

Review Article

The Application of Fiber-Reinforced Materials in Disc Repair

Bao-Qing Pei, Hui Li, Gang Zhu, De-Yu Li, Yu-Bo Fan, and Shu-Qin Wu

Key Laboratory for Biomechanics and Mechanobiology of Ministry of Education, School of Biological Science and Medical Engineering, Beihang University, Beijing 100191, China

Correspondence should be addressed to Bao-Qing Pei; pbq@buaa.edu.cn and Shu-Qin Wu; wushuqin@nuc.edu.cn

Received 16 October 2013; Accepted 18 November 2013

Academic Editor: Xiaoming Li

Copyright © 2013 Bao-Qing Pei et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The intervertebral disc degeneration and injury are the most common spinal diseases with tremendous financial and social implications. Regenerative therapies for disc repair are promising treatments. Fiber-reinforced materials (FRMs) are a kind of composites by embedding the fibers into the matrix materials. FRMs can maintain the original properties of the matrix and enhance the mechanical properties. By now, there are still some problems for disc repair such as the unsatisfied static strength and dynamic properties for disc implants. The application of FRMs may resolve these problems to some extent. In this review, six parts such as background of FRMs in tissue repair, the comparison of mechanical properties between natural disc and some typical FRMs, the repair standard and FRMs applications in disc repair, and the possible research directions for FRMs' in the future are stated.

1. Introduction

The intervertebral disc (IVD) is a heterogeneous, cartilaginous structure which contributes to the flexibility and load support in the spine. It consists of three parts: the nucleus pulposus (NP) in the center, the annulus fibrosus (AF) peripherally, and the cartilaginous endplates (CE) [1]. The fluid NP delivers loads to the AF in the form of swelling pressure while the multilayer and angle-ply AF wraps NP like a net bag guarded against the excessive expansion of NP (Figure 1). The direction of the fibers was angled varying from 40° to 70° to the vertical axis [1]. The intervertebral disc is in contact with the vertebral bodies through CE which is responsible for the exchange of substance through the microporous structure [2]. Anyone off normal of these three parts may cause the disc degeneration.

The spine disease which is related to the disc degeneration affects human health and normal life. The treatment and regeneration of the degenerated disc is one of the most urgent current clinical problems. There are complex mechanical and structures requirements for the disc repair. The fluid nature of the NP ensures the compressive loading applied to a disc to generate a tensile hoop stress (T) in the annulus (Figure 1) [3, 4]. Meanwhile, since the IVD joint possess 6-degree freedom

of motion, AF needs to bear axial and radial compression and stretch as well [5]. According to the *in vivo* and *vitro* results, the interdiscal pressure under 300 N loading is approximately 1.3 MPa, the circumference stretch stress of the AF is 12.7 MPa [6], and the shear modulus is 25~110 KPa [7]. In this view, high mechanical properties and complex load bearing capacity are necessary for IVD repair materials.

Many methods for disc restoration are aimed at the reinforcement of disc mechanical properties, such as injection of the polymethylmethacrylate for the traditional intervertebral injury and fracture [8], the usage of biocompatible cage and metal internal fixations for bone fusion, and preventing the intervertebral collapse and failure restoration [9]. Some chemistry technologies such as doubly cross-linked microgels can make the strength enhanced by 3 to 5 times compared to original materials and dynamic properties similar to the health disc [10]. The fiber-reinforced technology is commonly used for increasing the mechanical properties of materials and some researchers are trying to use FRMs into the field of disc repair.

Many biocompatible and biodegradable fiber-reinforced polymers combined with matrix form new mechanical enhanced FRMs and have mature application in bone repair field. The calcium phosphate cement (CPC) reinforced with

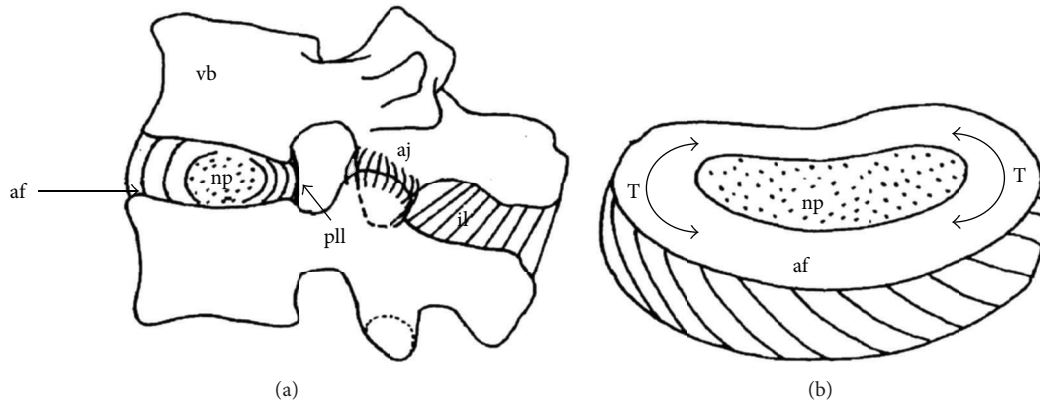


FIGURE 1: Schematic structure of the IVD and NP-AF interaction under compression [4]. vb: vertebral body; af: annulus fibrosus; np: nucleus pulposus; aj: apophyseal joint; pll: posterior longitudinal ligament; il: interspinous ligaments; T: tensile stress.

