [58,84,85] and that these cells react to pathological insults with sex-specific inflammatory, migratory and phagocytic responses [8].

5. Role of Neuroglobin in the Neuroprotective Actions of Natural and Synthetic Compounds with Estrogenic Activity

Health benefits have been associated with the use of several natural compounds with estrogenic activity, such as the isoflavone genistein, which is known to exert neuroprotective actions in animal models of neurodegeneration [86–88]. It has been reported that genistein—and another isoflavone, formononetin—increases neuroglobin expression in primary neurons [89], suggesting that neuroglobin may participate in the neuroprotective actions of phytoestrogens and that these molecules may be used as pharmacological inductors of neuroglobin.

Synthetic steroids with estrogenic activity have also been shown to induce neuroglobin expression in neural cells. Thus, it has been reported that the synthetic steroid diarylpropionitrile (DPN), a selective agonist of ER β that exerts neuroprotective actions [90–92], induces the expression of neuroglobin in SK-N-BE neuroblastoma cells, while propyl pyrazole triol (PTT), a selective agonist of ER α , does not [73].

Another synthetic steroid with estrogenic actions that has been shown to upregulate neuroglobin in neural cells is tibolone. Tibolone is a selective tissue estrogenic activity regulator currently used in clinical practice for the treatment of osteoporosis and climacteric symptoms. Following administration, tibolone is converted into 3α -hydroxytibolone, 3β -hydroxytibolone and $\delta 4$ -tibolone [93–96]. These metabolites are highly lipophilic, able to cross plasma membranes, hence penetrating the BBB, and act as agonists of androgen (AR), ERs and progesterone (PGR) receptors. Thus, 3α and 3β metabolites preferentially bind to ERs, mainly ER α , while $\delta 4$ -tibolone binds to AR and PGR [97]. In the nervous system, tibolone exerts neuroprotective actions and promotes cognition [98,99], probably by acting in part over glial cells through the attenuation of the reactive activation of astrocytes and microglia after traumatic brain injury [100], and the regulation of astrocyte phagocytosis of brain cellular debris [101].

The induction of neuroglobin signaling is one of the neuroprotective mechanisms activated by tibolone (Figure 1). Upon binding to tibolone metabolites (α , β , and δ), ER, ARs and PGR are activated, migrate to the nucleus and interact with steroid hormone response elements localized in the promotor or upstream of Ngb gene. Although this genomic mechanism may stimulate the expression of neuroglobin in both glial cells and neurons, this protein can also be regulated by non-genomic signaling through activation of PI3k/AKT and p38/MAPK. Once activated, neuroglobin may interact with G protein alpha subunit (Gα) and phosphatidylinositol 3,4,5-trisphosphate 3-phosphatase (PTEN) leading to the activation of survival signaling in neurons and glial cells. Estrogenic signaling not only increases neuroglobin expression, but also induces its translocation to the mitochondria [50]. Then, neuroglobin interacts in the mitochondria with cytochrome c, mitochondrial complex proteins and scavenge ROS and NOS. Collectively, these actions carried out by neuroglobin indicate its antioxidative, anti-inflammatory, and antiapoptotic properties in the brain. Finally, by regulating these pathways, tibolone reduces the inflammatory response and increases the viability of microglia cells exposed to a metabolic insult by a mechanism involving ERβ activation, NF-κB inhibition and neuroglobin upregulation [85]. A similar mechanism, involving mainly ER β , but also ER α mediated neuroglobin upregulation, is activated by tibolone to protect astrocytes from glucose deprivation [62]. The neuroprotective actions of tibolone are of interest, given that this compound is currently used in clinical practice. However, systemic off-target effects of tibolone may limit its use as a treatment for specific brain diseases.

Cells **2021**, 10, 1907 7 of 12

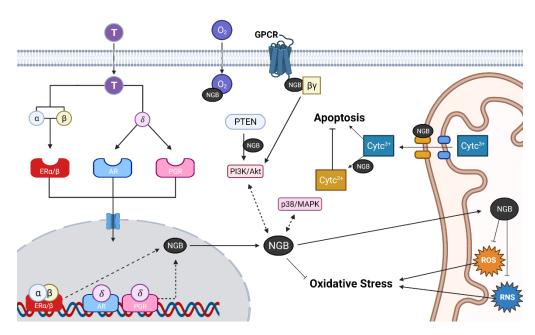


Figure 1. Transcriptional and post-translational regulation of neuroglobin (NGB) and biological functions. Abbreviations: T, tibolone; α , 3α -hydroxytibolone; β , 3β -hydroxytibolone; δ , δ 4-tibolone; AR, androgen receptor; ER α , estrogen receptor alpha; ER β , estrogen receptor beta; PGR, progesterone receptor; cytc, cytochrome c; $\beta\gamma$, G protein beta and gamma subunits; PTEN, phosphatidylinositol 3,4,5-trisphosphate 3-phosphatase; ROS, reactive oxygen species; RNS, reactive nitrogen species.

