Effects of particulate matter exposure on blood 5-hydroxymethylation: results from the Beijing truck driver air pollution study

Marco Sanchez-Guerra^{1,*}, Yinan Zheng², Citlalli Osorio-Yanez³, Jia Zhong¹, Yana Chervona⁴, Sheng Wang⁵, Dou Chang⁶, John P McCracken⁷, Anaite Díaz⁷, Pier Alberto Bertazzi⁸, Petros Koutrakis¹, Choong-Min Kang¹, Xiao Zhang⁹, Wei Zhang⁹, Hyang-Min Byun¹, Joel Schwartz¹, Lifang Hou^{9,#,*}, and Andrea A Baccarelli^{1,#}

¹Department of Environmental Health; Harvard T.H. Chan School of Public Health; Boston, MA USA; ²Institute for Public Health and Medicine; Northwestern University Feinberg School of Medicine; Chicago, IL USA; ³Department of Epidemiology; Harvard T.H Chan School of Public Health; Boston, MA USA; ⁴Department of Environmental Medicine; New York University School of Medicine; Tuxedo, NY USA; ⁵Department of Occupational and Environmental Health; Peking University Health Science Center; Beijing, China; ⁶Department of Safety Engineering; China Institute of Industrial Relations; Beijing, China; ⁷Center for Health Studies; Universidad del Valle de Guatemala; Guatemala; Guatemala; ⁸Department of Clinical Sciences and Community Health—DISCO, Università degli Studi di Milano; Milan, Lombardy, Italy; ⁹Department of Preventive Medicine; Northwestern University Feinberg School of Medicine; Chicago, IL USA

*These author contributed equally to this work.

Keywords: DNA methylation, Epigenetics, Particulate Matter, 5-hydroxymethylcytosine, 5-methylcytosine

Abbreviations: 5hmC, 5-hydroxymethylcytosine; 5mC, 5-methylcytosine; Al, aluminum; BC, black carbon; BMI, body mass index; Ca, calcium; CI, confidence interval; ELISA, enzyme-linked immunosorbent assay; Fe, iron; FWER, family-wise error rate; HPLC, high-performance liquid chromatography; K, potassium; PM, particulate matter; PM_{2.5}, particulate matter $\leq 2.5 \ \mu$ m; PM₁₀, particulate matter $\leq 10 \ \mu$ m; S, sulfur; Si: silicon; TET, ten-eleven translocation enzymes; Ti, titanium and Zn: zinc.

Previous studies have reported epigenetic changes induced by environmental exposures. However, previous investigations did not distinguish 5-methylcytosine (5mC) from a similar oxidative form with opposite functions, 5hydroxymethylcytosine (5hmC). Here, we measured blood DNA global 5mC and 5hmC by ELISA and used adjusted mixed-effects regression models to evaluate the effects of ambient PM_{10} and personal PM_{25} and its elemental components—black carbon (BC), aluminum (AI), calcium (Ca), potassium (K), iron (Fe), sulfur (S), silicon (Si), titanium (Ti), and zinc (Zn)—on blood global 5mC and 5hmC levels. The study was conducted in 60 truck drivers and 60 office workers in Beijing, China from The Beijing Truck Driver Air Pollution Study at 2 exams separated by one to 2 weeks. Blood 5hmC level (0.08%) was ~83-fold lower than 5mC (6.61%). An inter-guartile range (IQR) increase in same-day PM_{10} was associated with increases in 5hmC of 26.1% in office workers (P = 0.004), 20.2% in truck drivers (P = 0.014), and 21.9% in all participants combined (P < 0.001). PM₁₀ effects on 5hmC were increasingly stronger when averaged over 4, 7, and 14 d preceding assessment (up to 132.6% for the 14-d average in all participants, P < 0.001). PM₁₀ effects were also significant after controlling for multiple testing (family-wise error rate; FWER < 0.05). 5hmC was not correlated with personal measures of $PM_{2.5}$ and elemental components (FWER > 0.05). 5mC showed no correlations with PM_{10} , $PM_{2.5}$, and elemental components measures (FWER > 0.05). Our study suggests that exposure to ambient PM₁₀ affects 5hmC over time, but not 5mC. This finding demonstrates the need to differentiate 5hmC and 5mC in environmental studies of DNA methylation.

Introduction

Urban air pollution is a major public health concern that, according to the World Health Organization, is responsible for \sim 3.7 million deaths annually, representing 6.7% of all deaths worldwide.¹ Governments and regulatory agencies have made

concerted efforts to abate air pollution emissions, specifically by regulating particulate matter (PM) levels.²⁻⁴ However, further abatements are constrained by high marginal costs.^{5,6} Moreover, limited resources are available to identify individuals who are affected by air pollution and ascertain their risks. Biomarkers that reflect adverse exposures could be vital for identifying people

© Marco Sanchez-Guerra, Yinan Zheng, Citlalli Osorio-Yanez, Jia Zhong, Yana Chervona, Sheng Wang, Dou Chang, John P McCracken, Anaite Díaz, Pier Alberto Bertazzi, Petros Koutrakis, Choong-Min Kang, Xiao Zhang, Wei Zhang, Hyang-Min Byun, Joel Schwartz, Lifang Hou, and Andrea A Baccarelli

*Correspondence to: Marco Sanchez-Guerra: E-mail: msanchez@hsph.harvard.edu; Lifang Hou: E-mail: I-hou@northwestern.edu

Submitted: 02/25/2015; Revised: 04/24/2015; Accepted: 05/06/2015

http://dx.doi.org/10.1080/15592294.2015.1050174

This is an Open Access article distributed under the terms of the Creative Commons Attribution-Non-Commercial License (http://creativecommons.org/licenses/ by-nc/3.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The moral rights of the named author(s) have been asserted. who are affected by PM exposure before it becomes clinically evident.

Epigenetic features like DNA methylation are increasingly being implicated as sensitive to environmental exposures, and therefore could serve as important biomarkers for exposure effects.⁷⁻¹² DNA methylation modifications, both in specific genes and in non-coding repetitive elements, have been shown to be altered following PM exposure.¹³⁻¹⁸ Traditional DNA methylation involves the transfer of a methyl group to the carbon 5 position of cytosine to produce 5-methylcytosine (5mC). However, research has shown that the Tet family of cytosine oxygenase enzymes are involved in oxidizing 5methylcytosine into 5-hydroxymethylcytosine (5hmC). In contrast to 5mC, which is often associated with suppressed gene expression, 5hmC has been found to be specifically enriched in expressed genes, and may play a role in activating and/or maintaining gene expression.¹⁹⁻²¹ Therefore, distinguishing these 2 forms of methylation is critical for understanding the potential effects of methylation changes.

Because the standard technologies for DNA methylation analysis, including those based on bisulfite treatment of DNA, fail to separate 5hmC from 5mC²²⁻²⁴ (providing instead a composite measure of the sum of 5mC and 5hmC), most epigenetic studies have not differentiated 5hmC from 5mC. This differentiation, however, is particularly critical for air pollution studies, because PM exposures can introduce oxidative stress. Oxidative stress can alter the activity of the Ten-11 translocation (TET) protein family members, which catalyze enzymatic reactions that generate 5hmC from 5mC.²⁵ Further, Chia et al. suggested that pro-oxidant environmental exposures can increase the global genomic content of 5hmC.²⁵ However—while previous studies have reported that PM exposure is negatively associated with 5mC or total methylation levels^{13,26-28}—no study to date has evaluated whether PM exposure changes 5hmC levels in human DNA.

