Magnetic Resonance Insights



REVIEW

OPEN ACCESS

Full open access to this and thousands of other papers at http://www.la-press.com.

Magnetic Resonance in the Detection of Breast Cancers of Different Histological Types

Rebecca M. Mayrhofer¹, Hsiao Piau Ng¹, Thomas C. Putti² and Philip W. Kuchel³

¹Mechanistic Systemsbiology NMR Group, Singapore Bioimaging Consortium, Agency for Science Technology and Research, Singapore. ²Department of Pathology, National University Hospital, Singapore. ³School of Molecular Bioscience, University of Sydney, NSW 2006, Australia. Corresponding author email: philip.kuchel@sydney.edu.au

Abstract: Breast cancer incidence is increasing worldwide. Early detection is critical for long-term patient survival, as is monitoring responses to chemotherapy for management of the disease. Magnetic resonance imaging and spectroscopy (MRI/MRS) has gained in importance in the last decade for the diagnosis and monitoring of breast cancer therapy. The sensitivity of MRI/MRS for anatomical delineation is very high and the consensus is that MRI is more sensitive in detection than x-ray mammography. Advantages of MRS include delivery of biochemical information about tumor metabolism, which can potentially assist in the staging of cancers and monitoring responses to treatment. The roles of MRS and MRI in screening and monitoring responses to treatment of breast cancer are reviewed here. We rationalize how it is that different histological types of breast cancer are differentially detected and characterized by MR methods.

Keywords: ¹H nuclear magnetic resonance spectroscopy, ³¹P nuclear magnetic resonance spectroscopy, breast cancer, diffusion weighted imaging, dynamic contrast enhanced imaging, magnetic resonance imaging

Magnetic Resonance Insights 2013:6 33-49

doi: 10.4137/MRI.S10640

This article is available from http://www.la-press.com.

© the author(s), publisher and licensee Libertas Academica Ltd.

This is an open access article published under the Creative Commons CC-BY-NC 3.0 license.



Introduction

Breast cancer has dramatic emotional and physical impacts on the person afflicted with it, and from a public health perspective it is a disease of accelerating socioeconomic importance. Approximately 10% of all new cancers diagnosed are female breast cancer. This makes detection and treatment of the disease of high priority world-wide. The incidence of breast cancer is increasing in almost all countries although mortality is decreasing, due to earlier detection of tumors and advances in treatment. The map in Appendix 1, Figure 1 shows the regional incidence of female breast cancer.

We aimed to increase our understanding of the success rates in clinical detection of breast cancer using magnetic resonance (MR) methods by inspecting the structural features of the cancer's various types. We surmised that detection of the different types of breast cancer by MR varies because of differences in the organization and structures of the cells and of the interstitial mucin and fibers. There are likely to be systematic differences in MR relaxation times, spectral line broadening, and diffusional mobility of water. Therefore, we begin this explanation of breast cancer MR-detectability with an inspection of the micro-anatomical structures in these cancers.

Photomicrographs obtained at the National University Hospital, Singapore, of the six "archetypal forms" of invasive breast cancers are shown in Figure 1. The cancers can be classified into six histological types: (1) Invasive ductal carcinoma (IDC; Fig. 1A and B); (2) invasive lobular carcinoma (ILC; Fig. 1C); (3) medullary carcinoma (MC; Fig. 1D); (4) tubular carcinoma (TC; Fig. 1E); (5) mucinous carcinoma (MuC; Fig. 1F); and (6) inflammatory breast carcinoma (IBC; Fig. 1G). An image of healthy breast tissue (Fig. 1H) is given for comparison.

IDC is the most common type of all breast carcinomas, accounting for 70%–80%.^{4,5} The cells are often small and structurally similar, or can be large and varying in size and shape (pleomorphic). The tumor masses formed by IDCs are usually multinodular or stellate (star-like).⁴⁻⁶ The non-invasive form of ductal carcinoma is known as 'ductal carcinoma in situ' (DCIS), which is confined within the secretory ducts. ILCs (8%–14%)^{4,5} are the second most common type of breast cancer followed by MCs (7%).⁴ ILC is characterized by small, round cells forming single files and breaking into the connective tissue. They often do

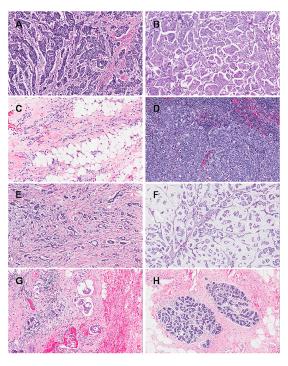


Figure 1. Photomicrographs of various histological types of breast cancers provided by National University Hospital, Singapore. (A) Haematoxylin and eosin (H&E) 40×, invasive ductal carcinoma. Note that the invasive tumor exhibits no features of any special histological type. The tumor is arranged in cords, clusters and trabeculae with scanty tubule formation and several mitoses per high-power image field. (B) H&E 40×, invasive micropapillary carcinoma (a rare variant of invasive ductal carcinoma): the tumor clusters show irregular central spaces, surrounded by artefactually retracted stromal spaces. Some of the clusters have reversed polarity (an "inside out" morphology). (C) H&E 40x, lobular carcinoma: the tumor shows uniform tumor cells mostly arranged in a single file. Some of the tumor cells show intracytoplasmic luminae. (D) H&E 40×, medullary carcinoma: the tumor is composed of a syncytial sheet of pleomorphic cells with no glandular differentiation. The stroma contains prominent lymphoplasmacytic infiltrate. (E) H&E 40x, tubular carcinoma: the tumor is highlighted by irregularly distributed rounded and angulated tubules with open luminae. The tubules are lined by a single layer of epithelial cells and are surrounded by desmoplastic stroma. (F) H&E 40×, mucinous carcinoma: clusters of uniform tumor cells floating in lakes of mucin. (G) H&E 40×, inflammatory breast cancer: characteristic dermal lymphatic invasion by carcinoma associated with surrounding lymphoplasmacytic inflammatory infiltrate. (H) healthy breast tissue: terminal duct-lobular units surrounded by fibro-myxoid stroma. These are lined by an inner epithelial and an outer myoepithelial cell layer. The full time scale is ~10 min.

not form well-defined masses that can be diagnosed easily by physical examination or mammography, making early diagnosis difficult.^{5,7} MCs form a nearly spherical mass of lymphocytes and poorly differentiated epithelial cells.^{6,8} Less common types of breast cancer include tubular (TC), MuC and IBC. The mass of a TC is small is typically stellate; it is characterized by low grade tube-shaped cells.⁹ MuCs are characterized by uniform neoplastic cells arranged in clusters seemingly "floating" in a large amount of mucin, produced by the cells. The mucus and carcinoma cells form a jelly-like tumor. There are two subtypes



of MuCs: mucinous A (MuC-A) and mucinous B (MuC-B). MuC-As are hypocellular while MuC-Bs are hypercellular.¹⁰ Inflammatory breast cancer is aggressive and the cells often block the lymphangioles of the breast, causing macroscopic reddening and swelling of the whole organ, as well as a classical 'peau d'orange' effect on the skin. The prognosis for this aggressive form of cancer is poor.¹¹

MR characteristics of any tissue will depend on the histology, and this will be true for the various specific types of breast cancer. For example, Figure 1A and B are photomicrographs of IDC; the extent of fibrosis, cell density and tumor vasculature varies considerably across the tumor and between tumors, thus leading to a range of patterns on contrast enhanced MR images. 12 Knopp et al¹³ noted a more rapid enhancement and elimination of contrast agent in contrast-enhanced magnetic resonance imaging (MRI) in IDC compared to ILC. This is because IDC tends to have a higher proliferative activity while ILC (Fig. 1C) is typified by diffuse infiltrative growth and does not exhibit extensive neovascularization, resulting in poor MRI enhancement.¹³ MC tends to have high signal intensity on T_2 -weighted images with a heterogeneous distribution of contrast enhancement due to the particular architecture (Fig. 1D) of the tumor. 14 For small TCs (Fig. 1E) on MRI, the presence of spicules in the images is pathognomonic of this type of cancer. The spicules are the stroma that extend out of the tumor and are lined by neoplastic cells. Small lesions have Type I kinetic wash out curves for the contrast agent (steady enhancement) in dynamic contrast enhancement MRI (DCE-MRI) and can show low-density enhancement. However, larger lesions tend to contain hyper-enhancing areas in DCE-MRI images and Type III kinetic curves (indicating washout of signal intensity). This is because of the increased vasculature of the tumor. Figure 2 illustrates the characteristics of the various types of kinetic curves. Figure 1F is a photomicrograph of an MuC; the abundant mucin is evident in the optical image. The mucin has a variable protein concentration and because of its high water content it has dramatically prolonged T_1 values of the water; T_1 -weighted MRI signal intensity is enhanced. High signal intensity in T₂-weighted images, compared to those from other histological types of IDC, is predicted to be a result of the large amount of mucin present. 15,16 For this particular type of cancer, the signal-intensity versus time curves in DCE-MRI are characterized by a Type I kinetic curve

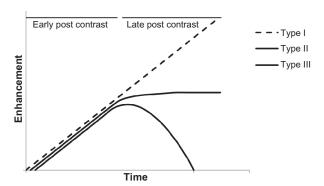


Figure 2. Illustration of the different types of kinetic curves from DCE MRI. **Notes:** Type I is a progressive enhancement pattern that shows a continuous increase in signal intensity over time. The Type II curve has a plateau pattern where there is initial contrast-molecule uptake followed by a plateau phase and the result is suggestive of malignant tumor. The Type III curve is a washout pattern where there is initial rapid uptake followed by reduction in signal enhancement and it is strongly suggestive of malignancy.

that is likely to be from the high extracellular volume fraction in these malignant tumors. ^{16,17} Figure 1G shows the extensive infiltration of inflammatory cells in IBC. This cancer is typically associated with thickening of the overlying skin as can often be seen on MRI images; and this radiological sign constitutes a marker for diagnosing IBC. The abnormal skin also typically has a large water signal. ¹⁸ Diffuse signal enhancement is expected for this type of cancer as the associated inflammation has related edema; this is evident in the photomicrograph as 'separation artifacts' in the tissue slice.