polycaprolactone polymer (PCL) fibers in different weight increased the modulus nearly twofold from 85 to 155 Mpa [16]. CPC power matrix addition to poly(coglycolide) (PLGA) fibers makes the compressive and bending strength close to the bone [17]. 15 wt% self-reinforced poly-L-lactide (PLLA) fiber coated with hydroxyapatite (HA) composite makes the original bending modulus increase from 8.3 Gpa to 9.7 Gpa [18]. Multi-fiber-reinforced CPC compound with the compressive strength from 0.9 to 69 Mpa [19] and the energy to fracture increase by 390X compared to pure CPC, with bending strength range from 1.2 to 60 Mpa. Chitosan fiber-reinforced composite which possesses well-rheological behavior [20] and biodegradability [21] repairs 40 mm goat shank bone defect successfully after 15 weeks, with the chitosan FRM group recovering to the same level BMD and bone mechanical strength to the intact group [22–26]. The porous three-dimensional poly(L-lactic acid) scaffold reinforced by the chitin fibers with link is an appropriate scaffold for tissue engineering [27]. As for the cell level, the material microstructure after fiber being reinforced is more suitable for cell adhesion and reproduction. Poly(3-caprolactone) (PCL)/b-tricalcium phosphate (TCP) nanofibre-reinforced hierarchical collagen scaffolds cell seeding efficiency increased from 55% for pure collagen scaffolds to 78% after fiber reinforced with tensile modulus increased by 7 times as well [28]. The carbon nanotube reinforced materials have a better ability to adsorb proteins and better cell attachment, proliferation, and differentiation than graphite [29], so they could also induced more bone formation [30]. Hydrogel is widely used for many soft tissues and is supposed to be the most promising disc repair materials [31]. However, pure hydrogel is proved to be insufficient for disc mechanical restoration [32]. Elastin fiber-reinforced hydrogel makes a better performance and can achieve cartilage-like properties [33]. The hyaluronic acid matrix reinforced by cellulose and elastin-like polypeptide fibers with adding collagen, hyaluronic acid, and chondroitin sulfate can obtain a better collision property [34–36]. Although these researches have not reached the stage of tissue fabrication or the *in vivo* and *vitro* experiments, the composites' properties got enhancement through the fiber-reinforced technology.

Scaffolds play fatal roles in both disc arthroplasty and whole disc replacement tissue regeneration [37–40]. The FRMs are becoming the ideal material for kinds of hard and soft tissue regeneration. A combination of matrix materials and different fibers arrays may help address the weakness of each component as a scaffold and supply easy grown mechanical and structure environments. So far, FRMs have been widely used in bone repair and bone tissue engineering [41]. A great many of fibers may be used for fiber-reinforced material fabrication such as polymer fibers such as polyethylene (PE), polylactic acid (PLA), and chitosan [42], biological ceramic fibers like bioglass, HA and calcium polyphosphate [43], and metallic fibers such as stainless steel and titanium [44].

This review addresses the mechanical requirement for disc tissue repair, compares the mechanical properties of FRM and disc tissue, and summarizes typical FRMs applications in disc tissue engineering. The adjustable mechanical properties by different spatial configuration of the fibers and their impacts on the cell growth have also been discussed preliminarily. Currently, the definition and summary about the FRMs for disc repair are not that much. The developments of FRMs in the field of disc tissue repair have very important practical applications and biomimetic meanings for the clinical spinal surgery.

2. Comparison of Mechanical Properties between Nature Disc and FRMs

2.1. The Mechanical Properties of Healthy Human Disc. The properties of the disc as a whole are determined by the mechanical behavior of the NP. NP is the most active element of the physiological hydrodynamical system of the intervertebral disc extradiscal space [56]. The most important characteristic of this system is the intradiscal pressure (IP) which depends on the external loads and the degree of hydration of the nucleus. The IP when being in latericumbent position is 0.3234 Mpa and 0.8428 Mpa when sitting. The standing IP is 60~80% sitting pressure. In young people aged from 20 to 30, the vertical stretch strength is about

TABLE 1: Compressive stiffness and modulus of healthy human discs.