In the present study, we determined the genomic content of both 5hmC and 5mC in a study of 2 highly exposed groups with different occupations, i.e., office workers and truck drivers, in Beijing, China. Beijing has one of the highest air pollution levels worldwide.²⁹ We used measures of ambient PM less than 10 μ m in diameter (PM₁₀) mass from ambient monitors across Beijing City, as well as personal measures of both PM_{2.5} and elemental components using personal samplers. We hypothesized that PM exposure—by facilitating the oxidation of 5mC into 5hmC would result in increased blood levels of 5hmC. The findings underscore the necessity for distinguishing these 2 types of DNA methylation to achieve a better understanding of the effects of environmental exposures and the roles of epigenetic mechanisms in non-communicable diseases development due to pro-oxidant exposures.

Results

Characteristics of the study participants

The participants in the Beijing Truck Driver Air Pollution Study have been described elsewhere.^{13,30-32} Truck drivers were moderately but significantly older than office workers. Truck drivers had higher BMI, reported more pack-years of smoking, and smoked more cigarettes during the examination time.

Ambient PM_{10} mass and personal $\mathrm{PM}_{2.5}$ mass and elemental components

Office workers and truck drivers were examined on days with average ambient PM_{10} mass of 118.1 \pm 52.8 and 124.9 \pm 51.2 µg/m³ for the 2 groups, respectively (**Table 1**, *P*=0.33 for

Table 1. Levels of ambient PM₁₀ on and before the examination days, and personal PM_{2.5} and elemental components of PM during work hours

		Office workers [*]							Truck drivers [*]						
Time window	Mean \pm SD	10pct	25pct	50pct	75pct	90pct	Mean \pm SD	10pct	25pct	50pct	75pct	90pct	p-value †		
PM ₁₀ (μ	ug/m³) from ambi	ent monit	ors on th	e examina	ation days	s (24-h av	erage) and multi	-day avera	ages on a	nd before	the exam	nination d	ays		
24-h mean	118.1 ± 52.8	60.0	82.0	114.0	150.0	186.0	124.9 \pm 51.2	72.0	88.0	118.0	168.0	190.0	0.33		
4-d mean	119.0 ± 32.0	78.8	89.8	114.5	136.5	165.0	$120.5\ \pm 29.8$	83.8	101.3	117.0	136.5	164.0	0.59		
7-d mean	117.9 ± 24.1	83.3	98.3	115.1	137.4	147.7	119.0 ± 22.4	94.4	99.9	116.6	137.4	147.7	0.76		
14-d mean	119.0 ± 19.3	93.4	99.7	124.7	139.0	141.4	$121.8\ \pm\ 18.4$	97.1	104.2	127.0	140.1	143.1	0.26		
	Particle and elem	ental con	nponent n	neasures	(μg/m ³) c	luring the	8 work h on the	examina	tion days,	from per	sonal mor	nitors			
PM _{2.5}	94.3 ± 68.2	22.4	43.3	84.5	131.6	185.9	128.6 \pm 70.9	45.4	75.3	118.9	162.9	215.8	< 0.001		
BC	$13.0\ \pm 4.2$	7.1	10.0	13.1	15.9	18.4	$17.1\ \pm 6.9$	8.9	12.2	16.6	20.7	26.2	< 0.001		
AI	0.53 ± 0.25	0.23	0.37	0.49	0.70	0.85	1.35 ± 0.94	0.42	0.59	1.29	1.85	2.28	< 0.001		
Ca	$0.32\ \pm 0.17$	0.16	0.19	0.28	0.41	0.52	$2.07\ \pm 2.23$	0.29	0.39	1.59	3.02	4.50	< 0.001		
Fe	0.38 ± 0.19	0.18	0.24	0.34	0.44	0.67	$1.00\ \pm 0.63$	0.37	0.49	0.82	1.33	1.75	< 0.001		
К	0.72 ± 0.74	0.18	0.26	0.52	0.74	1.68	$1.33 \ \pm 1.05$	0.34	0.46	0.93	2.10	2.77	< 0.001		
S	6.28 ± 5.25	0.75	1.63	5.31	8.64	14.37	8.67 ± 4.97	2.82	5.02	7.26	12.70	16.29	< 0.001		
Si	$0.79 \ \pm 0.55$	0.28	0.43	0.68	1.01	1.42	2.37 ± 1.78	0.63	0.82	2.10	3.54	4.12	< 0.001		
Ti	$0.02\ \pm 0.01$	0.01	0.01	0.02	0.03	0.04	$0.06\ \pm 0.04$	0.02	0.03	0.05	0.08	0.10	< 0.001		
Zn	$0.15\ \pm 0.17$	0.02	0.05	0.09	0.23	0.36	$0.27\ \pm 0.22$	0.07	0.11	0.17	0.40	0.68	< 0.001		

*Number of observations used for analysis was 103 for office workers and 109 for truck drivers (missing values were from missing %5hmC or %5mC measures, since we did not have samples from some individuals).

[†]*P*-values were obtained from Wilcoxon-Mann-Whitney tests.



Figure 1. Heat map for pair-wise correlations of the levels of ambient PM_{10} (24-h, 4-d, 7-d, and 14-d averages) and personal levels of $PM_{2.5}$, BC, and elemental components of $PM_{2.5}$ measured during 8 work h. The four ambient PM_{10} measures were correlated with each other. Measures from personal monitors (i.e., $PM_{2.5}$, BC, and elemental components of $PM_{2.5}$) tend to be correlated with each other. Abbreviations: PM_{10} , Particulate matter $\leq 10 \ \mu$ m; $PM_{2.5}$, Particulate matter $\leq 2.5 \ \mu$ m; BC, Black Carbon.

difference in PM_{10} mass between the 2 groups). Table 1 also reports the multi-day averages of ambient PM_{10} mass on and before the examination days (including means of 4, 7, and 14 d).

Personal levels of $PM_{2.5}$ mass, and elemental components (BC, Al, Ca, Fe, K, S, Si, Ti, and Zn) measured during 8 work h on examination days were significantly higher in truck drivers than in office workers (P < 0.001 for all exposures). However, all personal exposure metrics showed largely overlapping distributions between the 2 groups.

Ambient PM_{10} mass and personal exposure measures ($PM_{2.5}$ mass, BC, and elemental components) showed variable degrees of correlation (Fig. 1). In particular, the 4 time-window averages of ambient PM_{10} mass showed strong-to-moderate correlations with each other. As expected, correlations were higher for the closer averages (i.e., 24-h and 4-d average, or 4-d and 7-d average). Personal exposures, which all were measured on particles sampled during 8 work h on the days of examination, mostly showed medium to high correlations, but had moderate or little correlation with ambient PM_{10} mass.

Descriptive statistics of 5hmC and 5mC levels

The characteristics of the 60 office workers and 60 truck drivers by blood levels of 5-hydroxymethylcytosine (5hmC) and 5-methylcytosine (5mC) are presented in Table 2.

The geometric mean of 5hmC (0.08%; 95% CI: 0.08 - 0.09) was ~83-fold lower than the geometric mean of 5mC (6.61%, 95% CI: 6.24 - 7.01). The 5hmC values ranged between 0.02% (min) and 0.29% (max); 5mC ranged between 3.29% (min) and 10.19% (max). The levels of 5hmC and 5mC in blood DNA were significantly correlated with each other (Pearson's

correlation=0.35, P=0.001 on the first examination day; Pearson's correlation=0.22, P=0.021 on the 2nd examination day) (Fig. 2).

The geometric means of 5hmC levels (Table 2) did not differ between office workers (0.09%, 95% CI: 0.08 – 0.10) and truck drivers (0.08%, 95% CI: 0.07 – 0.09, P=0.28). Males had significantly higher 5hmC than females (0.10%, 95% CI: 0.09 – 0.11, and 0.06%, 95% CI: 0.06; 0.07 respectively, P < 0.001). Further, 5hmC decreased moderately and non-significantly with participant age (P=0.094). Former smokers and current smokers had marginally higher 5hmC than never smokers (P=0.082). 5hmC tended to fluctuate across the days of the week (P=0.035) and increased with higher dew point (P=0.048), whereas BMI (P=0.58) and temperature (P=0.40) were not related with 5hmC.

