

# Synergistic cardioprotective effects of Danshensu and hydroxysafflor yellow A against myocardial ischemia-reperfusion injury are mediated through the Akt/Nrf2/HO-1 pathway

TIANXIN HU, GUO WEI, MIAOMIAO XI, JIAJIA YAN, XIAOXIAO WU,  
YANHUA WANG, YANRONG ZHU, CHAO WANG and AIDONG WEN

Department of Pharmacy, Xijing Hospital, The Fourth Military Medical University, Xi'an, Shaanxi 710032, P.R. China

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**Abstract.** In clinical practice, the traditional Chinese medicinal herbs, *Radix Salvia Miltiorrhiza* and *Carthamus tinctorius L.*, are usually prescribed in combination due to their significant cardioprotective effects. However, the mechanisms responsible for these combined effects remain unknown. Thus, in this study, we investigated the mechanisms responsible for the combined effects of Danshensu (DSS) and hydroxysafflor yellow A (HSYA) by establishing a rat model of myocardial ischemia/reperfusion (MI/R), as well as a model of hypoxia/reoxygenation (H/R) using H9c2 cells. The combination index (CI) was calculated using the median-effect method. DSS and HSYA in combination led to a CI value of <1 as regards infarct size *in vivo* and cell viability *in vitro*. The rats with MI/R injury that were treated with DSS and/or HSYA were found to have significantly lower levels of creatine kinase-MB (CK-MB) and cardiac troponin I (cTnI) and malondialdehyde (MDA), and a lower expression of 8-hydroxydeoxyguanosine (8-OHdG), and markedly enhanced superoxide dismutase (SOD) activity. Our *in vitro* experiments revealed that the cells treated with DSS and/or HSYA had a reduced lactate dehydrogenase (LDH)

activity and a decreased percentage of cell apoptosis (increased Bcl-2/Bax ratio, decreased expression of cleaved caspase-3). DSS and HSYA increased the expression of heme oxygenase-1 (HO-1), the phosphorylation of Akt and the translocation of nuclear factor erythroid 2-related factor 2 (Nrf2). Furthermore, the Akt inhibitor, LY294002, partially hampered the expression of Nrf2 and HO-1. The HO-1 inhibitor, zinc protoporphyrin IX (ZnPP-IX), did not decrease the expression of p-Akt and Nrf2, although it abolished the anti-apoptotic and antioxidant effects of DSS and HSYA. The findings of our study thus demonstrate that DSS and HSYA confer synergistic cardioprotective effects through the Akt/Nrf2/HO-1 signaling pathway, to certain extent, by enhancing the antioxidant defense system and exerting anti-apoptotic effects.

## Introduction

Myocardial infarction is a leading cause of mortality and morbidity in developed countries. Although prompt reperfusion therapy has been shown to reduce the infarct size and improve left ventricular function in ST-elevation myocardial infarction, reperfusion itself may cause lethal tissue injury and a series of cellular events termed myocardial ischemia-reperfusion (MI/R) injury (1-4).

The molecular mechanisms of reperfusion injury appear to be multifactorial with various consequences on cellular function (1). It has been previously noted that apoptosis occurs shortly after myocardial infarction and it is markedly enhanced during reperfusion (5,6). The production of reactive oxygen species (ROS) in the ischemic region and the surrounding myocardium is also generated, which directly triggers cell death, including apoptosis. Inflammatory responses, disruptions of energy metabolism and calcium overload have also been proposed to underlie the pathology of MI/R injury (7). Thus, a single therapeutic agent may be incapable of hitting multiple targets to achieve therapeutic efficacy.

Comprehensive investigations have been focusing on combination drugs in order to optimize or amplify the therapeutic effects (8,9). In traditional Chinese medicine (TCM), a number of herbs are paired together in order to attenuate toxicity, as well as to enhance the therapeutic effects (10). *Radix Salvia miltiorrhiza* (*S. miltiorrhiza*) and *Carthamus tin-*

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*Correspondence to:* Dr Aidong Wen, Department of Pharmacy, Xijing Hospital, The Fourth Military Medical University, 15 Changle West Road, Xi'an, Shaanxi 710032, P.R. China  
E-mail: aidongwen2014@163.com

**Abbreviations:** MI/R, myocardial ischemia-reperfusion; Nrf2, nuclear factor erythroid 2-related factor 2; CI, combination index; DSS, Danshensu; SOD, superoxide dismutase; HSYA, hydroxysafflor yellow A; ROS, reactive oxygen species; Akt, protein kinase B; HO-1, heme oxygenase-1; H/R, hypoxia/reoxygenation; CK-MB, creatine kinase-MB; cTnI, cardiac troponin I; MDA, malondialdehyde; LDH, lactate dehydrogenase; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling; 8-OHdG, 8-hydroxydeoxyguanosine

**Key words:** myocardial ischemia/reperfusion, Danshensu, hydroxysafflor yellow A, synergy, anti-apoptosis, antioxidant, protein kinase B/nuclear factor erythroid 2-related factor 2/heme oxygenase-1

*torius* L. (*C. tinctorius*; also known as Flos Carthami) are usually used as a combination herbal formulation, known as Danhong injection, which can relieve the symptoms of angina pectoris, attenuate myocardial ischemia and promotes atherosclerotic plaque regression (11). Due to their complex constituents, studies have mainly focused on their combination effects. However, previous studies have confirmed that Danshensu (DSS) and hydroxysafflor yellow A (HSYA) are the main active ingredients of Radix *Salvia miltiorrhiza* and Flos Carthami, respectively (12,13). Hence, evaluating the combination effects of these two active compounds may be of importance in understanding the rationale for the combined use of the two herbs in TCM.

There is evidence to indicate that *S. miltiorrhiza* and *C. tinctorius* inhibit cellular apoptosis and oxidative stress induced by MI/R (14,15). It would be of interest to investigate the mechanisms responsible for the combined effects of DSS and HSYA. Thus, the present study aimed to evaluate the cardioprotective effects of combination therapy with DSS and HSYA in order to elucidate the mechanisms responsible for their combined antioxidant and anti-apoptotic effects *in vivo* and *in vitro*.

## Materials and methods

**Chemicals and drugs.** DSS was purchased from the National Institute for the Control of Pharmaceutical and Biological Products (Beijing, China) as an amorphous powder. Its purity (>98%) was determined using high-performance liquid chromatography (HPLC). The molecular formula of DSS is  $C_9H_{10}O_5$  and its molecular weight is 162.14. HSYA was obtained from *C. tinctorius* as a yellow amorphous powder. Its purity (>99%) was determined using HPLC. Being soluble in water, it has a molecular formula of  $C_{27}H_{32}O_{16}$  and a molecular weight of 611.1614.

The creatine kinase-MB (CK-MB) and cardiac troponin I (cTnI) kits were purchased from Roche Molecular Biochemicals (Mannheim, Germany). The lactate dehydrogenase (LDH), superoxide dismutase (SOD) and malondialdehyde (MDA) kits were obtained from Jiancheng Bioengineering Institute (Nanjing, China).

Dulbecco's modified Eagle's medium (DMEM) and other cell culture supplies were purchased from Gibco-BRL (Grand Island, NY, USA). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), zinc protoporphyrin IX [ZnPP-IX; a heme oxygenase-1 (HO-1) inhibitor], 2,3,5-triphenyltetrazolium chloride (TTC), Evans blue, and hematoxylin and eosin were the products of Sigma Chemical Co. (St. Louis, MO, USA). The phosphoinositide 3-kinase (PI3K) inhibitor, LY294002 (#9901), and rabbit polyclonal antibodies specific for Bcl-2 (#2870), Bax (#2772),  $\beta$ -actin (#4970), total protein kinase B (Akt; t-Akt, serine 473; #9272), phosphorylated Akt (p-Akt; #9271), cleaved caspase-3 (#9661) and caspase-3 (#9662) were obtained from Cell Signaling Technology (Beverly, MA, USA). Rabbit polyclonal antibodies specific for HO-1 (# sc-10789), 8-hydroxydeoxyguanosine (8-OHdG; #sc-66036) and nuclear factor erythroid 2-related factor 2 (Nrf2; #sc-722) were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). Rabbit polyclonal antibody specific for histone 3 (H3; #ab4729) was purchased from Abcam (Cambridge, UK). All materials for sodium dodecyl sulfate-polyacrylamide

gel electrophoresis (SDS-PAGE) were obtained from Bio-Rad Laboratories (Hercules, CA, USA).

## Experimental protocols

***In vivo* experiments.** In consideration of the failures which may occur when performing coronary ligation, such as no infarcts or death, 10 rats were randomly selected in each group so that a sufficient number of animals (at least 6 rats) was maintained. The concentrations of DSS and HSYA were 10, 20, 30, 45 and 60 mg/kg, and 10, 20, 30, 50 and 70 mg/kg, respectively. They were combined at concentrations of 10, 20, 30, 50 and 70 mg/kg (at a ratio of 1:1 according to each  $IC_{50}$  value) to analyze the synergistic effects on myocardial infarct size. In the subsequent experiments, the rats were randomly divided into 5 groups as follows: i) the sham-operated group, ii) the MI/R group, iii) the MI/R + DSS (60 mg/kg) group, iv) the MI/R + HSYA (70 mg/kg) group, and v) the MI/R + DSS + HSYA (DH; 35 mg/kg DSS + 35 mg/kg HSYA) group. All drugs were administered via tail vein injection at the time of reperfusion. The concentrations of DSS and HSYA were selected on the basis of the reported literature and our preliminary dose selection experiments (15,16).