6. Conclusions and Perspectives

Since its discovery and until the present day, neuroglobin has shown enormous potential as a therapy against inflammation and oxidative stress due to its common pathological features across many neurological diseases, including stroke, Alzheimer's disease and traumatic brain injury, among others. The fact that in some CNS diseases, such as Alzheimer's, the intracellular levels of neuroglobin decrease exponentially with time of injury suggests the intrinsic participation of this protein in the pathological mechanism. It should be noted that the expression of neuroglobin and its downstream signaling might be different depending on the sex, and therefore further studies exploring the sex dimorphic role of neuroglobin is expected.

Perhaps one of the greatest current challenges is to seek strategies that aim to increase both the levels and the cellular expression of neuroglobin. One strategy that has been explored in the last few years is the delivery of neuroglobin to increase its brain levels, especially in those diseases characterized by a significant reduction of endogenous neuroglobin. Considering the inability of this protein to cross membranes after brain delivery, it seems that the effect is transient and limited as its intracellular levels do not remain for a long time inside cells post-injury. Even if delivery is successful and steady, further studies are needed to delve into whether a rise in neuroglobin is directly correlated with an increased neuroprotection, so future work to unravel this possible relationship is warranted.

Finally, endogenously, neuroglobin can be stimulated by genomic and nongenomic pathways, where a wide range of pharmacological compounds have the ability to promote its expression. One of these groups capable of stimulating the transcriptional activity of Ngb gene in neurons and glial cells are compounds with estrogenic activity, which can be those of endogenous origin such as estradiol or of synthetic origin such as tibolone. In both cases, brain cells treated with these estrogenic compounds have a better outcome against an inflammatory stimulus, possibly by activating the endogenous synthesis of neuroglobin and ERs, since the blockade of the estrogen receptors (i.e., ER β) is able to reduce the protection mediated by this protein. A limiting factor for a long-term therapy with estrogenic compounds is related to the side effects that can occur in other peripheral organs. Therefore, the discovery and design of more specific ligands using high-throughput

Cells 2021, 10, 1907 8 of 12

screening and artificial intelligence/machine learning [102] targeting estrogen receptors only in the brain is desirable with further studies going forward in this direction.

In conclusion, regardless of the approach that may favor and stimulate neuroglobin, the neuroprotective potential of this protein is yet far from being fully explored in males and females. Unravelling which other biological functions this protein has in the brain upon injury merits to be the subject of future studies.

Author Contributions: L.M.G.-S.; writing—original draft preparation. G.E.B. and A.J.M.; writing—review and editing. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Azcoitia, I.; Barreto, G.E.; Garcia-Segura, L.M. Molecular mechanisms and cellular events involved in the neuroprotective actions of estradiol. Analysis of sex differences. *Front. Neuroendocrinol.* **2019**, *55*, 100787. [CrossRef]

