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RESEARCH ARTICLE

Methacholine-Induced Variations in Airway Volume and the Slope of the Alveolar Capnogram Are Distinctly Associated with Airflow Limitation and Airway Closure

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Abstract

Mechanisms driving alteration of lung function in response to inhalation of a methacholine aerosol are incompletely understood. To explore to what extent large and small airways contribute to airflow limitation and airway closure in this context, volumetric capnography was performed before (n = 93) and after (n = 78) methacholine provocation in subjects with an intermediate clinical probability of asthma. Anatomical dead space (VDaw), reflecting large airway volume, and the slope of the alveolar capnogram (slope3), an index of ventilation heterogeneity linked to small airway dysfunction, were determined. At baseline, VDaw was positively correlated with lung volumes, FEV₁ and peak expiratory flow, while slope3 was not correlated with any lung function index. Variations in VDaw and slope3 following methacholine stimulation were correlated to a small degree ($R^2 = -0.20$). Multivariate regression analysis identified independent associations between variation in FEV1 and variations in both VDaw (Standardized Coefficient-SC = 0.66) and Slope3 (SC = 0.35). By contrast, variation in FVC was strongly associated with variations in VDaw (SC = 0.8) but not Slope3. Thus, alterations in the geometry and/or function of large and small airways were weakly correlated and contributed distinctly to airflow limitation. While both large and small airways contributed to airflow limitation as assessed by FEV1, airway closure as assessed by FVC reduction mostly involved the large airways.

Introduction

Non-specific bronchial hyperresponsiveness (BHR) is defined by an exaggerated decrease in forced expiratory flows in response to inhalation of agents inducing contraction of airway smooth muscle. Airway smooth muscle is present along the whole length of the bronchial tree [1]. The muscarinic agonist methacholine is the most commonly used such agent. BHR is considered a feature of bronchial asthma, although it is not specific to asthma since the prevalence of BHR is approximately twice that of asthma in the general population [2][3]. Better understanding of the factors underlying methacholine-induced airflow reductions may lead to refinements in bronchial challenge testing.

Because methacholine drives smooth muscle contraction in vitro and induces reductions in airway caliber that are detectable in vivo [4], and because subjects with BHR have an increased variability of peak expiratory flow rate as compared to subjects without BHR suggesting increased ease of airways to constrict [5], it is recognized that excessive methacholine-induced airway smooth muscle constriction and subsequent changes in airway geometry play key roles. Several lines of evidence, however, suggest that airway responses to methacholine are complex.

Earlier studies consistently show poor correlations between reductions in forced expiratory flows induced by methacholine inhalation and changes in airway resistance as measured during tidal ventilation, regardless of whether resistance was measured by plethysmography [6], forced oscillations [7][8], or the interrupter technique [9]. This discrepancy stresses that decreases in the forced expiratory volume in 1 second (FEV_1) can be related to either reduction in the calibre of large and small airways, thus increasing airway resistance, or to focal airway closure which may decrease mobilizable lung volumes without inducing detectable changes in airway resistance, or to changes in elastic lung recoil. Airway imaging studies show either absent or poor correlations between changes in FEV₁ and changes in airway calibre assessed by computed tomography (CT) scanning [10], while some studies demonstrate focally distributed paradoxical methacholine-induced bronchodilation [11], the latter finding supported by increases in Bohr's anatomical dead space in asthmatics following methacholine provocation [12]. Measurements of ventilation distribution using ventilation scintigraphy and SPECT imaging [13], positron emission tomography (PET) imaging [14], Xe-enhanced CT scanning [15] and ³Heenhanced MRI [16] suggest that focal airway closure is a key feature of methacholine-evoked responses. Finally, computed tomography measurements of air trapping [17] or oscillometryderived indices of small airway function [18][19][20] show that dysfunction of small airways is a feature of methacholine-induced responses in asthmatics.

Overall, available evidence suggests that methacholine-induced airflow reduction is associated with alterations in the geometry and function of both large and small airways. It remains to define, however, to what extent constriction of large and small airways contribute to airflow limitation and airway closure. To address this question, we used volumetric capnography to assess the strength of relationships between, on the one hand, variations in Fletcher's dead space, an indicator of large airway volume [21][22], and variations in the slope of the alveolar capnogram, an index associated with small airway dysfunction [23][24], and on the other hand, with variations in FEV₁ assessing airflow reduction and variations in forced vital capacity (FVC) assessing airway closure.

The slope of the alveolar capnogram reflects ventilation/perfusion mismatching in the distal lung due to regional variations in ventilatory time constants or perfusion. Because of the lack of effects of methacholine on the vasculature of the respiratory system [25][26], we assumed that the probability that alterations in lung perfusion determine variations in the slope of the alveolar capnogram in subjects with mild symptoms and without previous respiratory disease would be very low. Because slope3 is associated with the extent of small airway disease in

resected lungs [23][24], as well as with disease severity in conditions where alterations of small airways play key roles, such as COPD [27] or cystic fibrosis [28], and because Slope3 is associated with airway resistance in humans [29], this index has been proposed as an indicator of small airway dysfunction.

Materials and Methods

Study design

Subjects eligible for enrollment into the study were patients over 18 years of age referred for bronchial provocation testing to the lung function department of a university hospital in Paris, France, because of suspected asthma as appreciated by the referring physician. Patients were included if they had no history of previous chronic respiratory disease, had not taken inhaled or oral steroids for three weeks prior to the visit, and had normal lung function at rest. Internal review board (IRB) approval was obtained (CEPRO, ethics committee of the French Language Pulmonology Society, #2013-011). In accordance with French law with regard to research into common clinical practice (Law #2004-806), subjects gave oral informed consent and were handed an explanatory leaflet, and their participation to the study was recorded in their clinical file. Participant consent was also recorded by the investigator in the main study folder. IRB approval was obtained for the consent procedure. After baseline forced spirometry, patients with normal lung volumes and FEV_1/FVC ratio were invited to participate in the study. Plethysmography was performed before capnography and methacholine provocation testing. Volumetric capnography was performed before provocation testing and immediately after the last spirometry measures, less than 2 minutes after the last dose of methacholine, before albuterol was given. The presence of chronic cough (cough present over 3 or more consecutive months), exertional dyspnea (mMRC scale 1 or more), paroxysmal dyspnea or wheezing was recorded.

