

RESEARCH

Open Access



Ketogenic diet in a patient with congenital hyperinsulinism: a novel approach to prevent brain damage

Arianna Maiorana^{1*}, Lucilla Manganozzi¹, Fabrizio Barbetti², Silvia Bernabei³, Giorgia Gallo¹, Raffaella Cusmai⁴, Stefania Caviglia⁵ and Carlo Dionisi-Vici¹

Abstract

Background: Congenital hyperinsulinism (CHI) is the most frequent cause of hypoglycemia in children. In addition to increased peripheral glucose utilization, dysregulated insulin secretion induces profound hypoglycemia and neuroglycopenia by inhibiting glycogenolysis, gluconeogenesis and lipolysis. This results in the shortage of all cerebral energy substrates (glucose, lactate and ketones), and can lead to severe neurological sequelae. Patients with CHI unresponsive to medical treatment can be subjected to near-total pancreatectomy with increased risk of secondary diabetes. Ketogenic diet (KD), by reproducing a fasting-like condition in which body fuel mainly derives from beta-oxidation, is intended to provide alternative cerebral substrates such ketone bodies. We took advantage of known protective effect of KD on neuronal damage associated with GLUT1 deficiency, a disorder of impaired glucose transport across the blood-brain barrier, and administered KD in a patient with drug-unresponsive CHI, with the aim of providing to neurons an energy source alternative to glucose.

Methods: A child with drug-resistant, long-standing CHI caused by a spontaneous *GCK* activating mutation (p.Val455Met) suffered from epilepsy and showed neurodevelopmental abnormalities. After attempting various therapeutic regimes without success, near-total pancreatectomy was suggested to parents, who asked for other options. Therefore, we proposed KD in combination with insulin-suppressing drugs.

Results: We administered KD for 2 years. Soon after the first six months, the patient was free of epileptic crises, presented normalization of EEG, and showed a marked recover in psychological development and quality of life.

Conclusions: KD could represent an effective treatment to support brain function in selected cases of CHI.

Keywords: Congenital hyperinsulinism, Ketogenic diet, Hypoglycemia, Epilepsy, Neurodevelopment

Background

Congenital hyperinsulinism (CHI) is the most frequent cause of hypoglycemia in children [1, 2]. Mutations that affect insulin secretion regulation by the three main classes of energy substrates, i.e. glucose, aminoacids, and free fatty acids (FFA) [2, 3], can cause CHI that requires rapid diagnosis and treatment to limit/avoid neuronal damage [4] and the irreversible neurological sequelae consequent to prolonged, severe hypoglycemia. Neurons in the superficial layers of cerebral cortex and hippocampus

are those preferentially affected by lack of glucose, followed by neurons in basal ganglia and thalamus [5]. However, mild, recurrent hypoglycemia can cause hippocampal synaptic dysfunction even in absence of neuronal damage [6, 7]. These experimental findings explain why memory, learning, intelligence and attention are the cognitive domains most vulnerable to hypoglycemia in children with type 1 diabetes [8, 9]. Although glucose is the main energy source for neurons, human brain can also utilize ketone bodies from FFA, lactate, pyruvate, glycerol and some aminoacids, as alternative substrate [10]. The protective effect of ketone bodies on hypoglycemia-induced neuronal damage has been demonstrated in animal studies [11, 12] and also in patients

* Correspondence: arianna.maiorana@opbg.net

¹Metabolic Unit, Department of Pediatric Medicine, Bambino Gesù Children's Hospital, piazza S. Onofrio 4, 00165 Rome, Italy