- 2. Elkabes, S.; Nicot, A.B. Sex steroids and neuroprotection in spinal cord injury: A review of preclinical investigations. *Exp. Neurol.* **2014**, 259, 28–37. [CrossRef]
- 3. Labandeira-Garcia, J.L.; Rodriguez-Perez, A.I.; Valenzuela, R.; Costa-Besada, M.A.; Guerra, M.J. Menopause and Parkinson's disease. Interaction between estrogens and brain renin-angiotensin system in dopaminergic degeneration. *Front. Neuroendocrinol.* **2016**, *43*, 44–59. [CrossRef] [PubMed]
- 4. Engler-Chiurazzi, E.B.; Brown, C.M.; Povroznik, J.M.; Simpkins, J.W. Estrogens as neuroprotectants: Estrogenic actions in the context of cognitive aging and brain injury. *Prog. Neurobiol.* **2017**, *157*, 188–211. [CrossRef]
- 5. Céspedes Rubio, Á.E.; Pérez-Alvarez, M.J.; Lapuente Chala, C.; Wandosell, F. Sex steroid hormones as neuroprotective elements in ischemia models. *J. Endocrinol.* **2018**, 237, R65–R81. [CrossRef]
- 6. Bourque, M.; Morissette, M.; Di Paolo, T. Repurposing sex steroids and related drugs as potential treatment for Parkinson's disease. *Neuropharmacology* **2019**, *147*, 37–54. [CrossRef]
- 7. Almey, A.; Milner, T.A.; Brake, W.G. Estrogen receptors in the central nervous system and their implication for dopamine-dependent cognition in females. *Horm. Behav.* **2015**, *74*, 125–138. [CrossRef]
- 8. Chowen, J.A.; Garcia-Segura, L.M. Role of glial cells in the generation of sex differences in neurodegenerative diseases and brain aging. *Mech. Ageing Dev.* **2021**, *196*, 111473. [CrossRef] [PubMed]
- 9. Marino, M.; Galluzzo, P.; Ascenzi, P. Estrogen signaling multiple pathways to impact gene transcription. *Curr. Genom.* **2006**, *7*, 497–508. [CrossRef]
- 10. Marin, R.; Casañas, V.; Pérez, J.A.; Fabelo, N.; Fernandez, C.E.; Diaz, M. Oestrogens as modulators of neuronal signalosomes and brain lipid homeostasis related to protection against neurodegeneration. *J. Neuroendocrinol.* **2013**, 25, 1104–1115. [CrossRef] [PubMed]
- 11. Yang, S.H.; Liu, R.; Perez, E.J.; Wen, Y.; Stevens, S.M., Jr.; Valencia, T.; Brun-Zinkernagel, A.M.; Prokai, L.; Will, Y.; Dykens, J.; et al. Mitochondrial localization of estrogen receptor beta. *Proc. Natl. Acad. Sci. USA* 2004, 101, 4130–4135. [CrossRef]
- 12. Gaudet, H.M.; Cheng, S.B.; Christensen, E.M.; Filardo, E.J. The G-protein coupled estrogen receptor, GPER: The inside and inside-out story. *Mol. Cell. Endocrinol.* **2015**, *418 Pt 3*, 207–219. [CrossRef]
- 13. Vail, G.; Roepke, T.A. Membrane-initiated estrogen signaling via Gq-coupled GPCR in the central nervous system. *Steroids* **2019**, 142, 77–83. [CrossRef]
- 14. Arevalo, M.A.; Azcoitia, I.; Garcia-Segura, L.M. The neuroprotective actions of oestradiol and oestrogen receptors. *Nat. Rev. Neurosci.* 2015, *16*, 17–29. [CrossRef]
- 15. Fiocchetti, M.; Ascenzi, P.; Marino, M. Neuroprotective effects of 17β-estradiol rely on estrogen receptor membrane initiated signals. *Front. Physiol.* **2012**, *3*, 73. [CrossRef]
- 16. Ascenzi, P.; di Masi, A.; Leboffe, L.; Fiocchetti, M.; Nuzzo, M.T.; Brunori, M.; Marino, M. Neuroglobin: From structure to function in health and disease. *Mol. Asp. Med.* **2016**, *52*, 1–48. [CrossRef]
- 17. Fabrizius, A.; Andre, D.; Laufs, T.; Bicker, A.; Reuss, S.; Porto, E.; Burmester, T.; Hankeln, T. Critical re-evaluation of neuroglobin expression reveals conserved patterns among mammals. *Neuroscience* **2016**, 337, 339–354. [CrossRef] [PubMed]
- 18. Van Acker, Z.P.; Luyckx, E.; Dewilde, S. Neuroglobin Expression in the Brain: A Story of Tissue Homeostasis Preservation. *Mol. Neurobiol.* **2019**, *56*, 2101–2122. [CrossRef] [PubMed]
- 19. Fiocchetti, M.; Cracco, P.; Montalesi, E.; Solar Fernandez, V.; Stuart, J.A.; Marino, M. Neuroglobin and mitochondria: The impact on neurodegenerative diseases. *Arch. Biochem. Biophys.* **2021**, *701*, 108823. [CrossRef] [PubMed]
- 20. Gorabi, A.M.; Aslani, S.; Barreto, G.E.; Báez-Jurado, E.; Kiaie, N.; Jamialahmadi, T.; Sahebkar, A. The potential of mitochondrial modulation by neuroglobin in treatment of neurological disorders. *Free Radic. Biol. Med.* **2021**, 162, 471–477. [CrossRef] [PubMed]

Cells 2021, 10, 1907 9 of 12

21. Antao, S.T.; Duong, T.T.; Aran, R.; Witting, P.K. Neuroglobin overexpression in cultured human neuronal cells protects against hydrogen peroxide insult via activating phosphoinositide-3 kinase and opening the mitochondrial K(ATP) channel. *Antioxid. Redox Signal.* **2010**, *13*, 769–781. [CrossRef]

