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PRRT2 gene mutations associated with infantile convulsions induced by sucking and the genotype-phenotype correlation

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Introduction: *PRRT2* is a major causative gene for self-limited familial neonatal-infantile epilepsy, paroxysmal kinesigenic dyskinesia, and paroxysmal kinesigenic dyskinesia with infantile convulsions. Voluntary movement trigger is prominent in adolescence and adulthood, but the triggers are unknown in infants.

Methods: A gene panel designed for targeted next-generation sequencing (NGS) was used to screen genetic abnormalities in a cohort of 45 cases with infantile convulsions. The copy number variation was detected by a computational method based on the normalized depth of coverage and validated by a quantitative real-time polymerase chain reaction (RT-qPCR) method. The genotype-phenotype correlation of the *PRRT2* mutation gene was analyzed.

Results: A *de novo* heterozygous *PRRT2* deletion was identified in a child who had infantile convulsions induced by vigorous sucking. Seizures happened during the change of feeding behavior from breast to formula, which led to hungry and vigorous sucking. Ictal electroencephalograms recorded seizures with focal origination, which provided direct evidence of epileptic seizures in infants with *PRRT2* mutations. Seizures stopped soon after the feeding behavior was changed by reducing feeding interval time and extending feeding duration. Data reanalysis on our previously reported cases with *PRRT2* mutations showed that six of 18 (33.3%) patients had infantile convulsions or infantile non-convulsion seizures during feeding. The mutations included two truncating mutations (c.579dupA/p.Glu194Argfs*6, and c.649dupC/p.Arg217Profs*8) that were identified in each of the three affected individuals.

Conclusions: This study suggests that feeding, especially vigorous sucking, is potentially a trigger and highlights the significance of feeding behavior in

preventing seizures in infants with *PRRT2* mutations. Identification of *PRRT2* haploinsufficiency mutations in the patients with infantile convulsions induced by sucking suggested a potential genotype-phenotype correlation.

KEYWORDS

PRRT2, infantile convulsion, seizures, sucking-induced, genotype-phenotype correlation

Introduction

The PRRT2 gene (OMIM: 614386) encodes a transmembrane protein containing a proline-rich domain in the N-terminal half and being involved in synaptic transmission (1). PRRT2-associated diseases are mainly paroxysmal kinesigenic dyskinesia (PKD) and paroxysmal hypnogenic dyskinesia (PHD) in adults and self-limited familial neonatal-infantile epilepsy or infantile convulsion and choreoathetosis (ICCA) in infants. Infantile convulsion (IC) is the most common feature in early life among the wide spectrum of PRRT2 phenotypes (2). A previous study demonstrated that self-limited familial neonatal-infantile epilepsy accounted for 41.7%, PKD accounted for 38.7%, and ICCA accounted for 14.3% of the PRRT2-associated phenotypes (3). The PRRT2 mutations previously reported included missense, non-sense, splicing, deletions, insertions, indels, and complex rearrangement; and the majority are truncating mutations that lead to haploinsufficiency. Voluntary movement triggering is a prominent feature in patients of adolescence and adulthood, which is highlighted in the diagnostic criteria for PKD (2). However, the triggers of attacks in early childhood were unknown. We previously observed several patients with PRRT2 mutations that had attacks during feeding, suggesting sucking as a possible kinesigenic trigger (4). In this study, we screened a cohort of 45 cases with infant seizures and identified a PRRT2 heterozygous deletion mutation in a child with frequent seizures induced by vigorous sucking and became seizure-free after changing feeding behavior. We analyzed the clinical and genetic features of the cases who had attacks during feeding and found that their PRRT2 mutations were all truncating or deletion mutations that would lead to haploinsufficiency, which suggested a potential genotype-phenotype correlation and clinical significance.