The geometric mean of 5mC levels (**Table 2**) also did not differ between office workers (6.47%, 95% CI: 6.19 – 6.77) and truck drivers (6.75%, 95% CI: 6.46 – 7.06, P=0.19). As for 5hmC, 5mC levels were higher in males (6.93%, 95% CI: 6.67 – 7.21) than females (6.10%, 95% CI: 5.80 – 6.41, P < 0.001). 5mC also decreased slightly with age (P=0.069), but was not associated with BMI, smoking, day of week, temperature, or dew point (P > 0.1). Based on our results, 5hmC measurements exhibited less day-to-day variability (about 28 times lower median intra-individual variability), compared to 5mC (Supplemental Figure S1). These findings suggested that global 5hmC in our study population was likely to be a relatively stable DNA modification compared to 5mC, which is in line with previous studies.³³

Effects of ambient and personal exposures on blood genomic levels of 5hmC and 5mC

We used mixed-effects regression models adjusted for age, gender, BMI, smoking status, day of week, temperature, and dew point to evaluate the associations of 5hmC with ambient and personal exposure levels (Table 3). An IQR increase in ambient PM₁₀ levels on the examination days was associated with an increase in blood 5hmC equal to 26.1% in office workers (95% CI: 7.9 - 47.4, P=0.004), 20.2% in truck drivers (95% CI: 4.0 - 39.0, P=0.014), and 21.9% in all participants combined (95% CI: 10.3 - 34.7, P < 0.001). The correlations between 5hmC and ambient PM₁₀ mass increased in strength with increasing number of days included in multiday averages. Among office workers, an IQR increase ambient PM₁₀ mass was associated with increased 5hmC of 27.7% (95% CI: 9.0 - 49.5, P=0.003) with the 4-d average, 43.2% (95% CI: 15.6 - 77.4, P=0.001) with the 7-d average, and 152.1% (95% CI: 59.8 - 297.8, P < 0.001) with the 14-d average. In truck drivers, an IQR increase of ambient PM₁₀ mass was associated with increased 5hmC of 34.1% (95% CI: 13.3 - 58.6, P=0.001) with the 4-d average, 39.9% (95% CI: 11.6 - 75.5, P=0.005) with the 7-d average, and 122.6% (95% CI: 49.9 - 230.6, P < 0.001) with the 14-d average. In all participants combined, an IQR increase in ambient PM₁₀ mass was associated with increased 5hmC of 28.1% (95% CI: 15.3 - 42.2, P < 0.001) with the 4-d average, 37.3% (95%)

Table 2. Participant characteristics and blood levels	of 5-hydroxymethylcytosine	(5hmC) and 5-methylcytosine (5mC)
--	----------------------------	-----------------------------------

		5h	1mC (%C [*])		5mC (%C [*])					
Variables	Obs†	Mean [‡]	(95% CI) [‡]	<i>P</i> -value [§]	Obs†	Mean [‡]	(95% CI) [‡]	<i>P</i> -value [§]		
Group										
Office Worker	103	0.09	(0.08; 0.10)	0.28	102	6.47	(6.19; 6.77)	0.19		
Truck Driver	109	0.08	(0.07; 0.09)		106	6.75	(6.46; 7.06)			
Sex										
Male	136	0.10	(0.09; 0.11)	< 0.001	131	6.93	(6.67; 7.21)	< 0.001		
Female	76	0.06	(0.06; 0.07)		77	6.10	(5.80; 6.41)			
Age (Quartiles)										
Q1 [18-27 years]	49	0.10	(0.08; 0.11)	0.094	46	6.96	(6.51; 7.43)	0.069		
Q2 [28-32 years]	54	0.09	(0.08; 0.10)		56	6.85	(6.45; 7.27)			
Q3 [33–37 years]	54	0.08	(0.07; 0.09)		52	6.43	(6.04; 6.84)			
Q4 [39–46 years]	55	0.07	(0.06; 0.09)		54	6.27	(5.90; 6.67)			
BMI										
14.8-24.9 kg/m ²	148	0.08	(0.08; 0.09)	0.58	146	6.51	(6.27; 6.76)	0.27		
25-29.9 kg/m ²	55	0.08	(0.07; 0.10)		54	6.83	(6.42; 7.26)			
30-32.4 kg/m ²	9	0.10	(0.07; 0.15)		8	7.15	(6.09; 8.40)			
Smoking status										
Never-smoker	125	0.08	(0.07; 0.09)	0.082	124	6.44	(6.18; 6.71)	0.14		
Former smoker	8	0.09	(0.06; 0.13)		8	6.77	(5.77; 7.94)			
Current smoker	79	0.09	(0.08; 0.10)		76	6.88	(6.53; 7.25)			
Day of week										
Monday	28	0.08	(0.05; 0.08)	0.035	29	6.79	(5.73; 6.79)	0.40		
Tuesday	26	0.09	(0.06; 0.09)		25	7.57	(6.31; 7.57)			
Wednesday	28	0.11	(0.08; 0.11)		28	7.49	(6.30; 7.49)			
Thursday	34	0.11	(0.07; 0.11)		35	7.07	(6.06; 7.07)			
Friday	35	0.11	(0.08; 0.11)		34	7.03	(6.01; 7.03)			
Saturday	30	0.11	(0.07; 0.11)		29	7.61	(6.42; 7.61)			
Sunday	31	0.10	(0.07; 0.10)		28	6.91	(5.81; 6.91)			
Temperature (Median)**										
Low [20–25 °C]	91	0.08	(0.07; 0.09)	0.40	89	6.74	(6.42; 7.07)	0.30		
High [26–29 °C]	121	0.09	(0.08; 0.09)		119	6.52	(6.25; 6.79)			
Dew point (Median)**			*				/			
Low [16–20 °C]	86	0.08	(0.07; 0.09)	0.048	84	6.76	(6.43; 7.10)	0.27		
High [21–24 °C]	126	0.09	(0.08; 0.10)		124	6.51	(6.25; 6.79)			

^{*}Percent of hydroxymethylated (for 5hmC) or methylated (for 5mC) cytosines in blood genomic DNA.

[†]Number of non-missing 5hmC or 5mC measures.

[‡]Geometric means and 95% confident intervals (95% Cl) of 5hmC or 5mC.

 $^{\$}$ P-values were calculated using mixed-effects regression models with log transformed measures of 5hmC or 5mC.

**Temperature and dew point were measured on the study examination day.

CI: 18.8 - 58.6, P < 0.001) with the 7-d average, and 132.6% (95% CI: 75.6 – 208.0, P < 0.001) with the 14-d average. To account for the multiple dependent statistical tests conducted in our analysis, we calculated permutation-based Pvalues adjusted for multiple comparisons using FWER. At FWER < 0.05, all the different time windows of ambient PM10 mass remained highly significant for office workers, truck drivers, and all participants combined. The increasingly larger cumulative effects of PM₁₀ on 5hmC with more days before the examination days suggest that the effect of PM_{10} on 5hmC tend to build up in time. Although an IQR increase BC was significantly associated with increased 5hmC of 27.0% (95% CI: 6.5 - 51.4, P=0.009) in office workers and 14.6% (95% CI: 3.5 - 26.9, P=0.010) in truck drivers, at the same FWER threshold, none of the personal measures of exposures (PM2.5 mass, BC, and elemental components during 8 work h on examination days) was associated with 5hmC.

We conducted the same set of statistical analyses in relation to 5mC (Table 4). 5mC did not show significant associations with ambient PM_{10} mass, personal $PM_{2.5}$ mass, BC, or elemental components at FWER < 0.05.