Lung function testing at baseline

Forced spirometry and plethysmographic measurement of lung volumes and airway resistances were performed on the Jaeger Masterscreen system (Carefusion, Rolle, Switzerland), according to European Respiratory Society guidelines [30]. Lung function was considered normal if FEV_1 and FEV_1/VC were greater than the lower limit of normal as described by the mean predicted value– 1.64 standard deviation according to the ECCS1993 reference equations for European populations [31].

Methacholine provocation testing

Provocation testing was performed using the automated APS tidal ventilation aerosolization system (Jaeger) and methacholine chloride in phosphate buffered saline (Assistance Publique-Hôpitaux de Paris, France). The initial dose was 100 µg (0.625 µmol), followed by doses of 0.1mg, 0.3mg (1.250020µmol), 0.4mg (2.5 µmol) and 0.8mg (5µmol). Forced spirometry was performed 90 sec after each dose. Increasing doses of methacholine were administered until FEV₁ fell by 20% in comparison to the baseline value, or until a total dose of 1.6 mg (10 µmol) was delivered. BHR was defined as a FEV₁ decrease \geq 20% in response to a maximal methacholine dose of 1.6 mg [32]. Airway reactivity was quantitated in all subjects by the FEV₁-dose-response slope (DRS-FEV₁) and the FVC-dose-response slope (DRS-FVC), defined by the respective reduction in FEV₁ and FVC expressed as a percentage divided by the methacholine dose. The dose inducing a 20% fall in FEV1 (PD₂₀FEV₁) was calculated by linear interpolation in subjects with BHR.

Volumetric capnography

Volumetric capnography was performed during exhalations from total lung capacity (TLC) using the FE141 differential pressure spirometer and the ML-206 gas analyser, both from AD Instruments (Oxford, UK). Signals were synchronized and digitized at a sampling frequency of 500 Hz using LabChart software v7.3.1 (AD Instruments). Patients were connected to the capnography apparatus through a non-resistive filter (PF30, Pall, Fribourg, Switzerland). Every five tidal breaths, patients were instructed to inhale to TLC, and to immediately exhale passively. Subjects were asked to maintain their expiratory flow at the level observed during tidal breathing by visual feedback of flow measurements. Expiratory volume and the expired fraction of CO₂ (FeCO₂) were exported to a spreadsheet and airway volume (VDaw) was calculated according to Fletcher's method for determination of anatomical dead space using equal area determination [33]. To assess ventilation heterogeneity related to small airway dysfunction [23][24][27][28], the slope of the alveolar (third) phase of the volumetric capnogram (slope3) was determined [33]. Five consecutive exhalations from TLC were acquired. Because distribution of VDaw and slope3 values was not normal in all subjects, the median airway volume and phase 3 slope values were retained for analysis. Changes in airway volume and ventilation heterogeneity were expressed relative to the total methacholine dose in mg (Δ VDaw/Mch and Δ Slope3/Mch). In addition, airway volume and slope 3 were measured during the first five tidal breaths taken before the first inspiration to TLC, and their changes relative to the methacholine dose were calculated as described above ($\Delta VDaw_{tidal}/Mch$ and $\Delta Slope3_{tidal}/Mch$).

Statistical analysis

Data were expressed as mean +/- standard deviation, with the exception of PD₂₀ which was expressed as mean and range because it is log-distributed. Relationship between baseline capnographic indices and baseline lung function parameters were determined by linear regression. To assess the association between capnographic indices with airflow reduction, we explored relationships between the FEV1 and FVC dose-response slopes (dependent variables) and both the baseline values in VDaw and slope3 and the fractional changes induced by methacholine in these indices (Δ VDaw/Mch and Δ Slope3/Mch) expressed as %/mg methacholine (independent variables). Univariate linear regression was performed, followed by multiple regression testing variables showing a significant association by univariate analysis. Differences between subjects without and with BHR were determined by student's t-test for continuous variables and the Chi-square test for categorical variables. All statistical analyses were performed with Statview 5.0 software (SAS institute, Cary, North Carolina, USA). A p-value <0.05 was considered significant. Because multivariate analysis with 2 variables was scheduled, we aimed to recruit at least 20 patients with BHR. Because the expected prevalence of BHR is close to 25% among patients referred for provocation testing at our center, we thus planned to recruit at least 80 subjects. Original data are available as an Excel file in the online supplement (S1 Original Data).

Results

Baseline characteristics of subjects

A total of 93 subjects were included into the study between June 3^{rd} , 2013 and October 8^{th} , 2014 (<u>Table 1</u>). Of these, 25 (27%) had BHR defined by $PD_{20}FEV_1 \le 1.6mg$, with a mean PD_{20} of 0.69 mg (range: 0.09–1.52). Patients with BHR showed signs of airflow limitation at baseline in comparison with those without BHR, as indicated by lower FEV_1 , FEF_{25-75} , FEF_{25-75}/FVC , FEV_1/FVC , and higher airway resistance. By contrast, neither airway volume nor slope3 were

Table 1. Characteristics of subjects before methacholine testing.FEV1: Forced expiratory volume in 1second. FVC: Forced vital capacity. FEF25-75: Mean forced expiratory flow between 25% and 75% of FVC.PEF: Peak expiratory flow. TLC: Total lung capacity. RV: Residual volume. Raw: Airway resistance. sRaw:Specific airway resistance. VDaw: Airway volume. Slope3: Slope of the alveolar capnogram.