Materials and methods

Patients

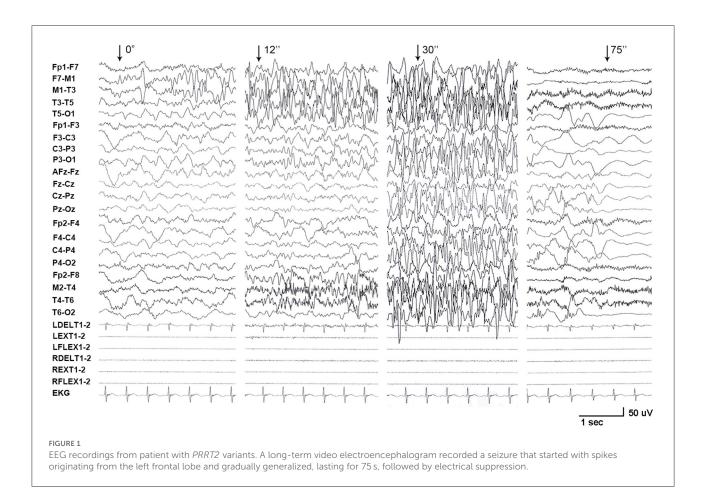
A total of 45 patients with infant convulsion were recruited from the Epilepsy Center of the Second Affiliated Hospital of Guangzhou Medical University and the First People's Hospital of Foshan. Epileptic seizures and epilepsy syndromes were diagnosed and classified according to the criteria of the Commission on Classification and Terminology of the International League Against Epilepsy (1981, 1989, 2001, 2010, and 2017). The collected clinical data included detailed semiology, onset age, seizure type, the evolution of disorder and frequency, family history, response to antiseizure medications, general and neurological examination, and brain magnetic resonance imaging (MRI). Long-term video-EEG monitoring records that included open-close eyes test, hyperventilation, intermittent photic stimulation, and sleep recording were obtained.

This study complied with the principles of the International Committee of Medical Journal Editors about patient consent for research or participation. Written informed consent was obtained from the individuals or legal guardians. Ethical approval had been obtained from the Ethics Committee of the Second Affiliated Hospital of Guangzhou Medical University.

Targeted sequencing and filtering

Peripheral blood samples were collected from the probands, their parents, and other family members if available. Genomic DNA (gDNA) was extracted using the Qiagen Flexi Gene DNA Kit (Qiagen, Hilden, Germany). The *PRRT2* mutations in the early six cases in our previous study were identified by direct Sanger sequencing, targeting the coding regions (4). A customtargeted NGS gene panel was performed in recent years to screen gene abnormalities associated with episodic disorders (5), including *PRRT2*, *CHRNA4*, *KCNA1*, *KCNQ2*, *KCNMA1*, *PNKD*, *SCN2A*, and *SLC2A1* genes that are associated with BFIS and PKD. Trios-based exome paired-end sequencing was performed on the Illumina Nextseq 500 platform by capture probe NeuMet_V1. The mean read depth for each sample

Abbreviations: PKD, paroxysmal kinesigenic dyskinesia; PHD, paroxysmal hypnogenic dyskinesia; ICCA, infantile convulsion and choreoathetosis; IC, infantile convulsion; EEG, electroencephalogram; MRI, brain magnetic resonance imaging; CNVs, copy number variations; qPCR, quantitative real-time-polymerase chain reaction.



was 208-fold. The raw data were aligned to the reference genome (GRCh37/hg19). Single nucleotide variants (SNVs) and insertions/deletions (indels) filtering were performed by using the Genome Analysis Toolkit (6). Stepwise filtering was applied to the derived candidate causative variants as we previously described (7). The copy number variations (CNVs) were performed by a computational method (CNVkit) based on the normalized depth of coverage, which used both the targeted reads and the non-specifically captured off-target reads to infer copy number (8). Sanger sequencing was applied to confirm the candidate variants.

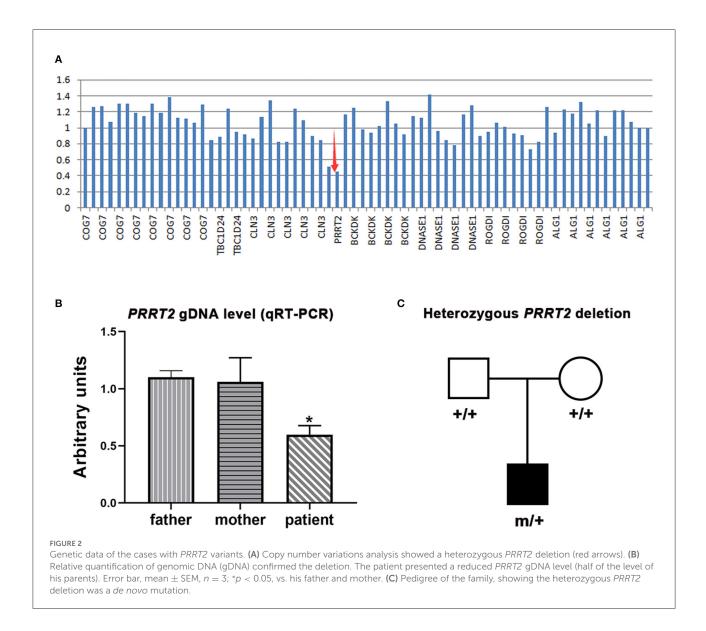
Quantitative real-time PCR (QPCR) analysis