The complexity of ¹H MR spectra varies according to the cancer and the grade of each specific type. Taking the biochemical variation between cancer types with the heterogeneity of the physical-chemical environment of the water in them leads to MR outcomes that are listed in Table 1; this lists predicted relative values of the MR parameters for the different types of breast cancer. Not all relative values are given in the literature, especially for the less common breast cancer types. Values not found in the literature were inferred from histological characteristics such as extent of cellularity, regularity of the arrangement of cells, amount of edema, and variations of fat (adipocyte) content. For example, MuCs have high water content and are therefore expected to have longer T₂ values, as was shown by Santamaria et al. ¹⁹ TCs, however, have little if any edema and are therefore expected to have shorter T_2 values than MuCs. IBC is an aggressive cancer and is expected to have a high concentration of metabolites due to a high density of "aggressive" or "reactive" cells that are involved in



Table 1. Relative values of NMR parameters of different breast cancer types

Type of cancer	T ₁ ¹	T ₂ ²	T ₂ *3	D ⁴	$\Delta \chi^5$	δ metabolites ⁶			References	
						Α	В	С	D	
Invasive ductal carcinoma (IDC)	+	+	+	_	+	+	++	+	+	87 ⁶ , 103 ² , 104 ⁴ ,105 ⁴
Lobular carcinoma (LC)	-	_		_	++	+	++	-	+	19¹, 106⁴
Medullary carcinoma (MC)	+	+	+			++	+++	++	++	107 ² , 50 ⁴
Tubular carcinoma (TC)		_	_		_	_	+	_	_	106⁴
Mucinous carcinoma (MuC)	++	++	+	++	++	_	++	_	+++	19 ² , 50 ⁴ , 106 ⁴
Inflammatory breast carcinoma (IBC)	+	+	+	_	++	++	+++	++	+	108 ¹ , 109 ¹
Healthy breast tissue (non lactating)	_	++	+	+	+				_	87 ⁶ , 110 ⁴ , 65 ⁶

Notes: Predicted relative values of the MR parameters for the different types of breast cancer. Not all relative values are given in the literature; values not found in the literature were inferred from histological characteristics such as extent of cellularity, regularity of the arrangement of cells, amount of edema, and variations of fat (adipocyte) content. The superscripts under the reference column refer to the corresponding superscripts of the MR parameters in the columns, where the Ts denote the MR relaxation times of water, D its diffusion coefficient, χ^s heterogeneity in magnetic susceptibility in the sample, and Δ the chemical shift of the various metabolites. The metabolites, A,B,C,D refer to: A, Phosphate and its esters/phospholipid membrane metabolites: GPCho, GPEth, PCho, PCr, PDE, PEth, PME; B, Glycolytic metabolites: glycogen, lactate; C, redox: GSH; D, Protein synthesis: amino acids.

the inflammatory process than exist in a more slowly growing cancer, such as TC.

In order to "metrify" the digital images in Figure 1 we generated red, blue and green pixel intensity histograms, and the results are given in Figure 3. The color in each pixel of the image is formed by an additive color model consisting of 'red', 'green' and 'blue'. The x-axis ranges from 0 to 256 with 0 denoting pure black and 256 denoting pure red, green or blue. The y-axis indicates the number of times that a particular value of red, green or blue has occurred in the image when traversing from left to right and top to bottom. There were substantial qualitative differences in the histograms of the different types of cancer; this implies that further research into this method could be very rewarding. The analysis of pixel intensity was carried out on the histological images of the various types of breast cancer and characteristic shapes of the color-histograms were very striking. Thus, correlations of the pixel intensity histograms and breast cancer types warrant further study as a potential diagnostic methodology. However, in the present context, the metrification was used to correlate the images with MR analysis of the corresponding cancer types.

In addition to histological metrification, a variety of biomarkers (biochemicals/metabolites) can also be assessed in cancers. The biomarkers provide a means of conferring a more convincing diagnosis, monitoring treatment, and potentially aiding the design of a treatment plan. Table 1 in Appendix 2 describes the various major biomarkers and their clinical uses; we do not discuss these particular biomarkers here. And for reviews of biomarkers and diagnosis and treatments see Weigel et al,²⁰ Dowsett et al,²¹ and Vivanco et al.²²

In screening for breast cancer, x-ray mammography (MMG) has been the technique of choice for several decades. Women at high risk of hereditary breast cancer are recommended to have a semi-annual clinical examination (palpation) and an annual mammogram beginning between the ages of 25–35 years. ^{23,24} Mammography is a diagnostically limited technique as it does not provide sufficiently detailed information, such as vascularity, especially when breast tissue is dense. However, MMG can detect microcalcifications that MRI cannot readily reveal, and the images provide important indicators for cancer.

Over the past 10 years, increased research efforts have helped to establish breast MRI as an important tool for the detection and characterization of breast cancer, especially for naturally dense breast tissue.²⁵ Although MRI has a very high sensitivity for detecting tissue pathology, it suffers from low specificity that also depends on the protocol used (pulse sequences) and the level of experience of the



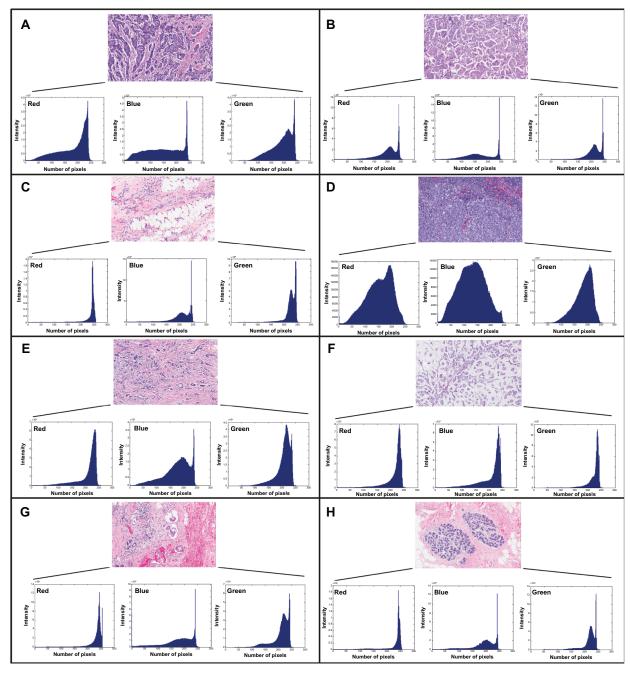


Figure 3. Analyses of the red, blue, and green pixel intensities in the different breast cancer types for the microphotographs provided by National University Hospital, Singapore: (A) invasive ductal carcinoma (IDC). (B) invasive micropapillary carcinoma (a form of IDC). (C) invasive lobular carcinoma (ILC). (D) medullary carcinoma (MC). (E) tubular carcinoma (TC). (F) mucinous carcinoma (MuC). (G) inflammatory breast cancer '(IBC). (H) healthy breast tissue. Note: The unique shapes of the histograms for the different colors and breast cancer types.

technician using it. This results in the need to perform significantly more biopsies of lesions after MRI scanning than for MMG.^{25,26} Nevertheless, Van Goethem et al, in reviewing more than 80 papers concluded that MR mammography has the highest sensitivity to detect IDC (Fig. 1A).²⁵ Thus, MR is an excellent tool for determining tumor extent and is likely to benefit women with dense breast tissue.²⁷

MRI and magnetic resonance spectroscopy (MRS) can be used for early detection of breast lesions and then both MR and MMG can be used to follow responses to treatment. Health-planning and clinical analysis of whether MRI is more beneficial for women at high risk is influenced by a concern with high cost. Figure 2 in Appendix 3 shows the trends of research in the diagnosis of



breast cancer over the past 10 years. The heightened global awareness of breast cancer has increased research efforts for early detection and monitoring breast cancers using many new physical methods such as biophotonics, but the most mature technologies are MRI and MRS.

Detection of Breast Cancer by MRI/MRSDCE-MRI

This is most commonly used for diagnosing and assessing progression of breast lesions and has shown great promise in differentiating between malignant and benign tumors. It can also be used to monitor neovascularization of tumors and this can be further quantified using pharmacokinetic modeling of tracer distributions.^{28–30} In the past, the specificity of DCE-MRI ranged from 37%-91%.31,32 This has not improved significantly in recent years and the specificity continues to range widely due to varying criteria used to differentiate between benign and malignant tumors.33 Lipnick et al34 improved the specificity for diagnosis by combining DCE-MRI and 2-dimensional (2D) MRS. However, the protocol is protracted and more work is required to validate it for breast cancer detection.34 Diffusion weighted (DW)-MRI can also be used to differentiate between malignant and benign lesions.35,36 Response to treatment can be evaluated by studying vascular density or vascular permeability as well as tumor size and water-signal enhancement. The apparent diffusion coefficient (ADC) of tissue water that is associated with changes in tissue and intracellular structure (plasma membrane permeability) can be measured by using DW-MRI.^{29,37-39}

DCE-MRI (introduced above) provides information on changes in vascularity, vascular permeability and the relative volume of the extracellular space. 40,41 It involves the rapid acquisition of images before, during, and after the injection of a contrast agent (CA); such agents increase the contrast between different tissues by altering the relaxation times, T_1 and T_2 . The most common MRI CAs are gadolinium based chelates. To analyze the relaxation data in clinical studies and trials, semi-quantitative measures are commonly used; these are typically the 'maximum change in image intensity' (slope of the wash out curve), and the initial area under the curve (AUC) of contrast agent concentration versus time as it is washed

out of the system.⁴³ These methods reflect the tracer kinetics of the blood exchange rather than the cellphysiological parameters like membrane transport. Another method that is more complicated and provides information on cell-physiological parameters is kinetic modeling. There are several model-based methods as well as model-free algorithms that are used, and several empirical quantification methods involving enhancement patterns that include the two measurements noted above. 44,45 For parametric contrast-enhanced imaging, a mathematical model is used to relate the kinetic changes in signal intensity in the presence of the contrast agent to the values of the kinetic and relaxation parameters that describe the tissue properties. However, the various models provide overall estimates of many interrelated parameters. Empirical-parametric methods are not standardized and vary for different experimental protocols.46 Specifically, Kelcz et al⁴⁶ performed a clinical test of a high-spatial-resolution parametric method for detecting breast lesions using DCE-MRI. They did this to establish standards for diagnostic breast MRI protocols. In their study, 57 women with 45 masses and 23 clusters of micro-calcifications were imaged. Three images were obtained at various times; one unenhanced image was followed by two CE images. Parametric maps were generated and the sensitivity and specificity for the 45 solid lesions were found to be 96% and 82%, respectively. In the kinetic modeling case, it is noted that there was greater variability as not only the mathematical modeling affected the results but also the choice of arterial input rate and volume. Overall, this method provides valuable information on physiological changes in the tumor that the AUC method does not.