	No BHR (n = 68)	BHR (n = 25)	All (n = 93)
Gender: male	42%	32%	40%
Age (years)	41 +/- 14	45 +/- 15	42 +/- 14
Height (cm)	169 +/- 8	166 +/- 9	168 +/- 9
Body Mass Index	24 +/- 4	27 +/- 6*	25 +/- 6
Tobacco (Pack-years)	7 +/- 14	4 +/- 10	6 +/- 13
FEV1 (% pred)	100 +/- 14	92 +/- 16*	97 +/- 15
FVC (% pred)	100 +/- 18	103 +/- 19	101 +/- 18
FEV1/FVC	0.82 +/- 0.08	0.78 +/- 0.13	0.81 +/- 0.1
FEF25-75 (% pred)	85 +/- 24	61 +/- 18 ***	79 +/- 25
FEF25-75/FVC	0.86 +/-0.27	0.63 +/-0.24 **	0.80 +/- 0.28
PEF (% pred)	104 +/- 17	91 +/- 24	101 +/- 18
TLC (% pred)	102 +/- 14	93 +/- 31	108 +/- 14
RV (% pred)	100 +/- 26	102 +/- 23	101 +/- 25
RV/TLC (%)	31 +/- 8	37 +/- 17	32 +/- 12
Raw (kPa.L ⁻¹ .s ⁻¹)	0.24 +/- 0.1	0.36 +/- 0.27*	0.27 +/- 0.15
VDaw (mL)	209 +/- 69	185 +/- 69	202 +/- 69
VDaw/TLC (mL/L)	35 +/- 11	33 +/- 8	34 +/- 10
Slope3 (%CO ₂ /L)	0.63 +/- 0.27	0.70 +/- 0.29	0.65 +/- 0.28
Cough	65%	60%	63%
Wheezing	28%	40%	32%
Paroxysmal dyspnea	43%	60%	48%
Exertional dyspnea	37%	40%	38%

*: p < 0.05,

**: p<0.001,

***: p<0.0001 between subjects with and without BHR.

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different at baseline. Symptoms were not different in subjects with BHR in comparison with subjects without BHR, consistent with an intermediate pre-test probability of asthma for all patients.

VDaw, but not Slope3, was associated with lung volumes

Associations between baseline capnographic indices and baseline lung function were explored by linear regression. As shown in <u>Table 2</u>, VDaw was positively associated with TLC, FVC, FEV_1 , the peak expiratory flow rate (PEF) and FEF25-75, consistent with the expected correlation between VDaw and lung size, but neither with residual volume (RV) nor the maximum midexpiratory flow (FEF₂₅₋₇₅), nor FEV1/FVC. By contrast, no significant association was observed between slope3 and any lung function index.

Decreased airway volume and increased ventilation heterogeneity were independently associated with methacholine-induced changes in FEV₁

Post-methacholine capnography data were available for 78 subjects (n = 55 without BHR and n = 23 with BHR). To characterize relationships between variations in capnographic variables

	VDaw		slope3	
	R ²	р	R ²	р
FEV1 (L)	0.16	<0.001	0.01	0.8
FVC(L)	0.26	<0.0001	0.07	0.52
PEF (L.s ⁻¹)	0.3	<0.0001	4E-04	0.8
FEF25-75 (L.s ⁻¹)	0.06	0.02	0.07	0.24
RV (L)	2E-04	0.91	0.05	0.17
TLC (L)	0.21	<0.0001	0.02	0.27
Raw (kPa.L ⁻¹ .s ⁻¹)	0.1	0.12	0.03	0.26
FEV1/FVC	0.003	0.65	0.012	0.39

Table 2. Correlation between baseline lung function parameters and baseline airway volume (VDaw) and the slope of the alveolar capnogram (slope3) in 93 subjects. Linear regression was performed with VDaw and slope3 as dependent variables.

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and airflow limitation, univariate and multivariate regression analyses were performed. FEV₁ and FVC dose-responses were strongly correlated ($R^2 = 0.79$, p<0.0001). Baseline VDaw and Δ VDaw/Mch were not correlated, nor were baseline Slope3 and Δ Slope3/Mch. Δ VDaw/Mch and Δ Slope3/Mch were correlated, although the strength of the correlation was weak ($R^2 = -0.20$, p<0.001, Fig 1). There was no correlation between DRS-FEV1 and either baseline VDaw/TLC ($R^2 = 0.002 \text{ p} = 0.70$) or baseline Slope3 (R^2 <0.0001, p = 0.54), nor between DRS-FVC and either baseline VDaw/TLC (R^2 <0.0001 p = 0.97) or baseline Slope3 (R^2 <0.0001, p = 0.47).

<u>Table 3</u> shows univariate relationships between the methacholine dose-response of FEV_1 and FVC, and the methacholine dose-response of capnographic variables. The DRS-FEV₁ was strongly associated with reductions in airway volume in subjects both without and with BHR (Fig 2A). The relationship between variations in Slope3 adjusted for methacholine dose and DRS-FEV₁, although significant, was weaker (Fig 2B). Likewise, univariate associations were



Fig 1. Relationships between methacholine-induced variations in airway volume and the slope of the alveolar capnogram, in patients without BHR (empty circles) and in patients with BHR (filled grey circles). Scales are linear to a value of 100 and exponential thereafter. ΔVDaw / Mch: Fractional change in aiway volume reported to the methacholine dose, expressed as -% / mg. Δslope3 / Mch: Fractional change in the slope of the alveolar capnogram reported to the methacholine dose, expressed as % / mg.