The qPCR was used to verify the heterozygous *PRRT2* deletion at the gDNA level. The parents were regarded as healthy controls. Primers F1 (5'-GGATGCAGAGGGAGTGGAATG-3') and R1 (5'-CCCAGCACCCCAAAATCAAC-3') were designed to amplify the fore part of *PRRT2*, primers F2 (5'-GAAGGCACCCAGAAACCTCG-3') and R2 (5'-ACAGCATAAGCGAAGGCCAC-3') were designed to

amplify the middle part of PRRT2, and primers F3 (5'-AACACCTCCCCAACTCTGCG-3') (5'and R3 CCAGGGTGGAGGTCCAGAGAAT-3') were designed to amplify the whole length of PRRT2. The total volume of each sample reaction was 10 µl, including 1.5 µl (15 ng gDNA), 1 μ l forward primer (2 μ M), 1 μ l reverse primer (2 μ M), 2 μ l ROX Reference Dye (50×), 5 μ l SYBR Premix Ex Taq (2×), and 1.3 µl double distilled water. Each sample was run in triplicate on a 7300 Realtime PCR System (Applied Biosystems) by using the SYBR^(R) Premix Ex TaqTM Reagent (Takara). The expression of POLR2A as a reference gene was used for data normalization. Relative ratios of the gene expression were calculated by formula: $r = 2^{-\Delta\Delta Ct}$ with $\Delta\Delta Ct = (Ct_{PRRT2})$ - Ct_{POLR2A}) ind tested - (Ct_{PRRT2} - Ct_{POLR2A}) ind ref. Values were calculated about the father and arbitrarily attributed to the value of 1.0. SPSS 19.0 software (Chicago, USA) was used for statistical analysis.

Results

One *de novo PRRT2* mutation was identified in a child with infant seizures. The child was a full-term boy who was born



to healthy non-consanguineous parents. His early development was normal. The seizures began at the age of 4 months when breastfeeding was switched to formula feeding. He presented his first convulsion when he started sucking eagerly and vigorously. The seizure began with a deviation of head and eyes to the right, then cyanosis, and followed by the loss of consciousness and generalized hypertonic lasting about 1 min. The attacks repeated 2-10 times per day and mostly occurred during vigorous sucking. The ictal EEGs recorded a seizure that started with spikes from the left frontal lobe and gradually generalized, lasting for 75 s, followed by electrical suppression (Figure 1). Interictal EEGs revealed low voltage spikes and waves in the left frontal lobe with a normal background. Brain MRI was normal. Blood routine and biochemical tests were normal. Valproate (16 mg/kg/d) was applied on the third day after onset but did not stop the seizures. Vigorous sucking was subsequently avoided by shortening interval time, extending the duration of feeding, and adding the rice paste. The child was seizure-free since then (followed-up for 2 years until now). EEG became normal in the third month after being seizure-free. Valproate was discontinued after a year. The neurodevelopment was normal at 3 years of follow-up. Using the gene panel, we failed to find any pathogenic point mutations in this child. Heterozygous *PRRT2* gene deletion was identified by the computational method of CNVs (Figure 2A) and validated by the qPCR method (Figure 2B). The mutation was a heterozygous whole deletion of one allele of the *PRRT2* gene, which led to haploinsufficiency. Co-segregation analysis showed that the mutation was a *de novo* mutation (Figure 2C).

We reanalyzed the data from the cohort of 34 individuals with *PRRT2* mutations (Figure 2 in that article) (4). Six patients were found to have experienced feeding-associated infantile