Discussion

In this study of truck drivers and office workers in Beijing, China, we found that exposure to ambient PM_{10} mass was associated with increased levels of 5hmC, while no association was observed with 5mC. The effects of ambient PM_{10} mass levels on 5hmC were progressively higher in relation to longer time windows of exposure, possibly due to accumulation of effects over time. Consistent with this time-dependent dynamics, short-term (8-h) personal exposures to $PM_{2.5}$, BC, Al, C, K, Fe, S, Si, Ti,



Figure 2. Scatter plot of blood levels of 5-hydroxymethylcytosine (5hmC) and 5-methylcytosine (5mC). Pearson's correlation coefficient=0.35 (*P*=0.001) on the first examination day. Pearson's correlation coefficient=0.22 (*P*=0.021) on the 2nd examination day.

and Zn were not associated with either blood global 5hmC or 5mC.

As expected based on previous analysis of human blood,³⁴ we found that the levels of 5hmC were much lower than those of 5mC. We found higher global 5mC in males than females, consistent with previous studies conducted in Chinese populations

and in a Belgian population; 26,35,36 findings were similar for global 5hmC. However, the ratio of 5mC/5hmC was higher in our data compared to previous studies. In our study, 5hmC was \sim 83-fold lower than 5mC. In 48 participants in the US Strong Heart Study, 5hmC levels were just ~2.5-fold lower than 5mC, with median values of 0.12% for 5hmC and 0.32% for 5mC in DNA isolated from frozen buffy coat, and 0.15 for 5hmC and 0.32 for 5mC in frozen whole blood.³⁴ In a study of 48 healthy men from Spain, 5hmC levels were 10-fold lower than 5mC in DNA from whole blood (0.09% for 5hmC vs. 0.90% for 5mC). We note that 5hmC levels in our Beijing population (0.08%) were remarkably similar to those reported in these previous studies in Spain (0.09%) and the United States (0.12%). Although, the 5mC (6.61%) levels differed between 7-fold and 21-fold across the 3 studies; the levels found in this study are more similar to those reported by De Prins et al. (4.3%).²⁶

Previous investigations have suggested that DNA methylation is highly sensitive to environmental exposures,⁷⁻¹¹ including air pollution. To the best of our knowledge, the present study is the first human investigation of both 5hmC and 5mC in relation to PM exposure. Our results show that 5hmC may reflect PM-related epigenetic changes that were not captured by previous studies only examining 5mC. The positive correlation between ambient PM₁₀ and 5hmC is in line with 5hmC biology. PM exposure results in generation of reactive oxygen species (ROS),³⁷⁻³⁹ which are chemically reactive molecules that can induce oxidation of 5mC into 5hmC.²⁵ Our findings are also consistent with recent *in vitro* results by Coulter et al,⁴⁰ who showed that hydroquinone—a benzene metabolite that has been shown to heighten ROS levels—increased

Table 3. Percent changes on 5-hydroxymethylcytosine associated with an interquartile range (IQR) increase in exposure levels*

	Office Workers [†]					Truck [Drivers [†]		All participants †			
	%	95% CI	<i>P</i> -value [‡]	Adj.	%	95% Cl	<i>P</i> -value [‡]	Adj.	%	95% CI	<i>P</i> -value [‡]	Adj.
Study group				<i>P</i> -value [§]				<i>P</i> -value [§]				<i>P</i> -value [§]
PM ₁₀ (μg/m³) fi	rom ambient mo	nitors on th	e examina	tion days	(24-h average)	and multi-da	ay averages	on and	before the exa	mination da	ys
24-h average	26.1	(7.9; 47.4)	0.004	0.025	20.2	(4.0; 39.0)	0.014	0.042	21.9	(10.3; 34.7)	<0.001	< 0.001
4-d average	27.7	(9.0; 49.5)	0.003	0.019	34.1	(13.3; 58.6)	0.001	0.002	28.1	(15.3; 42.2)	< 0.001	< 0.001
7-d average	43.2	(15.6; 77.4)	0.001	0.011	39.9	(11.6; 75.5)	0.005	0.014	37.3	(18.8; 58.6)	< 0.001	< 0.001
14-d average	152.1	(59.8; 297.8)	< 0.001	0.001	122.6	(49.9; 230.6)	< 0.001	< 0.001	132.6	(75.6; 208.0)	< 0.001	< 0.001
		Particle me	easures (μg/	m ³) during	g the 8 w	ork h on the exa	amination da	ays, from pe	ersonal m	nonitors		
PM _{2.5}	10.0	(-3.7; 25.6)	0.16	0.79	10.0	(-6.8; 29.8)	0.26	0.92	8.9	(-0.7; 19.5)	0.072	0.44
BC	27.0	(6.5; 51.4)	0.009	0.18	3.1	(-9.5; 17.4)	0.65	0.99	7.9	(-2.1; 18.9)	0.13	0.60
Al	1.1	(-29.5; 44.9)	0.95	0.99	8.3	(-3.8; 22.0)	0.19	0.83	5.7	(-4.0; 16.4)	0.26	0.85
Ca	-1.9	(-57.1; 124.3)	0.96	0.99	3.2	(-3.7; 10.6)	0.38	0.98	3.2	(-3.0; 9.8)	0.32	0.91
Fe	3.0	(-24.9; 41.3)	0.85	0.99	4.7	(-6.3; 16.9)	0.42	0.99	2.9	(-5.9; 12.6)	0.53	0.99
К	10.0	(-3.9; 25.9)	0.17	0.80	3.3	(-8.1; 16.0)	0.59	0.99	4.3	(-3.6; 12.7)	0.30	0.89
S	14.3	(-1.1; 32.2)	0.074	0.56	9.5	(-7.8; 30.1)	0.31	0.95	9.1	(-1.2; 20.5)	0.087	0.49
Si	-18.4	(-38.9; 9.1)	0.17	0.99	9.5	(-2.3; 22.7)	0.12	0.69	4.2	(-4.9; 14.1)	0.38	0.96
Ti	-12.6	(-32.9; 13.7)	0.32	0.99	14.6	(3.5; 26.9)	0.010	0.16	8.2	(-0.7; 18.0)	0.075	0.45
Zn	-0.4	(14.4; 15.8)	0.96	0.99	0.3	(-12.3; 14.7)	0.96	0.99	-1.4	(-9.5; 7.5)	0.75	0.99

*Adjusted for age, sex, BMI, smoking status, day of week, and average of temperature and dew point values corresponding to the average of exposure metrics.

[†]Number of observations used for analysis was 103 for office workers, 109 for truck drivers, and 212 for all participants.

[‡]*P*-values were calculated using mixed-effects regression models.

[§]Adjusted *P*-values were estimated by a resampling-based approach, which takes dependent structures among exposures into consideration. This approach controls family-wise error rate (FWER), i.e., probability of having at least one false positive among the whole set of comparisons at a given significance level.

Table 4. Percent changes on 5-methylcytosine associated with an interquartile range (IQR) increase in air particle levels*