Full list of author information is available at the end of the article

with type 1 diabetes, in whom the ingestion of medium-chain triglycerides prevented the cognitive deficit induced by hypoglycemia by elevating blood levels of 3-hydroxybutyrate [13]. Ketogenic diet (KD), which provides FFA as alternative fuel to carbohydrates for neuronal energy metabolism, has therefore a strong potential neuroprotective effect. The main indication of KD in children is the treatment of refractory epilepsy, but it is also the causal therapy of GLUT1 deficiency, a metabolic disorder characterized by epilepsy, developmental delay and movement disorders [14, 15]. In GLUT1 deficiency, neuroglycopenia that ensues as consequence of the impaired glucose transport across the blood-brain barrier [14, 15] is effectively improved by KD that provides ketone bodies as alternative energy source for the brain. In CHI, excessive insulin secretion not only induces severe neuroglycopenia, but also halts, by inhibiting gluconeogenesis, glycogenolysis and lipolysis, the use of other metabolic pathways that provide energetic substrates to the neurons. Other inherited metabolic diseases, such as mitochondrial fatty oxidation defects, share the same neurological risk of hypoglycemia because of lack of ketones [16]. Therefore, developing brain of patients with CHI is more vulnerable than other forms of hypoglycemia. Based on the similarities of brain metabolism perturbation shared by GLUT1 deficiency and CHI, we attempted to tackle neuroglycopenic symptoms and outcome by administering KD in a patient with severe, drug-resistant form of CHI.

Methods

We report a case of severe persistent hyperinsulinemic hypoglycemia due to a “de novo” mutation in *GCK*. The girl presented with hypoglycemic seizures since the first years of life. Family history was positive for type 2 diabetes. From the age of 2 years, she was admitted to the hospital for recurrent episodes of severe hypoglycemia (up to 1.5 mmol/L, normal value 3.9–5.5 mmol/L [17]) associated to cold sweating, seizures and hypotonia. At the age of 3 years laboratory tests were performed and showed hypoglycemia with hyperinsulinemia (blood glucose 1.95–2.3 mmol/L; plasma insulin 5.5–10.2 μ UI/ml). EEG showed abnormal waves in the left hemisphere and epileptogenic abnormalities, and treatment with valproate was started. Genetic investigation for *ABCC8* and *KNCJ11* performed elsewhere was negative. At 9 years of age the patient was referred to our hospital, and the heterozygous p.Val455Met mutation was found at *GCK* gene sequencing [18]; parents showed wild type *GCK* sequence. At admission, the child was receiving a combination of high-dose diazoxide (12 mg/kg/day) and octreotide (15 μ g/kg/day), with no apparent clinical response. Previously, a trial with nifedipine (up to 0.75 mg/kg/day) had been attempted without benefit. We gradually increased