***In vitro* experiments.** To further explore the mechanisms responsible for the combined effects of DSS and HSYA, H9c2 cardiomyocytes were used. The concentrations of DSS and HSYA were 1, 10, 35, 60 and 80  $\mu$ M. DSS and HSYA at concentrations of 1, 10, 35, 60 and 80  $\mu$ M were used in combination (at a ratio of 1:1 according to each  $IC_{50}$  value) to analyze the synergistic effects on cell viability. In the subsequent experiments, the H9c2 cells were randomly divided into the following groups: i) the control (Con) group, ii) the hypoxia/reoxygenation (H/R) group, iii) the H/R + DSS (80  $\mu$ M) group, iv) the H/R + HSYA (80  $\mu$ M) group, v) the H/R + DH (40  $\mu$ M DSS + 40  $\mu$ M HSYA) group, vi) the H/R + DH + ZnPP-IX (10  $\mu$ M) group, and vii) the H/R + DH + LY294002 (50  $\mu$ M) group. The doses of each agent were selected according to the published literature and our preliminary experiments (17,18).

**Animals and the establishment of an animal model of MI/R injury.** Adult male Sprague-Dawley rats (n=220, approximately 2 months old, weighing  $250 \pm 20$  g) were purchased from the Experimental Animal Research Center at the Fourth Military Medical University (Xi'an, China). The experimental protocols involving animals were performed in adherence with Institutional Animal Care and were approved by the Animal Care and Use Committee of the Fourth Military Medical University.

The rats were anesthetized with sodium pentobarbital (40 mg/kg intraperitoneally) and the tracheas were cannulated with a polyethylene-90 (PE-90) tube connected to a rodent ventilator with a tidal volume of 1.2 l/kg (75 breaths/min). A left thoracotomy was performed between the fourth and fifth ribs. The pericardium was removed and the left anterior descending (LAD) artery was visualized and ligated with a 6-0 Prolene suture, as previously described (19). The appearance of myocardial pallor was confirmed as ischemia. After 30 min of LAD ligation, the ligature was removed to allow for 180 min of reperfusion. In the sham-operated group, Prolene was drilled underneath the LAD, but not ligated.

**Assessment of myocardial infarct size.** After the completion of 180 min of reperfusion, the myocardial infarct size was assessed by a double-staining technique using 2% TTC and 3% Evans blue as previously described (20). Briefly, the LAD was re-ligated and 2 ml of 3% Evans blue dye was retrogradely infused into the carotid artery to demarcate the area at risk (AAR; area not perfused with blue dye) from the area not at risk (stained with blue dye). After the dye was uniformly distributed, the hearts were rapidly excised and frozen at  $-20^{\circ}\text{C}$ , and subsequently the heart tissue was cut into 5 transverse slices. The sections were incubated in 2% TTC solution in phosphate buffer (pH 7.4) at  $37^{\circ}\text{C}$  in the dark for 15 min and then stored in 4% paraformaldehyde overnight to delineate the infarct size (IS; pale area). The IS and AAR were measured using Image-Pro Plus software (Media Cybernetics, Inc., Silver Spring, MD, USA) after capturing images. The myocardial infarct size was calculated as a percentage of the infarct size over the total AAR. The dose-effect curve and fraction versus combination index (Fa-CI) curve were analyzed using CompuSyn software (MIT, Cambridge, MA, USA).

**Determination of CK-MB and cTnI release in serum.** After being anesthetized, the rats were subjected to MI/R surgery. Following 3 h of reperfusion, blood samples were collected from the abdominal aorta of the rats using a 10 ml syringe. The blood of the experimental rats was collected and serum was separated by centrifugation and then kept at  $-20^{\circ}\text{C}$ . The levels of CK-MB, and cTnI were estimated using a commercially available enzyme-linked immunosorbent assay kit with a microplate reader (Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer's instructions.

**Assessment of markers for oxidative stress.** The serum and the cell culture supernatant were used to assay the MDA content and SOD activity using a microplate reader (Multiskan GO; Thermo Fisher Scientific) according to the instructions provided by the manufacturer.

**Immunohistochemical assay for the evaluation of 8-OHdG expression.** The paraffin-embedded tissue samples were deparaffinized in xylene and then dehydrated with ethanol. Subsequently, the samples were subjected to antigen retrieval and then incubated in 3%  $\text{H}_2\text{O}_2$  in 0.01 M phosphate-buffered saline (PBS) and in 5% bovine serum albumin (BSA) successively. The sections were incubated overnight at  $4^{\circ}\text{C}$  with a primary antibody anti-8-OHdG (1:100) and then incubated for 1 h with a secondary antibody (Boster Biological Technology, Wuhan, China). The reaction was visualized with a solution of diaminobenzidine (DAB) and counterstained with hematoxylin. For quantification, the number of 8-OHdG-stained positive cells was calculated using Image-Pro Plus 6.0 software (Media Cybernetics, Inc.).

**Cell culture and H/R injury.** The H9c2 cardiomyocyte cell line was purchased from the Chinese Academy of Sciences Cell Bank (Shanghai, China) and maintained in DMEM supplemented with 10% (v/v) fetal bovine serum at  $37^{\circ}\text{C}$  in a  $\text{CO}_2$  incubator. The medium was replaced every 2-3 days, and the cells were subcultured or subjected to experimental procedures at 80-90% confluence.

To mimic ischemic injury *in vivo*, the procedures for inducing H/R injury were modified from a previously described method (21). Briefly, the cells were maintained in serum-free DMEM (glucose-free) instead of routine culture medium. Hypoxic conditions were established by equilibrating a humidified chamber containing the cells with 95%  $\text{N}_2$  and 5%  $\text{CO}_2$  via a gas transfusion apparatus (Billups-Rothenberg, Del Mar, CA, USA). Following 4 h of incubation, the cells were transferred to normal conditions in a  $\text{CO}_2$  incubator and the medium was replaced with routine culture medium to achieve re-oxygenation. The drugs were administered at the beginning of re-oxygenation. Following 20 h of re-oxygenation, the culture medium was collected and stored at  $-80^{\circ}\text{C}$  until analysis. In the control group, the H9c2 cells were cultured under normal conditions for 24 h.

**Analysis of cell viability and LDH activity.** The cells in the exponential phase were seeded at  $1 \times 10^4$  cells/well in 96-well plates. After being subjected to the different treatments, 20  $\mu\text{l}$  of MTT solution were added to the medium (0.5 mg/ml final concentration in medium) and the cells were incubated for an additional 4 h at  $37^{\circ}\text{C}$ . The supernatants were removed and the formazan crystals were dissolved in 150  $\mu\text{l}$  DMSO. The absorbance was read at 490 nm using a microplate reader (Multiskan GO; Thermo Fisher Scientific). A reduction in optical density reduction was considered to indicate a decrease in cell viability. The cells in the control group were considered 100% viable. The results were also assessed using CompuSyn software to calculate the CI value.

In order to confirm the degree of cardiomyocyte injury in H/R, the release of LDH was measured. After being subjected to the different treatments, the cell culture supernatants were collected to assay LDH activity immediately according to the manufacturer's instructions using a microplate reader (Multiskan GO; Thermo Fisher Scientific) at 450 nm.

**Protein extraction and western blot analysis.** The cardiomyocytes were resuspended in radioimmunoprecipitation assay (RIPA) lysis buffer (Beyotime Institute of Biotechnology, Jiangsu, China) on ice for 30 min. They were then centrifuged at  $4^{\circ}\text{C}$  for 20 min at 10,000 rpm to separate the supernatant before being stored at  $-80^{\circ}\text{C}$ . The nuclear proteins were extracted separately from cultured myocytes using NE-PER nuclear and cytoplasmic extraction reagents according to the manufacturer's instructions (Thermo Fisher Scientific). A Mitochondria/Cytosol Fractionation kit (BioVision, San Francisco, CA, USA) was used to prepare mitochondrial protein. Protein concentrations were measured using the Bradford method with the Bio-Rad protein assay kit (Bio-Rad Laboratories). Denatured protein was separated by SDS-PAGE and then electrotransferred onto polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). After being blocked with 5% (w/v) non-fat milk at  $37^{\circ}\text{C}$  for 30 min, the membranes were incubated overnight at  $4^{\circ}\text{C}$  with primary antibodies including p-Akt, t-Akt, Bcl-2, Bax, cleaved caspase-3 and caspase-3, HO-1, Nrf2,  $\beta$ -actin and H3 (1:1,000), respectively. After washing in TBST 3 times, the membranes were incubated at room temperature for 1 h with secondary antibody diluted in TBST. The labeled protein bands were detected using chemiluminescent reagents and were exposed to film. The band intensity was determined

using an image analyzer (Quantity One System; Bio-Rad, Richmond, CA, USA).