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	All (n = 78)	
	R ²	р
DRS-FEV1		
Baseline VDaw	0.02	0.7
Baseline Slope3	0.06	0.54
ΔVDaw/Mch	0.62	<0.0001
ΔSlope3/Mch	0.42	<0.0001
DRS-FVC		
Baseline VDaw	<0.01	0.97
Baseline Slope3	0.09	0.48
ΔVDaw/Mch	0.69	<0.0001
∆Slope3/Mch	0.23	<0.0001

Table 3. Univariate correlation between the FEV1 and FVC dose-response, baseline airway volume and Slope3, and Mch-induced variations in Vdaw and Slope3. Linear regression was performed with FEV1 and FVC DRS as dependent variables.

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observed between the DRS-FVC and both reduction in VDaw (Fig 3A) and increases in slope3 (Fig 3B). By contrast, neither baseline VDaw nor baseline Slope3 were associated with methacholine-induced reductions in FEV1 or FVC. When analysis was restricted to patients with BHR, significant association remained between Δ VDaw/Mch and both DRS-FEV₁ (R² = 0.56, p<0.0001) and DRS-FVC (R² = 0.66, p<0.0001), and between Δ Slope3/Mch and DRS-FEV1 (R² = 0.33, p = 0.01).

Additional analyses tested whether capnographic parameters measured during tidal breathing correlated with DRS-FEV1 and DRS-FVC. Analysable three-phase capnograms were obtained in 49 patients. In contrast with measurements made from TLC, we did not observe any significant association between Δ VDaw_{tidal}/Mch and either DRS-FEV1 (p = 0.96,



Fig 2. Relationships between methacholine-induced variations in volumetric capnography variables and the FEV1 dose-response curve in patients without BHR (empty circles) and in patients with BHR (filled grey circles). A: Variations in airway volume (VDaw). B: Variation in the slope of the alveolar capnogram (slope3). Scales are linear to a value of 100 and exponential thereafter. ΔVDaw / Mch: Fractional change in aiway volume reported to the methacholine dose, expressed as -% / mg. Δslope3 / Mch: Fractional change in the slope of the alveolar capnogram reported to the methacholine dose, expressed as -% / mg.

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Fig 3. Relationships between methacholine-induced variations in volumetric capnography variables and the FVC dose-response curve in patients without BHR (empty circles) and in patients with BHR (filled grey circles). A: Variations in airway volume (VDaw). B: Variation in the slope of the alveolar capnogram (slope3). Scales are linear to a value of 100 and exponential thereafter. ΔVDaw / Mch: Fractional change in aiway volume reported to the methacholine dose, expressed as -% / mg. Δslope3 / Mch: Fractional change in the slope of the alveolar capnogram reported to the methacholine dose, expressed as % / mg.

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 $R^2 < 0.0001$) or DRS-FVC (p = 0.87, $R^2 = 0.001$), nor between $\Delta Slope3_{tidal}$ /Mch and either DRS-FEV1 (p = 0.53, $R^2 = 0.009$) nor DRS-FVC (p = 0.51, $R^2 = 0.009$).

Multivariate regression (Table 4) demonstrated the independent association of variations in both large airway volume and slope3 with FEV₁ reduction; the regression coefficient for the model was 0.77. By contrast, while reductions in FVC were quite strongly associated with variation in airway volume, no independent association was observed between FVC and variations in slope3. When analysis was restricted to patients with BHR, significant association remained between Δ VDaw/Mch and both DRS-FEV₁ (R² = 0.65, p = 0.0003) and DRS-FVC (R² = 0.81, p<0.0001), and between Δ Slope3/Mch and DRS-FEV1 (R² = 0.33, p = 0.03).

Discussion

The main results of this study are 1) that methacholine-induced variations in airway volume and the slope of the alveolar capnogram were weakly associated and 2) that variations in airway volume were associated with reduction in both FEV1 and FVC, while 3) variations in the slope of the alveolar capnogram were associated with reduction in FEV1 but not FVC. These data suggest that, in the context of methacholine provocation, changes in airflow (FEV₁) occur

Table 4. Multivariate analysis of relationships between the FEV1 and FVC dose-response and Mchinduced variations in capnographic variables. SC: Standardized coefficient.

	All (n = 78)	
	sc	р
DRS-FEV1		
ΔVDaw/Mch	0.66	<0.0001
∆Slope3/Mch	0.35	<0.0001
DRS-FVC		
ΔVDaw/Mch	0.80	<0.0001
∆Slope3/Mch	0.13	0.11

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across proximal and distal airways, whereas changes in lung volume (FVC) are associated with changes in large airways only, suggesting closure of large airways.

As expected, baseline VDaw was highly correlated with indices of lung and airway size. Of note, the strength of the relationship between VDaw and the peak expiratory flow rate in our study was strikingly similar with that reported for radiographic measurements of tracheal size [34], consistent with the association between VDaw and the cross-sectional area of proximal airways measured by acoustic reflexion [21]. Conversely, VDaw was not associated with indices of small airway dysfunction such as decreased FEF₂₅₋₇₅ and increased residual volume. Altogether, these data support the notion that VDaw mainly explores proximal large airways but not distal small airways. Although the delineation of proximal and distal airways in this context is imprecise, theoretical arguments suggest that airways with an internal diameter over 2 mm may be explored by VDaw [22].