Although DCE-MRI has the potential to detect breast cancer it can only be performed on patients without kidney disease or known reactions to contrast agents; these factors limit the number of patients who can undergo the investigation, ²⁹ and it is costly. Nevertheless, it is currently the standard method for high-risk patients as recommended by the American College of Radiology. ⁴⁷

DW-MRI

This is used to detect and differentiate between benign and malignant breast tumors.^{35,36,48,49} As the properties of malignant breast tissues vary between



benign and healthy tumors, the ADC can predict the presence of cancer.³⁷ Images and pulse sequences employing gradients of different durations and amplitudes are acquired along with various diffusion times; thus the ADC can be estimated in vivo. The ADC informs on the characteristics of diffusion within tissues of interest,⁴³ such as the extent to which there is 'restricted' or 'anisotropic' diffusion. Typically, the measured ADC reflects water diffusion in the extracellular space. Short diffusion paths are encountered when extracellular water molecules are hindered by structural interfaces such as in a highly cellular region of tissue. Thus correlations between the ADC values, tumor cellularity and tumor grade have been studied.⁴³

Differences in ADC values can be rationalized on the basis of the relative openness of tissue structures (see Fig. 1). ADC values are expected to be high in mucinous MuC due to the high water content of the extracellular mucin. Hatakenaka et al⁵⁰ studied the ADC of various breast tumors including MuC; significant differences were seen in different types of breast cancer compared with that of fibroadenoma that was slow at 1.7 ± 0.3 (SD) $\times 10^{-3}$ mm² s⁻¹. The mean ADC of MuC was $2.11 \pm 0.18 \times 10^{-3} \text{ mm}^2 \text{ s}^{-1}$. Woodhams et al51 reported a mean ADC value of $1.8 \pm 0.3 \times 10^{-3}$ mm² s⁻¹ for pure MuC while for MC, a lower ADC value is expected due to its higher cellularity and accordingly Hatakenaka et al⁵⁰ report a mean ADC value of $0.94 \pm 0.15 \times 10^{-3}$ mm² s⁻¹. The mean ADC value estimated by Woodhams et al⁵² for LC (among others) was $1.07 \pm 0.26 \times 10^{-3}$ mm² s⁻¹. The lower ADC value might be predicted from the appearances of tissue sections in Figure 1 because of the higher cellularity of this type of breast cancer. The mean ADC value of TC (of 43 carcinomas) reported by Woodhams et al⁵² was relatively low at $1.16 \pm 0.26 \times 10^{-3}$ mm² s⁻¹; although, no ADC values were determined for IBC, they are predicted be higher due to the large amount of interstitial watery edema.

In clinical practice, most DW studies are adjuncts to DCE-MRI. Changes in ADC that indicate proliferative cell activity are often recorded from regions of increased vascularity in DCE-MRI images. A combination of DW and DCE-MRI increases the specificity for detecting breast cancer with MRI. 33,53,54 Yabuuchi et al⁵³ evaluated the benefit of combining DCE-MRI with DW-MRI thus increasing the specificity of MRI

for breast-cancer detection in "non mass like" image enhancements. In their study of 45 lesions, significant indicators of MR malignancy were 'segmental distribution', 'clumped internal enhancement', and ADC values less than 1.3×10^{-3} mm² s⁻¹. In the validation study of 22 non mass-like image 'enhancements', the authors recorded 87% sensitivity and 86% specificity. Kul et al³³ recorded with DCE-MRI alone a specificity of 75.7% (97.9% sensitivity); and combined with DW-MRI a specificity of 89.2% without sacrificing sensitivity (95.7% sensitivity). Partridge et al⁵⁴ combined DW-MRI with DCE-MRI wash-out kinetics with strong indications that their method improved accuracy of making the diagnosis. The authors are in agreement that the combination of DW and DCE-MRI increases the specificity for detection.

In a recent study, Sharma et al⁵⁵ show the potential of using ADC values to identify viable tumor areas for voxel positioning in MRS without DCE-MRI data being available. The authors used ADC values to differentiate between necrotic and viable tumor regions in breast cancer patients. The ADC values of the necrotic regions were significantly higher than the regions of viable tumor. The authors attribute the lower ADC values in viable tumor tissue to the high cellularity of the highly proliferative regions, thus reducing water volume in the extracellular space, resulting in a decrease in the rate of diffusion of water. This study shows promise for using ADC values from MRS measurements without relying on DCE-MRI data.

There are pros and cons for the use of DW and ADC values in characterizing breast lesions. Detection of small tumors is limited because of the low geometrical resolution of MRI. DW-MRI requires echo-planar imaging (EPI) that places high demands on the performance of magnetic field gradient amplifiers, including the uniformity of the field gradients. However, EPI enables short imaging times so DW-MRI images can be obtained rapidly.

MRS

MRS provides biochemical information about tumor metabolism, which can potentially be used along with the tumor nodes metastasis (TNM) staging system (see Appendix 3) and monitoring responses to treatment.^{4,56,57} MRS studies of breast cancer typically use ³¹P and ¹H MRS. With ¹H MRS, total choline



(including free choline, its mono-phosphate ester and phosphodiesters; denoted by tCho) has been the most commonly assessed group of metabolites^{4,25,58–62}; they are readily detected even at low concentrations because of the high intensity signal from the quaternary ammonium -N+(CH₃)₃ of the choline moiety with its nine equivalent ¹H-atoms. The water-to-fat (W-F) ratio is another characteristic that has been explored for cancer diagnosis, but it is less sensitive than using tCho.63 For 31P MRS studies, resonances from the phosphate esters of metabolites are prominent; these are nucleoside phosphates (NTP), inorganic phosphate (Pi), and phosphocreatine (PCr). The activity of lipid metabolism can be inferred from the steady-state concentrations of phosphomonoesters (PMEs) and phosphodiesters (PDEs). The PME peaks are readily assigned to phosphoethanolamine (PEth) and phosphocholine (PCho), while the PDE peaks are assigned to glycerophosphoethanolamine (GPEth), glycerophosphocholine (GPCho). Therefore these metabolites have been used to monitor tumor responses in neoadjuvant chemotherapy. 4,64-66 This has been done by using the tCho signal intensity and the W-F ratio from ¹H MRS, while the phosphorus metabolites of cellular metabolism have been estimated, although less precisely, via ³¹P MRS.

Choline

The ¹H MRS signal-envelope from tCho has a peak maximum at ~3.2 ppm; as noted above the related molecules include choline, PCho, and GPCho.⁵⁹ Some resonances from ethanolamine, PEth and GPEth also contribute to the envelope of signals because the protons of the —CH₂-groups are neighbors of the amine groups at ~3.2 ppm. In addition, resonances from taurine contribute to the tCho envelope in low resolution ¹H spectra.⁶⁷ Higher tCho concentrations are correlated with the size of the tumor and negatively correlated with the age of the patient, since younger patients tend have more aggressive cancers.^{4,60}

The specificity and sensitivity for choline detection has been reported to be 0.92.⁶⁸ Regarding specificity, false negatives have been reported in several studies including those of Chen et al, Jagannathan, and Sardanelli et al.^{59,60,68–71} Failure to detect choline in some breast cancers has been attributed to patient motion, contamination by hemorrhage, and inclusion of lipids interspersed with stromal and

inflammatory cells. Yeung et al⁷² conducted ¹H MRS in contrast-enhanced breast lesions for choline detection and reported a significant choline resonance in IDC. A high resolution magic angle ¹H MR spectrum of IDC is presented in Figure 1A by Cheng et al.⁷³ More work is required to determine the biological factors that affect choline detectability for characterizing breast cancer by ¹H MRS.⁶⁰

Choline-family resonances have been detected in some benign tumors as well as in normal breast tissue; and attempts have been made towards the accurate quantification of choline for differentiating between benign and malignant tumors.⁷⁴ Baik et al⁷⁵ suggest that using the 'fully-relaxed' water resonance as an internal intensity reference increases the specificity of ¹H MRS. Thus, choline concentration is calculated by comparing the choline and water peak integrals and correcting for differences in relaxation times of the two molecular species. A study of breast in vivo using ¹H MRS at 1.5 T by Bolan et al⁷⁴ involved 45 patients with biopsy-confirmed breast cancer. Absolute quantification of choline concentrations was made for the malignant tumors. The relaxation times of water and total choline in malignant breast tumors in four patients were measured and the absolute choline concentrations were estimated to be from 0.76-21.20 mmol kg⁻¹, from 34 spectra. In a recent study, Dorrius et al55 also showed that choline is a good discriminator between benign and malignant breast lesions. Other ¹H MRS studies have used an external reference ('phantom') of known concentration for absolute quantification of choline in vivo. 69,76

Water-fat (W-F) Ratio by ¹H MRS

Sijens et al⁷⁷ were the first to report that the W-F ratio estimated by using a surface coil was higher in breast cancer patients than in healthy volunteers. Normal breast tissue has relatively high fat content while malignant tissue that is mostly derived from glandular tissue has higher water content (see Fig. 1A–H). This was confirmed in single-voxel spectroscopy^{63,78} with similar findings for the W-F ratio. The W-F ratio is calculated by using the peak areas of water at δ = 4.7 ppm and the major lipid peak area at δ = 1.33 ppm.^{63,78} Thomas et al⁷⁹ also report that differences in lipid content can be seen during tumor development and the value depends on the size of the neoplasm. However, the W-F ratio cannot be used to differentiate between



malignant and benign tumors; it does not differ significantly between the two.