**Detection of apoptotic cell death.** Cell apoptosis was determined by terminal deoxy-nucleotidyl transferase-mediated dUTP nick-end labeling (TUNEL) assay, as previously described (22). The H9c2 cells grown on a 6-mm plate were fixed with 4% paraformaldehyde solution for 30 min at room temperature. The cells were then treated with permeation solution. Subsequently, the samples were incubated with TUNEL reagent. The cells were also stained with 1  $\mu$ g/ml 4',6-diamidino-2-phenylindole (DAPI) for 30 min to detect cell nuclei (blue). The number of TUNEL-positive cells was presented as a percentage of the total cardiomyocytes and was evaluated at x400 magnification.

**Determination of combined effects.** The manner in which DSS and HSYA act with regard to myocardial infarct size and cell viability was determined by a median-effect method proposed by Chou (23). Synergism or antagonism was determined with CI values. CompuSyn software (MIT) was used to determine the CI value. The CI was plotted as the fractional inhibition (Fa) by computer simulation from 0.10 to 0.95. In this analysis, the combined effect at the 50% fractional inhibition (CI<sub>50</sub>) was reported as synergistic, antagonistic or additive when the CI<sub>50</sub> value was <1, >1 and equal to 1, respectively.

**Statistical analysis.** All values are expressed as the means  $\pm$  standard deviation (SD). Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by a least significant difference (LSD) test for multiple comparisons, using SPSS 19.0 software for Windows (SPSS Inc., Chicago, IL, USA). A value of P<0.05 was considered to indicate a statistically significant difference.

## Results

**DSS and HSYA synergistically alleviate myocardial injury in rats with MI/R injury.** Infarct size and the release of cTnI, and CK-MB in serum were measured to determine the mechanisms through which DSS and HSYA influence MI/R injury. The myocardial infarct size was significantly reduced in the groups treated with the compounds compared with that in the MI/R group (Fig. 1A and B). Combined treatment (DH) enhanced this effect to a greater extent than treatment with either DSS or HSYA alone. In order to determine whether DSS and HSYA have a synergistic effect on reducing the infarct size, the dose-effect curves of the single or combined treatment were analyzed by the median-effect method. We found that DSS, HSYA and combination therapy (Fig. 1C-1) at concentrations of 10-60, 10-70 and 10-70 mg/kg yielded CI<sub>50</sub> values <1 (Fig. 1C-2), indicating synergistic effects between the agents. As these effects occurred in a dose-dependent manner, the highest doses were selected for use in subsequent experiments. The numbers of rats that failed the coronary ligation procedure in each group shown in Fig. 1 are listed in Table I.

As markers of cardiomyocyte injury, the serum levels of CK-MB and cTnI were 104.68 $\pm$ 9.93 and 8.47 $\pm$ 0.80 ng/ml in the sham-operated group and were significantly increased in the MI/R group to 220.78 $\pm$ 10.63 and 21.52 $\pm$ 2.79 ng/ml, respectively.

Table I. Number of rats that failed the coronary ligation procedure in each group shown in Fig. 1.

Group	No.	Group	No.
Sham	1	MI/R + HSYA30	1
MI/R	3	MI/R + HSYA50	1
MI/R + DSS10	2	MI/R + HSYA70	3
MI/R + DSS20	3	MI/R + DH10	1
MI/R + DSS30	2	MI/R + DH20	2
MI/R + DSS45	3	MI/R + DH30	6
MI/R + DSS60	2	MI/R + DH50	2
MI/R + HSYA10	1	MI/R + DH70	1
MI/R + HSYA20	2		

MI/R, myocardial ischemia-reperfusion; HSYA, hydroxysafflor yellow A; DSS, Danshensu; DH, HSYA + DSS.

Table II. Number of rats that failed the coronary ligation procedure in each group shown in Fig. 2.

Group	Sham	MI/R	MI/R + DSS	MI/R + HSYA	MI/R + DH
No.	1	3	3	1	2

MI/R, myocardial ischemia-reperfusion; HSYA, hydroxysafflor yellow A; DSS, danshensu; DH, HSYA + DSS.

Following treatment with 60 mg/kg DSS or 70 mg/kg HSYA, the CK-MB and cTnI levels were significantly decreased (P<0.01 and P<0.05 vs. MI/R). The DH regimen (35+35 mg/kg) further enhanced these effects compared to treatment with each agent alone (Fig. 2). The numbers of rats that failed the coronary ligation procedure in each group shown in Fig. 2 are listed in Table II.

**DSS and HSYA exert antioxidant effect on rats with MI/R injury.** The rats in the MI/R group exhibited a significant increase in the level of MDA to 17.01 $\pm$ 3.58 nmol/ml (P<0.01 vs. sham-operated group) and a significant decrease in SOD activity to 69.66 $\pm$ 16.32 U/ml (P<0.01 vs. sham-operated group). The administration of DSS and HSYA in combination to the rats with MI/R injury exerted more significant cardioprotective effects (P<0.01 vs. MI/R) than the administration of each agent alone (P<0.05 vs. MI/R), alleviating the parameters of oxidative stress (Table III). 8-OHdG is regarded as a hallmark of oxidative DNA damage (24); thus, nuclear oxidative stress was assessed using 8-OHdG immunohistochemical staining, which was significantly increased (P<0.01 vs. sham-operated group) in the MI/R group, while it was significantly reduced (P<0.01 vs. MI/R) in the combined treatment group. The protocol used in the DH group further potentiated the cardioprotective effects compared with those observed in the individual treatment groups (Fig. 3). Additionally, HSYA exerted a more potent antioxidant effect than DSS. The number of rats that failed the coronary ligation procedure in each group shown Fig. 3 are listed in Table IV.