Mutation	Inheritance/sex	Seizure	Onset age	Frequency	Duration	Remission age	EEG	ASMs	Evolution/ onset age
c.579dupA/p.Glu194Argfs*	6 Familial/Male	IC	5 mon.	2–3 times/day	2 min.	12 mon.	Normal	None	PKD/7 yr.
c.579dupA/p.Glu194Argfs*	6 Familial/Female	IC	5 mon.	0–1 time/week	20 sec.	12 mon.	Normal	None	PKD/13 yr.
c.579dupA/p.Glu194Argfs*	6 Familial/Female	IC	5 mon.	1–2 times/day	20 sec.	12 mon.	Normal	None	PKD/5 yr.
c.649dupC/p.Arg217Profs*	8 Familial/Male	IC	6 mon.	2-3 times/day	1 min.	6 mon.	Normal	PB	PKD/13 yr.
c.649dupC/p.Arg217Profs*	8 Familial/Female	INCS-IC	8 mon.	4 times-once	1-2 min.	9 mon.	RTSW	VPA	NA
c.649dupC/p.Arg217Profs*	8 Familial/Male	INCS	10 mon.	0-8/day	${\sim}10$ sec.	3 yr.	RF-Sharp	PB	PD/9 yr.
Entire PRRT2	De novo/Male	IC	4 mon.	2-10 times/day	1 min.	5 mon.	Spikes	VPA	Follow-up
heterozygote deletion									

TABLE 1 Genetic and clinical characteristics of patients with feeding-induced infantile convulsion/non-convulsion seizure caused by PRRT2 mutations.

ASMs, anti-seizure medications; IC, infantile convulsion; INCS, infantile non-convulsion seizure; NA, no not available; PB, phenobarbital; PD, Paroxysmal dyskinesias; PKD, paroxysmal kinesigenic dyskinesia; RF, right frontal; RTSW, right temporal slow wave; VPA, Valproate.

convulsions or infantile non-convulsion seizures. The clinical and genetic features of the patients are summarized in Table 1. All patients had heterozygous deleterious mutations, including two truncating mutations, c.579dupA/p.Glu194Argfs*6, and c.649dupC/p.Arg217Profs*8, that were identified in each of the three affected individuals. Mutation c.649dupC/p.Arg217Profs*8 has previously been reported as a hot-spot mutation (3). Except for the deletion mutation that was de novo, the others were of familial inheritance. The average age at onset was 6.1 \pm 2.1 months (range 4–10 months). Five patients exhibited classical convulsions. Two patients presented non-convulsive seizures that manifested as a stop of suction, loss of responses, and then cyanosis without a generalized tonic seizure. All patients had normal EEG backgrounds. Three of them had focal (frontal/ temporal) spikes and sharp or slow waves. Six were in remission before 12 months and one was in remission at age of 3 years old. Five patients developed paroxysmal disorders or paroxysmal dyskinesias later in adolescence.

Discussion and conclusions

The *PRRT2* has been identified as the most common causative gene of PKD (9) and other paroxysmal disorders. Loss of function of the gene results in a delay of neuronal migration and a marked decrease in synaptic density, which were suggested to be the mechanism underlying *PRRT2*-associated disease (1, 10). Paroxysmal movement attack induced by triggering is a prominent feature of the disorders associated with *PRRT2* mutations. In this study, we reported seven cases with *PRRT2* mutations who presented feeding-induced infantile seizures. Ictal EEGs recorded a seizure with focal origination in the case induced by vigorous sucking. This study suggests feeding, especially sucking, as a potential trigger factor. The seven cases had deletion or truncating mutations that resulted in

haploinsufficiency of the *PRRT2* gene, suggesting a potential genotype-phenotype correlation. To date, 155 *PRRT2* mutations have been identified in patients with episodic disorders (Human Gene Mutation Database, www.hgmd.cf.ac.uk/ac/index.php). Mutations of haploinsufficiency accounted for 60.6%, including truncating, deletion, insertion, and splice-site mutations. The other mutations were missenses. The hot spot mutation c.649dupC/p.Arg217Profs*8 alone even contributed to 73.8% of PKD/IC etiology in a previous study (3). In our early cohort, six of the 18 (33.3%) patients with *PRRT2* variant had IC and infantile no-convulsion seizures during feeding (4). Therefore, feeding potentially triggers movement in infants with *PRRT2* mutations, especially in those with mutations of haploinsufficiency.

Previously, it was not determined whether the infant convulsion in patients with PRRT2 mutations was epileptic seizures in some cases. Marini et al. (11) reported a recorded focal seizure in a child with PRRT2 mutation. The reported case and the case presented in this study provided direct evidence of epileptic seizures in infants with PRRT2 mutations. Therefore, attention should be paid to triggers in infants with convulsions or seizures. The child in this study got seizure-free since the change of feeding behavior, highlighting the significance of feeding behavior in the early prevention of epileptic seizures and the possible secondary brain injury due to frequent seizures. Vigorous sucking behavior, such as feeding in hunger, should be avoided by maneuvers like reducing feeding interval time and extending feeding duration in infants with PRRT2 mutations. This case also suggests sucking-induced seizures as a reminder of PRRT2 mutations and further genetic tests. Although some patients do not have seizures after a year or two, frequent seizures are harmful. It is crucial to identify the trigger and use appropriate antiepileptic drugs, such as carbamazepine, to stop the seizures.