PME/PDE by 31P MRS

In malignant breast tissue, the PMEs and PDEs have higher concentrations than in healthy tissue.4 Park et al65 quantified the various phosphorus metabolites, PME, PME/PCr, PDE/PCr, total adenosine triphosphate(tATP)/PCr, and PCr/tATP, in benign, malignant and healthy breast tissue. Their study included three groups: (1) 17 patients with untreated primary malignant breast cancers; (2) 8 with untreated benign breast tumors; and (3) 7 with healthy breasts. Only PME and PME/PCr were significantly different between malignant and healthy tissues, and were not significantly different between the malignant and benign lesions. In hindsight, PCr concentrations were measured in the chest-wall muscle rather than the glandular tissue, resulting in a questionable significance of the PME/PCr ratio.

The tATP signal is lower in healthy tissue than in benign tumors, and other studies have confirmed that ³¹P MRS is capable of differentiating between benign and malignant neoplasms. ^{64,80} Variations in the conclusions from these reports are probably due to instrument-calibration artifacts, variable sensitivity of surface RF coils, as well as imperfect analysis of the phosphorus-containing metabolites due to a low signal-to-noise ratio. ⁸¹ Overall, ³¹P MR with today's clinical MRI scanner technology is still not sufficiently sensitive for routine breast cancer diagnosis.

PCho

PCho is a valuable biomarker in breast cancer diagnosis as 10-fold higher concentrations occur relative to normal breast tissue. Res. The largest contributor to the Res. PMR tCho peak in breast tissue is PCho (free choline that lacks a Res. Patom is not observable) so it is a useful biomarker for breast cancer diagnosis. Normal mammary epithelial cells, primary tumor cell lines, and a metastatic cell line from a single patient have been studied. The PCho concentrations were increased by 16- to 19-fold and 27-fold, respectively, in the primary tumor lines and metastatic cells compared to normal epithelial cells. Eliyahu et al Res. Showed that PCho is 5- to 17-fold higher than normal mammary epithelial cells. The higher concentrations

of PCho in breast cancer cells imply enhanced choline kinase activity in cancer cells as well as probable increased choline transport into the cells.

High resolution magic angle spinning MR has been used to study biopsies of many breast cancers⁸⁵: glycine concentrations tend to be lower in patients who have a good prognosis and are higher in patients with a poor prognosis, as judged clinically. High resolution magic angle spinning MR has been used to investigate IDC.86 Tumor grades correlate with the relative intensities of PCho/Cho and lactate (Lac)/ Cho, with a significant increase in Lac/Cho ratio from Grade II to Grade III IDC. 89 Leibfritz et al87 studied the metabolites present in various breast cancers, including IDC and reported that the metabolic profiles in the NMR spectra vary considerably. In a spectrum from healthy tissue there is a higher content of adipocytes decreasing the number of water-soluble metabolites compared to malignant tissue, thus making the metabolites relatively more abundant. A marked increase in PCho signal in cancerous tissue was observed in all three studies.88-90

Responses to Neoadjuvant Chemotherapy by MRI/MRS

Neoadjuvant chemotherapy is increasingly used with patients who have operable breast cancer and who require complementary therapy.88 Imaging techniques that provide information on the response to treatment are of great importance in this context, in addition to physical examination. The original criteria for assessing solid tumor response to therapy were developed in 1981 by the World Health Organization (WHO).42 The current standard method of responseevaluation in solid tumors (RECIST) uses 1-D measurements with x-rays, ultrasound, x-ray-computed tomography (CT), and/or MRI. The lesions identified on a scan are measured along their greatest dimension and the corresponding perpendicular short axis. For lymph node lesions, the short axis is used as the 'composite tumor burden score' and for 'non-lymph node lesions' the long axis is used; these measurements are then added to give the baseline sum diameters (BSDs). The fractional change in BSDs is used to classify responses to treatment into the following categories: (1) complete response (CR); (2) partial response (PR); (3) progressive disease (PD); and (4) stable disease (SD). Complete response is defined



as the disappearance of the lesion, while partial response is defined as more than a 30% decrease in BSDs. Progressive disease is defined as more than 20% increase in the BSD or the appearance of new lesions. While stable disease is defined as changes that do not fall into one of the above three categories.⁴²

Water-fat ratio by ¹H MRS—therapy responses

The W-F ratio estimated from ¹H MR spectra is high in breast tumors but low in the normal breast.⁷⁷ Jagannathan⁷⁸ and colleagues studied 44 patients with IDC (Fig. 1A). The spectrum of healthy tissue is dominated by resonances from fat while in the malignant breast the water resonance dominates. The W-F ratio was 6.0 ± 6.9 and a post-chemotherapy one of 1.3 ± 1.5 . Tumor regression can be verified with mammography and clinical examination. These findings posited the use of the W-F ratio in assessing outcomes in neoadjuvant chemotherapy, and this was verified in a later study.^{28,63} Although the W-F ratio is prognostic of tumor response, many researchers are now focusing on measurements of choline-containing compounds using water-suppressed ¹H MRS, as choline is a more sensitive marker of cancer aggressiveness (see above). The reluctance to use the W-F ratio for assessing tumor response could be because it depends strongly on which region of normal breast tissue is selected as the control.

Choline ¹H MRS—therapy responses

As concluded above, there is considerable potential for using the tCho signal intensity and tumor volume to assess tumor response in neoadjuvant chemotherapy. Several groups report no tCho or significantly reduced tCho in those patients on chemotherapy who have a "good response"; this was evidenced by clinical and histopathological evaluation. 59,62,63,89,90 In the study by Tozaki, et al, 91 tumor response to chemotherapy after the second cycle was studied in 16 patients (8 pathological responders and 8 non-responders) with breast cancer. The normalized tCho signal ranged from 0.40 to 2.8 and a significant decrease in the signal occurred, with its total disappearance in 6 cases. Overall, it is concluded that the change in tCho signal is more sensitive than changes in tumor volume for detecting a positive therapeutic response. 92 Hence, changes in tCho signal provides a means of assessing

response to neoadjuvant chemotherapy. This is an incredibly encouraging finding and deserves immediate further study.

PME/PDE ³¹P MRS—therapy responses

In human breast tumors, large signals from PME and PDE are present in the ³¹P MRS spectra (see above).⁴ PME and PDE peaks are not only present but they are of much greater intensity in spectra from breast tumors than normal breast tissue. According to Leach et al,4 a decrease in PME correlates with a stable or responding disease, while an increase in PME occurs with progressive disease (as classified by the WHO criteria; see Appendix 2). The PME peak in spectra from lactating breast is increased relative to nonlactating pre-menopausal breast.92 In an early study by Glaholm et al, 93 a significant reduction in PME occurred after chemotherapy in a patient with locally advanced carcinoma of the breast; this suggested that PME concentrations may be used to monitor tumor regression. In a later study by Redmond et al,66 pre- and post-chemotherapy ³¹P MRS signals were recorded from 16 patients with large cancers. Peak areas and ratios of ³¹P metabolite peaks were estimated. A clinically assessed "good response" to chemotherapy was correlated with decreases in PME, PME/PDE, PME/ PCr, PME/NTP, PDE/PCr and tumor pH, as well as increases in the ratios of Pi/PME and Pi/PDE. It was also noted that the ratio Pi/PME shows potential for separating partial responders from complete responders; a higher initial Pi/PME ratio is observed in complete responders compared with partial responders. The authors conclude that ³¹P MRS is potentially useful for monitoring responses to chemotherapy but there is as yet a lack of routine clinical application.

DW-MRI—therapy responses

The mean ADC estimated with DW-MRI can be used as another means of monitoring the response of patients to neoadjuvant chemotherapy (see above);^{37,94,95} it is especially useful in assessing early responses. Pickles et al⁹⁶ showed that changes in ADC precede tumor size reduction after neoadjuvant treatment; changes are seen as early as after the first cycle of chemotherapy. This was confirmed by Sharma et al³⁸: measurements were made of mean ADC values on four occasions on 56 patients with locally advanced breast cancer. ADC values were also measured in 10 benign tumors and



15 normal breasts in volunteers. The fractional change in ADC, as well as the tumor volume and diameter, were measured after each cycle of chemotherapy. Significant increases in ADC values occurred after the first cycle of chemotherapy compared with changes in tumor volume and main diameter. The sensitivity and specificity of ADC to detect responders was 68% and 100%, respectively. The sensitivity to detect responders using tumor volume and diameter were both 89%; and the specificity was 50% for volume, and 70% for diameter. From these results, it is concluded that DW is promising for predicting therapeutic responses in breast cancer patients.

DCE-MRI—therapy responses

DCE-MRI has been used to record physiological and morphological changes during neoadjuvant chemotherapy in breast tumors.⁹⁷ As noted above, Pickles and colleagues98 reported that DCE-MRI differentiates responders from non-responders early in chemotherapy. The kinetic parameters, transfer constant $(k_{\rm trans})$, and rate constant $(k_{\rm ep})$, were estimated by using a two compartment pharmacokinetic model. The transfer constant reflects tumor-blood-vessel flow and permeability, while the rate constant is that of contrast agent efflux from the extracellular extravascular space back into the blood compartment. Although the authors report a significant decrease in the kinetic parameters k_{trans} and k_{ep} for responders compared to non-responders in early treatment, they observed that the fractional change in volume of the breast tumor was the most significant outcome. However, they attributed this to acquiring the early treatment time point too late in the study. Nevertheless, these results confirmed those of their previous study.99

Martincich et al⁹⁹ conducted DCE-MRI studies on 30 patients with breast cancer who were receiving primary chemotherapy. Images were acquired before chemotherapy, after two cycles, and after four cycles of treatment. A reduction in the early enhancement ratio (ECU) after two cycles of chemotherapy correlated with tumor regression; and the reduction of tumor volume was the most significant change.