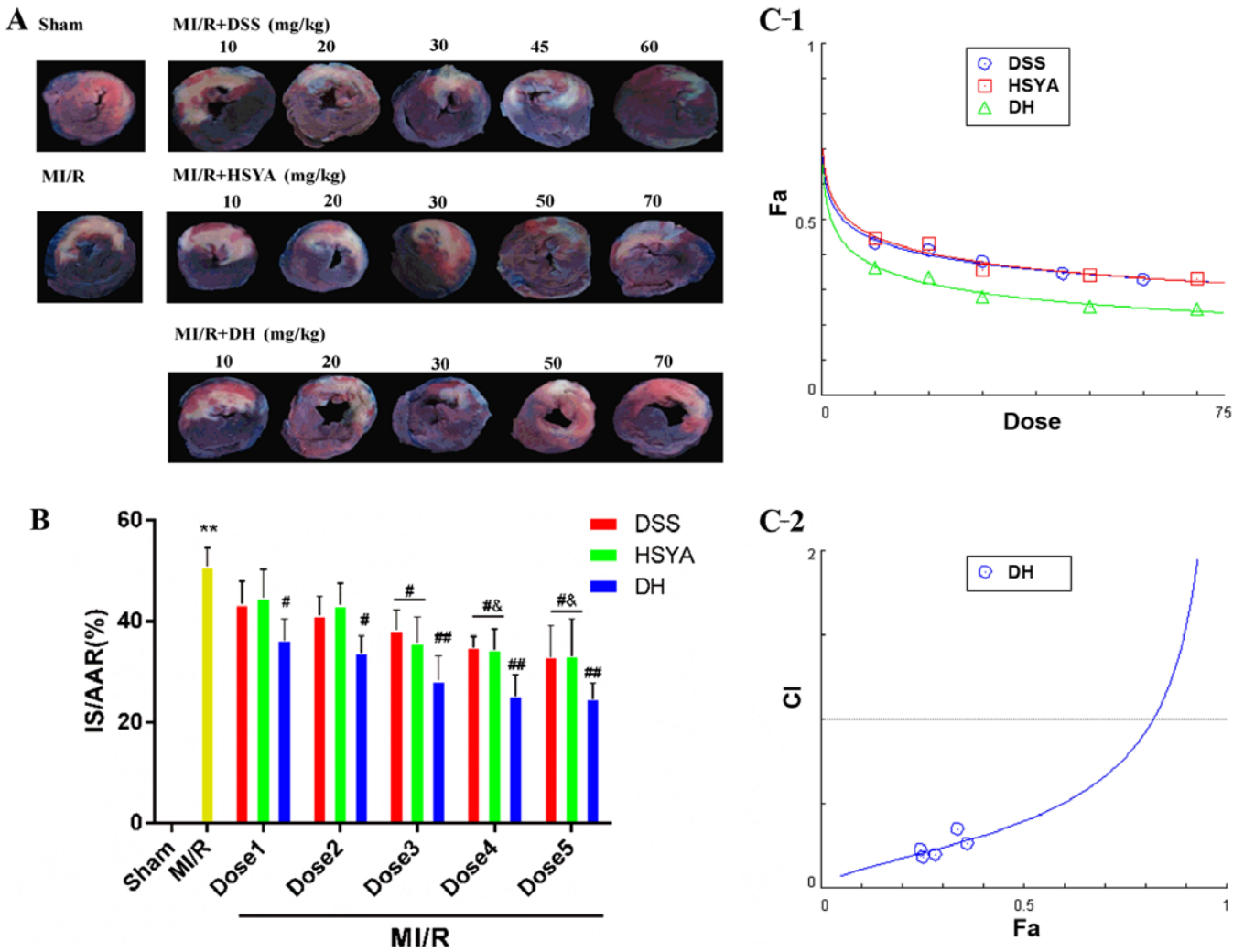


Figure 1. Danshensu (DSS) and hydroxysafflor yellow A (HSYA) administered individually and in combination (DH) reduce myocardial infarct size in rats subjected to myocardial ischemia-reperfusion (MI/R) injury, and in combination exert a synergistic effect. (A) A double-staining technique using TTC and Evans blue assessed the effect of DSS and HSYA, individually and in combination at different doses on infarct size and myocardial risk area. (B) Myocardial infarct size expressed as a percentage of the area at risk (IS/AAR%). Dose 1 indicates DSS (10 mg/kg), HSYA (10 mg/kg), DH (10 mg/kg); Dose 2 indicates DSS (20 mg/kg), HSYA (20 mg/kg), DH (20 mg/kg); Dose 3 indicates DSS (30 mg/kg), HSYA (30 mg/kg), DH (30 mg/kg); Dose 4 indicates DSS (45 mg/kg), HSYA (50 mg/kg), DH (50 mg/kg); Dose 5 indicates DSS (60 mg/kg), HSYA (70 mg/kg), DH (70 mg/kg). (C-1) The dose-effect curves of the single or combined drug treatment. (C-2) The fraction versus combination index (Fa-Cl) curve of DSS and HSYA in combination reveals that they exert a synergistic effect ( $CI_{50} < 1$ ) as reflected by the median-effect method. The dashed line at  $CI=1$  represents the additive effect. Values are expressed as the means  $\pm$  SD (n=7-9 rats in each group). \*\*P<0.01 vs. sham-operated (sham) group; ##P<0.01 and #P<0.05 vs. MI/R group; &P<0.05 vs. MI/R + DSS or MI/R + HSYA group.

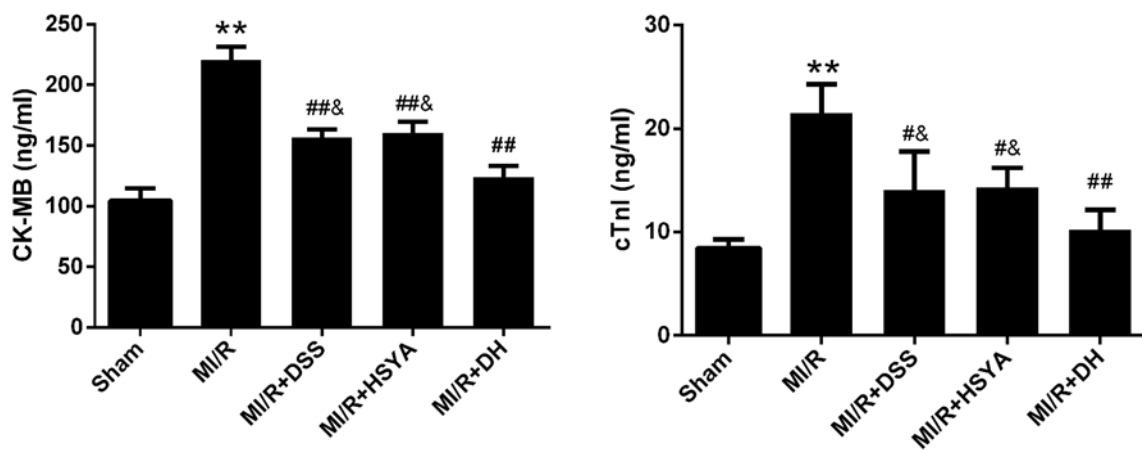


Figure 2. Levels of creatine kinase-MB (CK-MB) and cardiac troponin I (cTnI) in the serum of rats in the sham-operated (sham), myocardial ischemia-reperfusion (MI/R), MI/R + Danshensu (DSS; 60 mg/kg), MI/R + hydroxysafflor yellow A (HSYA; 70 mg/kg) and MI/R + DSS + HSYA (35+35 mg/kg) groups. Values are expressed as the means  $\pm$  SD (n=7-9 rats in each group). \*\*P<0.01 vs. sham group; ##P<0.01 and #P<0.05 vs. MI/R group; &P<0.05 vs. MI/R + DH group.



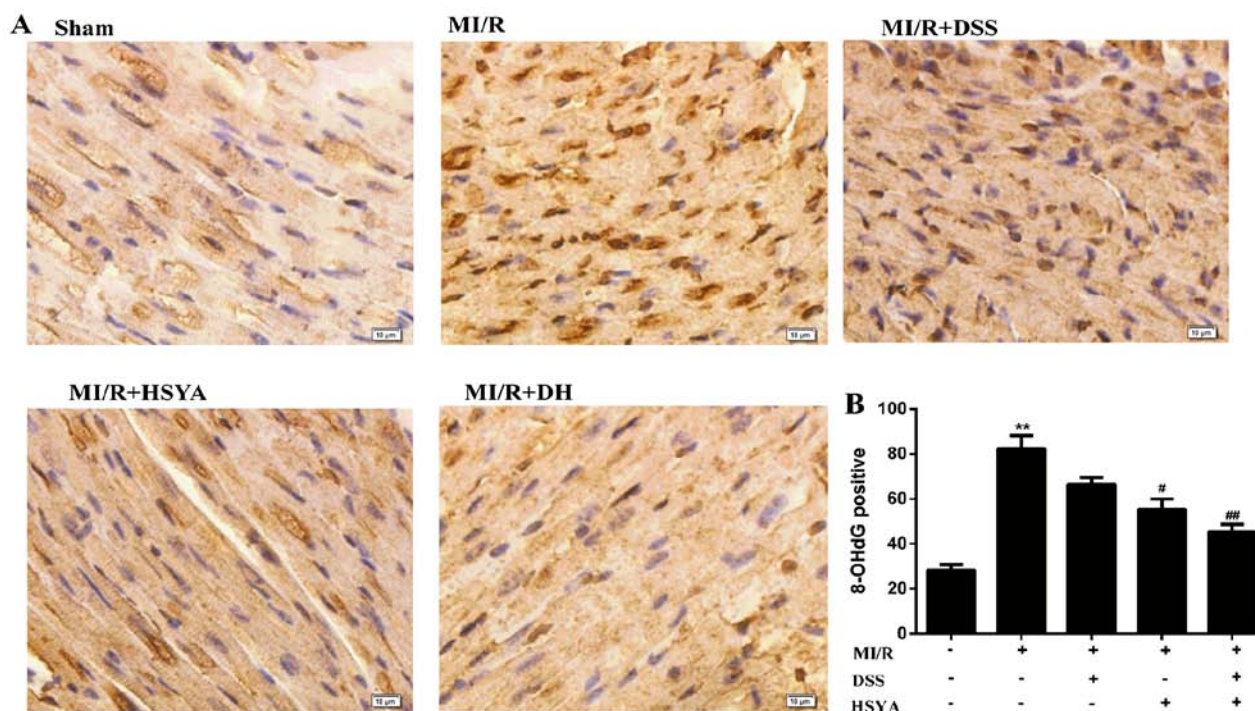


Figure 3. Effects of Danshensu (DSS) and hydroxysafflor yellow A (HSYA) administered individually or in combination evaluated by 8-hydroxydeoxyguanosine (8-OHdG) immunohistochemical staining in the sham-operated (sham), myocardial ischemia-reperfusion (MI/R), MI/R + DSS (60 mg/kg), MI/R + HSYA (70 mg/kg) and MI/R + DSS + HSYA (35+35 mg/kg) groups. (A) Representative images of 8-OHdG staining. (B) The quantitative bar graph was generated from the number of 8-OHdG-positive cells in the myocardium (positive nuclei brown and negative nuclei blue). Values are expressed as the means  $\pm$  SD; scale bar, 20  $\mu$ m ( $n=7-9$  rats in each group). \*\* $P<0.01$  vs. sham group; # $P<0.05$  and ### $P<0.01$  vs. MI/R group.

Table III. Levels of malondialdehyde (MDA) and superoxide dismutase (SOD) activity in the serum of rats.

Treatment	MDA (nmol/ml)	SOD (U/ml)
Sham	7.17 $\pm$ 2.61	186.10 $\pm$ 16.23
MI/R	17.01 $\pm$ 3.58 <sup>a</sup>	69.66 $\pm$ 16.32 <sup>a</sup>
MI/R + DSS	11.26 $\pm$ 2.03 <sup>b</sup>	110.60 $\pm$ 14.75 <sup>b</sup>
MI/R + HSYA	9.05 $\pm$ 2.48 <sup>b</sup>	121.80 $\pm$ 21.44 <sup>b</sup>
MI/R + DH	7.73 $\pm$ 1.62 <sup>c</sup>	152.5 $\pm$ 12.73 <sup>c</sup>

The MDA and SOD activity levels were examined in the serum of rats in the sham-operated, MI/R, MI/R + DSS (60 mg/kg), MI/R + HSYA (70 mg/kg) and MI/R + DSS + HSYA (35+35 mg/kg) groups. Values are expressed as the means  $\pm$  SD ( $n=7-9$  rats in each group). <sup>a</sup> $P<0.01$  vs. sham-operated (sham) group; <sup>b</sup> $P<0.05$  and <sup>c</sup> $P<0.01$  vs. MI/R group. MI/R, myocardial ischemia-reperfusion; HSYA, hydroxysafflor yellow A; DSS, danshensu.