The absence of a significant association between baseline proximal airway volume and airway responsiveness in the present study is consistent with our previous observation that tracheal volume is not associated with airway responsiveness in patients with nasal polyposis [35], and suggests that interindividual variation in the baseline volume of large airways is not an important determinant of BHR. This result contrasts with the known association between BHR and low forced expiratory flows at low lung volume, and in particular the FEF₂₅₋₇₅/FVC ratio, which was interpreted as an indication for the involvement of anatomical variation of small airways in BHR [36]. This discrepancy suggests that proximal and distal airway sizes may not be strongly correlated in patients with respiratory symptoms. Based on theoretical grounds, the progressive reduction of airway calibre at each generation can be described by a single constant factor, the homothety ratio, due to the fractal nature of the bronchial tree [35]. Thus, one may hypothesize that patients with BHR could have a reduced homothety ratio, which would not impact the volume of proximal airways but would increase the propensity for distal airflow limitation to occur. Whether the hypothesized decrease in the homothety ratio could be related to underlying anatomy or related to a remodelling process, as suggested for COPD [37], remains to be studied.

The absence of a relationship between baseline Slope3 and airway responsiveness contrasts with findings where ventilation heterogeneity assessed by the multiple breath nitrogen washout technique at baseline was associated with the dose response ratio to methacholine, which is analogous to DRS-FEV₁. In a group of 40 asthmatics, airway responsiveness was significantly associated with Scond, an index of ventilation heterogeneity at the level of conducting airways [<u>38</u>]. In another series of 19 asthmatics, airway responsiveness was correlated with Sacin, an index of ventilation heterogeneity in the diffusing regions of the lung [<u>39</u>]. A possible explanation for the lack of an association between baseline capnographic indices and airway responsiveness may be the high interindividual variability in the former [<u>29</u>].

Previous studies showed correlation between decrements of FEV1 and FVC under methacholine stimulation, suggesting that airway closure is a key factor in airflow reduction in this context, as well as an independent but likely weaker association of FEV1 decrements with reduction in the FEV1/FVC ratio consistent with a role for airway narrowing in non-occluded lung regions [40]. Furthering these studies, we observed that decreases in FVC were strongly associated with decreases in both FEV₁ and VDaw, but not with the slope of the alveolar capnogram which reflects ventilation/perfusion mismatching in lung regions participating to ventilation but does not provide information about lung regions excluded from ventilation. These data support a model where complete closure of large airways reduces FVC and FEV₁ almost proportionally, as suggested by the formation of large and complete ventilation defects in the lungs of asthmatics, the topography of which is consistent with segmental or subsegmental localization [15][16][41]. Whether closure of large airways occurs focally, independent of small airway narrowing, or in association with distal airway closure propagating up the airway tree, remains to clarify. The hypothesis that large airway closure occurs focally is supported by imaging studies directly showing focal stenosis of large airways following methacholine provocation [42]. Conversely, studies relying on the combination of computational modelling and PET imaging of ventilation defects suggest that narrowing of both large and small airway drives the constitution of ventilation defects. Particularly, Tgavalekos and colleagues used a model combining PET and oscillatory mechanics to analyse data from 6 asthmatics in the aim of inferring the site of airway responses, and showed that either constriction of small (diameter <2.4mm) airways alone, or simultaneous constriction of large and small airways could explain both ventilation defects and mechanical alterations induced by inhaled methacholine [43]. Multiple breath nitrogen washout studies in normal subjects and asthmatics showed association between airway closure and Scond [44][45]. Overall, available data suggest that a combination of large and small airway responses drives alterations in lung physiology in the setting of methacholine provocation.

Methacholine-induced variations in VDaw and in the slope of the alveolar capnogram were weakly associated in our study. In addition, the association of increases in slope3 with decreases in FEV1, consistent with participation of small airway contraction to methacholine-induced airflow limitation, was independent of variations in VDaw which reflects large airway geometry. This result suggests that variation in these two capnographic indices, and thus presumably involvement of large and small airways in the response to methacholine, may be driven in part by distinct mechanisms. This hypothesis is supported by a similar observation in the context of exacerbated asthma, where acinar ventilation heterogeneity and conducting airway heterogeneity measured by the multiple breath nitrogen washout technique were independent from each other [46]. Interestingly, in line with the differential expression of cholinergic receptors in large and small airways to a cholinergic stimulus may be different in dogs [48], rats [49] and mice [50].

Our results are consistent with previous work showing increases in KPIv, an index integrating changes in both the second and third phases of the volumetric capnogram, in asthmatic children following induced bronchoconstriction [51]. Importantly, capnography during exhalation from TLC in our work provided information complementary to previous studies where the impact of methacholine on capnography variables was determined during tidal breathing [52][22]. In particular, although we showed a tight relationship between changes in airway volume measured during tidal volume and FEV1 variation, such an association was not evidenced during tidal breathing in normal subjects [52], a finding replicated in our study. Measurements from TLC allow for the quantitation of the volume of the whole bronchial tree, and can thus detect changes related to localized airway closure, as well as to generalized bronchoconstriction. By contrast, measurements made during tidal breathing only explore airway segments participating in tidal ventilation, and may thus be biased by the preferential exploration of airway segments that are either unaffected by methacholine exposure, or subjected to methacholine-induced bronchodilation [11]. Such sampling bias may explain why variations in VDaw were not associated with variations in the resistance of the respiratory system as measured by the force oscillation technique in asthmatics, thus raising the possibly misled hypothesis that alterations in the function of small airways played key roles in methacholine-evoked responses in these subjects [22].