Ah-See et al¹⁰⁰ applied both DCE-MRI and dynamic susceptibility MRI (DSC-MRI) to assess tumor response to chemotherapy. They concluded that the value of $k_{\rm trans}$ (see above) was the "best" predictor of tumor regression, while changes in tumor size did

not predict histo-pathological changes. Controversy between the value of the peak enhancement and its correlation to treatment response is evident: while some authors suggest that a low initial value predicts good treatment response, ¹⁰¹ others conclude the opposite. ¹⁰² Although DCE-MRI shows considerable potential for monitoring responses to chemotherapy, further work in larger patient populations with appropriate validation of outcomes are needed to guide the establishment of clinical protocols.

Conclusions

Global breast cancer incidence has increased significantly in the past 10 years. Clinical attention has turned to early detection and monitoring early responses to chemotherapy. Both MRI and MRS are used as tools for detecting and monitoring cancer responses to chemotherapy. Future developments in MRI and MRS described above will speed up diagnostic breast cancer imaging and spectroscopy. However, much more work is required to lead to standardized protocols for various MRI methods, especially for detecting the rarer types of breast cancer. With further technical advances including the use of higher magnetic fields, many of these techniques such as MRS will replace current routine ones for detection and monitoring breast cancer. There is also the need to consider the cost-effectiveness of the MR-based methods of imaging relative to the older x-ray and newer PET-based methods. Because this review focuses on MR methods, the comparisons could be of interest to clinical planners but they are not "scientific" per se, so they are given in Appendix 4.

Acknowledgements

This review is part of emerging engagement with the development of diagnostic techniques that use molecular imaging, in the SBIC.

Funding Sources

This review was part of a project funded by an intramural grant in the SBIC.

Competing Interests

Author(s) disclose no potential conflicts of interest.

Author Contributions

Conceived and designed the literature review, histology, and image analysis: RMM, HPN, TCP, PWK.



Analyzed the histological-image data: RMM, HPN. Wrote the first draft of the manuscript: RMM, PWK. Contributed to the writing of the manuscript: RMM, HPN, TCP, PWK. Agree with manuscript results and conclusions: RMM, HPN, TCP, PWK. Jointly developed the structure and arguments for the paper: RMM, PWK. Made critical revisions and approved final version: RMM, PWK. All authors reviewed and approved of the final manuscript.

Disclosures and Ethics

As a requirement of publication author(s) have provided to the publisher signed confirmation of compliance with legal and ethical obligations including but not limited to the following: authorship and contributorship, conflicts of interest, privacy and confidentiality and (where applicable) protection of human and animal research subjects. The authors have read and confirmed their agreement with the ICMJE authorship and conflict of interest criteria. The authors have also confirmed that this article is unique and not under consideration or published in any other publication, and that they have permission from rights holders to reproduce any copyrighted material. Any disclosures are made in this section. The external blind peer reviewers report no conflicts of interest. Provenance: the authors were invited to submit this paper.

References

- Ferlay J, Héry C, Autier P, Sankaranarayanan R. Global burden of breast cancer. Breast Cancer Epidemiol. 2010:1–19.
- Parkin DM, Fernández LM. Use of statistics to assess the global burden of breast cancer. *Breast J.* 2006;12(Suppl 1):S70–80.
- Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int. J. Cancer*. 2010;127:2893–917.
- Leach M, Verrill M, Glaholm J, et al. Measurements of human breast cancer using magnetic resonance spectroscopy: a review of clinical measurements and a report of localized ³¹P measurements of response to treatment. *NMR Biomed*. 1998;11(7):314–40.
- Korkola J, DeVries S, Fridlyand J, et al. Differentiation of lobular versus ductal breast carcinomas by expression microarray analysis. *Cancer Res.* 2003;63(21):7167–75.
- Gallager HS. Pathologic types of breast cancer: their prognoses. *Cancer*. 1984;53(Suppl 3):623–9.
- Chen Y, Hwang E, Roy R, et al. Genetic and phenotypic characteristics of pleomorphic lobular carcinoma in situ of the breast. *Am J Surg Pathol*. 2009;33(11):1683–94.
- 8. Moore OS Jr, Foote FW Jr. The relatively favorable prognosis of medullary carcinoma of the breast. *Cancer.* 1949;2(4):635–42.
- Cooper HS, Patchefsky AS, Krall RA. Tubular carcinoma of the breast. Cancer. 1978;42(5):2334–42.
- Lacroix-Triki M, Suarez PH, MacKay A, et al. Mucinous carcinoma of the breast is genomically distinct from invasive ductal carcinomas of no special type. *J Pathol.* 2010;222(3):282–98.

- Renz D, Baltzer P, Böttcher J, et al. Magnetic resonance imaging of inflammatory breast carcinoma and acute mastitis. A comparative study. *Eur Radiol.* 2008;18(11):2370–80.
- Kitagawa K, Sakuma H, Ishida N, Hirano T, Ishihara A, Takeda K. Contrast-enhanced high-resolution MRI of invasive breast cancer: correlation with histopathologic subtypes. AJR Am J Roentgenol. 2004;183(6): 1805–9.
- Knopp MV, Weiss E, Sinn HP, et al. Pathophysiologic basis of contrast enhancement in breast tumors. J Magn Reson Imaging, 1999;10(3):260–6.
- Nunes LW, Schnall MD, Orel SG, et al. Correlation of lesion appearance and histologic findings for the nodes of a breast MR imaging interpretation model. *Radiographics*. 1999;19(1):79–92.
- Kawashima M, Tamaki Y, Nonaka T, et al. MR imaging of mucinous carcinoma of the breast. AJR Am J Roentgenol. 2002;179(1):179.
- Linda A, Zuiani C, Girometti R, et al. Unusual malignant tumors of the breast: MRI features and pathologic correlation. *Eur J Radiol.* 2010;75(2): 178–84
- Stack JP, Redmond OM, Codd MB, Dervan PA, Ennis JT. Breast disease: tissue characterization with Gd-DTPA enhancement profiles. *Radiology*. 1990;174(2):491–4.
- Renz DM, Baltzer PA, Böttcher J, et al. Inflammatory breast carcinoma in magnetic resonance imaging: a comparison with locally advanced breast cancer. Acad Radiol. 2008;15(2):209–21.
- Santamaría G, Velasco M, Bargalló X, Caparrós X, Farrús B, Luis Fernández P. Radiologic and pathologic findings in breast tumors with high signal intensity on T2-weighted MR images. *Radiographics*. 2010;30(2):533–48.
- Weigel MT, Dowsett M. Current and emerging biomarkers in breast cancer: prognosis and prediction. *Endocr Relat cancer*: 2010;17(4):R245–62.
- Dowsett M, Dunbier AK. Emerging biomarkers and new understanding of traditional markers in personalized therapy for breast cancer. *Clin Cancer Res.* 2008;14(24):8019–26.
- 22. Vivanco M. Biomarkers in breast cancer. *Methods Mol Biol.* 2010;593: 137–56.
- Warner E, Plewes D, Shumak R, et al. Comparison of breast magnetic resonance imaging, mammography, and ultrasound for surveillance of women at high risk for hereditary breast cancer. *J Clin Oncol.* 2001;19(15): 3524–31.
- 24. Warner E, Plewes D, Hill K, et al. Surveillance of BRCA1 and BRCA2 mutation carriers with magnetic resonance imaging, ultrasound, mammography, and clinical breast examination. *JAMA*. 2004;292(11):1317–25.
- Sinha S, Sinha U. Recent advances in breast MRI and MRS. NMR Biomed. 2009;22(1):3–16.
- Orel SG, Schnall MD. MR Imaging of the breast for the detection, diagnosis, and staging of breast cancer. *Radiology*. 2001;220(1):13–30.
- Saslow D, Boetes C, Burke W, et al. American Cancer Society guidelines for breast screening with MRI as an adjunct to mammography. CA Cancer J Clin. 2007;57(2):75–89.
- Manton D, Chaturvedi A, Hubbard A, et al. Neoadjuvant chemotherapy in breast cancer: early response prediction with quantitative MR imaging and spectroscopy. *Br J Cancer*. 2006;94(3):427–35.
- 29. Yabuuchi H, Matsuo Y, Sunami S, et al. Detection of non-palpable breast cancer in asymptomatic women by using unenhanced diffusion-weighted and T2-weighted MR imaging: comparison with mammography and dynamic contrast-enhanced MR imaging. *Eur Radiol*. 2010:11–7.
- Huang W, Tudorica LA, Li X, et al. Discrimination of benign and malignant breast lesions by using shutter-speed dynamic contrast-enhanced MR imaging. *Radiology*. 2011;261(2):394–403.
- Drew PJ, Chatterjee S, Turnbull LW, et al. Dynamic contrast enhanced magnetic resonance imaging of the breast is superior to triple assessment for the pre-operative detection of multifocal breast cancer. *Ann Surg Oncol*. 1999;6(6):599–603.
- Kelcz F, Santyr GE, Cron GO, Mongin SJ. Application of a quantitative model to differentiate benign from malignant breast lesions detected by dynamic, gadolinium enhanced MRI. *J Magn Reson Imaging*. 1996;6(5): 743–52.
- Kul S, Cansu A, Alhan E, Dinc H, Gunes G, Reis A. Contribution of diffusion-weighted imaging to dynamic contrast-enhanced MRI in the characterization of breast tumors. *Am J Roentgenol*. 2011;196(1):210–7.