- 22. Cai, B.; Li, W.; Mao, X.; Winters, A.; Ryou, M.G.; Liu, R.; Greenberg, D.A.; Wang, N.; Jin, K.; Yang, S.H. Neuroglobin overexpression inhibits AMPK signaling and promotes cell anabolism. *Mol. Neurobiol.* **2016**, *53*, 1254–1265. [CrossRef]
- 23. Yu, Z.; Cheng, C.; Liu, Y.; Liu, N.; Lo, E.H.; Wang, X. Neuroglobin promotes neurogenesis through Wnt signaling pathway. *Cell Death Dis.* **2018**, *9*, 945. [CrossRef] [PubMed]
- 24. Greenberg, D.A.; Jin, K.; Khan, A.A. Neuroglobin: An endogenous neuroprotectant. *Curr. Opin. Pharmacol.* **2008**, *8*, 20–24. [CrossRef] [PubMed]
- 25. Lechauve, C.; Augustin, S.; Cwerman-Thibault, H.; Bouaita, A.; Forster, V.; Célier, C.; Rustin, P.; Marden, M.C.; Sahel, J.A.; Corral-Debrinski, M. Neuroglobin involvement in respiratory chain function and retinal ganglion cell integrity. *Biochim. Biophys. Acta.* 2012, 1823, 2261–2273. [CrossRef]
- Luyckx, E.; Van Leuven, W.; Andre, D.; Quarta, A.; Reekmans, K.; Fransen, E.; Moens, L.; Hankeln, T.; Ponsaerts, P.; Dewilde, S. Loss of neuroglobin expression alters CDKN1A/CDK6-expression resulting in increased proliferation of neural stem cells. Stem Cells Dev. 2018, 27, 378–390. [CrossRef]
- 27. Li, L.; Liu, Q.R.; Xiong, X.X.; Liu, J.M.; Lai, X.J.; Cheng, C.; Pan, F.; Chen, Y.; Yu, S.B.; Yu, A.C.; et al. Neuroglobin promotes neurite outgrowth via differential binding to PTEN and Akt. *Mol. Neurobiol.* **2014**, 49, 149–162. [CrossRef] [PubMed]
- 28. Takahashi, N.; Onozuka, W.; Watanabe, S.; Wakasugi, K. Chimeric ZHHH neuroglobin acts as a cell membrane-penetrating inducer of neurite outgrowth. *FEBS Open Bio* **2017**, *7*, 1338–1349. [CrossRef]
- 29. Schneuer, M.; Flachsbarth, S.; Czech-Damal, N.U.; Folkow, L.P.; Siebert, U.; Burmester, T. Neuroglobin of seals and whales: Evidence for a divergent role in the diving brain. *Neuroscience* **2012**, 223, 35–44. [CrossRef] [PubMed]
- 30. Sun, Y.; Jin, K.; Mao, X.O.; Zhu, Y.; Greenberg, D.A. Neuroglobin is up-regulated by and protects neurons from hypoxic-ischemic injury. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 15306–15311. [CrossRef]
- 31. Khan, A.A.; Mao, X.O.; Banwait, S.; DerMardirossian, C.M.; Bokoch, G.M.; Jin, K.; Greenberg, D.A. Regulation of hypoxic neuronal death signaling by neuroglobin. *FASEB J.* **2008**, 22, 1737–1747. [CrossRef] [PubMed]
- 32. Watanabe, S.; Takahashi, N.; Uchida, H.; Wakasugi, K. Human neuroglobin functions as an oxidative stress-responsive sensor for neuroprotection. *J. Biol. Chem.* **2012**, *287*, 30128–30138. [CrossRef]
- 33. Yu, Z.; Liu, N.; Li, Y.; Xu, J.; Wang, X. Neuroglobin overexpression inhibits oxygen-glucose deprivation-induced mitochondrial permeability transition pore opening in primary cultured mouse cortical neurons. *Neurobiol. Dis.* **2013**, *56*, 95–103. [CrossRef] [PubMed]
- 34. Sun, Y.; Jin, K.; Peel, A.; Mao, X.O.; Xie, L.; Greenberg, D.A. Neuroglobin protects the brain from experimental stroke in vivo. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 3497–3500. [CrossRef] [PubMed]
- 35. Cai, B.; Lin, Y.; Xue, X.H.; Fang, L.; Wang, N.; Wu, Z.Y. TAT-mediated delivery of neuroglobin protects against focal cerebral ischemia in mice. *Exp. Neurol.* **2011**, 227, 224–231. [CrossRef]
- 36. Raida, Z.; Hundahl, C.A.; Nyengaard, J.R.; Hay-Schmidt, A. Neuroglobin over expressing mice: Expression pattern and effect on brain ischemic infarct size. *PLoS ONE* **2013**, *8*, e76565. [CrossRef]
- 37. Jin, K.; Mao, X.; Xie, L.; Greenberg, D.A. Interactions between vascular endothelial growth factor and neuroglobin. *Neurosci. Lett.* **2012**, *519*, 47–50. [CrossRef]
- 38. Luyckx, E.; Van Acker, Z.P.; Ponsaerts, P.; Dewilde, S. Neuroglobin Expression Models as a Tool to Study Its Function. *Oxid. Med. Cell Longev.* **2019**, 2019, 5728129. [CrossRef]
- 39. Zhao, S.; Yu, Z.; Zhao, G.; Xing, C.; Hayakawa, K.; Whalen, M.J.; Lok, J.M.; Lo, E.H.; Wang, X. Neuroglobin-overexpression reduces traumatic brain lesion size in mice. *BMC Neurosci.* **2012**, *13*, 67. [CrossRef]
- 40. Taylor, J.M.; Kelley, B.; Gregory, E.J.; Berman, N.E. Neuroglobin overexpression improves sensorimotor outcomes in a mouse model of traumatic brain injury. *Neurosci. Lett.* **2014**, *577*, 125–129. [CrossRef]
- 41. Lan, W.B.; Lin, J.H.; Chen, X.W.; Wu, C.Y.; Zhong, G.X.; Zhang, L.Q.; Lin, W.P.; Liu, W.N.; Li, X.; Lin, J.L. Overexpressing neuroglobin improves functional recovery by inhibiting neuronal apoptosis after spinal cord injury. *Brain Res.* **2014**, 1562, 100–108. [CrossRef]
- 42. Khan, A.A.; Mao, X.O.; Banwait, S.; Jin, K.; Greenberg, D.A. Neuroglobin attenuates beta-amyloid neurotoxicity in vitro and transgenic Alzheimer phenotype in vivo. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 19114–19119. [CrossRef]
- 43. Li, R.C.; Pouranfar, F.; Lee, S.K.; Morris, M.W.; Wang, Y.; Gozal, D. Neuroglobin protects PC12 cells against beta-amyloid-induced cell injury. *Neurobiol. Aging* **2008**, 29, 1815–1822. [CrossRef] [PubMed]
- 44. Li, Y.; Dai, Y.B.; Sun, J.Y.; Xiang, Y.; Yang, J.; Dai, S.Y.; Zhang, X. Neuroglobin attenuates beta amyloid-induced apoptosis through inhibiting caspases activity by activating PI3K/Akt signaling pathway. *J. Mol. Neurosci.* **2016**, *58*, 28–38. [CrossRef]
- 45. Szymanski, M.; Wang, R.; Fallin, M.D.; Bassett, S.S.; Avramopoulos, D. Neuroglobin and Alzheimer's dementia: Genetic association and gene expression changes. *Neurobiol. Aging* **2010**, *31*, 1835–1842. [CrossRef] [PubMed]
- 46. Cardinale, A.; Fusco, F.R.; Paldino, E.; Giampà, C.; Marino, M.; Nuzzo, M.T.; D'Angelo, V.; Laurenti, D.; Straccia, G.; Fasano, D.; et al. Localization of neuroglobin in the brain of R6/2 mouse model of Huntington's disease. *Neurol. Sci.* 2018, 39, 275–285. [CrossRef] [PubMed]