Methodological limitations of our study must be kept in mind. All patients had respiratory symptoms cautioning against translation of results to asymptomatic subjects. Only subjects not receiving asthma medication and without signs of airflow obstruction at baseline were studied, thus precluding any conclusion regarding mechanisms of methacholine-induced airflow limitation in patients with moderate or severe asthma, or in treated patients. In addition, we

ackowledge that, in the absence of a direct comparison between asthmatics and normal controls, our study could not contribute to the identification of capnographic patterns that may be of use for the diagnosis of asthma. The question whether bronchial inflammation was associated with capnographic indices and their variations under provocation was not addressed. Also, it must be acknowledged that determination of Fletcher's dead space by volumetric capnography relies on modelization of the interface of atmospheric and alveolar gas at the start of expiration, and is an estimation of airway volume rather than a direct measurement [53]. In addition, inspiration to TLC during capnography integrates the bronchoprotective and bronchodilatory effect of deep inspiration. It is possible that the well-demonstrated lack of such effects of deep inspirations in most asthmatics [54] may explain part of our results. In particular, one may ask whether airway volume and slope3 measured after methacholine inhalation, but before the deep inspirations associated with spirometry, would have been associated with airflow limitation and airway closure. In any case, we believe that performing capnography after spirometry controlled for the effect of deep inspirations and allowed to study relationships between spirometric and capnographic parameters in a majority of subjects.

In conclusion, our results show that variations in capnographic indices reflecting large airway volume and small airway dysfunction are independently associated with methacholineinduced reductions in FEV₁, indicating that both large and small airways contribute to airflow limitation in the context of methacholine provocation testing. This result supports the use of FEV1 as the outcome variable during methacholine provocation, as this parameters integrates alterations in the geometry and function of both proximal and distal airways. In addition, the observation that methacholine-induced reduction in FVC, which relates to airway closure, was associated with variation in VDaw, which reflects the volume of large airways, but not with variations in Slope3, which reflects small airway dysfunction, indicates that changes in FVC relate for a major part on the closure of large airways following methacholine stimulation.

Supporting Information

S1 Original Data. Data are supplied as a Microsoft Excel file. (XLS)

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Author Contributions

Conceived and designed the experiments: LP SMA LB CT CD. Performed the experiments: LP LB CD. Analyzed the data: LP SMA LB CT CD. Contributed reagents/materials/analysis tools: LP CD. Wrote the paper: LP SMA LB CT CD.

References

- 1. James A, Carroll N. Airway smooth muscle in health and disease; methods of measurement and relation to function. Eur Respir J. 2000; 15: 782–789. PMID: <u>10780774</u>
- Chinn S, Burney P, Jarvis D, Luczynska C. Variation in bronchial responsiveness in the European Community Respiratory Health Survey (ECRHS). Eur Respir J. 1997; 10: 2495–2501. PMID: <u>9426085</u>
- 3. Eder W, Ege MJ, von Mutius E. The asthma epidemic. N Engl J Med. 2006; 355: 2226–2235. PMID: 17124020

- Okazawa M, Müller N, McNamara AE, Child S, Verburgt L, Paré PD. Human airway narrowing measured using high resolution computed tomography. Am J Respir Crit Care Med. 1996; 154: 1557–1562. PMID: 8912780
- Laprise C, Boulet LP. Asymptomatic airway hyperresponsiveness: a three-year follow-up. Am J Respir Crit Care Med. 1997; 156: 403–409. PMID: <u>9279216</u>
- Clini E, Vitacca M, Scalvini S, Quadri A, Foglio K. Methacholine inhaled challenge: study of correlation among different indices expressing the result. Monaldi Arch Chest Dis. 1996; 51: 194–198. PMID: 8766192
- Bouaziz N, Beyaert C, Gauthier R, Monin P, Peslin R, Marchal F. Respiratory system reactance as an indicator of the intrathoracic airway response to methacholine in children. Pediatr Pulmonol. 1996; 22: 7–13. PMID: <u>8856798</u>
- Bohadana AB, Peslin R, Megherbi SE, Teculescu D, Sauleau EA, Wild P, et al. Dose-response slope of forced oscillation and forced expiratory parameters in bronchial challenge testing. Eur Respir J. 1999; 13: 295–300. PMID: 10065671
- Koopman M, Brackel HJL, Vaessen-Verberne AAPH, Hop WC, van der Ent CK, COMBO-Rint Research Group. Evaluation of interrupter resistance in methacholine challenge testing in children. Pediatr Pulmonol. 2011; 46: 266–271. doi: 10.1002/ppul.21362 PMID: 24081886
- King GG, Carroll JD, Müller NL, Whittall KP, Gao M, Nakano Y, et al. Heterogeneity of narrowing in normal and asthmatic airways measured by HRCT. Eur Respir J. 2004; 24: 211–218. PMID: <u>15332387</u>
- Kotaru C, Coreno A, Skowronski M, Muswick G, Gilkeson RC, McFadden ER. Morphometric changes after thermal and methacholine bronchoprovocations. J Appl Physiol 1985. 2005; 98: 1028–1036. PMID: <u>15542566</u>
- Koulouris NG, Latsi P, Stavrou E, Chroneou A, Gaga M, Jordanoglou J. Unevenness of ventilation assessed by the expired CO(2) gas volume versus V(T) curve in asthmatic patients. Respir Physiol Neurobiol. 2004; 140: 293–300. PMID: <u>15186790</u>
- Pellegrino R, Biggi A, Papaleo A, Camuzzini G, Rodarte JR, Brusasco V. Regional expiratory flow limitation studied with Technegas in asthma. J Appl Physiol 1985. 2001; 91: 2190–2198. PMID: <u>11641361</u>
- Harris RS, Fujii-Rios H, Winkler T, Musch G, Vidal Melo MF, Venegas JG. Ventilation defect formation in healthy and asthma subjects is determined by lung inflation. PloS One. 2012; 7: e53216. doi: <u>10.</u> <u>1371/journal.pone.0053216</u> PMID: 23285270
- Kim WW, Lee CH, Goo JM, Park SJ, Kim JH, Park E-A, et al. Xenon-enhanced dual-energy CT of patients with asthma: dynamic ventilation changes after methacholine and salbutamol inhalation. AJR Am J Roentgenol. 2012; 199: 975–981. doi: 10.2214/AJR.11.7624 PMID: 23096168
- Samee S, Altes T, Powers P, de Lange EE, Knight-Scott J, Rakes G, et al. Imaging the lungs in asthmatic patients by using hyperpolarized helium-3 magnetic resonance: assessment of response to methacholine and exercise challenge. J Allergy Clin Immunol. 2003; 111: 1205–1211. PMID: 12789218
- Cohen J, Douma WR, Ten Hacken NHT, Oudkerk M, Postma DS. Physiology of the small airways: A gender difference? Respir Med. 2008; 102: 1264–1271. doi: <u>10.1016/j.rmed.2008.04.007</u> PMID: <u>18617383</u>
- Boudewijn IM, Telenga ED, van der Wiel E, van der Molen T, Schiphof L, Ten Hacken NHT, et al. Less small airway dysfunction in asymptomatic bronchial hyperresponsiveness than in asthma. Allergy. 2013; 68: 1419–1426. doi: 10.1111/all.12242 PMID: 24128343
- Beretta E, Tana F, Grasso GS, Bartesaghi M, Novelli L, Pesci A, et al. Regional differences in bronchial reactivity assessed by respiratory impedance. Respir Physiol Neurobiol. 2014; 192: 23–29. doi: <u>10.</u> <u>1016/j.resp.2013.12.002</u> PMID: <u>24321278</u>
- Alfieri V, Aiello M, Pisi R, Tzani P, Mariani E, Marangio E, et al. Small airway dysfunction is associated to excessive bronchoconstriction in asthmatic patients. Respir Res. 2014; 15: 86. doi: <u>10.1186/s12931-014-0086-1</u> PMID: <u>25158694</u>
- Jackson AC, Loring SH, Drazen JM. Serial distribution of bronchoconstriction induced by vagal stimulation or histamine. J Appl Physiol. 1981; 50: 1286–1292. PMID: <u>7263390</u>
- Sekizawa K, Sasaki H, Shimizu Y, Takishima T. Dose-response effects of methacholine in normal and in asthmatic subjects. Relationship between the site of airway response and overall airway hyperresponsiveness. Am Rev Respir Dis. 1986; 133: 593–599. PMID: 3516037
- Cosio M, Ghezzo H, Hogg JC, Corbin R, Loveland M, Dosman J, et al. The relations between structural changes in small airways and pulmonary-function tests. N Engl J Med. 1978; 298: 1277–1281. PMID: 651978