- Lipnick S, Liu X, Sayre J, Bassett LW, DeBruhl N, Thomas MA. Combined DCE-MRI and single voxel 2D MRS for differentiation between benign and malignant breast lesions. NMR Biomed. 2010;23(8):922–30.
- Guo Y, Cai YQ, Cai ZL, et al. Differentiation of clinically benign and malignant breast lesions using diffusion weighted imaging. *J Magn Reson Imaging*. 2002;16(2):172–8.
- Lo GG, Ai V, Chan JK, et al. Diffusion-weighted magnetic resonance imaging of breast lesions: first experiences at 3 T. J Comput Assisted Tomogr. 2009;33(1):63–9.
- Nilsen L, Fangberget A, Geier O, Olsen DR, Seierstad T. Diffusion-weighted magnetic resonance imaging for pretreatment prediction and monitoring of treatment response of patients with locally advanced breast cancer undergoing neoadjuvant chemotherapy. *Acta Oncol.* 2010;49(3):354–60.
- Sharma U, Danishad KK, Seenu V, Jagannathan NR. Longitudinal study of the assessment by MRI and diffusion weighted imaging of tumor response in patients with locally advanced breast cancer undergoing neoadjuvant chemotherapy. NMR Biomed. 2009;22(1):104–13.
- 39. Tozaki M, Oyama Y, Fukuma E. Preliminary study of early response to neoadjuvant chemotherapy after the first cycle in breast cancer: comparison of ¹H magnetic resonance spectroscopy with diffusion magnetic resonance imaging. *Jpn J Radiol*. 2010;28(2):101–9.
- Padhani AR. Dynamic contrast enhanced MRI in clinical oncology: current status and future directions. J Magn Reson Imaging. 2002;16(4):407–22.
- Hayes C, Padhani AR, Leach MO. Assessing changes in tumour vascular function using dynamic contrast enhanced magnetic resonance imaging. NMR Biomed. 2002;15(2):154–63.
- Lori R. Current and future trends in magnetic resonance imaging assessments of the response of breast tumors to neoadjuvant chemotherapy. *J Oncol.* 2010;2010.
- Marcus C, Ladam-Marcus V, Cucu C, Bouche O, Lucas L, Hoeffel C. Imaging techniques to evaluate the response to treatment in oncology: current standards and perspectives. *Crit Rev Oncol Hematol.* 2009;72(3):217–38.
- Eyal E, Degani H. Model based and model-free parametric analysis of breast dynamic contrast enhanced MRI. NMR Biomed. 2009;22(1):40–53.
- 45. Furman-Haran E, Degani H. Parametric analysis of breast MRI. *J Comput Assist Tomogr.* 2002;26(3):376–86.
- Kelcz F, Furman-Haran E, Grobgeld D, Degani H. Clinical testing of highspatial-resolution parametric contrast-enhanced MR imaging of the breast. *AJR Am J Roentgenol*. 2002;179(6):1485–92.
- 47. Ojeda-Fournier H, Comstock CE. MRI for breast cancer: Current indications. *Indian J Radiol Imaging*. 2009;19(2):161–9.
- Charles-Edwards E, deSouza NM. Diffusion-weighted magnetic resonance imaging and its application to cancer. *Cancer Imaging*. 2006;6:135–43.
- Kuroki Y, Nasu K, Kuroki S, et al. Diffusion-weighted imaging of breast cancer with the sensitivity encoding technique: analysis of the apparent diffusion coefficient value. *Magn Reson Med Sci.* 2004;3(2):79–85.
- Hatakenaka M, Soeda H, Yabuuchi H, et al. Apparent diffusion coefficients of breast tumors: clinical application. *Magn Reson Med Sci.* 2008;7(1): 23–9.
- Woodhams R, Kakita S, Hata H, et al. Diffusion-weighted imaging of mucinous carcinoma of the breast: evaluation of apparent diffusion coefficient and signal intensity in correlation with histologic findings. *Am J Roentgenol*. 2009;193(1):260–6.
- Woodhams R, Matsunaga K, Kan S, et al. ADC mapping of benign and malignant breast tumors. Magn Reson Med Sci. 2005;4(1):35–42.
- Yabuuchi H, Matsuo Y, Kamitani T, et al. Non-mass-like enhancement on contrast-enhanced breast MR imaging: lesion characterization using combination of dynamic contrast-enhanced and diffusion-weighted MR images. *Eur Radiol.* 2010;75(1):e126–32.
- 54. Partridge S, Rahbar H, Murthy R, et al. Improved diagnostic accuracy of breast MRI through combined apparent diffusion coefficients and dynamic contrast-enhanced kinetics. *Magn Reson Med.* 2011;65(6):1759–67.
- 55. Sharma U, Sah RG, Parshad R, Sharma R, Seenu V, Jagannathan NR. Role of apparent diffusion coefficient values for the differentiation of viable and necrotic areas of breast cancer and its potential utility to guide voxel positioning for MRS in the absence of dynamic contrast-enhanced MRI data. Magn Reson Imaging. 2012;30(5):649–55.

- Cheng LL, Burns MA, Taylor JL, et al. Metabolic characterization of human prostate cancer with tissue magnetic resonance spectroscopy. *Cancer Res.* 2005;65(8):3030–4.
- Sitter B, Lundgren S, Bathen TF, Halgunset J, Fjosne HE, Gribbestad IS.
 Comparison of HR MAS MR spectroscopic profiles of breast cancer tissue with clinical parameters. NMR Biomed. 2006;19(1):30–40.
- 58. Daly PF, Cohen JS. Magnetic resonance spectroscopy of tumors and potential in vivo clinical applications: a review. *Cancer Res.* 1989;49(4): 770–9
- Jagannathan N, Kumar M, Seenu V, et al. Evaluation of total choline from in-vivo volume localized proton MR spectroscopy and its response to neoadjuvant chemotherapy in locally advanced breast cancer. *Br J Cancer*: 2001;84(8):1016–22.
- Chen J, Mehta R, Baek H, et al. Clinical characteristics and biomarkers of breast cancer associated with choline concentration measured by 1H MRS. NMR Biomed. 2011;24(3):316–24.
- Lenkinski R, Wang X, Elian M, Goldberg S. Interaction of gadolinium-based MR contrast agents with choline: Implications for MR spectroscopy (MRS) of the breast. *Magn Reson Med.* 2009;61(6): 1286–92.
- Tozaki M, Sakamoto M, Oyama Y, et al. Monitoring of early response to neoadjuvant chemotherapy in breast cancer with (1)H MR spectroscopy: Comparison to sequential 2-[18F]-fluorodeoxyglucose positron emission tomography. J Magn Reson Imaging, 2008;28(2):420–7.
- 63. Kumar M, Jagannathan NR, Seenu V, Dwivedi S, Julka P, Rath G. Monitoring the therapeutic response of locally advanced breast cancer patients: sequential in vivo proton MR spectroscopy study. *J Magn Reson Imaging*. 2006;24(2):325–32.
- Merchant T, Meneses P, Gierke L, Den Otter W, Glonek T. 31P magnetic resonance phospholipid profiles of neoplastic human breast tissues. Br J Cancer. 1991;63(5):693–8.
- Park JM, Park JH. Human in-vivo 31P MR spectroscopy of benign and malignant breast tumors. Korean J Radiol. 2001;2(2):80–6.
- Redmond O, Stack J, O'Connor N, et al. ³¹P MRS as an early prognostic indicator of patient response to chemotherapy. *Magn Reson Med*. 1992;25(1):30-44.
- 67. Li ML, Song Y, Cho N, et al. An HR-MAS MR metabolomics study on breast tissues obtained with core needle biopsy. *PloS one*. 2011;6(10):e25563.
- 68. Sardanelli F, Fausto A, Di Leo G, de Nijs R, Vorbuchner M, Podo F. In vivo proton MR spectroscopy of the breast using the total choline peak integral as a marker of malignancy. *Am J Roentgenol*. 2009;192(6):1608–17.
- Katz-Brull R, Lavin PT, Lenkinski RE. Clinical utility of proton magnetic resonance spectroscopy in characterizing breast lesions. *J Natl Cancer Inst.* 2002;94(16):1197–203.
- Tse GM, Cheung HS, Pang LM, et al. Characterization of lesions of the breast with proton MR spectroscopy: comparison of carcinomas, benign lesions, and phyllodes tumors. *Am J Roentgenol*. 2003;181(5): 1267–72.
- Jacobs MA, Barker PB, Bottomley PA, Bhujwalla Z, Bluemke DA. Proton magnetic resonance spectroscopic imaging of human breast cancer: a preliminary study. *J Magn Reson Imaging*. 2004;19(1):68–75.
- Yeung DK, Cheung HS, Tse GM. Human breast lesions: Characterization with contrast-enhanced in vivo proton MR spectroscopy—initial results. *Radiology*. 2001;220(1):40–6.
- Cheng LL, Chang I. Evaluating human breast ductal carcinomas with highresolution magic-angle spinning proton magnetic resonance spectroscopy. *J Magn Reson.* 1998;135(1):194–202.
- Bolan P, Meisamy S, Baker E, et al. In vivo quantification of choline compounds in the breast with ¹H MR spectroscopy. *Magn Reson Med.* 2003;50(6):1134–43.
- Baik H, Su MY, Yu H, Mehta R, Nalcioglu O. Quantification of cholinecontaining compounds in malignant breast tumors by 1 H MR spectroscopy using water as an internal reference at 1.5 T. MAGMA. 2006;19(2):96–104.
- Bakken I, Gribbestad I, Singstad T, Kvistad K. External standard method for the in vivo quantification of choline-containing compounds in breast tumors by proton MR spectroscopy at 1.5 Tesla. *Magn Reson Med.* 2001;46(1): 189–92