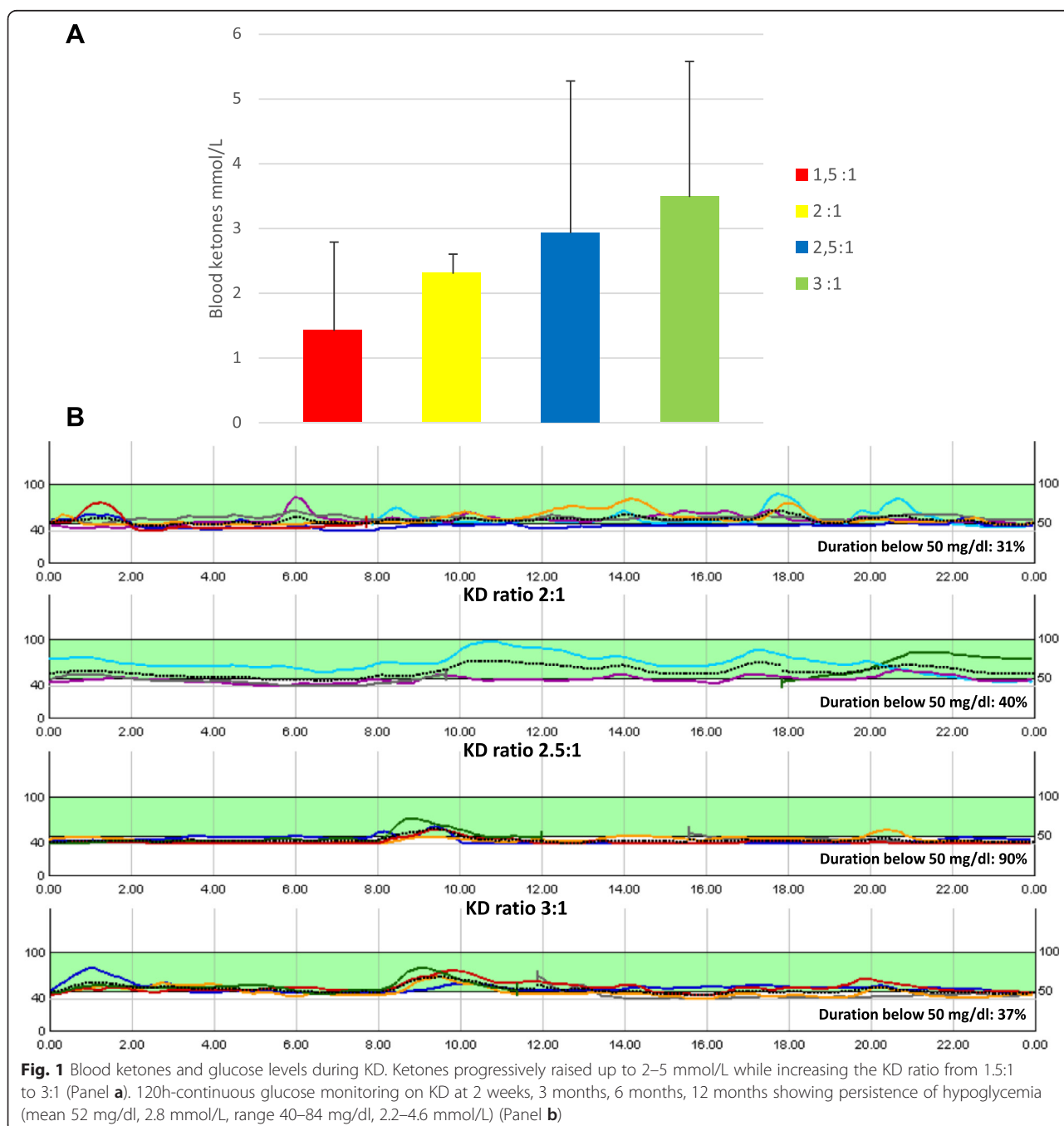
octreotide to 50 μ g/kg/day in combination with diazoxide (10 mg/kg/day) but observed persistence of hypoglycemia and epilepsy. A further association with slow-release carbohydrate to drugs did not elicit any clinical improvement, and the patient continued to present hypoglycemic episodes (0.5–1.6 mmol/L), seizures and absence epilepsy regardless of glycemic values, that required frequent hospitalizations. Despite a mild improvement of neurologic symptoms after switch from valproate therapy to ethosuccimide, absence epilepsy and evident EEG abnormalities persisted, even in the intercritical phases; her intellectual function was borderline. Furthermore, at the age of 10 years the patient quickly gained 11.5 kg within 12 months, becoming mildly obese (BMI z-score: >97th centile). Overall, her quality of life was very poor. The lack of response to drug therapy with risk of permanent and severe brain sequelae made us to consider a near-total pancreatectomy, that was discussed with parents with the warning of no guarantee to achieve normoglycemia and of the increased hazard of secondary diabetes. At parents' request to avoid surgery, we then proposed a trial with KD, explaining that it was aimed to prevent neuroglycopenic epilepsy and to improve neurological status by providing ketone bodies as an alternative energy source for neurons, as seen in GLUT1 deficiency. Consent was obtained from patient and parents after full explanation of the objective. With the new dietary regimen we provided 85 % of energy from lipids, 8 % from proteins, and 7 % from glucose of a normocaloric diet divided in 5 meals. Determination of blood ketones (Nova StatStrips[®] Glucose Ketone Meter, Nova Medical, Menarini Diagnostics, Florence, Italy) in the morning (after overnight fast of 10–12 h) and in the evening (after dinner) was utilized as guidance to establish the ratio of lipids: proteins plus carbohydrates. The ratio was progressively increased every three months from 1.5:1 to 3:1 during the first year of KD in order to obtain a blood ketone concentration close to 4 mmol/L [19]. First assessment of the KD outcome was performed as inpatient; auxological evaluation, hepatic and renal function, plasma glucose, insulin, lactate, FFA, blood ketones and lipid profile, along with renal tubular function and abdomen ultrasound were assessed every 6 months. Follow-up included 96 to 120-h long continue glucose monitoring (CGM- ipro2, Medtronic; data analysis by MiniMed software, Medtronic, MiniMed, Northridge, CA, USA[®]) [20] with EEG performed simultaneously. Weschsler Intelligence scale for Children (WISC III: third edition) and Vineland Adaptive Behavior Scale were serially administered to evaluate cognitive and adaptive abilities.