- Petty TL, Silvers GW, Stanford RE, Baird MD, Mitchell RS. Small airway pathology is related to increased closing capacity and abnormal slope of phase III in excised human lungs. Am Rev Respir Dis. 1980; 121: 449–456. PMID: <u>7416578</u>
- Petersson G, Svensjö E. Nasal mucosal permeability after methacholine, substance P, and capsaicin challenge in the rat. Int J Microcirc Clin Exp. 1990; 9: 205–212. PMID: <u>1692009</u>
- McLeod D, Parsons G, Gunther R, Quail A, Cottee D, White S. Differential effects of inhaled methacholine on circumferential wall and vascular smooth muscle of third-generation airways in awake sheep. J Appl Physiol 1985. 2012; 113: 1233–1242. doi: <u>10.1152/japplphysiol.00133.2012</u> PMID: <u>22898550</u>
- Romero PV, Rodriguez B, de Oliveira D, Blanch L, Manresa F. Volumetric capnography and chronic obstructive pulmonary disease staging. Int J Chron Obstruct Pulmon Dis. 2007; 2: 381–391. PMID: <u>18229577</u>
- 28. Veronez L, Moreira MM, Soares STP, Pereira MC, Ribeiro MA, Ribeiro JD, et al. Volumetric capnography for the evaluation of pulmonary disease in adult patients with cystic fibrosis and noncystic fibrosis bronchiectasis. Lung. 2010; 188: 263–268. doi: 10.1007/s00408-009-9213-z PMID: 20049470
- Babik B, Csorba Z, Czövek D, Mayr PN, Bogáts G, Peták F. Effects of respiratory mechanics on the capnogram phases: importance of dynamic compliance of the respiratory system. Crit Care. 2012; 16: R177. doi: <u>10.1186/cc11659</u> PMID: <u>23031408</u>
- Brusasco V, Crapo R, Viegi G, American Thoracic Society, European Respiratory Society. Coming together: the ATS/ERS consensus on clinical pulmonary function testing. Eur Respir J. 2005; 26: 1–2. PMID: <u>15994380</u>
- Quanjer PH, Tammeling GJ, Cotes JE, Pedersen OF, Peslin R, Yernault JC. Lung volumes and forced ventilatory flows. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. Eur Respir J Suppl. 1993; 16: 5–40. PMID: 8499054
- 32. Sterk PJ, Fabbri LM, Quanjer PH, Cockcroft DW, O'Byrne PM, Anderson SD, et al. Airway responsiveness. Standardized challenge testing with pharmacological, physical and sensitizing stimuli in adults. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. Eur Respir J Suppl. 1993; 16: 53–83. PMID: 8499055
- Fletcher R, Jonson B, Cumming G, Brew J. The concept of deadspace with special reference to the single breath test for carbon dioxide. Br J Anaesth. 1981; 53: 77–88. PMID: 6779846
- Osmanliev D, Bowley N, Hunter DM, Pride NB. Relation between tracheal size and forced expiratory volume in one second in young men. Am Rev Respir Dis. 1982; 126: 179–182. PMID: 7091902
- Bokov P, Chevalier-Bidaud B, Al Dandachi G, Londner C, Plantier L, Bonfils P, et al. Tracheal section is an independent predictor of asthma in patients with nasal polyposis. Respir Physiol Neurobiol. 2014; 203: 15–18. doi: 10.1016/j.resp.2014.08.017 PMID: 25174298
- Litonjua AA, Sparrow D, Weiss ST. The FEF25-75/FVC ratio is associated with methacholine airway responsiveness. The normative aging study. Am J Respir Crit Care Med. 1999; 159: 1574–1579. PMID: 10228129
- Bokov P, Mauroy B, Revel MP, Brun PA, Peiffer C, Daniel C, et al. Lumen areas and homothety factor influence airway resistance in COPD. Respir Physiol Neurobiol. 2010; 173: 1–10. doi: <u>10.1016/j.resp.</u> <u>2010.05.011</u> PMID: <u>20478416</u>
- Downie SR, Salome CM, Verbanck S, Thompson B, Berend N, King GG. Ventilation heterogeneity is a major determinant of airway hyperresponsiveness in asthma, independent of airway inflammation. Thorax. 2007; 62: 684–689. PMID: <u>17311839</u>
- Hardaker KM, Downie SR, Kermode JA, Berend N, King GG, Salome CM. Ventilation heterogeneity is associated with airway responsiveness in asthma but not COPD. Respir Physiol Neurobiol. 2013; 189: 106–111. doi: <u>10.1016/j.resp.2013.07.009</u> PMID: <u>23876741</u>
- 40. Chapman DG, Berend N, King GG, Salome CM. Increased airway closure is a determinant of airway hyperresponsiveness. Eur Respir J. 2008; 32: 1563–1569. doi: <u>10.1183/09031936.00114007</u> PMID: <u>18653648</u>
- Venegas JG, Winkler T, Musch G, Vidal Melo MF, Layfield D, Tgavalekos N, et al. Self-organized patchiness in asthma as a prelude to catastrophic shifts. Nature. 2005; 434: 777–782. PMID: <u>15772676</u>
- Wongviriyawong C, Harris RS, Zheng H, Kone M, Winkler T, Venegas JG. Functional effect of longitudinal heterogeneity in constricted airways before and after lung expansion. J Appl Physiol 1985. 2012; 112: 237–245. doi: <u>10.1152/japplphysiol.01400.2010</u> PMID: <u>21940845</u>
- 43. Tgavalekos NT, Tawhai M, Harris RS, Musch G, Mush G, Vidal-Melo M, et al. Identifying airways responsible for heterogeneous ventilation and mechanical dysfunction in asthma: an image functional modeling approach. J Appl Physiol 1985. 2005; 99: 2388–2397. PMID: <u>16081622</u>