- Sijens P, Wijrdeman H, Moerland M, Bakker C, Vermeulen J, Luyten P. Human breast cancer in vivo: H-1 and P-31 MR spectroscopy at 1.5 T. Radiology. 1988;169(3):615–20.
- Jagannathan NR, Singh M, Govindaraju V, et al. Volume localized in vivo proton MR spectroscopy of breast carcinoma: variation of water-fat ratio in patients receiving chemotherapy. NMR Biomed. 1998;11(8):414–22.
- Thomas M, Lipnick S, Velan S, et al. Investigation of breast cancer using two-dimensional MRS. NMR Biomed. 2009;22(1):77–91.
- Degani H, Horowitz A, Itzchak Y. Breast tumors: evaluation with P-31 MR spectroscopy. *Radiology*. 1986;161(1):53–5.
- Arias-Mendoza F, Payne GS, Zakian KL, et al. In vivo 31P MR spectral patterns and reproducibility in cancer patients studied in a multi institutional trial. NMR Biomed. 2006;19(4):504–12.
- 82. Singer S, Souza K, Thilly W. Pyruvate utilization, phosphocholine and adenosine triphosphate (ATP) are markers of human breast tumor progression: a ³¹P-and ¹³C-nuclear magnetic resonance (NMR) spectroscopy study. *Cancer Res.* 1995;55(22):5140–55.
- Aboagye E, Bhujwalla Z. Malignant transformation alters membrane choline phospholipid metabolism of human mammary epithelial cells. *Cancer Res.* 1999:59(1):80–4.
- Eliyahu G, Kreizman T, Degani H. Phosphocholine as a biomarker of breast cancer: molecular and biochemical studies. *Int J Cancer*. 2007;120(8): 1721–30
- Sitter B, Bathen TF, Singstad TE, et al. Quantification of metabolites in breast cancer patients with different clinical prognosis using HR MAS MR spectroscopy. NMR Biomed. 2010;23(4):424–31.
- Sitter B, Sonnewald U, Spraul M, Fjösne HE, Gribbestad IS. High resolution magic angle spinning MRS of breast cancer tissue. NMR Biomed. 2002;15(5):327–37.
- Beckonert O, Monnerjahn J, Bonk U, Leibfritz D. Visualizing metabolic changes in breast-cancer tissue using ¹H NMR spectroscopy and selforganizing maps. NMR Biomed. 2003;16(1):1–11.
- Fisher B, Bryant J, Wolmark N, et al. Effect of preoperative chemotherapy on the outcome of women with operable breast cancer. *J Clin Oncol*. 1998; 16(8):2672–85
- Danishad K, Sharma U, Sah R, Seenu V, Parshad R, Jagannathan N. Assessment of therapeutic response of locally advanced breast cancer (LABC) patients undergoing neoadjuvant chemotherapy (NACT) monitored using sequential magnetic resonance spectroscopic imaging (MRSI). NMR Biomed. 2010;23(3):233–41.
- Meisamy S, Bolan P, Baker E, et al. Neoadjuvant chemotherapy of locally advanced breast cancer: predicting response with in vivo (1)H MR spectroscopy—a pilot study at 4 T. *Radiology*. 2004;233(2):424–31.
- Tozaki M, Sakamoto M, Oyama Y, Maruyama K, Fukuma E. Predicting pathological response to neoadjuvant chemotherapy in breast cancer with quantitative ¹H MR spectroscopy using the external standard method. *J Magn Reson Imaging*. 2010;31(4):895–902.
- Twelves CJ, Porter DA, Lowry M, et al. Phosphorus-31 metabolism of post-menopausal breast cancer studied in vivo by magnetic resonance spectroscopy. *Br J Cancer*. 1994;69(6):1151–6.
- Glaholm J, Leach M, Collins D, et al. In-vivo ³¹P magnetic resonance spectroscopy for monitoring treatment response in breast cancer. *The Lancet*. 1989;1(8650):1326–7.
- 94. Park SH, Moon WK, Cho N, et al. Diffusion-weighted MR imaging: pretreatment prediction of response to neoadjuvant chemotherapy in patients with breast cancer. *Radiology.* 2010;257(1):56–63.

- 95. Yankeelov T, Lepage M, Chakravarthy A, et al. Integration of quantitative DCE-MRI and ADC mapping to monitor treatment response in human breast cancer: initial results. *Magn Reson Imaging*. 2007;25(1):1–13.
- 96. Pickles M, Gibbs P, Lowry M, Turnbull L. Diffusion changes precede size reduction in neoadjuvant treatment of breast cancer. *Magn Reson Imaging*. 2006;24(7):843–7.
- Türkbey B, Thomasson D, Pang Y, Bernardo M, Choyke P. The role of dynamic contrast-enhanced MRI in cancer diagnosis and treatment. *Diagn Interv Radiol.* 2010;16(3):186–92.
- 98. Pickles M, Lowry M, Manton D, Gibbs P, Turnbull L. Role of dynamic contrast enhanced MRI in monitoring early response of locally advanced breast cancer to neoadjuvant chemotherapy. *Breast Cancer Res Treat.* 2005;91(1):1–10.
- Martincich L, Montemurro F, De Rosa G, et al. Monitoring response to primary chemotherapy in breast cancer using dynamic contrast-enhanced magnetic resonance imaging. *Breast Cancer Res Treat*. 2004;83(1):67–76.
- 100. Ah-See M, Makris A, Taylor N, et al. Early changes in functional dynamic magnetic resonance imaging predict for pathologic response to neoadjuvant chemotherapy in primary breast cancer. Clin Cancer Res. 2008;14(20): 6580–9.
- 101. Hawighorst H, Knopp M, Debus J, et al. Pharmacokinetic MRI for assessment of malignant glioma response to stereotactic radiotherapy: initial results. *J Magn Reson Imaging*. 1998;8(4):783–8.
- Loncaster JA, Carrington BM, Sykes JR, et al. Prediction of radiotherapy outcome using dynamic contrast enhanced MRI of carcinoma of the cervix. *Int J Radiat Oncol Biol Phys.* 2002;54(3):759–67.
- Bradamante S, Barchiesi E, Pilotti S, Borasi G. High resolution 1H NMR spectroscopy in the diagnosis of breast cancer. *Magn Reson Med.* 1988; 8(4):440–9.
- 104. Lima M, Le Bihan D, Okumura R, et al. Apparent diffusion coefficient as an MR imaging biomarker of low-risk ductal carcinoma in situ: a pilot study. *Radiology*. 2011;260(2):364–72.
- Razek AA, Gaballa G, Denewer A, Nada N. Invasive ductal carcinoma: correlation of apparent diffusion coefficient value with pathological prognostic factors. NMR Biomed. 2010;23(6):619–23.
- 106. Woodhams R, Matsunaga K, Iwabuchi K, et al. Diffusion-weighted imaging of malignant breast tumors: the usefulness of apparent diffusion coefficient (ADC) value and ADC map for the detection of malignant breast tumors and evaluation of cancer extension. *J Comput Assist Tomogr*: 2005;29(5):644–9.
- Uematsu T, Kasami M. MR imaging findings of benign and malignant circumscribed breast masses: part 1. Solid circumscribed masses. *Jpn J Radiol.* 2009;27(10):395–404.
- Girardi V, Carbognin G, Camera L, et al. Inflammatory breast carcinoma and locally advanced breast carcinoma: characterisation with MR imaging. *Radiol Med.* 2011;116(1):1–13.
- Carbognin G, Calciolari C, Girardi V, Camera L, Pollini G, Pozzi Mucelli R. Inflammatory breast cancer: MR imaging findings. *Radiol Med.* 2010; 115(1):70–82.
- 110. Yoshikawa M, Ohsumi S, Sugata S, et al. Comparison of breast cancer detection by diffusion-weighted magnetic resonance imaging and mammography. *Radiat Med.* 2007;25(5):218–23.



Appendices

Appendix 1: Incidence of Breast Cancer World-Wide

Regional incidence of female breast cancer

For all regions, breast cancer is one of the top three most common cancers. Northern and western Europe, northern America, Australia, New Zealand and the southern countries of South America have the highest incidence, while Africa, Asia and most of Central America have the lowest incidences.¹

The incidence of breast cancer correlates with a variety of factors such as decreased rate of childbearing and breast-feeding, late age at first birth, late maternal age, level and duration of hormone exposure (such as contraceptives), detrimental dietary and lifestyle changes including obesity and low physical activity, as well as genetic predisposition.²⁻⁴

Appendix 2: Diagnostics

Disease staging commonly uses the globally recognized tumour-nodes-metastasis (TNM) system.⁵ This system consists of four clinical stages (I–IV) and is based on features that are primarily the size and extension of the tumour (T), regional lymph nodes involved (N), and the presence of metastases (M), to classify the extent of a cancer. Table A2.1 describes the four stages in more detail.⁵

Appendix 3: Publication Trends in Breast Cancer Research

The number of paper was estimated from the search engines Scopus and Web of Science using the key words as shown in the legend of Figure A3.2. The largest number of papers on the topic in each year contained the keywords "magnetic resonance imaging and breast cancer". This number nearly quadrupled over 10 years, from 205 articles in year 2000 to 795 articles in year 2010. Interest in MRS has also increased within the past 10 years; the number of papers increased from 27 to 142 from year 2000 to year 2010. From Figure A3.2, it can be seen that there has been an increase in the number of papers published every year for each of the keyword combinations described in the legend.

Table A2.1. Breast cancer biomarkers and diagnostic utility.

Biomarkers	Utility	References
Estrogen receptor (ER) progesterone receptor (PR), and HER2/neu	Indicates sensitivity to endocrine therapy	(13)
Glycosyltransferase enzymes and antigen CA15–3	Means of monitoring treatment	(14), (15)
Thymidine labeling index, BUdR labeling, KI-67 labeling index, percentage of cells in S-phase and ploidy	Means of assessing proliferation rate	(16), (17), (18), (19), (20)
TNF α , EGF, VEGF, IGF and estrogen receptor TNF β	A proxy for stimulated cell growth Inhibits cell growth	(21), (22), (23), (24), (25) (26)
Oncogene expression such as c-myc, c-Ha-ras, erbB2 and Int-2	Affect tumour growth and the regulation and activity of receptors and signaling pathways	(27), (28), (29)

Appendix 4: Cost Effectiveness of MRI versus Mammography in High Risk Population

According to the national comprehensive cancer network (NCCN) clinical practice guidelines in the United States, women at an increased risk of breast cancer are those with: (1) prior thoracic radiation therapy; (2) 5-year risk of invasive breast cancer $\geq 1.7\%$ in women who are over 35 years of age; (3) women who have a lifetime risk of \geq 20% as defined by models that are greatly dependent on family history; (4) a strong

Table A2.1A. Breast cancer staging using the TNM system.