*DSS and HSYA synergistically protect H9c2 cardiomyocytes against injury induced by H/R.* The results of MTT assay demonstrated that H/R significantly reduced ( $P<0.05$  vs. Con) cell viability to 35.26 $\pm$ 6.10%, while DSS and HSYA used alone or in combination protected the H9c2 cardiomyocytes against H/R injury, and the DH group exhibited a significant increase in viability at dose 3 to 68.38 $\pm$ 3.35% ( $P<0.05$  vs. H/R + DSS or H/R + HSYA) (Fig. 4A). The dose-effect curves of DSS and HSYA used alone or in combination were analyzed using the median-effect method to determine whether they acted

Table IV. Number of rats that failed the coronary ligation procedure in each group shown in Fig. 3.

Group	Sham	MI/R	MI/R + DSS	MI/R + HSYA	MI/R + DH
No.	2	2	3	3	1

MI/R, myocardial ischemia-reperfusion; HSYA, hydroxysafflor yellow A; DSS, danshensu; DH, HSYA + DSS.

synergistically to protect the H9c2 cardiomyocytes against H/R injury (Fig. 4B-1). We found that treatment with DSS and HSYA or in combination at concentrations of 1-80  $\mu$ M yielded  $CI_{50}$  values  $<1$  (Fig. 4B-2), indicating synergistic effects between the agents. Since the agents exerted cardioprotective effects in a dose-dependent manner, the highest doses were selected for use in subsequent studies. As an acknowledged marker of cell damage, the LDH levels in the cell supernatant significantly increased ( $P<0.01$  vs. Con) to 91.14 $\pm$ 9.71 U/l in the H/R group. The DSS or HSYA groups exhibited levels of 53.50 $\pm$ 12.54 U/l and 65.11 $\pm$ 5.51 U/l, respectively ( $P<0.05$  vs. H/R). These effects were markedly enhanced in the DH group compared with the H/R group ( $P<0.01$ ; Fig. 4C).

*DSS and HSYA increase HO-1 expression in H9c2 cardiomyocytes.* HO-1 expression was increased in the H/R + DSS group (80  $\mu$ M) and the H/R + HSYA group (80  $\mu$ M). HO-1 expression was further increased in the H/R + DH group (40+40  $\mu$ M) to a level higher than that in the single treat-

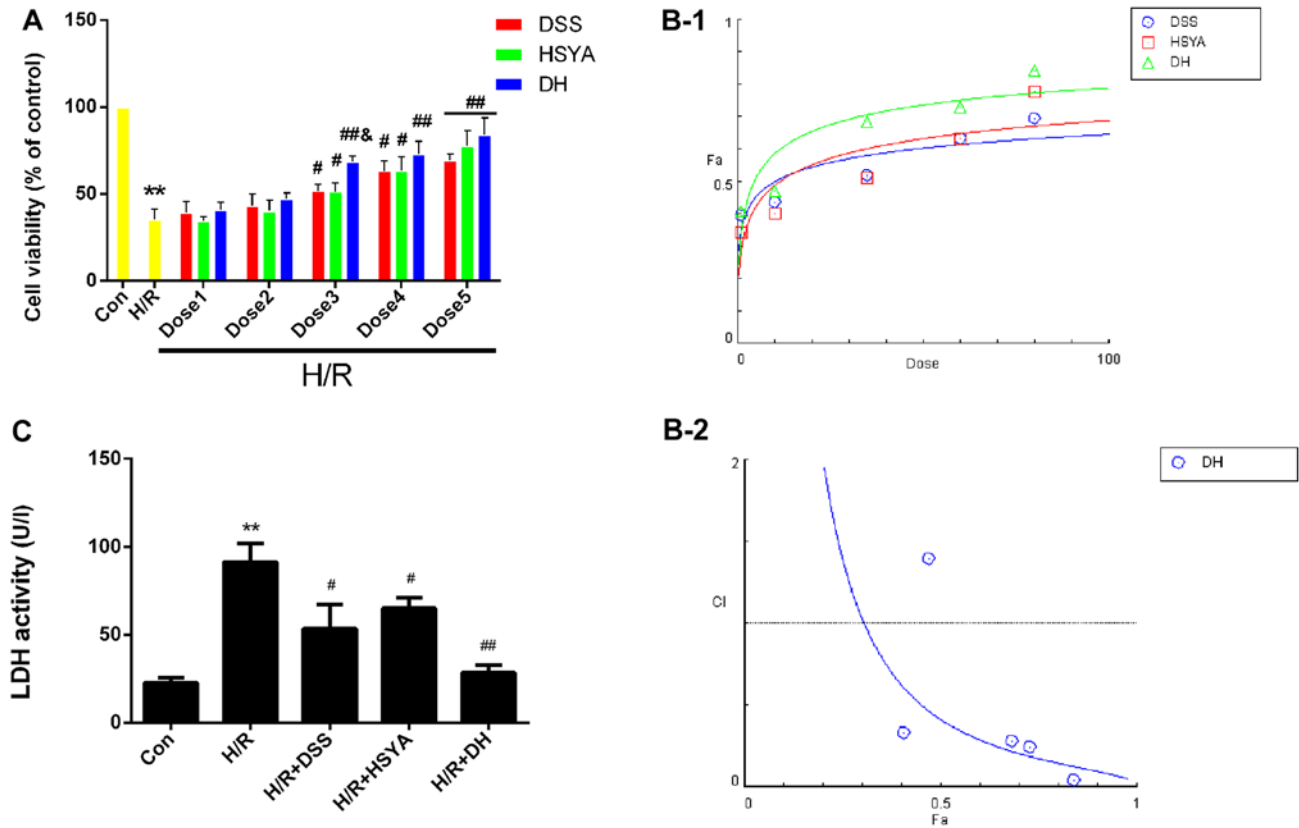


Figure 4. Danshensu (DSS), hydroxysafflor yellow A (HSYA) administered individually or in combination protect the H9c2 cardiomyocytes from hypoxia/reoxygenation (H/R) injury, and in combination, exert a synergistic effect. (A) Cell viability was detected by an MTT reduction assay. H9c2 cardiomyocytes in the control (Con) group were considered 100% viable. Dose 1 indicates DSS (1  $\mu$ M), HSYA (1  $\mu$ M), DH (1  $\mu$ M); Dose 2 indicates DSS (10  $\mu$ M), HSYA (10  $\mu$ M), DH (10  $\mu$ M); Dose 3 indicates DSS (35  $\mu$ M), HSYA (35  $\mu$ M), DH (35  $\mu$ M); Dose 4 indicates DSS (60  $\mu$ M), HSYA (60  $\mu$ M), DH (60  $\mu$ M); Dose 5 indicates DSS (80  $\mu$ M), HSYA (80  $\mu$ M), DH (80  $\mu$ M). (B-1) The dose-effect curves of the single or combined drug treatment. (B-2) The fraction versus combination index (Fa-CI) curve of DSS and HSYA in combination reveals that they exert a synergistic effect ( $CI_{50} < 1$ ) as reflected by the median-effect method. The dashed line at the combination index ( $CI=1$ ) represents the additive effect. (C) The effect of DSS and HSYA on LDH activity in the Con, H/R, H/R + DSS (80  $\mu$ M), H/R + HSYA (80  $\mu$ M) and H/R + DH (40+40  $\mu$ M) groups. Values are expressed as the means  $\pm$  SD (n=6). \*\*P<0.01 vs. Con group; #P<0.05 and ##P<0.01 vs. H/R group; \*P<0.05 vs. H/R + DSS or H/R + HSYA group.

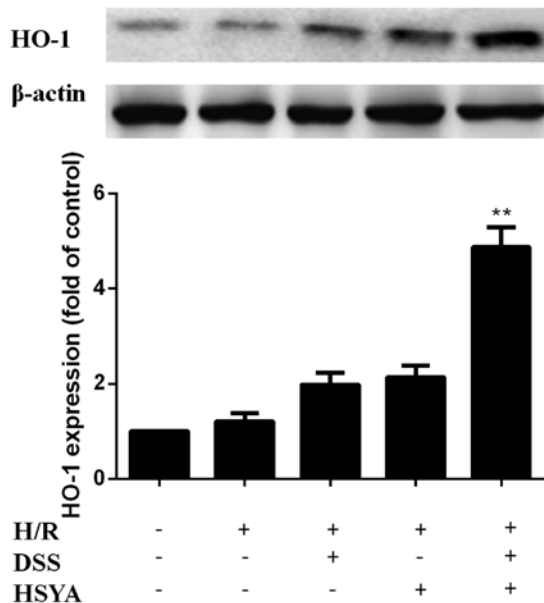


Figure 5. Heme oxygenase-1 (HO-1) expression in the control (Con), hypoxia/reoxygenation (H/R), H/R + Danshensu (DSS; 80  $\mu$ M), H/R + hydroxysafflor yellow A (HSYA; 80  $\mu$ M) and H/R + DSS + HSYA (40+40  $\mu$ M) groups. Data obtained from half-quantitative densitometry are presented as the means  $\pm$  SD of 3 independent experiments. \*\*P<0.01 vs. H/R + DSS or H/R + HSYA group. Note that cropped gel images are used in this figure and the gels were run under the same experimental conditions.

ment groups (P<0.01). There were no significant differences between the H/R group and the groups treated with each agent alone (Fig. 5).