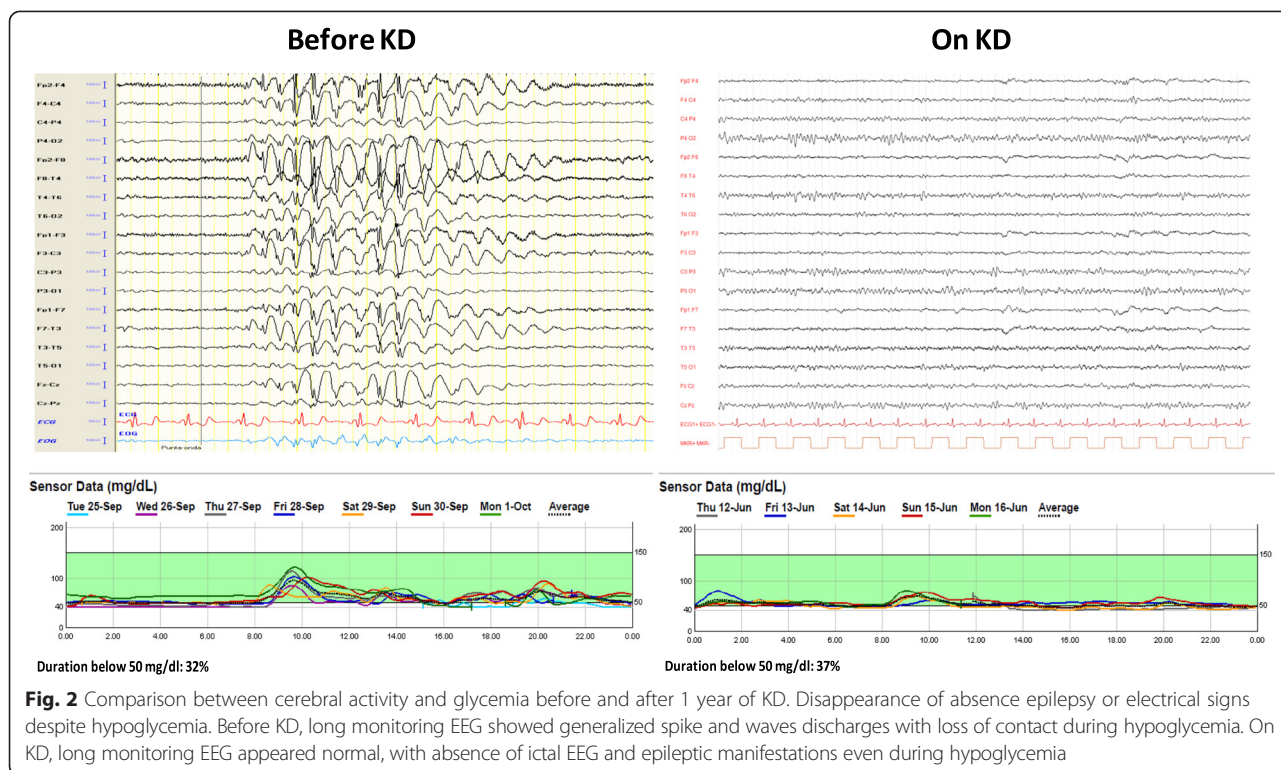
Results and discussion

While on KD the patient continued diazoxide and octreotide treatment (10 mg/kg/day and 35 μ g/kg/day, respectively). Six months after KD was started, maintenance of