		Office V	Vorkers [†]			Truck [Drivers [†]		All participants †			
	%	95% Cl	<i>P</i> -value [‡]	Adj.	%	95% CI	<i>P</i> -value [‡]	Adj.	%	95% Cl	<i>P</i> -value [‡]	Adj.
Study group	-			<i>P</i> -value [§]				<i>P</i> -value [§]				<i>P</i> -value [§]
PM ₁₀	μg/m³)	from ambient m	onitors on th	ne examina	ition day	rs (24-h average) and multi-c	lay average	es on and	d before the exa	mination da	iys
24-h average	-1.2	(-9.0; 7.2)	0.77	0.99	1.8	(-4.5; 8.6)	0.58	0.91	-0.4	(-5.2; 4.6)	0.87	0.99
4-d average	3.3	(-4.3; 11.5)	0.41	0.71	-2.7	(-9.9; 5.1)	0.49	0.99	-0.1	(-5.0; 5.1)	0.98	0.99
7-d average	6.0	(-4.2; 17.3)	0.26	0.52	-3.0	(-12.5; 7.5)	0.57	0.99	0.3	(-6.3; 7.3)	0.94	0.99
14-d average	2.5	(-17.9; 28.0)	0.83	0.99	1.9	(-16.4; 24.3)	0.85	0.99	-1.8	(-14.7; 13.1)	0.80	0.99
		Particle n	neasures (µg	/m ³) durin	g the 8 v	vork h on the ex	amination d	ays, from p	ersonal	monitors		
PM _{2.5}	8.6	(1.7; 16.0)	0.016	0.10	7.5	(0.1; 15.5)	0.050	0.25	3.9	(-0.5; 8.5)	0.085	0.35
BC	10.6	(1.4; 20.6)	0.025	0.15	2.3	(-3.2; 8.2)	0.42	0.94	3.0	(-1.6; 7.8)	0.21	0.69
Al	9.5	(-8.0; 30.4)	0.31	0.88	4.2	(-1.4; 10.1)	0.15	0.57	0.0	(-4.6; 4.8)	0.99	0.99
Ca	68.4	(5.6; 168.4)	0.031	0.18	1.3	(-2.0; 4.7)	0.45	0.96	-0.7	(-3.7; 2.5)	0.67	0.99
Fe	7.5	(-8.2; 25.9)	0.37	0.94	2.5	(-2.5; 7.6)	0.34	0.88	-0.4	(-4.6; 3.9)	0.84	0.99
К	6.7	(-0.5; 14.5)	0.074	0.34	4.6	(-0.6; 10.0)	0.085	0.39	2.3	(-1.4; 6.2)	0.23	0.74
S	6.1	(-1.4; 14.2)	0.12	0.48	4.1	(-3.5; 12.3)	0.30	0.85	2.1	(-2.6; 7.0)	0.39	0.93
Si	4.7	(-9.2; 20.7)	0.53	0.99	3.8	(-1.5; 9.3)	0.16	0.62	-0.1	(-4.4; 4.3)	0.96	0.99
Ti	6.2	(-6.5; 20.7)	0.36	0.92	4.7	(-0.1; 9.7)	0.061	0.30	1.3	(-2.8; 5.7)	0.54	0.99
Zn	3.3	(-4.1; 11.4)	0.39	0.95	1.0	(-4.8; 7.1)	0.75	0.99	0.3	(-3.6; 4.5)	0.88	0.99

*Adjusted for age, sex, BMI, smoking status, day of week, and average of temperature and dew point values corresponding to the average of exposure metrics.

[†]Number of observations used for analysis was 102 for office workers, 106 for truck drivers, and 208 for all participants.

[‡]*P*-values were calculated using mixed-effects regression models.

[§]Adjusted *P*-values were estimated by a resampling-based approach, which takes dependent structures among exposures into consideration. This approach controls family-wise error rate (FWER), i.e., probability of having at least one false positive among the whole set of comparisons at a given significant level.

5hmC content in HEK293 cells.⁴¹ Chia et al.²⁵ have proposed that ROS affects 5hmC patterns via metabolic alterations influencing the tricarboxylic acid cycle and thereby activating TET and other chromatic modifying proteins. We note that our study was based on global measures of genomic content of 5hmC and 5mC. It is plausible that PM and other environmental exposures may have different effects on specific genes and non-coding sequences across the human genome. Our results support further studies using emerging platforms for epigenome-wide scans of 5hmC, which will allow the mapping of environmental effects on 5hmC across the human genome.

In contrast to previous studies, we did not observe any robust significant effects on 5mC. In a study on healthy adults with an annual low PM₁₀ exposure (21.7 \pm 2.0 µg/m³), De Prins et al. showed that 5mC was negatively associated with PM10 average exposure in the 14 days before blood drawing.²⁶ Differences in analytical methods may help account for the differences in results compared to our study. While we used ELISA, De Prins et al. used high-performance liquid chromatography (HPLC) to measure 5mC. Head-to-head comparisons of the performances of HPLC and ELISA in measuring DNA methylation are not yet available, but previous reproducibility studies for other analytes have shown better precision for HPLC.^{42,43} ELISA, which we selected because of the higher sample throughput, may be sufficiently precise to detect environment-induced differences in 5hmC, but it might be inadequate to identify potentially smaller effects on 5mC. In addition, there is a big difference in concentration range of particulate matter air pollution between the

present study (range: $60-186 \ \mu g/m3$ in office workers and 72–190 $\ \mu g/m3$ in truck drivers) and De Prins et al. (range: 18.5–27.7 $\ \mu g/m3$). This could be another contributing factor that leads to the discrepancy in the conclusions.

Differences in the levels and patterns of 5hmC and 5mC are also expected. While the 2 marks often co-exist, 5hmC is significantly enriched at gene transcription start sites coinciding with TET1 on genes with high CpG-content.⁴⁴ Additionally, 5hmC exhibits lower levels in repetitive elements and minor satellites compared with 5mC.⁴⁴ Therefore, 5hmC and 5mC have distinct genomic distributions; the different localizations of 5hmC and 5mC reflect their distinct biological functions and might help to explain the differential effects of PM observed in our study.

In our study, we did not find any significant associations of personal measures of exposures to $PM_{2.5}$ mass and elemental components with 5hmC. Ambient PM_{10} mass measures and personal measures differed for the time windows over which they were evaluated. Ambient PM_{10} mass data were available as daily averages, while we conducted our personal measures for 8 h during the participants' work shifts. This difference in sampling times might have contributed to the differences in the results. In fact, our data show stronger effects of ambient PM_{10} with longer multi-day averages. The increasingly larger effects of PM_{10} on 5hmC with longer PM_{10} averages over the days preceding the exams suggest that the effect of PM_{10} on 5hmC tend to build up in time. Shorter hourly averages, such as the 8-h time window used for personal exposure, might

have comparatively weaker effects, explaining the lack of associations in our study. Taken together, since personal $PM_{2.5}$ measurement were only collected for 8-hr average, we are not able to draw a definite conclusion on the impact of short-term $PM_{2.5}$ exposure on 5hmC. However, it is possible that PM_{10} and $PM_{2.5}$ may have distinct biologic effects on 5hmC due to their differences in composition and size. Future toxicological studies are required to ascertain the mechanistic underpinning of the different effects of PM_{10} and $PM_{2.5}$ on 5hmC.

Our results suggested potential effect modification by occupation (truck driver/office workers), which might imply that office workers were more sensitive/vulnerable to the effect of BC on 5hmC, compared to truck drivers due to unknown physiological/sociological factors. However, due to limited sample size we did not have enough statistical power to detect effect modification.

We recognize that our study is subject to a number of limitations. We cannot exclude false negative findings or chance findings due to the relatively small sample size; however, all our findings were significant using a stringent cutoff of 0.05 for FWER, limiting the chance of false-positive results. While our study population may not be representative of the general population, consistent internal validity and adjustment for various lifestyle factors and demographic characteristics allow for moderate generalizability. Our main findings are based on stationary measures of ambient PM_{10} , which are used as a proxy of personal exposure. Based on the study operations, it is reasonable to assume that the measurement error occurred independent of the outcome (global 5mC and 5hmC in blood DNA). Therefore, the misclassification was likely non-differential and can only bias the results toward to null. Previous studies have also shown that the error introduced by using data from stationary monitors usually leads to underestimating of the effects of air pollution.⁴⁵ Moreover, we used multivariable models to control potential confounders, while residual confounding due to unmeasured variables is not unlikely, chances that the observed association reflected bias resulting from residual confounding are minimized. Finally, due to the lack of blood cell counts data, we cannot exclude actual epigenetic variation from exposurerelated differences in cellular composition. It is possible that the change in methylation status was partially due to changes in cell composition following PM₁₀ exposures.

Conclusion

In summary, our study suggests that exposure to ambient PM_{10} affects blood genomic content of 5hmC over time, but not of 5mC. If confirmed, this finding would indicate the need to differentiate 5hmC and 5mC in environmental studies of DNA methylation. Further studies are warranted to evaluate the patterns of 5hmC in relation to PM across the human epigenome and to identify relevant biological pathways for human health effects.