In conclusion, we identified a *de novo* heterozygous *PRRT2* deletion in a patient who presented infantile seizures induced by

sucking. Data reanalysis on our previously reported cases with *PRRT2* mutations showed that 33.3% of patients had infantile convulsions or infantile no-convulsion seizures during feeding. This study suggests sucking as a potential trigger movement in infants with *PRRT2* mutations, highlighting the significance of feeding behavior in the early prevention of epileptic seizures. All seven patients had *PRRT2* mutations of haploinsufficiency, suggesting a potential genotype-phenotype correlation.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found at: NCBI - PRJNA833568.

Ethics statement

All procedures performed were in accordance with the ethical standards of the institutional committee. The present study was approved by the Ethics Committee of the Second Affiliated Hospital of Guangzhou Medical University.

Author contributions

Substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data were made by SL, B-ZG, BL, L-HL, B-ML, and Z-GL. Drafting the article or revising it critically for important intellectual content were made by Y-HY, L-SX, D-TL, X-QT, and R-PW. Final approval of the version to be published was done by all authors.

References

1. Liu YT, Nian FS, Chou WJ, Tai CY, Kwan SY, Chen C, et al. PRRT2 mutations lead to neuronal dysfunction and neurodevelopmental defects. *Oncotarget.* (2016) 7:39184–96. doi: 10.18632/oncotarget.9258

2. Bruno MK, Hallett M, Gwinn-Hardy K, Sorensen B, Considine E, Tucker S, et al. Clinical evaluation of idiopathic paroxysmal kinesigenic dyskinesia: new diagnostic criteria. *Neurology.* (2004) 63:2280–7. doi: 10.1212/01.WNL.0000147298.05983.50

3. Ebrahimi-Fakhari D, Saffari A, Westenberger A, Klein C. The evolving spectrum of PRRT2-associated paroxysmal diseases. *Brain.* (2015) 138(Pt. 12):3476-95. doi: 10.1093/brain/awv317

4. Liu XR, Wu M, He N, Meng H, Wen L, Wang JL, et al. Novel PRRT2 mutations in paroxysmal dyskinesia patients with variant inheritance and phenotypes. *Genes Brain Behav.* (2013) 12:234–40. doi: 10.1111/gbb.12008

5. Zhou P, He N, Zhang JW, Lin ZJ, Wang J, Yan LM, et al. Novel mutations and phenotypes of epilepsy-associated genes in epileptic encephalopathies. *Genes Brain Behav.* (2018) 17:e12456. doi: 10.1111/gbb.12456

6. DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat Genet.* (2011) 43:491–8. doi: 10.1038/ng.806

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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7. Chen ZR, Liu DT, Meng H, Liu L, Bian WJ, Liu XR, et al. Homozygous missense TPP1 mutation associated with mild late infantile neuronal ceroid lipofuscinosis and the genotype-phenotype correlation. *Seizure.* (2019) 69:180–5. doi: 10.1016/j.seizure.2018.08.027

8. Talevich E, Shain AH, Botton T, Bastian BC. CNVkit: genomewide copy number detection and visualization from targeted DNA sequencing. *PLoS Comput Biol.* (2016) 12:e1004873. doi: 10.1371/journal.pcbi. 1004873

9. Chen WJ, Lin Y, Xiong ZQ, Wei W, Ni W, Tan GH, et al. Exome sequencing identifies truncating mutations in PRRT2 that cause paroxysmal kinesigenic dyskinesia. *Nat Genet*. (2011) 43:1252–5. doi: 10.1038/ng.1008

10. Tan G-H, Liu Y-Y, Wang L, Li K, Zhang Z-Q, Li H-F, et al. PRRT2 deficiency induces paroxysmal kinesigenic dyskinesia by regulating synaptic transmission in cerebellum. *Cell Res.* (2017) 28:90–110. doi: 10.1038/cr. 2017.128

11. Marini C, Conti V, Mei D, Battaglia D, Lettori D, Losito E, et al. PRRT2 mutations in familial infantile seizures, paroxysmal dyskinesia, and hemiplegic migraine. *Neurology.* (2012) 79:2109–14. doi: 10.1212/WNL.0b013e31 82752ca2