Cells 2021, 10, 1907 10 of 12

47. Yu, Z.; Xu, J.; Liu, N.; Wang, Y.; Li, X.; Pallast, S.; van Leyen, K.; Wang, X. Mitochondrial distribution of neuroglobin and its response to oxygen-glucose deprivation in primary-cultured mouse cortical neurons. *Neuroscience* 2012, 218, 235–242. [CrossRef]

- 48. Liu, J.; Yu, Z.; Guo, S.; Lee, S.R.; Xing, C.; Zhang, C.; Gao, Y.; Nicholls, D.G.; Lo, E.H.; Wang, X. Effects of neuroglobin overexpression on mitochondrial function and oxidative stress following hypoxia/reoxygenation in cultured neurons. *J. Neurosci. Res.* 2009, 87, 164–170. [CrossRef]
- 49. Yu, Z.; Liu, N.; Wang, Y.; Li, X.; Wang, X. Identification of neuroglobin-interacting proteins using yeast two-hybrid screening. Neuroscience 2012, 200, 99–105. [CrossRef] [PubMed]
- 50. De Marinis, E.; Fiocchetti, M.; Acconcia, F.; Ascenzi, P.; Marino, M. Neuroglobin upregulation induced by 17β-estradiol sequesters cytocrome c in the mitochondria preventing H₂O₂-induced apoptosis of neuroblastoma cells. *Cell Death Dis.* **2013**, *4*, e508. [CrossRef]
- Nuzzo, M.T.; Fiocchetti, M.; Totta, P.; Melone, M.A.B.; Cardinale, A.; Fusco, F.R.; Gustincich, S.; Persichetti, F.; Ascenzi, P.; Marino, M. Huntingtin polyQ Mutation Impairs the 17β-Estradiol/Neuroglobin Pathway Devoted to Neuron Survival. *Mol. Neurobiol.* 2017, 54, 6634–6646. [CrossRef]
- 52. Raychaudhuri, S.; Skommer, J.; Henty, K.; Birch, N.; Brittain, T. Neuroglobin protects nerve cells from apoptosis by inhibiting the intrinsic pathway of cell death. *Apoptosis* **2010**, *15*, 401–411. [CrossRef] [PubMed]
- 53. Wen, H.; Liu, L.; Zhan, L.; Liang, D.; Li, L.; Liu, D.; Sun, W.; Xu, E. Neuroglobin mediates neuroprotection of hypoxic postconditioning against transient global cerebral ischemia in rats through preserving the activity of Na⁺/K⁺ ATPases. *Cell Death Dis.* **2018**, *9*, 635. [CrossRef]
- 54. Xiong, X.X.; Pan, F.; Chen, R.Q.; Hu, D.X.; Qiu, X.Y.; Li, C.Y.; Xie, X.Q.; Tian, B.; Chen, X.Q. Neuroglobin boosts axon regeneration during ischemic reperfusion via p38 binding and activation depending on oxygen signal. *Cell Death. Dis.* **2018**, *9*, 163. [CrossRef] [PubMed]
- 55. Sugitani, K.; Koriyama, Y.; Sera, M.; Arai, K.; Ogai, K.; Wakasugi, K. A novel function of neuroglobin for neuroregeneration in mice after optic nerve injury. *Biochem. Biophys. Res. Commun.* **2017**, 493, 1254–1259. [CrossRef]
- 56. DellaValle, B.; Hempel, C.; Kurtzhals, J.A.; Penkowa, M. In vivo expression of neuroglobin in reactive astrocytes during neuropathology in murine models of traumatic brain injury, cerebral malaria, and autoimmune encephalitis. *Glia* **2010**, *58*, 1220–1227. [CrossRef] [PubMed]
- 57. De Marinis, E.; Acaz-Fonseca, E.; Arevalo, M.A.; Ascenzi, P.; Fiocchetti, M.; Marino, M.; Garcia-Segura, L.M. 17beta-Oestradiol anti-inflammatory effects in primary astrocytes require oestrogen receptor beta-mediated neuroglobin up-regulation. *J. Neuroendocrinol.* **2013**, 25, 260–270. [CrossRef]
- 58. Li, W.D.; Sun, Q.; Zhang, X.S.; Wang, C.X.; Li, S.; Li, W.; Hang, C.H. Expression and cell distribution of neuroglobin in the brain tissue after experimental subarachnoid hemorrhage in rats: A pilot study. *Cell. Mol. Neurobiol.* **2014**, *34*, 247–255. [CrossRef]