Methods

Study population and design

The Beijing Truck Driver Air Pollution Study, 13, 30-32 conducted between June 15 and July 27, 2008, included 60 truck drivers and 60 indoor office workers. All study participants worked and lived in the Beijing metropolitan area and had held their current jobs for at least 2 years. Truck drivers and indoor office workers were similarly distributed by age, sex, smoking, and education level. In-person interviews using a detailed questionnaire were conducted to collect information on demographics, lifestyle, and other exposures. Information on timevarying factors, including alcohol consumption and smoking status, was obtained for past usual exposure as well as for each examination day. Because PM levels vary on a day-to-day basis, we examined all participants on 2 workdays separated by a 1-2 week period. Individual written informed consent was obtained from all participants before enrollment in the study. Institutional Review Board approval was obtained at all participating institutions before study participant recruitment.

Sample collection and DNA global methylation analysis (5hmC and 5mC)

All blood draw procedures were conducted at approximately the same time of the day between 4-6 pm (after participants completed a full work-day) to eliminate confounding due to diurnal variation. Their whole blood was collected in EDTA tubes and processed within 2 h. The samples were centrifuged at 2,500 rpm to separate the buffy coat and stored at -80° C until analysis. Wizard Genomic DNA Purification Kit (Promega, Madison, WI) was used to extract total DNA from 200 µL buffy coat.³² One hundred nanograms of genomic DNA in each reaction were used to measure global %5hmC and %5mC by specific ELISA. 5hmC was measured using the Quest 5hmCTM DNA ELISA Kit. 5mC was measured using the 5mC DNA ELISA Kit (both kits from Zymo Research, Orange, CA, USA). Analyses were conducted according to the manufacturer's protocols, and all experiments were run in triplicate. Blood DNA samples were randomized across plates to limit potential bias from plate effects, and laboratory personnel were blinded to exposure groups and exposure study. Both kits are highly sensitive and specific to the mC modification of concern, with no cross-reactivity for the other mC modification. The within- and between-assay coefficients of variation were 3.9% and 10.8%, respectively, for 5hmC; and 3.01% and 14.5%, respectively, for 5mC. The detection limit for per 100 ng input DNA was 0.02% for 5hmC and 0.5% for 5mC.

Ambient PM₁₀ mass data

Ambient PM_{10} data during the study period were obtained from the Beijing Municipal Environmental Protection Bureau (http://www.bjepb.gov.cn/). To determine individual-level PM_{10} exposures, we used the aggregate measures (the daily averages of PM_{10} levels across all the 27 monitoring stations) as surrogate measures of individual-level exposure in Beijing on that specific day. We then constructed the exposure matrix for each individual based on the time window of interest. In detail, we calculated the individual-level exposure for 1-day mean (PM_{10} on examination day), 4-d mean (average of PM_{10} mass on the examination day and on the 3 d before examination), 7-d mean (average of PM_{10} on the examination day and on the 6 days before examination), and 14-d mean (average of PM_{10} on the examination day and on the 13 d before examination). We obtained daily outdoor temperature data for Beijing city from the National Oceanic and Atmospheric Administration online database.³⁰

Personal $PM_{2.5}$ mass and elemental component measurements

We measured average personal PM2.5 on both examination days using gravimetric samplers worn by the study participants during 8 h of work. The air sampler was carried in a belt pack with the inlet clipped near the breathing zone. Each air sampler setup included an Apex pump (Casella Inc., Bedford, UK), a Triplex Sharp-Cut Cyclone (BGI Inc., Waltham, Massachusetts, USA), and a 37-mm Teflon filter placed on top of a drain disc and inside a metal filter holder. The filters were kept under atmosphere-controlled conditions before and after sampling and were weighed with a microbalance (Mettler-Toledo Inc., Columbus, Ohio, USA). A time-weighted average of PM_{2.5} concentrations was calculated by dividing the change in filter weight before and after sampling by the volume of air sampled. We found high reproducibility of $PM_{2,5}$ measures (r=0.944) in replicate samples on a subset of 24 participants who wore 2 monitors at the same time (data not shown). The blackness of the same filters was used to measure PM_{2.5} and was assessed using an EEL Model M43D smoke stain reflectometer, applying the standard black-smoke index calculations of the absorption coefficients based on reflectance.⁴⁶ We assumed a factor of 1.0 for converting the absorption coefficient to black carbon (BC) mass,^{47,48} which was then divided by the sampled air volume to calculate average BC exposure concentration.⁴⁶ BC is a combustion by -product contained in PM that has been used as a surrogate measure for PM from gasoline- and, especially, diesel-powered motor vehicles,47 and others like coal combustion and biomass burning.49

Elemental components of PM were measured from the PM collected on the filters using an XRF PANanalytical Epsilon 5 analyzer (Almelo, Netherlands), as described previously.^{50,51} We selected the 8 elements, i.e., potassium (K), sulfur (S), iron (Fe), silicon (Si), aluminum (Al), zinc (Zn), calcium (Ca), and titanium (Ti), that showed the highest reproducibility (r>0.75) in replicate samples from the subset of 24 participants who wore 2 monitors at the same time (data not shown).

Statistical analysis

In the present study, we examined each participant on 2 different days. Therefore, measures may lack independence within participants. To account for this data feature, we used mixedeffects regression models with random intercept in all analyses (PROC MIXED in SAS 9.3, SAS Institute Inc., Cary, NC).

To evaluate the association of short-term variations on air particle with 5hmC and 5mC, the following mixed-effects model was used:

$$Y_{ij} = \beta_0 + \beta_1 (\text{Airparticle})_{ij} + \beta_2 X_{2ij} + \dots + \beta_n X_{nij} + \zeta_j + e_{ij}$$

where Y_{ii} is the measures of 5hmC or 5mC for the *j*th participant measured on the *i*th examination day; β_0 is the overall intercept; β_1 is the regression coefficient for exposure variable, i.e., air particle level; $\beta_2 \dots \beta_n$ are the regression coefficients for the time-dependent covariates (i.e., temperature and dew point) or time-independent covariates (i.e., occupation group, gender, age, BMI, and smoking status) included in multivariate models; ξ_i is the random effect for the participant, and e_{ij} is the residual error term. A two-sided p level of less than 0.05 was considered significant. To account for multiple hypothesis testing where the underlying tests are dependent, we conducted a single-step permutation max T procedure⁵² within the 4 ambient PM_{10} measures, and within the other personal monitor measures (PM2.5, BC, and elemental component), respectively. The conventional used Benjamini-Hochberg False Discovery Rate correction assumes that all hypotheses should be independent, which was not the case in our study. In fact, the 4 PM₁₀ time windows are inherently inter-related, as the shorter moving averages are embedded in the longer moving averages. The personal monitor measures (PM_{2.5}, BC, and constituents) are not independent as BC and constituents are all analyzed on the same PM_{2.5} sample collected from the same device and same filter. Adjusted p-values were calculated from 1,000 permutations in a way that matches the null hypothesis. This method strongly controls FWER (family-wise error rate), which is the probability of having at least one false positive among the whole set of comparisons. All analyses were performed in SAS 9.3 (SAS Institute Inc., Cary, NC).

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Funding

This work has been supported by the NIEHS (R21ES020984, R21ES020010, R01ES021733). MSG has been financially supported by the Fundación México en Harvard, A.C and Consejo Nacional de Ciencia y Tecnología (CONACYT-Mexico). Additional support was received from center grants from the NIEHS (ES00002) and the US EPA (R-82735301).

Supplemental Material

Supplemental data for this article can be accessed on the publisher's website.

Authors' Contributions

SW, CD, PAB, JS, WZ, LH, and AAB designed the study and supervised the study operations. CD, JPM, AD, and XZ prepared the study protocols and oversaw their implementation. MSG, COY, YC, and HMB developed and/or supervised the epigenetic analyses. JM, CMK, and PK were responsible for personal exposure measures. YZ performed the statistical analyses. MSG, COY, JZ, LH, and AAB wrote the manuscript. All authors read and approved the final manuscript.