blood ketones between 2–5 mmol/L (Fig. 1, panel a) fully resolved neuroglycopenic signs with parallel disappearance of both epileptic crisis and absence epilepsy, despite blood glucose levels permanently below 5.5 mmol/L even after meal, and close to 2.2–2.7 mmol/L most of time (Fig. 1, panel b). EEG improved and became normal within the first year on KD, showing no alteration even during episodes of hypoglycemia (Fig. 2). During the first 6 months of KD the patient lost 9 kg and her BMI

normalized. Psychological evaluation revealed a strengthening of social, cognitive and verbal capacities (Fig. 3). The child and her family reported an improvement of physical and psychosocial well-being, reduction of fear of hypoglycemic symptoms and awareness of a lower risk of neurological injury, with an overall amelioration of the quality of life related to the management of disease. Diazoxide was discontinued, and currently the patient is given octreotide, reduced to 25 µg/kg/day, without

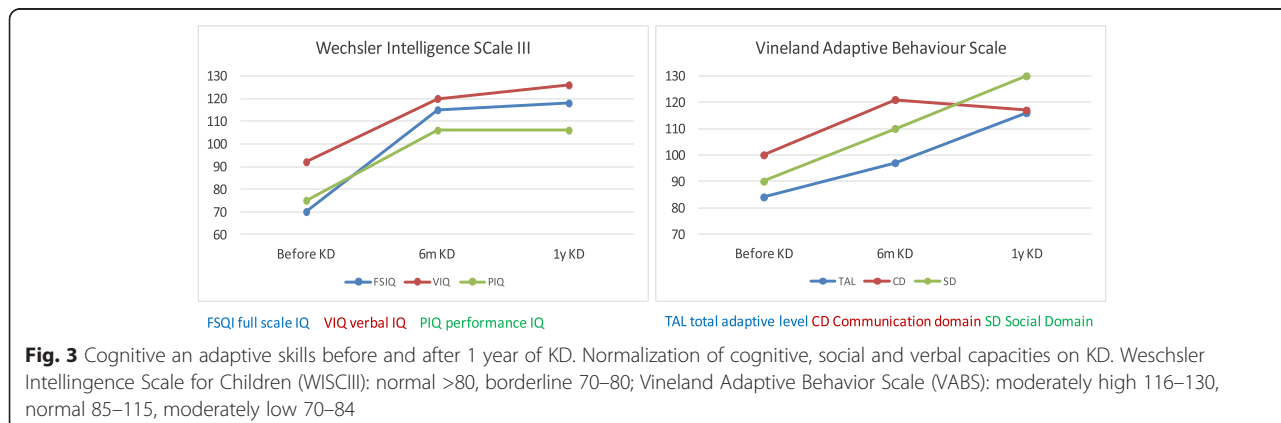




any neuroglycopenic symptoms. KD was well tolerated over a period of 24 months, with no side-effects and no changes in laboratory tests.

GCK activation caused by gain-of-function mutations determines an increased affinity for glucose and lowers the glucose thresholds for insulin release in the pancreatic beta cell, giving rise to CHI [18, 20–23]. Most patients bearing a *GCK* mutation have relatively mild disease, easily managed with diazoxide, but a few cases with a severe phenotype, who were subjected to near-total pancreatectomy to avoid neurological sequelae, have been described [24, 25]. Although V455M mutation has been previously associated with a mild phenotype [18], our patient displayed a drug-resistant form, with a

steep increase of BMI in peripubertal age and a clinical course resembling that described in a patient with the *GCK/A456V* mutation [26]. Prompted by patient’s long-standing symptoms, drug resistance and the preference of parents to avoid demolitive surgery, we opted for a KD trial to provide an energy source to central nervous system (CNS) alternative to glucose, on the bases of its proven efficacy for the treatment of GLUT1 deficiency [14]. We thus challenged the central tenet of treatment of CHI patients, which is to maintain normoglycemia to avoid irreversible brain damage [4]. As a matter of fact, neurological sequelae are more prevalent in CHI than in other forms of hypoglycemia because of the inhibition of lipolysis and ketogenesis by inappropriately high insulin