- 59. Acaz-Fonseca, E.; Duran, J.C.; Carrero, P.; Garcia-Segura, L.M.; Arevalo, M.A. Sex differences in glia reactivity after cortical brain injury. *Glia* **2015**, *63*, 1966–1981. [CrossRef] [PubMed]
- 60. Tun, S.B.B.; Barathi, V.A.; Luu, C.D.; Lynn, M.N.; Chan, A.S.Y. Effects of Exogenous Neuroglobin (Ngb) on retinal inflammatory chemokines and microglia in a rat model of transient hypoxia. *Sci. Rep.* **2019**, *9*, 18799. [CrossRef]
- 61. Amri, F.; Ghouili, I.; Amri, M.; Carrier, A.; Masmoudi-Kouki, O. Neuroglobin protects astroglial cells from hydrogen peroxide-induced oxidative stress and apoptotic cell death. *J. Neurochem.* **2017**, *140*, 151–169. [CrossRef]
- 62. Avila-Rodriguez, M.; Garcia-Segura, L.M.; Hidalgo-Lanussa, O.; Baez, E.; Gonzalez, J.; Barreto, G.E. Tibolone protects astrocytic cells from glucose deprivation through a mechanism involving estrogen receptor beta and the upregulation of neuroglobin expression. *Mol. Cell. Endocrinol.* **2016**, 433, 35–46. [CrossRef]
- 63. Cabezas, R.; Vega-Vela, N.E.; González-Sanmiguel, J.; González, J.; Esquinas, P.; Echeverria, V.; Barreto, G.E. PDGF-BB Preserves Mitochondrial Morphology, Attenuates ROS Production, and Upregulates Neuroglobin in an Astrocytic Model Under Rotenone Insult. *Mol. Neurobiol.* 2018, 55, 3085–3095. [CrossRef] [PubMed]
- 64. Baez-Jurado, E.; Vega, G.G.; Aliev, G.; Tarasov, V.V.; Esquinas, P.; Echeverria, V.; Barreto, G.E. Blockade of Neuroglobin Reduces Protection of Conditioned Medium from Human Mesenchymal Stem Cells in Human Astrocyte Model (T98G) Under a Scratch Assay. *Mol. Neurobiol.* 2018, 55, 2285–2300. [CrossRef] [PubMed]
- 65. Baez-Jurado, E.; Guio-Vega, G.; Hidalgo-Lanussa, O.; González, J.; Echeverria, V.; Ashraf, G.M.; Sahebkar, A.; Barreto, G.E. Mitochondrial Neuroglobin Is Necessary for Protection Induced by Conditioned Medium from Human Adipose-Derived Mesenchymal Stem Cells in Astrocytic Cells Subjected to Scratch and Metabolic Injury. *Mol. Neurobiol.* **2019**, *56*, 5167–5187. [CrossRef] [PubMed]
- 66. Venturini, A.; Passalacqua, M.; Pelassa, S.; Pastorino, F.; Tedesco, M.; Cortese, K.; Gagliani, M.C.; Leo, G.; Maura, G.; Guidolin, D.; et al. Exosomes From Astrocyte Processes: Signaling to Neurons. *Front. Pharmacol.* **2019**, *10*, 1452. [CrossRef] [PubMed]
- 67. Chen, F.; Lu, J.; Chen, F.; Lin, Z.; Lin, Y.; Yu, L.; Su, X.; Yao, P.; Cai, B.; Kang, D. Recombinant neuroglobin ameliorates early brain injury after subarachnoid hemorrhage via inhibiting the activation of mitochondria apoptotic pathway. *Neurochem. Int.* **2018**, 112, 219–226. [CrossRef] [PubMed]
- 68. Zhang, C.; Hao, X.; Chang, J.; Geng, Z.; Wang, Z. Mn-TAT PTD-Ngb attenuates oxidative injury by an enhanced ROS scavenging ability and the regulation of redox signaling pathway. *Sci. Rep.* **2019**, *9*, 20103. [CrossRef] [PubMed]
- 69. Mendoza, V.; Klein, D.; Ichii, H.; Ribeiro, M.M.; Ricordi, C.; Hankeln, T.; Burmester, T.; Pastori, R.L. Protection of islets in culture by delivery of oxygen binding neuroglobin via protein transduction. *Transplant. Proc.* 2005, 37, 237–240. [CrossRef] [PubMed]

Cells **2021**, 10, 1907 11 of 12

70. Peroni, D.; Negro, A.; Bähr, M.; Dietz, G.P. Intracellular delivery of Neuroglobin using HIV-1 TAT protein transduction domain fails to protect against oxygen and glucose deprivation. *Neurosci. Lett.* **2007**, *421*, 110–114. [CrossRef]