References

- World Health Organization. Burden of disease from Ambient Air Pollution for 2012 - Summary of Results. Geneva, Switzerland: WHO, 2014.
- EPA. Identification of Nonattainment Classification and Deadlines for Submission of State Implementation Plan (SIP) Provisions for the 1997 Fine Particle (PM2.5) National Ambient Air Quality Standard (NAAQS) and 2006 PM2.5 NAAQS Federal register, 2014.
- Johnson PR, Graham JJ. Fine particulate matter national ambient air quality standards: public health impact on populations in the northeastern United States. Environ Health Perspect 2005; 113:1140-7; PMID:16140618; http://dx.doi.org/10.1289/ehp.7822
- Johnson PR, Graham JJ. Analysis of primary fine particle national ambient air quality standard metrics. J Air Waste Manag Assoc 2006; 56:206-18; PMID:16568804; http:// dx.doi.org/10.1080/10473289.2006.10464447
- Li CZ, Budh E. Environmental Objectives, Cost Efficiency, and Multivariate Stochastic Control. Environ Model Assess 2008; 13:315-25; http://dx.doi.org/ 10.1007/s10666-007-9095-5
- Amann M, Asman A, Bertok I, Cofala J, Heyes C, Klimont Z, Rafaj P, Schöpp W, Wagner F. Cost-optimized reductions of air pollutant emissions in the EU Member States to address the environmental objectives of the Thematic Strategy on Air Pollution. Laxenburg, Austria: International Institute for Applied Systems Analysis (IIASA), 2007.
- Bollati V, Baccarelli A. Environmental epigenetics. Heredity 2010; 105:105-12; PMID:20179736; http:// dx.doi.org/10.1038/hdy.2010.2
- Cortessis VK, Thomas DC, Levine AJ, Breton CV, Mack TM, Siegmund KD, Haile RW, Laird PW. Environmental epigenetics: prospects for studying epigenetic mediation of exposure-response relationships. Hum Genet 2012; 131:1565-89; PMID:22740325; http://dx.doi.org/10.1007/s00439-012-1189-8
- Hou L, Zhang X, Wang D, Baccarelli A. Environmental chemical exposures and human epigenetics. Int J Epidemiol 2012; 41:79-105; PMID:22253299; http:// dx.doi.org/10.1093/ijc/dyr154
- Alegria-Torres JA, Baccarelli A, Bollati V. Epigenetics and lifestyle. Epigenomics 2011; 3:267-77; PMID:22122337; http://dx.doi.org/10.2217/epi.11.22
- Baccarelli A, Ghosh S. Environmental exposures, epigenetics and cardiovascular disease. Curr Opin Clin Nutr Metab Care 2012; 15:323-9; PMID:22669047; http:// dx.doi.org/10.1097/MCO.0b013e328354bf5c
- Fleisch AF, Wright RO, Baccarelli AA. Environmental epigenetics: a role in endocrine disease? J Mol Endocrinol 2012; 49:R61-7; PMID:22798698; http://dx.doi. org/10.1530/JME-12-0066
- Guo L, Byun HM, Zhong J, Motta V, Barupal J, Zheng Y, Dou C, Zhang F, McCracken JP, Diaz A, et al. Effects of short-term exposure to inhalable particulate matter on DNA methylation of tandem repeats. Environ Mol Mutagen 2014; 55:322-35; PMID:24436168; http://dx.doi. org/10.1002/em.21838
- Tarantini L, Bonzini M, Apostoli P, Pegoraro V, Bollati V, Marinelli B, Cantone L, Rizzo G, Hou L, Schwartz J, et al. Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation. Environ Health Perspect 2009; 117:217-22; PMID:19270791; http://dx.doi.org/10.1289/ ehp.11898
- Baccarelli A, Wright R, Bollati V, Litonjua A, Zanobetti A, Tarantini L, Sparrow D, Vokonas P, Schwartz J. Ischemic heart disease and stroke in relation to blood DNA methylation. Epidemiology 2010; 21:819-28; PMID:20805753; http://dx.doi.org/10.1097/ EDE.0b013e3181f20457

- Bind MA, Lepeule J, Zanobetti A, Gasparrini A, Baccarelli A, Coull BA, Tarantini L, Vokonas PS, Koutrakis P, Schwartz J. Air pollution and gene-specific methylation in the Normative Aging Study: Association, effect modification, and mediation analysis. Epigenetics 2014; 9:448-58; PMID:24385016; http://dx. doi.org/10.4161/epi.27584
- Hou L, Zhang X, Zheng Y, Wang S, Dou C, Guo L, Byun HM, Motta V, McCracken J, Diaz A, et al. Altered methylation in tandem repeat element and elemental component levels in inhalable air particles. Environ Mol Mutagen 2014; 55:256-65; PMID:24273195; http://dx. doi.org/10.1002/em.21829
- Byun HM, Motta V, Panni T, Bertazzi PA, Apostoli P, Hou L, Baccarelli AA. Evolutionary age of repetitive element subfamilies and sensitivity of DNA methylation to airborne pollutants. Part Fibre Toxicol 2013; 10:28; PMID:23855992; http://dx.doi.org/10.1186/ 1743-8977-10-28
- Pastor WA, Pape UJ, Huang Y, Henderson HR, Lister R, Ko M, McLoughlin EM, Brudno Y, Mahapatra S, Kapranov P, et al. Genome-wide mapping of 5-hydroxymethylcytosine in embryonic stem cells. Nature 2011; 473:394-7; PMID:21552279; http://dx.doi.org/ 10.1038/nature10102
- Branco MR, Ficz G, Reik W. Uncovering the role of 5hydroxymethylcytosine in the epigenome. Nat Rev Genet 2012; 13:7-13; PMID:NOT_FOUND
- Wu H, D'Alessio AC, Ito S, Wang Z, Cui K, Zhao K, Sun YE, Zhang Y. Genome-wide analysis of 5-hydroxymethylcytosine distribution reveals its dual function in transcriptional regulation in mouse embryonic stem cells. Gen Dev 2011; 25:679-84; PMID:21460036; http://dx.doi.org/10.1101/gad.2036011
- Huang Y, Pastor WA, Shen Y, Tahiliani M, Liu DR, Rao A. The behaviour of 5-hydroxymethylcytosine in bisulfite sequencing. PloS one 2010; 5:e8888; PMID:20126651; http://dx.doi.org/10.1371/journal. pone.0008888
- Nestor C, Ruzov A, Meehan R, Dunican D. Enzymatic approaches and bisulfite sequencing cannot distinguish between 5-methylcytosine and 5-hydroxymethylcytosine in DNA. BioTechniques 2010; 48:317-9; PMID:20569209; http://dx.doi.org/10.2144/ 000113403
- Booth MJ, Branco MR, Ficz G, Oxley D, Krueger F, Reik W, Balasubramanian S. Quantitative sequencing of 5-methylcytosine and 5-hydroxymethylcytosine at single-base resolution. Science 2012; 336:934-7; PMID:22539555; http://dx.doi.org/10.1126/science. 1220671
- Chia N, Wang L, Lu X, Senut MC, Brenner C, Ruden DM. Hypothesis: environmental regulation of 5hydroxymethylcytosine by oxidative stress. Epigenetics 2011; 6:853-6; PMID:21617369; http://dx.doi.org/ 10.4161/epi.6.7.16461
- De Prins S, Koppen G, Jacobs G, Dons E, Van de Mieroop E, Nelen V, Fierens F, Int Panis L, De Boever P, Cox B, et al. Influence of ambient air pollution on global DNA methylation in healthy adults: a seasonal follow-up. Environ Int 2013; 59:418-24; PMID:23917442; http://dx.doi.org/10.1016/j.envint. 2013.07.007
- Madrigano J, Baccarelli A, Mittleman MA, Wright RO, Sparrow D, Vokonas PS, Tarantini L, Schwartz J. Prolonged exposure to particulate pollution, genes associated with glutathione pathways, and DNA methylation in a cohort of older men. Environ Health Perspect 2011; 119:977-82; PMID:21385671; http://dx.doi. org/10.1289/ehp.1002773
- Baccarelli A, Wright RO, Bollati V, Tarantini L, Litonjua AA, Suh HH, Zanobetti A, Sparrow D, Vokonas PS, Schwartz J. Rapid DNA methylation changes after

exposure to traffic particles. Am J Respir Crit Care Med 2009; 179:572-8; PMID:19136372; http://dx. doi.org/10.1164/rccm.200807-1097OC