*The antioxidant effects of DSS and HSYA in H9c2 cardiomyocytes are negated by ZnPP-IX.* The analysis of the MDA content and SOD activity revealed that H/R deteriorated the parameters of oxidative stress significantly compared with the control group (P<0.01). The MDA content markedly decreased to  $2.47 \pm 0.44$  nmol/ml in the DSS group and to  $2.05 \pm 0.53$  nmol/ml in the HSYA group (P<0.05 vs. H/R), and the activity of SOD increased to  $11.06 \pm 1.64$  and  $13.53 \pm 2.30$  nmol/ml in the DSS and the HSYA group, respectively. These protective effects were more significantly enhanced in the DH group compared with the H/R group (MDA, P<0.01; SOD, P<0.05). HSYA also exerted a more potent antioxidant effect than DSS. The antioxidant effects were markedly abrogated by ZnPP-IX (P<0.05 vs. H/R + DH) (Table V).

*ZnPP-IX inhibits the anti-apoptotic effect of DSS and HSYA in H9c2 cardiomyocytes.* TUNEL staining of apoptotic cells demonstrated that H/R led to a significant augmentation in cell apoptosis to  $56.41 \pm 14.23\%$

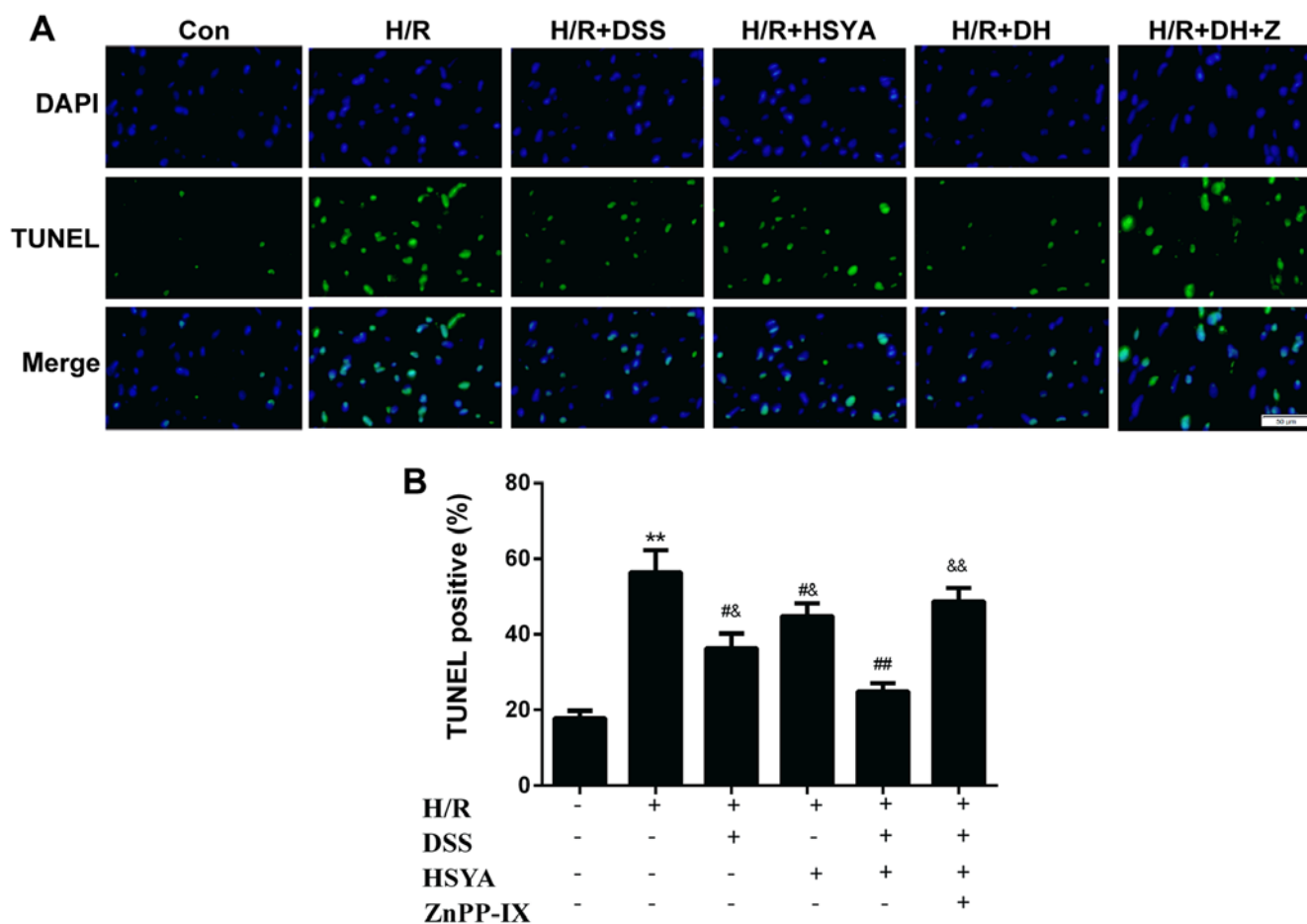


Figure 6. Representative images of the TUNEL staining in the control (Con), hypoxia/reoxygenation (H/R), H/R + Danshensu (DSS; 80  $\mu$ M), H/R + hydroxysafflor yellow A (HSYA; 80  $\mu$ M) and H/R + DSS + HSYA (40+40  $\mu$ M) groups. (A) (Top panel) DAPI staining (blue) indicates total nuclei, (middle panel) apoptotic nuclei detected by TUNEL staining (green), and (bottom panel) overlay of both types of staining. (B) The number of TUNEL-positive myocytes was expressed as a percentage of total nuclei detected by DAPI staining (fluorescence microscopy, magnification,  $\times 40$ , scale bar, 50  $\mu$ m) Data are presented as the means  $\pm$  SD of 3 independent experiments. \*\* $P < 0.01$  vs. Con group; # $P < 0.05$  and ## $P < 0.01$  vs. H/R group; & $P < 0.05$  and && $P < 0.01$  vs. H/R + DH group.

Table V. MDA content and SOD activity in culture supernatants of cardiomyocytes.

Treatment	MDA (nmol/ml)	SOD (U/ml)
Con	1.72 $\pm$ 0.70	17.98 $\pm$ 2.84
H/R	4.01 $\pm$ 0.85 <sup>a</sup>	7.40 $\pm$ 2.90 <sup>a</sup>
H/R + DSS	2.47 $\pm$ 0.44	11.06 $\pm$ 1.64
H/R + HSYA	2.05 $\pm$ 0.53 <sup>b</sup>	13.53 $\pm$ 2.30
H/R + DH	1.74 $\pm$ 0.63 <sup>c</sup>	15.58 $\pm$ 3.04 <sup>b</sup>
H/R + DH + Z	3.62 $\pm$ 0.42 <sup>d</sup>	7.11 $\pm$ 2.64 <sup>d</sup>

The MDA content and SOD activity were calculated in the cardiomyocytes in the Con, H/R, H/R + DSS (80  $\mu$ M), H/R + HSYA (80  $\mu$ M) and H/R + DSS + HSYA (40+40  $\mu$ M) groups. Values are expressed as the means  $\pm$  SD (n=6). <sup>a</sup> $P < 0.01$  vs. Con group; <sup>b</sup> $P < 0.05$  and <sup>c</sup> $P < 0.01$  vs. H/R group; <sup>d</sup> $P < 0.05$  vs. H/R + DH group. H/R, hypoxia/reoxygenation, HSYA, hydroxysafflor yellow A; DSS, danshensu; MDA, malondialdehyde; SOD, superoxide dismutase; Z, zinc protoporphyrin IX; Con, control.

respectively ( $P < 0.05$  vs. H/R). Apoptosis was decreased in the DH group to 24.95 $\pm$ 5.02% ( $P < 0.05$  vs. H/R + DSS or H/R + HSYA). ZnPP-IX abrogated the anti-apoptotic effect of DSS and HSYA ( $P < 0.01$  vs. H/R + DH) (Fig. 6). Following 20 h of reoxygenation, the Bcl-2/Bax ratio decreased to 0.58 $\pm$ 0.34 in the H/R-exposed cardiomyocytes. DSS or HSYA reversed the ratio to 2.86 $\pm$ 0.95 and 1.93 $\pm$ 0.49, respectively ( $P < 0.01$  and  $P < 0.05$  vs. H/R). In addition, treatment with DH increased the Bcl-2/Bax ratio to 4.93 $\pm$ 0.60 ( $P < 0.01$  vs. H/R + DSS or H/R + HSYA). The Bcl-2/Bax ratio was downregulated in the group treated with ZnPP-IX to 1.41 $\pm$ 0.32 ( $P < 0.01$  vs. H/R + DH) (Fig. 7A). H/R evoked a marked increase in the cleaved caspase-3 levels to 10.74 $\pm$ 1.39 ( $P < 0.01$  vs. Con), while DSS or HSYA decreased cleaved caspase-3 expression to 3.74 $\pm$ 1.34 and 5.46 $\pm$ 1.90, respectively ( $P < 0.01$  vs. H/R). Treatment with DH decreased the cleaved caspase-3 level to 1.80 $\pm$ 0.85 ( $P < 0.01$  vs. H/R + DSS or H/R + HSYA), and in the group co-incubated with ZnPP-IX, this effect was abolished (8.87 $\pm$ 2.82;  $P < 0.01$  vs. H/R + DH) (Fig. 7B). Furthermore, we found that DSS exerted a more potent anti-apoptotic effect than HSYA.