- 71. Lin, Y.; Cai, B.; Xue, X.H.; Fang, L.; Wu, Z.Y.; Wang, N. TAT-mediated delivery of neuroglobin attenuates apoptosis induced by oxygen-glucose deprivation via the Jak2/Stat3 pathway in vitro. *Neurol. Res.* **2015**, *37*, 531–538. [CrossRef]
- 72. Blanco, S.; Peralta, S.; Morales, M.E.; Martínez-Lara, E.; Pedrajas, J.R.; Castán, H.; Peinado, M.Á.; Ruiz, M.A. Hyaluronate Nanoparticles as a Delivery System to Carry Neuroglobin to the Brain after Stroke. *Pharmaceutics* **2020**, 12, 40. [CrossRef]
- 73. De Marinis, E.; Ascenzi, P.; Pellegrini, M.; Galluzzo, P.; Bulzomi, P.; Arevalo, M.A.; Garcia-Segura, L.M.; Marino, M. 17β-Estradiol—A new modulator of neuroglobin levels in neurons: Role in neuroprotection against H₂O₂-induced toxicity. *Neurosignals* **2010**, *18*, 223–235. [CrossRef]
- 74. Fiocchetti, M.; De Marinis, E.; Ascenzi, P.; Marino, M. Neuroglobin and neuronal cell survival. *Biochim. Biophys. Acta* 2013, 1834, 1744–1749. [CrossRef]
- 75. Cutrupi, S.; Ferrero, G.; Reineri, S.; Cordero, F.; De Bortoli, M. Genomic lensa on neuroglobin transcription. *IUBMB Life* **2014**, *66*, 46–51. [CrossRef]
- 76. Guglielmotto, M.; Reineri, S.; Iannello, A.; Ferrero, G.; Vanzan, L.; Miano, V.; Ricci, L.; Tamagno, E.; De Bortoli, M.; Cutrupi, S. E2 Regulates Epigenetic Signature on Neuroglobin Enhancer-Promoter in Neuronal Cells. *Front. Cell. Neurosci.* **2016**, *10*, 147. [CrossRef] [PubMed]
- 77. Karachitos, A.; Grobys, D.; Kulczyńska, K.; Sobusiak, A.; Kmita, H. The Association of VDAC with Cell Viability of PC12 Model of Huntington's Disease. *Front. Oncol.* **2016**, *6*, 238. [CrossRef]
- 78. Arevalo, M.A.; Santos-Galindo, M.; Bellini, M.J.; Azcoitia, I.; Garcia-Segura, L.M. Actions of estrogens on glial cells: Implications for neuroprotection. *Biochim. Biophys. Acta* **2010**, *1800*, 1106–1112. [CrossRef]
- 79. Acaz-Fonseca, E.; Sanchez-Gonzalez, R.; Azcoitia, I.; Arevalo, M.A.; Garcia-Segura, L.M. Role of astrocytes in the neuroprotective actions of 17beta-estradiol and selective estrogen receptor modulators. *Mol. Cell. Endocrinol.* **2014**, *389*, 48–57. [CrossRef] [PubMed]
- 80. Brocca, M.E.; Garcia-Segura, L.M. Non-reproductive Functions of Aromatase in the Central Nervous System Under Physiological and Pathological Conditions. *Cell. Mol. Neurobiol.* **2019**, *39*, 473–481. [CrossRef] [PubMed]
- 81. Acaz-Fonseca, E.; Castelló-Ruiz, M.; Burguete, M.C.; Aliena-Valero, A.; Salom, J.B.; Torregrosa, G.; García-Segura, L.M. Insight into the molecular sex dimorphism of ischaemic stroke in rat cerebral cortex: Focus on neuroglobin, sex steroids and autophagy. *Eur. J. Neurosci.* 2020, 52, 2756–2770. [CrossRef]
- 82. Jiang, M.; Ma, C.; Li, H.; Shen, H.; Li, X.; Sun, Q.; Chen, G. Sex Dimorphisms in Ischemic Stroke: From Experimental Studies to Clinic. *Front. Neurol.* **2020**, *11*, 504. [CrossRef] [PubMed]
- 83. Di Pietro, V.; Lazzarino, G.; Amorini, A.M.; Tavazzi, B.; D'Urso, S.; Longo, S.; Vagnozzi, R.; Signoretti, S.; Clementi, E.; Giardina, B.; et al. Neuroglobin expression and oxidant/antioxidant balance after graded traumatic brain injury in the rat. *Free Radic. Biol. Med.* 2014, 69, 258–264. [CrossRef] [PubMed]
- 84. Fernando, M.S.; Simpson, J.E.; Matthews, F.; Brayne, C.; Lewis, C.E.; Barber, R.; Kalaria, R.N.; Forster, G.; Esteves, F.; Wharton, S.B.; et al. MRC Cognitive Function and Ageing Neuropathology Study Group. White matter lesions in an unselected cohort of the elderly: Molecular pathology suggests origin from chronic hypoperfusion injury. *Stroke* **2006**, *37*, 1391–1398. [CrossRef] [PubMed]
- 85. Hidalgo-Lanussa, O.; Ávila-Rodriguez, M.; Baez-Jurado, E.; Zamudio, J.; Echeverria, V.; Garcia-Segura, L.M.; Barreto, G.E. Tibolone Reduces Oxidative Damage and Inflammation in Microglia Stimulated with Palmitic Acid through Mechanisms Involving Estrogen Receptor Beta. *Mol. Neurobiol.* **2018**, *55*, 5462–5477. [CrossRef] [PubMed]
- 86. Azcoitia, I.; Moreno, A.; Carrero, P.; Palacios, S.; Garcia-Segura, L.M. Neuroprotective effects of soy phytoestrogens in the rat brain. *Gynecol. Endocrinol.* **2006**, 22, 63–69. [CrossRef] [PubMed]
- 87. Duan, X.; Li, Y.; Xu, F.; Ding, H. Study on the neuroprotective effects of Genistein on Alzheimer's disease. *Brain Behav.* **2021**, 11, e02100. [CrossRef] [PubMed]
- 88. Rumman, M.; Pandey, S.; Singh, B.; Gupta, M.; Ubaid, S.; Mahdi, A.A. Genistein Prevents Hypoxia-Induced Cognitive Dysfunctions by Ameliorating Oxidative Stress and Inflammation in the Hippocampus. *Neurotox. Res.* **2021**. [CrossRef]
- 89. Liu, N.; Yu, Z.; Gao, X.; Song, Y.S.; Yuan, J.; Xun, Y.; Wang, T.; Yan, F.; Yuan, S.; Zhang, J.; et al. Establishment of Cell-Based Neuroglobin Promoter Reporter Assay for Neuroprotective Compounds Screening. *CNS Neurol. Disord. Drug Targets* **2016**, *15*, 629–639. [CrossRef]
- 90. Carswell, H.; Macrae, I.M.; Gallagher, L.; Harrop, E.; Horsburgh, K.J. Neuroprotection by a selective estrogen receptor beta agonist in a mouse model of global ischemia. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, 287, H1501–H1504. [CrossRef]
- 91. Wisdom, A.J.; Cao, Y.; Itoh, N.; Spence, R.D.; Voskuhl, R.R. Estrogen receptor-beta ligand treatment after disease onset is neuroprotective in the multiple sclerosis model. *J. Neurosci. Res.* **2013**, *91*, 901–908. [CrossRef] [PubMed]
- 92. Pietranera, L.; Correa, J.; Brocca, M.E.; Roig, P.; Lima, A.; Di Giorgio, N.; Garcia-Segura, L.M.; De Nicola, A.F. Selective Oestrogen Receptor Agonists Rescued Hippocampus Parameters in Male Spontaneously Hypertensive Rats. *J. Neuroendocrinol.* **2016**, 28, 10. [CrossRef] [PubMed]
- 93. Kloosterboer, H.J. Tissue-selective effects of tibolone on the breast. Maturitas 2004, 49, S5–S15. [CrossRef] [PubMed]
- 94. Kloosterboer, H.J. Tissue-selectivity: The mechanism of action of tibolone. *Maturitas* **2004**, *48* (Suppl. S1), S30–S40. [CrossRef] [PubMed]