levels [16]. Ketogenic diet, by restricting the amount of carbohydrates along with a significant increase of fat intake, reproduces a fasting-like condition in which metabolism is shifted from glycolysis to beta-oxidation of FFA and ketone bodies formation for ATP synthesis [27]. In our patient we observed that after the beginning of KD neurological symptoms, psychological development and epilepsy manifestations all improved, despite the persistence of hyperinsulinemic hypoglycemia. The neuroprotective effect of ketone bodies has been associated with the activation of a several endogenous metabolic and genetic programs that stabilize and/or enhance cellular metabolism [27], increasing cerebral ATP and reducing neuronal excitability. In addition, KD decreases mTOR activity in neuronal cells, thus conferring anticonvulsant effect [28]. Of note, KD reverted EEG pathologic patterns in our patient even during hypoglycemia (Fig. 2). This result highlights the protective effect of KD on CNS despite persistence of neuroglycopenia. Cognitive and adaptive development also improved after the first six months of KD, further confirming its positive effect. Bearing in mind that ketone bodies have additional neuroprotective effects acting as ROS scavengers [11], and by inducing PPAR α and UCP2 expression [29], and inhibiting glutamate uptake into synaptic vesicles [30], we hypothesize that KD might have prolonged positive effects on brain function.

CNS protective action of 3-hydroxybutyrate has been successfully exploited to treat hypoglycemic coma in rats [31] and multiple acyl-CoA dehydrogenase deficiency in humans [32]. Consequently, we suggest that the utilization of 3-hydroxybutyrate may be a safer option and an area of investigation in specific patients, such neonates with CHI, in whom administration of KD might be hazardous.

Conclusions

The clinical and neurological improvement observed in our patient with CHI suggests that KD could have a neuroprotective effect despite persistence of neuroglycopenia. Additional studies are needed to confirm the efficacy of this novel therapeutic approach in selected CHI cases to support brain function by providing an alternative energy source to CNS.

Abbreviations

CHI: Congenital hyperinsulinism; KD: Ketogenic diet; GCK: Glucokinase; GLUT1: Glucose transporter 1; EEG: Electroencephalography; FFA: Free fatty acids; ROS: Radical oxygen species; CNS: Central nervous system.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AM conceptualized and designed the study, collected, coordinated and supervised data collection and analysis, drafted the initial manuscript, and approved the final manuscript as submitted. LM contributes to data collection, drafted the initial manuscript, reviewed and revised the

manuscript, and approved the final manuscript as submitted. FB carried out genetic analysis, drafted the initial manuscript, critically revised the manuscript and approved the final manuscript as submitted. SB and GG designed the nutritional regimen, drafted the initial manuscript, and approved the final manuscript as submitted. RC performed the neurophysiological studies, drafted the initial manuscript, and approved the final manuscript as submitted. SC performed cognitive evaluation, drafted the initial manuscript, and approved the final manuscript as submitted. CDV conceptualized the study, gave a substantial contribution in data analysis, drafted the initial manuscript, critically reviewed the manuscript and approved the final manuscript as submitted. All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work. Questions related to the accuracy or integrity of any part of the work were appropriately investigated and resolved from all authors.

Author details

¹Metabolic Unit, Department of Pediatric Medicine, Bambino Gesù Children's Hospital, piazza S. Onofrio 4, 00165 Rome, Italy. ²Department of Experimental Medicine, University of Tor Vergata and Bambino Gesù Children's Hospital, Rome, Italy. ³Clinical Nutrition, Gastroenterology Department, Bambino Gesù Children's Hospital, Rome, Italy. ⁴Neurology, Neuroscience Department, Bambino Gesù Children's Hospital, Rome, Italy. ⁵Psychology Unit, Neuroscience Department, Bambino Gesù Children's Hospital, Rome, Italy.