( $P < 0.01$  vs. Con). Apoptosis was alleviated in the group treated with DSS or HSYA to 36.38 $\pm$ 9.46 and 44.88 $\pm$ 8.10%,

*DSS and HSYA modulate the activation of the Akt/Nrf2/HO-1 signaling pathway in H9c2 cardiomyocytes.* Following 20 h



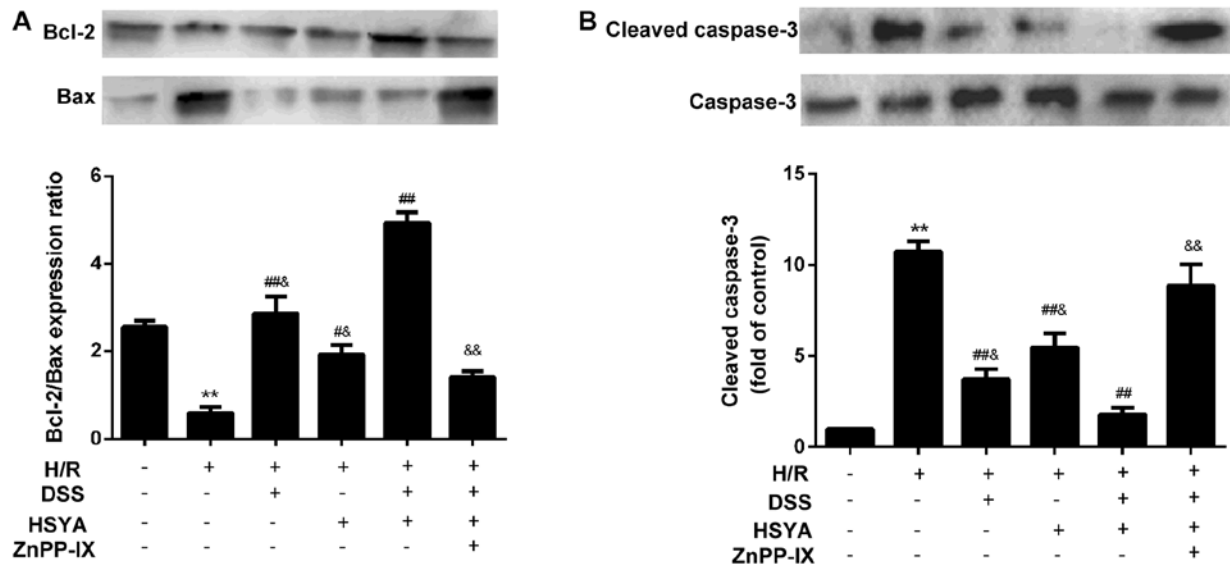


Figure 7. Bcl-2, Bax and cleaved caspase-3 levels in the control (Con), hypoxia/reoxygenation (H/R), H/R + Danshensu (DSS; 80  $\mu$ M), H/R + hydroxysafflor yellow A (HSYA; 80  $\mu$ M), H/R + DSS + HSYA (40+40  $\mu$ M) and H/R + DH + zinc protoporphyrin IX (ZnPP-IX; 10  $\mu$ M) groups. (A) Bcl-2/Bax was calculated and data are presented as the means  $\pm$  SD of 3 independent experiments; (B) cleaved caspase-3 levels obtained from half-quantitative densitometry are presented as the means  $\pm$  SD of 3 independent experiments. \*\* $P$ <0.01 vs. Con group; # $P$ <0.05 and ## $P$ <0.01 vs. H/R group; & $P$ <0.05 and && $P$ <0.01 vs. H/R + DH group. Note that cropped gel images are used in this figure and the gels were run under the same experimental conditions.

of reoxygenation, treatment with DSS or HSYA increased the expression of p-Akt, HO-1 and nuclear Nrf2, and combined treatment exerted even more significant effects than treatment with each agent alone ( $P$ <0.01). LY294002 markedly abolished the effects of DSS and HSYA on p-Akt expression ( $P$ <0.01 vs. H/R + DH), and partly blocked the expression of nuclear Nrf2 and HO-1 ( $P$ <0.05 and  $P$ <0.01 vs. H/R + DH). ZnPP-IX had no significant effect on p-Akt and nuclear Nrf2 expression compared with the H/R + DH group; however, it markedly negated HO-1 expression ( $P$ <0.01 vs. H/R + DH) (Fig. 8).

## Discussion

A number of studies have illustrated the protective effects of DSS or HSYA in cardiovascular diseases (15,16,18); however, limited attention has been paid to their use as a combination therapy and the mechanisms responsible for their combined effects have yet to be elucidated. In the present study, we investigated the synergistic protective effects of DSS and HSYA on myocardial injury through *in vivo* experiments using rats and *in vitro* experiments using H9c2 cardiomyocytes. To the best of our knowledge, this is the first study to demonstrate the role of DSS and HSYA in combination to protect against cellular injury through antioxidant and anti-apoptotic effects, which are associated with the activation of the Akt/Nrf2/HO-1 signaling pathway.

Infarct size is regarded as the gold standard in assessing the severity of MI/R injury (25). Thus, the first investigations of the combination effects examined infarct size. Following the administration of a series of doses of DSS and/or HSYA, the infarct size was reduced in a dose-dependent manner. The  $CI_{50}$  value of the infarct size (<1) determined by the median-effect method verified the combination effects to be synergistic between the agents. Additionally, the sensitive cardiac injury markers (26-28) CK-MB and cTnI, were measured to determine

whether DSS and HSYA are capable of alleviating the degree of myocardial injury caused by MI/R. The decreased release of CK-MB and cTnI in the treatment groups demonstrated the protective effects of DSS and HSYA. Furthermore, combined treatment exerted a more potent protective effect compared to treatment with either agent alone. To further confirm the combination effects, an MTT assay was performed. The  $CI_{50}$  value of cell viability was revealed to be <1, indicating a synergistic effect between DSS and HSYA. LDH is one of the specific enzymes (29) present in the myocardial cytoplasm, and its values may indirectly reflect the degree of damage of the myocardium exposed to H/R. In the present study, the LDH levels in the cell supernatant were significantly decreased following treatment with DH. These results indicated that DSS and HSYA used in combination exerted a synergistic cardioprotective effect *in vitro* and *in vivo*.

Having determined that DSS and HSYA exerted a synergistic cardioprotective effect, we then proceeded to elucidate the possible mechanisms responsible for these effects. The increased production of oxygen-free radicals in conjunction with the decreased activity of antioxidant defenses are considered to be a significant factor for myocardial reperfusion injury (30). SOD is the first line of cellular defense against oxidative injury, which decomposes  $O_2$  and  $H_2O_2$  before they interact to form the more reactive hydroxyl radical (31,32). MDA is a product of lipid peroxidation that may cause the crosslinking polymerization of proteins, nucleic acids and some macromolecules, and thus, it has been found that the amount of MDA often reflects the degree of lipid peroxidation (33). 8-OHdG appears to be a sensitive and integral marker of oxidative damage to DNA due to any imbalance between  $OH^{\bullet}$  generation, antioxidant defenses and the repair of damaged DNA sequences (34). Using these three important markers of oxidative stress, we confirmed that DSS and HSYA significantly alleviated the parameters of oxidative stress and HSYA exerted a profound protective effect

