



Chloroplast Genome Sequence of *Lagerstroemia guilinensis* (Lythraceae, Myrtales), a Species Endemic to the Guilin Limestone Area in Guangxi Province, China

Cuihua Gu,^a Luke R. Tembrock,^b Zhiqiang Wu^b

School of Landscape and Architecture, Zhejiang Agriculture and Forestry University, Lin'an, Hangzhou, Zhejiang, People's Republic of China^a; Department of Biology, Colorado State University, Fort Collins, Colorado, USA^b

We announce here the first complete chloroplast genome sequence of *Lagerstroemia guilinensis* (Lythraceae, Myrtales), a species endemic to the Guilin limestone area, along with its genome structure and functional gene annotations. The plant was collected from Guilin, Guangxi, China, and deposited as a germplasm accession of the Zhejiang Agriculture and Forestry University Collection (ZAFU 1507144). This genome will provide valuable information for future research of the *Lagerstroemia* genus and its relatives.

Received 11 March 2016 Accepted 8 April 2016 Published 19 May 2016

Citation Gu C, Tembrock LR, Wu Z. 2016. Chloroplast genome sequence of *Lagerstroemia guilinensis* (Lythraceae, Myrtales), a species endemic to the Guilin limestone area in Guangxi Province, China. Genome Announc 4(3):e00341-16 doi:10.1128/genomeA.00341-16.

Copyright © 2016 Gu et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license. Address correspondence to Zhiqiang Wu, wuzhiqiang.1020@gmail.com.

Lagerstroemia is the most economically valuable genus in Lythraceae due to its utility as an ornamental plant. The genus is composed of about 55 species (1–3). Lagerstroemia guilinensis is a 2-m-tall shrub with a distribution documented only around Xishan Park in Guilin City, China. Due to its narrow and limited distribution, L. guilinensis is at a higher risk of extinction than other broadly distributed Lagerstroemia species. L. guilinensis is only found growing on limestone mountains and blooms from May until July. Molecular research has been done to identify Lagerstroemia cultivars and interspecific hybrids (4, 5), but there is a lack of complete genome-level research on Lagerstroemia. We acquired L. guilinensis (ZAFU 1507144) samples from Xishan Park of Guilin City, Guangxi Province, China, to finish its chloroplast (cp) genome.

Chloroplast genomes have a highly conserved circular DNA quadruplet structure ranging from 120 to 165 kb, with conserved gene order, similarity of sequence across the land plants, uniparental inheritance, and low recombination rates (6–8) compared to nuclear genomes. Plant cp genomes provide a valuable resource of markers in phylogenetics (9–11), DNA barcoding (12), and biogeography among populations (13). With the dramatically reduced cost of next-generation sequencing, it has become more convenient to sequence whole cp genomes (14). More than 900 land plant complete cp genomes can be accessed at the NCBI database (15).

The raw Illumina reads generated for this report were trimmed by quality score using Trimmomatic version 0.3 (16). The *de novo* assembly of reads from *L. guilinensis* were finished using CLC Genomics Workbench version 7, with the default settings (CLC bio). *De novo* assembly was used (17) to construct the assemblies. After merging Illumina and Sanger sequence data, the whole cp genome for *L. guilinensis* was found to be 152,074 bp. The final cp genome was annotated by DOGMA (http://dogma.ccbb.utexas .edu/) with manual adjustment of the exon-intron junctions (17).

We elucidated the genomic characteristics of this species: the cp genome was 152,074 bp in length, with 37.6% overall G+C content. The genome structure was highly similar to that of land plants, consisting of two inverted regions (IRs) (25,677 bp), a large single copy (LSC) (83,811 bp), and a small single copy (SSC) (16,909 bp). Of the 112 unique genes (78 protein-coding genes, 4 rRNAs, and 30 tRNAs), 82 genes are located in the LSC region (60 protein-coding genes and 22 tRNA genes), 13 genes are located in the SSC region (12 protein-coding genes and 1 tRNA gene), and 17 genes are located in both IR regions (6 coding genes, 4 rRNA genes, and 7 tRNA genes). Sixteen genes were found to have introns, with 5 tRNA genes having a single intron each (trnA-GUC, trnG-UCC, trnI-GAU, trnK-UUU, trnL-UAA, and trnV-UAC), eight protein-coding genes having a single intron each (atpF, ndhA, ndhB, petB, petD, rpl16, rpoC1, and rps16), and three protein-coding genes having two introns each (clpP, rps12, and *ycf3*).