- Farrow CE, Salome CM, Harris BE, Bailey DL, Bailey E, Berend N, et al. Airway closure on imaging relates to airway hyperresponsiveness and peripheral airway disease in asthma. J Appl Physiol 1985. 2012; 113: 958–966. doi: <u>10.1152/japplphysiol.01618.2011</u> PMID: <u>22837168</u>
- 45. King GG, Downie SR, Verbanck S, Thorpe CW, Berend N, Salome CM, et al. Effects of methacholine on small airway function measured by forced oscillation technique and multiple breath nitrogen washout in normal subjects. Respir Physiol Neurobiol. 2005; 148: 165–177. PMID: <u>15950553</u>
- 46. Thompson BR, Douglass JA, Ellis MJ, Kelly VJ, O'Hehir RE, King GG, et al. Peripheral lung function in patients with stable and unstable asthma. J Allergy Clin Immunol. 2013; 131: 1322–1328. doi: <u>10.1016/j.jaci.2013.01.054</u> PMID: <u>23561802</u>
- Barnes PJ, Basbaum CB, Nadel JA. Autoradiographic localization of autonomic receptors in airway smooth muscle. Marked differences between large and small airways. Am Rev Respir Dis. 1983; 127: 758–762. PMID: <u>6305241</u>
- Chitano P, Sigurdsson SB, Halayko AJ, Stephens NL. Relevance of classification by size to topographical differences in bronchial smooth muscle response. J Appl Physiol 1985. 1993; 75: 2013–2021. PMID: 8307854
- **49.** Siddiqui S, Jo T, Tamaoka M, Shalaby KH, Ghezzo H, Bernabeu M, et al. Sites of allergic airway smooth muscle remodeling and hyperresponsiveness are not associated in the rat. J Appl Physiol 1985. 2010; 109: 1170–1178. doi: <u>10.1152/japplphysiol.01168.2009</u> PMID: <u>20651225</u>
- 50. Donovan C, Royce SG, Esposito J, Tran J, Ibrahim ZA, Tang MLK, et al. Differential effects of allergen challenge on large and small airway reactivity in mice. PloS One. 2013; 8: e74101. doi: <u>10.1371/journal.pone.0074101</u> PMID: <u>24040180</u>
- Strömberg NO, Gustafsson PM. Ventilation inhomogeneity assessed by nitrogen washout and ventilation-perfusion mismatch by capnography in stable and induced airway obstruction. Pediatr Pulmonol. 2000; 29: 94–102. PMID: <u>10639199</u>
- Olsson K, Greiff L, Karlefors F, Johansson S, Wollmer P. Changes in airway dead space in response to methacholine provocation in normal subjects. Clin Physiol. 1999; 19: 426–432. PMID: <u>10516894</u>
- 53. Drummond GB, Fletcher R. Deadspace: invasive or not? Br J Anaesth. 2006; 96: 4–7. PMID: <u>16357115</u>
- Skloot G, Permutt S, Togias A. Airway hyperresponsiveness in asthma: a problem of limited smooth muscle relaxation with inspiration. J Clin Invest. 1995; 96: 2393–2403. PMID: <u>7593627</u>