Cells 2021, 10, 1907 12 of 12

95. Reed, M.J.; Kloosterboer, H.J. Tibolone: A selective tissue estrogenic activity regulator (STEAR). *Maturitas* **2004**, *48* (Suppl. S1), S4–S6. [CrossRef]

- 96. Steckelbroeck, S.; Jin, Y.; Oyesanmi, B.; Kloosterboer, H.J.; Penning, T.M. Tibolone is metabolized by the 3alpha/3beta-hydroxysteroid dehydrogenase activities of the four human isozymes of the aldo-keto reductase 1C subfamily: Inversion of stereospecificity with a delta5(10)-3-ketosteroid. *Mol. Pharmacol.* 2004, 66, 1702–1711. [CrossRef]
- 97. Escande, A.; Servant, N.; Rabenoelina, F.; Auzou, G.; Kloosterboer, H.; Cavaillès, V.; Balaguer, P.; Maudelonde, T. Regulation of activities of steroid hormone receptors by tibolone and its primary metabolites. *J. Steroid Biochem. Mol. Biol.* **2009**, *116*, 8–14. [CrossRef]
- 98. Pinto-Almazán, R.; Segura-Uribe, J.J.; Farfán-García, E.D.; Guerra-Araiza, C. Effects of Tibolone on the Central Nervous System: Clinical and Experimental Approaches. *Biomed. Res. Int.* **2017**, 2017, 8630764. [CrossRef]
- 99. Del Río, J.P.; Molina, S.; Hidalgo-Lanussa, O.; Garcia-Segura, L.M.; Barreto, G.E. Tibolone as Hormonal Therapy and Neuroprotective Agent. *Trends Endocrinol. Metab.* **2020**, *31*, 742–759. [CrossRef]
- 100. Crespo-Castrillo, A.; Yanguas-Casás, N.; Arevalo, M.A.; Azcoitia, I.; Barreto, G.E.; Garcia-Segura, L.M. The Synthetic Steroid Tibolone Decreases Reactive Gliosis and Neuronal Death in the Cerebral Cortex of Female Mice After a Stab Wound Injury. *Mol. Neurobiol.* **2018**, *55*, 8651–8667. [CrossRef] [PubMed]
- 101. Crespo-Castrillo, A.; Garcia-Segura, L.M.; Arevalo, M.A. The synthetic steroid tibolone exerts sex-specific regulation of astrocyte phagocytosis under basal conditions and after an inflammatory challenge. *J. Neuroinflamm.* 2020, 17, 37. [CrossRef] [PubMed]
- 102. Kashyap, K.; Siddiqi, M.I. Recent trends in artificial intelligence-driven identification and development of anti-neurodegenerative therapeutic agents. *Mol. Divers.* **2021**. [CrossRef] [PubMed]