Received: 23 June 2015 Accepted: 10 September 2015

Published online: 24 September 2015

References

1. Arnoux JB, de Lonlay P, Ribeiro MJ, Hussain K, Blankenstein O, Mohnike K, et al. Congenital hyperinsulinism. *Early Hum Dev.* 2010;86(5):287–94.
2. Hussain K, De Lonlay P. Hyperinsulinism. In: Blau N, Duran M, Gibson KM, Dionisi-Vici C, editors. *Physician's Guide to the Diagnosis, Treatment, and Follow-Up of Inherited Metabolic Diseases.* Berlin Heidelberg: Springer-Verlag; 2014. p. 323–36.
3. Li C, Buettger C, Kwagh J, Matter A, Daikhin Y, Nissim IB, et al. A signaling role of glutamine in insulin secretion. *J Biol Chem.* 2004;279(14):13393–401.
4. Menni F, de Lonlay P, Sevin C, Touati G, Peigne C, Barbier V, et al. Neurologic outcomes of 90 neonates and infants with persistent hyperinsulinemic hypoglycemia. *Pediatrics.* 2001;107(3):476–9.
5. Auer RN, Hugh J, Cosgrove E, Curry B. Neuropathologic findings in three cases of profound hypoglycemia. *Clin Neuropathol.* 1989;8(2):63–8.
6. McNay EC, Williamson A, McCrimmon RJ, Sherwin RS. Cognitive and neural hippocampal effects of long-term moderate recurrent hypoglycemia. *Diabetes.* 2006;55(4):1088–95.
7. Yamada KA, Rensing N, Izumi Y, De Erausquin GA, Gazit V, Dorsey DA, et al. Repetitive hypoglycemia in young rats impairs hippocampal long-term potentiation. *Pediatr Res.* 2004;55(3):372–9.
8. Ryan C, Gurtunca N, Becker D. Hypoglycemia: a complication of diabetes therapy in children. *Pediatr Clin North Am.* 2005;52(6):1705–33.
9. Bassetti A, Chiuri RM, Tocco AM, Di Giulio C, Mattei PA, Ballone E, et al. The effect of recurrent severe hypoglycemia on cognitive performance in children with type 1 diabetes: a meta-analysis. *J Child Neurol.* 2011;26(11):1383–91.
10. Suh SW, Hamby AM, Swanson RA. Hypoglycemia, brain energetics, and hypoglycemic neuronal death. *Glia.* 2007;55(12):1280–6.
11. Haces ML, Hernandez-Fonseca K, Medina-Campos ON, Montiel T, Pedraza-Chaverri J, Massieu L. Antioxidant capacity contributes to protection of ketone bodies against oxidative damage induced during hypoglycemic conditions. *Exp Neurol.* 2008;211(1):85–96.
12. Yamada KA, Rensing N, Thio LL. Ketogenic diet reduces hypoglycemia-induced neuronal death in young rats. *Neurosci Lett.* 2005;385(3):210–4.
13. Page KA, Williamson A, Yu N, McNay EC, Dzuzira J, McCrimmon RJ, et al. Medium-chain fatty acids improve cognitive function in intensively treated type 1 diabetic patients and support in vitro synaptic transmission during acute hypoglycemia. *Diabetes.* 2009;58(5):1237–44.
14. Veggiotti P, De Giorgis V. Dietary Treatments and New Therapeutic Perspective in GLUT1 Deficiency Syndrome. *Curr Treat Options Neurol.* 2014;16(5):291.
15. Pearson TS, Akman C, Hinton VJ, Engelstad K, De Vivo DC. Phenotypic spectrum of glucose transporter type 1 deficiency syndrome (Glut1 DS). *Curr Neurol Neurosci Rep.* 2013;13(4):342.