- 29. Bank TW. The World Bank. 2012. World Development Indicators. Washington, D.C., 2012.
- Baccarelli A, Barretta F, Dou C, Zhang X, McCracken JP, Diaz A, Bertazzi PA, Schwartz J, Wang S, Hou L. Effects of particulate air pollution on blood pressure in a highly exposed population in Beijing, China: a repeated-measure study. Environ Health 2011; 10:108; PMID:22188661; http://dx.doi.org/10.1186/1476-069X-10-108
- 31. Hou L, Wang S, Dou C, Zhang X, Yu Y, Zheng Y, Avula U, Hoxha M, Diaz A, McCracken J, et al. Air pollution exposure and telomere length in highly exposed subjects in Beijing, China: a repeated-measure study. Environ Int 2012; 48:71-7; PMID:22871507; http://dx.doi.org/10.1016/j.envint.2012.06.020
- 32. Hou L, Zhang X, Dioni L, Barretta F, Dou C, Zheng Y, Hoxha M, Bertazzi PA, Schwartz J, Wu S, et al. Inhalable particulate matter and mitochondrial DNA copy number in highly exposed individuals in Beijing, China: a repeated-measure study. Part Fibre Toxicol 2013; 10:17; PMID:23628000; http://dx.doi.org/ 10.1186/1743-8977-10-17
- Bachman M, Uribe-Lewis S, Yang X, Williams M, Murrell A, Balasubramanian S. 5-Hydroxymethylcytosine is a predominantly stable DNA modification. Nat Chem 2014; 6:1049-55; PMID:25411882; http://dx. doi.org/10.1038/nchem.2064
- 34. Tellez-Plaza M, Tang WY, Shang Y, Umans JG, Francesconi KA, Goessler W, Ledesma M, Leon M, Laclaustra M, Pollak J, et al. Association of Global DNA Methylation and Global DNA Hydroxymethylation with Metals and other Exposures in Human Blood DNA Samples. Environ Health Perspect 2014; 122 (9):946-54; PMID:24769358; http://dx.doi.org/ 10.1289/ehp.1306674
- Sarter B, Long TI, Tsong WH, Koh WP, Yu MC, Laird PW. Sex differential in methylation patterns of selected genes in Singapore Chinese. Hum Genet 2005; 117:402-3; PMID:15928902; http://dx.doi.org/ 10.1007/s00439-005-1317-9
- Kim M, Long TI, Arakawa K, Wang R, Yu MC, Laird PW. DNA methylation as a biomarker for cardiovascular disease risk. PloS one 2010; 5:e9692; PMID:20300621; http://dx.doi.org/10.1371/journal. pone.0009692
- Gurgueira SA, Lawrence J, Coull B, Murthy GG, Gonzalez-Flecha B. Rapid increases in the steady-state concentration of reactive oxygen species in the lungs and heart after particulate air pollution inhalation. Environ Health Perspect 2002; 110:749-55; PMID:12153754; http://dx.doi.org/10.1289/chp.02110749
- Kim JY, Mukherjee S, Ngo LC, Christiani DC. Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates. Environ Health Perspect 2004; 112:666-71; PMID:15121508; http://dx.doi.org/10.1289/ehp.6827
- Ren C, Park SK, Vokonas PS, Sparrow D, Wilker E, Baccarelli A, Suh HH, Tucker KL, Wright RO, Schwartz J. Air pollution and homocysteine: more evidence that oxidative stress-related genes modify effects of particulate air pollution. Epidemiology 2010; 21:198-206; PMID:20110814; http://dx.doi.org/ 10.1097/EDE.0b013e3181cc8bfc
- Kolachana P, Subrahmanyam VV, Meyer KB, Zhang L, Smith MT. Benzene and its phenolic metabolites produce oxidative DNA damage in HL60 cells in vitro and in the bone marrow in vivo. Cancer Res 1993; 53:1023-6; PMID:8439949
- Coulter JB, O'Driscoll CM, Bressler JP. Hydroquinone increases 5-hydroxymethylcytosine formation through

ten eleven translocation 1 (TET1) 5-methylcytosine dioxygenase. J Bio Chem 2013; 288:28792-800; PMID:23940045; http://dx.doi.org/10.1074/jbc.M113. 491365

- 42. Pirestani A, Tabatabae SN, Fazeli MH, Antikchi M, Baabaei B. Comparison of HPLC and Elisa for Determination of Aflatoxin Concentration in the Milk and Feeds of Dairy Cattle. J Res Agricultural Sci 2011; 7:71-8.
- Fischer JB, Michael JL. Use of ELISA immunoassay kits as a complement to HPLC analysis of imazapyr and triclopyr in water samples from forest watersheds. Bull Environ Contam Toxicol 1997; 59:611-8; PMID:9307427; http://dx.doi.org/10.1007/ s001289900523
- Williams K, Christensen J, Pedersen MT, Johansen JV, Cloos PA, Rappsilber J, Helin K. TET1 and hydroxymethylcytosine in transcription and DNA methylation fidelity. Nature 2011; 473:343-8; PMID:21490601; http://dx.doi.org/10.1038/nature10066
- Zeger SL, Thomas D, Dominici F, Samet JM, Schwartz J, Dockery D, Cohen A. Exposure measurement error in time-series studies of air pollution: concepts and consequences. Environ Health Perspect 2000; 108:419-26; PMID:10811568; http://dx.doi.org/10.1289/ehp. 00108419
- ISO. ISO 9835, Ambient air Determination of a black smoke index. Geneva: International Organization for Standardization. 1993.
- Kinney PL, Aggarwal M, Northridge ME, Janssen NA, Shepard P. Airborne concentrations of PM(2.5) and diesel exhaust particles on Harlem sidewalks: a community-based pilot study. Environ Health Perspect 2000; 108:213-8; PMID:10706526
- Janssen NA vVP, Aarts F, Harssema H, Brunekree B. Assessment of exposure to traffic related air pollution of children attending schools near motorways. Atmospheric Environment 2001; 35:9; http:// dx.doi.org/10.1016/S1352-2310(01)00144-3
- Schleicher N, Norra S, Fricker M, Kaminski U, Chen Y, Chai F, Wang S, Yu Y, Cen K. Spatio-temporal variations of black carbon concentrations in the Megacity Beijing. Environ Pollut 2013; 182:392-401; PMID:23978522; http://dx.doi.org/10.1016/j.envpol. 2013.07.042
- Watson GJ, Chow JC, Frazier CA. X-ray fluorescence analysis of ambient air samples. In: Landsberger SC M , ed. Elemental analysis of airborne particles. Amsterdam, The Netherlands Gordon and Breach Science Publishers 1999:67-96.
- 51. Chow JC, Watson JG. Guideline on speciated particulate monitoring. Draft report 3 prepared for the US Environmental Protection Agency, Office of Air Quality Planning and Standards. Research Triangle Park, NC: Desert Research Institute, Reno, NV, 1998.
- Westfall P, Young S. Resampling-Based Multiple Testing: Examples and Methods for p-Value Adjustment (Wiley Series in Probability and Statistics). Wiley-Interscience, 1993.