Stage	Т	N	M
0	Tis	N0	MO
IA	T1, T1mi	N0	MO
IB	T0,̈T1,T1mi	N1 mi	MO
IIA	T0,T1,T1mi,T2	N1, N0	MO
IIB	T2,T3	N1,N0	MO
IIIA	T0, T1,T1mi,T2,T3	N2,N1,N2	MO
IIIB	T4	N0,N1,N2	MO
IIIC	Any T	N3 [°]	MO
IV	Any T	Any N	M1

Adapted from Sobin et al.5



Table A2.1B. Definitions of the TNM system.

Tis	carcinoma <i>in situ</i>
T1	≤2 cm
T1mi	≤0.1 cm
T1a	≥0.1 cm to 0.5 cm
T1b	≥0.5 cm to 1.0 cm
T1c	>1.0 cm to 2.0 cm
T2	>2 cm to 5 cm
T3	>5 cm
T4	Chest wall or skin ulceration, skin nodules, inflammatory >4 mm, no ulceration
T4a	Chest wall
T4b	Skin ulceration, satellite skin nodules,
	skin oedema
T4c	T4a and T4b combined
T4d	Inflammatory carcinoma
N0	no regional lymph node metastasis
N1	1 node
(movable	
axillary)	
N1mi	Microscmetastasis >0.2 mm to 2 mm
N1a	1-2 axillary nodes
N1b	Internal mammary nodes with microscopic/ macroscopic metastasis by sentinel node biopsy, not clinically detected
N1c	1–3 axillary nodes and internal mammary nodes with microscopic/macroscopic metastasis by sentinel node biopsy, not clinically detected
N2a, pN2a	2–3 nodes macroscopic 2–3 nodes
(fixed axillary)	microscopic, 4-9 axillary nodes
N2b,pN2b	Internal mammary clinically apparent, internal mammary clinically apparent without axillary nodes
N3a, pN3a	Infraclavicular, ≥10 axillary nodes or infraclavicular
N3b, pN3b	Internal mammary and axillary, internal mammary nodes, clinically detected, with does >3 axillary nodes and internal axillary mammary nodes with microscopic metastasis by sentinel node biopsy but not clinically detected
N3c, pN3c M0 M1	Supraclavicular No distant metastasis Metastasis beyond regional lymph nodes

family history, or predisposition; (5) lobular carcinoma, or atypical hyperplasia; (6) and prior history of breast cancer. Women with a strong family history of breast cancer are those with BRCA1 or BRCA2 mutations, or a personal family history of breast cancer and one of several other familial risk categories. These categories include: (1) being diagnosed before the age of 40; (2) being diagnosed before the age of 50 y with one or more close blood relatives with breast cancer;

and (3) a close family member meeting any of the above criteria.⁷

The aim to detect cancer at its earliest stages for a good clinical outcome needs to be balanced by the ability of communities to provide breast cancer screening technology.8 Although MR is an excellent tool for early detection of breast cancer, breast MRI screening is at least 10 times more expensive than x-ray MMG screening, and it generates higher associated diagnostic costs. Plevritis et al⁹ developed a continuous time Monte Carlo model to estimate the health and economic outcomes of BRCA1 and BRCA2 mutation carriers, incorporating the effects of MMG and breast MRI. From their simulations, the authors conclude that breast MRI screening is more cost effective for BRCA1 than BRCA2 mutation carriers. They also conclude that the cost effectiveness of combining MRI with MMG is highly dependent on the age of the patient. Moore et al¹⁰ argue that breast MRI does not appear to be a cost effective screening option for high-risk women. Using a Markov decision model to compare annual breast cancer screening with either breast MRI or MMG over a period of 25 years in high risk women, they determined that it does not appear to be cost effective at a willingness-to-paythreshold of \$USD50,000/QALY (quality-adjustedlife-years). Others conclude that although MRI is more sensitive diagnostically than MMG in a high risk population, it has not been shown to reduce mortality and it leads to increased diagnostic costs due to its lower specificity for detecting cancer. 11,12 Although current screening guidelines recommend consideration of MRI screening in high-risk women, the use of MR remains controversial.

References

- Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer. 2010.
- 2. Parkin D, Fernández L. Use of statistics to assess the global burden of breast cancer. *Breast J.* 2006;12(s1):S70–80.
- Leach M, Verrill M, Glaholm J, et al. Measurements of human breast cancer using magnetic resonance spectroscopy: a review of clinical measurements and a report of localized 31P measurements of response to treatment. NMR Biomed. 1998;11(7):314–40.
- Mettlin C. Global breast cancer mortality statistics. A Cancer Journal for Clinicians. 1999;49(3):138–44.
- 5. Sobin LH. TNM classification of malignant tumours: Wiley-Blackwell; 2009.
- Bevers T, Anderson B, Bonaccio E, et al. Breast cancer screening and diagnosis. *Journal of the National Comprehensive Cancer Network*. 2009;7(10):1060.
- Network NCC. Practice guidelines in oncology—genetic/familial high-risk assessment: breast and ovarian. National Comprehensive Cancer Network, Inc.; 2005:I.

48



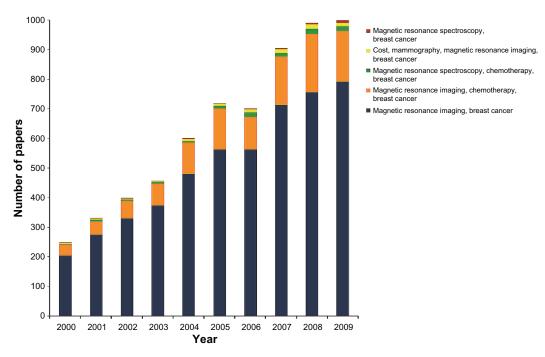


Figure A3.2. Number of papers published per year on breast cancer diagnosis with MRI/MRS. **Note:** The search engine Scopus was used with the keywords as indicated in the figure.

- Sarvazyan A, Egorov V, Son J, Kaufman C. Cost-effective screening for breast cancer worldwide: current state and future directions. *Breast Cancer: Basic and Clinical Research*. 2008;1:91.
- Plevritis S, Kurian A, Sigal B, et al. Cost-effectiveness of screening BRCA1/2 mutation carriers with breast magnetic resonance imaging. The Journal of the American Medical Association. 2006;295(20):2374.
- Moore S, Shenoy P, Fanucchi L, Tumeh J, Flowers C. Cost-effectiveness of MRI compared to mammography for breast cancer screening in a high risk population. *BMC Health Services Research*. 2009;9(1):9.
- Kriege M, Brekelmans C, Boetes C, Besnard P, et al. Efficacy of MRI and mammography for breast-cancer screening in women with a familial or genetic predisposition. New Engl J Med. 2004;351(5):427.
- 12. Elmore J, Armstrong K, Lehman C, Fletcher S. Screening for breast cancer. The Journal of the American Medical Association. 2005;293(10):1245.
- Gruvberger S, Ringnér M, Chen Y, et al. Estrogen receptor status in breast cancer is associated with remarkably distinct gene expression patterns. *Cancer Res.* 2001;61(16):5979.
- Brockhausen I, Yang J, Burchell J, Whitehouse C, Taylor-Papadimitriou J. Mechanisms underlying aberrant glycosylation of MUC1 mucin in breast cancer cells. Eur J Biochem. 1995;233(2):607–17.
- Safi F, Kohler I, Beger H, Röttinger E. The value of the tumor marker CA 15–3 in diagnosing and monitoring breast cancer. A comparative study with carcinoembryonic antigen. *Cancer*. 1991;68(3):574–82.
- Meyer J, Province M. Proliferative index of breast carcinoma by thymidine labeling: prognostic power independent of stage, estrogen and progesterone receptors. *Breast Cancer Res Treat*. 1988;12(2):191–204.
- Tubiana M, Pejovic M, Koscielny S, Chavaudra N, Malaise E. Growth rate, kinetics of tumor cell proliferation and long term outcome in human breast cancer. *Int J Cancer*. 1989;44(1):17–22.
- 18. Crispino S, Brenna A, Colombo D, et al. Ki-67 labeling index in breast cancer. *Tumori*. 1989;75(6):557.
- Hedley D, Rugg C, Gelber R. Association of DNA index and S-phase fraction with prognosis of nodes positive early breast cancer. *Cancer Res*. 1987;47(17):4729.

- Elston C, Ellis I. Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term follow-up. *Histopathology*. 1991;19(5):403–10.
- Leek R, Landers R, Fox S, Ng F, Harris A, Lewis C. Association of tumour necrosis factor alpha and its receptors with thymidine phosphorylase expression in invasive breast carcinoma. *Br J Cancer*. 1998;77(12): 2246.
- Sainsbury J, Sherbet G, Farndon J, Harris A. Epidermal-growth-factor receptors and oestrogen receptors in human breast cancer. *The Lancet*. 1985;325(8425):364–6.
- 23. Price D, Miralem T, Jiang S, Steinberg R, Avraham H. Role of vascular endothelial growth factor in the stimulation of cellular invasion and signaling of breast cancer cells. Cell Growth & Differentiation: The Molecular Biology Journal of the American Association for Cancer Research. 2001;12(3):129.
- Sachdev D, Yee D. The IGF system and breast cancer. Endocrine-Related Cancer. 2001;8(3):197.
- Weigel R. Estrogen and breast cancer. Annals of Surgery. 1997;225(1):
 122.
- Lentz M. The role of therapeutic apheresis in the treatment of cancer: a review. Therapeutic Apheresis. 1999;3(1):40–9.
- Dubik D, Dembinski T, Shiu R. Stimulation of c-myc oncogene expression associated with estrogen-induced proliferation of human breast cancer cells. *Cancer Res.* 1987;47(24 Part 1):6517.
- Garcia I, Dietrich P, Aapro M, Vauthier G, Vadas L, Engel E. Genetic alterations of c-myc, c-erbB-2, and c-Ha-ras protooncogenes and clinical associations in human breast carcinomas. *Cancer Res.* 1989;49(23):6675.
- Sato T, Tanigami A, Yamakawa K, et al. Allelotype of breast cancer: cumulative allele losses promote tumor progression in primary breast cancer. *Cancer Res.* 1990;50(22):7184.