16. Gataullina S, Dellatolas G, Perdry H, Robert JJ, Valayannopoulos V, Touati G, et al. Comorbidity and metabolic context are crucial factors determining neurological sequelae of hypoglycaemia. *Dev Med Child Neurol*. 2012;54(11):1012–7.
17. Thornton PS, Stanley CA, De Leon DD, Harris D, Haymond MW, Hussain K, et al. Recommendations from the Pediatric Endocrine Society for Evaluation and Management of Persistent Hypoglycemia in Neonates, Infants, and Children. *J Pediatr*. 2015;167(2):238–45.
18. Glaser B, Kesavan P, Heyman M, Davis E, Cuesta A, Buchs A, et al. Familial hyperinsulinism caused by an activating glucokinase mutation. *N Engl J Med*. 1998;338(4):226–30.
19. Gilbert DL, Pyzik PL, Freeman JM. The ketogenic diet: seizure control correlates better with serum beta-hydroxybutyrate than with urine ketones. *J Child Neurol*. 2000;15(12):787–90.
20. Maiorana A, Barbetti F, Boiani A, Rufini V, Pizzoferro M, Francalanci P, et al. Focal congenital hyperinsulinism managed by medical treatment: a diagnostic algorithm based on molecular genetic screening. *Clin Endocrinol (Oxf)*. 2014;81(5):679–88.
21. Christesen HB, Tribble ND, Molven A, Siddiqui J, Sandal T, Brusgaard K, et al. Activating glucokinase (GCK) mutations as a cause of medically responsive congenital hyperinsulinism: prevalence in children and characterisation of a novel GCK mutation. *Eur J Endocrinol*. 2008;159(1):27–34.
22. Meissner T, Marquard J, Cobo-Vuilleumier N, Maringa M, Rodriguez-Bada P, Garcia-Gimeno MA, et al. Diagnostic difficulties in glucokinase hyperinsulinism. *Horm Metab Res*. 2009;41(4):320–6.
23. Dullaart RP, Hoogenberg K, Rouwe CW, Stulp BK. Family with autosomal dominant hyperinsulinism associated with A456V mutation in the glucokinase gene. *J Intern Med*. 2004;255(1):143–5.
24. Sayed S, Langdon DR, Odili S, Chen P, Buettger C, Schiffman AB, et al. Extremes of clinical and enzymatic phenotypes in children with hyperinsulinism caused by glucokinase activating mutations. *Diabetes*. 2009;58(6):1419–27.
25. Cuesta-Munoz AL, Huopio H, Otonkoski T, Gomez-Zumaquero JM, Nanto-Salonen K, Rahier J, et al. Severe persistent hyperinsulinemic hypoglycemia due to a de novo glucokinase mutation. *Diabetes*. 2004;53(8):2164–8.
26. Christesen HB, Jacobsen BB, Odili S, Buettger C, Cuesta-Munoz A, Hansen T, et al. The second activating glucokinase mutation (A456V): implications for glucose homeostasis and diabetes therapy. *Diabetes*. 2002;51(4):1240–6.
27. Bough KJ, Rho JM. Anticonvulsant mechanisms of the ketogenic diet. *Epilepsia*. 2007;48(1):43–58.
28. McDaniel SS, Rensing NR, Thio LL, Yamada KA, Wong M. The ketogenic diet inhibits the mammalian target of rapamycin (mTOR) pathway. *Epilepsia*. 2011;52(3):e7–11.
29. Diano S, Matthews RT, Patrylo P, Yang L, Beal MF, Barnstable CJ, et al. Uncoupling protein 2 prevents neuronal death including that occurring during seizures: a mechanism for preconditioning. *Endocrinology*. 2003;144(11):5014–21.
30. Juge N, Gray JA, Omote H, Miyaji T, Inoue T, Hara C, et al. Metabolic control of vesicular glutamate transport and release. *Neuron*. 2010;68(1):99–112.
31. Schutz PW, Struys EA, Sinclair G, Stockler S. Protective effects of d-3-hydroxybutyrate and propionate during hypoglycemic coma: clinical and biochemical insights from infant rats. *Mol Genet Metab*. 2011;103(2):179–84.
32. Van Hove JL, Grunewald S, Jaeken J, Demaerel P, Declercq PE, Bourdoux P, et al. D, L-3-hydroxybutyrate treatment of multiple acyl-CoA dehydrogenase deficiency (MADD). *Lancet*. 2003;361(9367):1433–5.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