Nucleotide sequence accession number. The accession number for the DNA sample of *L. guilinensis* was stored at Zhejiang Agriculture and Forestry University. The complete cp genome sequence with its annotation has been submitted to GenBank under the accession no. KU885923.

ACKNOWLEDGMENTS

This study was supported by The National Natural Science Foundation of China (grant 31300581).

FUNDING INFORMATION

This work, including the efforts of Cuihua Gu, was funded by National Natural Science Foundation of China (NSFC) (31300581).

REFERENCES

- Qin HN, Graham SA. 2007. Lagerstroemia. Flora of China 13:277–281. http://www.efloras.org/florataxon.aspx?flora_id=2&taxon_id=117489.
- 2. Koehne E. 1884. Lythraceae monographice describuntur. Morphologie de

Vegetationsorgane. Botanical Jahrbücher für Systematik, Pflanzengeschichte und Pflanzengeographie 4: Beiblatt 2, Heft 1:95–132.

- 3. Furtado CX, Srisuko M. 1969. A revision of *Lagerstroemia* L. (*Lythraceae*). Gdns Bull 24:185–335.
- Pooler MR. 2003. Molecular genetic diversity among 12 clones of *Lagerstroemia fauriei* revealed by AFLP and RAPD markers. HortScience 38: 256–259.
- Pounders C, Rinehart T, Sakhanokho H. 2007. Evaluation of interspecific hybrids between *Lagerstroemia indica* and *L. speciosa*. HortScience 42:1317–1322.
- Palmer JD. 1985. Comparative organization of chloroplast genomes. Annu Rev Genet 19:325-354. http://dx.doi.org/10.1146/ annurev.ge.19.120185.001545.
- Ravi V, Khurana JP, Tyagi AK, Khurana P. 2008. An update on chloroplast genomes. Plant Syst Evol 271:101–122. http://dx.doi.org/10.1007/ s00606-007-0608-0.
- Wicke S, Schneeweiss GM, DePamphilis CW, Müller KF, Quandt D. 2011. The evolution of the plastid chromosome in land plants: gene content, gene order, gene function. Plant Mol Biol 76:273–297. http:// dx.doi.org/10.1007/s11103-011-9762-4.
- Moore MJ, Bell CD, Soltis PS, Soltis DE. 2007. Using plastid genomescale data to resolve enigmatic relationships among basal angiosperms. Proc Natl Acad Sci USA 104:19363–19368. http://dx.doi.org/10.1073 /pnas.0708072104. http://dx.doi.org/10.1073/pnas.0708072104.
- 10. Wang L, Qi XP, Xiang QP, Heinrichs J, Schneider H, Zhang XC. 2010.

Phylogeny of the paleotropical fern genus *Lepisorus* (Polypodiaceae, Polypodiopsida) inferred from four chloroplast DNA regions. Mol Phylogenet Evol 54:211–225. http://dx.doi.org/10.1016/j.ympev.2009.08.032.

- Wu ZQ, Ge S. 2012. The phylogeny of the BEP clade in grasses revisited: evidence from the whole-genome sequences of chloroplasts. Mol Phylogenet Evol 62:573–578. http://dx.doi.org/10.1016/j.ympev.2011.10.019.
- Day A, Goldschmidt-Clermont M. 2011. The chloroplast transformation toolbox: selectable markers and marker removal. Plant Biotechnol J 9:540–553. http://dx.doi.org/10.1111/j.1467-7652.2011.00604.x.
- Wang W, Messing J. 2011. High-throughput sequencing of three *Lemnoideae* (duckweeds) chloroplast genomes from total DNA. PLoS One 6:e24670. http://dx.doi.org/10.1371/journal.pone.0024670.
- Soltis DE, Gitzendanner MA, Stull G, Chester M, Chanderbali A, Chamala S, Jordon-Thaden I, Soltis PS, Schnable PS, Barbazuk WB. 2013. The potential of genomics in plant systematics. Taxon 62:886–898.
- Wu ZQ, Tembrock LR. 2015. Two complete chloroplast genomes of white campion (*Silene latifolia*) from male and female individuals. Mitochondrial DNA 1736:1–2. http://dx.doi.org/10.3109/19401736.2015.1126829.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. http:// dx.doi.org/10.1093/bioinformatics/btu170.
- Wu ZQ, Tembrock LR, Ge S. 2015. Are differences in genomic data sets due to true biological variants or errors in genome assembly: an example from two chloroplast genomes. PLoS One 10:e0118019. http://dx.doi.org/ 10.1371/journal.pone.0118019.