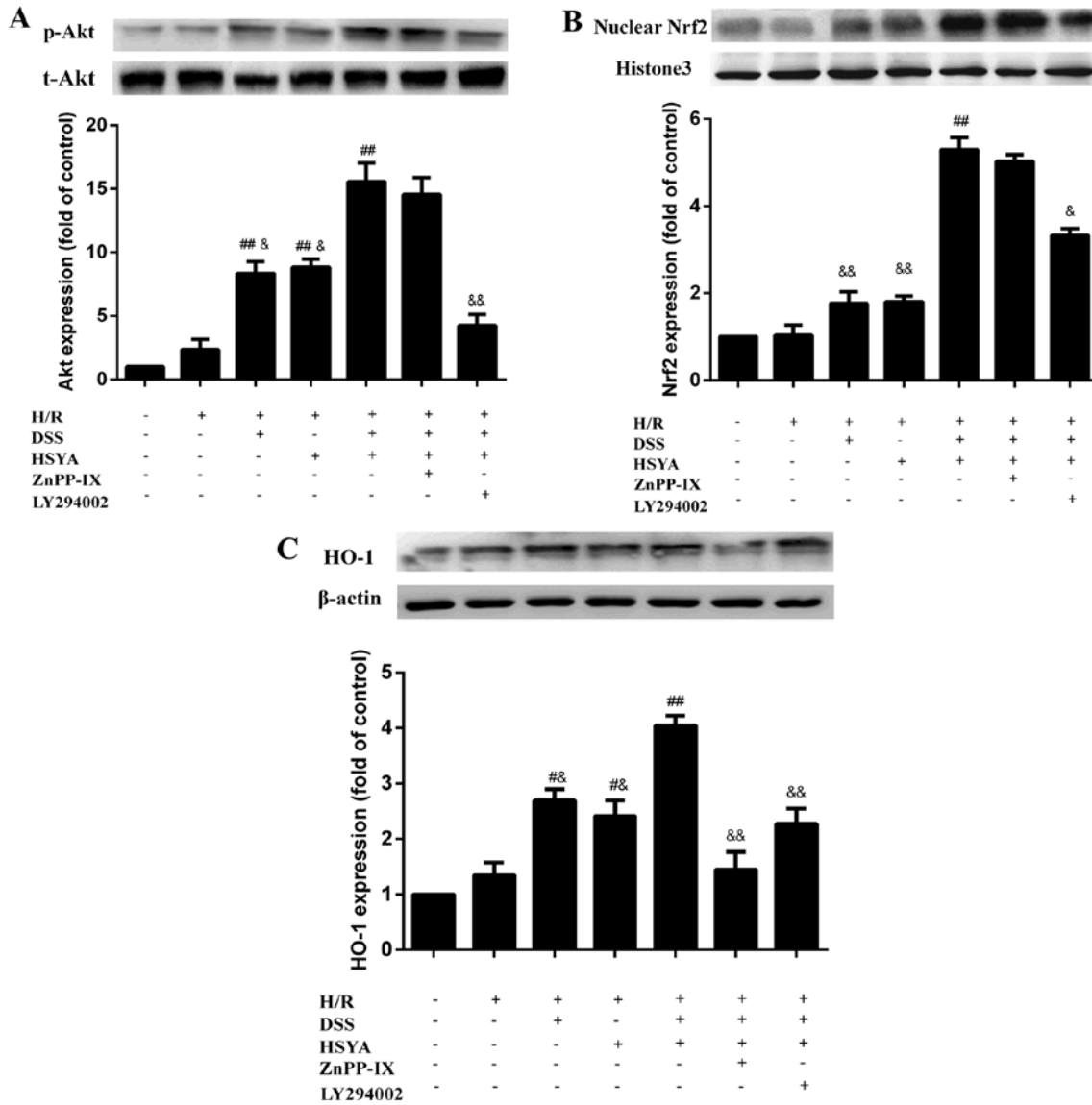


Figure 8. (A) Protein kinase B (Akt), (B) nuclear factor erythroid 2-related factor 2 (Nrf2) and (C) heme oxygenase-1 (HO-1) levels in the control (Con), hypoxia/reoxygenation (H/R), H/R + Danshensu (DSS; 80  $\mu$ M), H/R + hydroxysafflor yellow A (HSYA; 80  $\mu$ M), H/R + DSS + HSYA (40+40  $\mu$ M), H/R + DH + zinc protoporphyrin IX (ZnPP-IX; 10  $\mu$ M) and H/R + DH + LY294002 (50  $\mu$ M) groups. Data obtained from half-quantitative densitometry are presented as the means  $\pm$  SD of 3 independent experiments. <sup>#</sup>P<0.05 and <sup>##</sup>P<0.01 vs. H/R group; <sup>&</sup>P<0.05 and <sup>&&</sup>P<0.01 vs. H/R + DH group. Note that cropped gel images are used in this figure and the gels were run under the same experimental conditions. t-AKT, total AKT; p-AKT, phospho-AKT.

*in vitro* and *in vivo*. This finding indicated that DSS and HSYA played a pivotal role in ameliorating the oxidative stress injury caused by MI/R or H/R.

HO-1 is an inducible enzyme with potent antioxidant activities due to its ability to degrade heme into biliverdin or bilirubin, carbon monoxide and free iron (35,36). Furthermore, it has been demonstrated that plant-derived chemical substances may act as inducers of the HO-1, and these substances have been found to exert cardioprotective effects against oxidative injury (37). Thus, we hypothesized that DSS and HSYA may protect myocardial tissue and H9c2 cardiomyocytes from oxidative stress-induced injury in association with the upregulation of HO-1. Western blot analysis revealed that HO-1 expression was profoundly enhanced in the combined treatment group compared with the groups treated with either agent alone. This result not only verified the above-mentioned hypothesis, but also indicated that combination therapy may be more effective

in increasing HO-1 expression than monotherapy. It has been noted that ZnPP-IX, which is a competitive inhibitor of the HO-1 enzyme, conjugates to HO-1 and reduces the conjugation between HO-1 and heme (38,39). Following treatment with ZnPP-IX (10  $\mu$ M), the decreased MDA content and the increased activity of SOD were both revoked. These results illustrated that DSS and HSYA upregulated HO-1 expression and that the antioxidant effects were partly dependent on HO-1 expression.

Apoptosis plays an important role in the loss of cardiomyocytes in a variety of pathologies, including H/R injury (18,40). It has been demonstrated that the mitochondrial apoptotic pathway may be activated when a number of pro-apoptotic factors are released (41). Bax is a pro-apoptotic protein, whereas Bcl-2 is an anti-apoptotic protein. It has been noted that the ratio of Bcl-2 to Bax acts as an essential element in determining the threshold of apoptosis (18,42). Caspases transduce and execute

apoptotic signaling. Caspase-3 has been found to be activated by the apoptotic pathway and then autocatalyzed, and processed into activated fragments such as cleaved caspase-3, which are considered as an index of apoptosis (43). In the present study, DSS and HSYA increased the Bcl-2/Bax ratio in the mitochondria, downregulated cleaved caspase-3 expression and reduced TUNEL staining compared with the H/R group. DH therapy enhanced the effect to a greater extent compared with DSS or HSYA alone. The anti-apoptotic effects were markedly negated by ZnPP-IX. Additionally, DSS exerted a more potent anti-apoptotic effect than HSYA. These results confirmed that the overexpression of HO-1 induced by the agents and the combined regimen led to an enhanced anti-apoptotic effect.

It has been proven that a specific molecular pathway is important for compound-induced HO-1 expression. The PI3K/Akt pathway has been demonstrated to be associated with increased HO-1 expression (44). In this study, Akt phosphorylation was significantly enhanced in the H9c2 cardiomyocytes treated with DSS and HSYA compared with that observed in the H/R group. The Akt inhibitor, LY294002, significantly blocked this effect, while ZnPP-IX had no obvious impact on Akt phosphorylation induced by the two agents. Nrf2 is a key regulator of HO-1 expression in cells (45). It has been demonstrated that the nuclear factor Nrf2 binds to Kelch-like ECH-associated protein-1 (Keap1) to form the Keap1-Nrf2 complex and limit Nrf2-mediated gene expression in the cytoplasm under normal physiological circumstances. Nrf2 is released from Keap1 under conditions of oxidative stress or other potential damage and then translocates to the nucleus, where it binds to antioxidant response element (ARE) sequences, leading to the transcriptional activation of anti-apoptotic genes, including HO-1 (18,46-49). In the present study, the two agents enhanced the expression of nuclear Nrf2. LY294002 partly blocked the increase in Nrf2 translocation to the nucleus and the upregulation of HO-1 expression, while ZnPP-IX had no notable effect on nuclear Nrf2 expression induced by DSS and HSYA. These results indicated that the DSS- and HSYA-mediated upregulation of Akt phosphorylation and nuclear Nrf2 did not rely on the upregulation of HO-1 expression. On the contrary, the upregulation of HO-1 expression was dependent on Akt phosphorylation. Akt phosphorylation, as well as nuclear Nrf2 and HO-1 expression were enhanced in the DH-treated group compared with that observed in the groups treated with either agent alone. As we confirmed above, treatment with DH exerted a synergistic effect. However, it would be interesting to determine the possible mechanisms underlying these synergistic effects. It has been pointed out that each single constituent of a combination affects several targets, such as enzymes, substrates, receptors and transport proteins (8,50). Most notably, DSS mainly exerted anti-apoptotic effects, while HSYA exerted significant antioxidant effects. They acted on apoptosis-related proteins and oxidative-related enzymes, respectively; thus, in combination they exerted complementary and synergistic effects. Our results revealed that the combination of DSS and HSYA conferred a synergistic effect on the activation of the Akt/Nrf2/HO-1 pathway, which is a potential mechanism for enhancing the anti-apoptotic and antioxidant effects.

In the present study, we only investigated the antioxidant and anti-apoptotic effects involving the Akt/Nrf2/HO-1

pathway. It would be of interest to determine whether other aspects contribute to these synergistic effects. Further studies are warranted to clarify the synergistic effects of DSS and HSYA on the regulation of other pathways.

In conclusion, we found that DSS and HSYA acted synergistically to significantly attenuate myocardial injury *in vitro* and *in vivo* by exerting antioxidant and anti-apoptotic effects through the Akt/Nrf2/HO-1 signaling pathway. The results from the present study provide insight into the effects and the mechanisms responsible for these synergistic effects. This may lead to the development of effective combined therapeutic TCM regimens so as to combat myocardial complications in clinical practice.

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