Load (N)	Stiffness (N/mm)	Modulus (Mpa) [45]
490	392–490	16.366
490–1470	666.4–3479	8.624~26.166
1470–9800	1400–7791	9.604~21.364

0.196 Mpa, and the horizontal one is 0.294 Mpa. The vertical tensile strength of the outer layer AF is 15.68 Mpa and that of the inner layer is 6.664 Mpa. The horizontal outer layer is 7.84 Mpa and inner layer is 4.4 Mpa [45, 57]. The compressive stiffness is increased with loading while the modulus is not increased as significantly as the stiffness. The compressive stiffness and modulus are shown in Table 1. The *in vitro* human lumbar torsional stiffness is 2.0 Nm/deg [58], and the compressive stiffness is 2~14 KN [59]. The unconfined compressive elastic modulus of human disc is 5 kpa, and the Poisson ratio is 0.62, and relaxation rate is 65% [60]. The confined compressive elastic modulus is 0.14 Mpa, effective modulus is 1 Mpa, and permeability is $0.9E - 15$ m⁴/Ns [61]. The torsional shear modulus is from 7.4 to 19.8 Kpa; phase angle is between 23° and 30° [60]. The disc system is a typical MKC oscillatory system, with inherent frequency about 4~5 Hz [62].

The NP mechanical properties are related to the content of glycosaminoglycan (GAG) and water. The normal NP swelling pressure is about 0.138 ± 0.029 MPa; effective aggregate modulus is 1.01 ± 0.43 MPa [63]. The average vertical strain is 2~8% and radial strain is 1~4% of human lumbar AF under neutral compression [64]. Healthy NP compressive strain is -10%~+10% [65]. The elastic modulus is 3~6 Kpa [66]. The single axial compression of the AF has obvious compliance, but after the strain increased to 20%~40%, the stiffness grows spurt. The horizontal shear compliance is smaller than the vertical ones. When the strain loads over 60%, the AF fracture not happened, but irreversible deformation happened [67].

2.2. Mechanical Properties of FRMs. The mechanical properties of FRMs increase with the content of fibers mostly, but the low density indicates a desired reinforcing storage and loss modulus of nanofibers [49]. It is possible to control the porosity and porous size through fiber-reinforced method, and the critical length of the fibers could be calculated using empirical formula, and so does the compressive strength of the scaffold using the fiber length [68]. Higher density fibers may result in a morphological change of the gel structure where the occurrence of nanofibers disrupts the continuity of the gel network and overall weakening of the construct. Meanwhile, the fiber processing technique may bring in bubble; the more the fibers added, the more the bubbles taken in, and this may be an important factor of modulus weakening. Table 2 shows some FRMs for disc and disc-like joint and cartilage tissue engineering. The average compressive stiffness of the human lumbar when loaded by 577 ~ 2058 N was 1400 N/mm [69], while the biomimetic artificial IVD with fiber-reinforced annulus structure got a 2-fold stiffness

of the normal disc [70]. Using 3D printed technique for the rat tissue engineered-total disc replacement (TE-TDR) with cell-seeded alginate and cell-seeded collage fibers was found a similar dynamic modulus (235 ± 51 kPa) with the native disc (238 ± 68 kPa) [71].

These FRMs are characterized by the following features. (1) Properties tunableness: a wide range of properties could be obtained through adjustment of fiber scales, processing technology, and geometry space configuration. (2) Good viscoelastic properties: body tissues are viscoelastic materials to some extent. The IVD restoration needs more significant viscosity properties than other tissues and load bearing strength as well. (3) Different fibers and matrix mix and match increase material diversity: as for this point, some researchers suggested that the focus should be on the deep boost of several certain mature compounds instead of keep searching for different materials with a smattering of knowledge of each one such as only culture cells for only days.

The main structure of the FRMs is the fiber. A study reported the diameters of the individual collage fibrils which are the main structural components of the NP and found that the nanoscale (with a mean diameter of 92.1 ± 26.54 nm) of the collage fibrils had a mild linear correlation with the compressive modulus of the NP [72]. The human fibrous tissue (FT) was compared with annulus and nucleus in relaxation and dynamic properties [60]. The percent relaxation of the FT was 90% included both AF and NP (70%~80%). The storage modulus of the fibrous tissue was also larger than that of the AF and NP. The FT is not proved to be a substitute for native tissue within the disc space.

Many researches regarded the constitutive equation of FRMs under different loading as very important research fields for FRMs tissue engineering. The models included anisotropic viscoelastic behavior under finite deformation [73, 74] and large-strain deformation [75] of soft composites. The stress transfer in collagen fibrils reinforcing tissues was infected by the fibril slenderness significantly. A large slenderness value led to high stress in a fibril and it is beneficially provided since they do not exceed the fracture stress of collagen. Fibers with taper-type shape are better than fibers with uniform cylindrical shape in against fiber fracture [76].

2.3. FRMs and Cell Culture. The small porosity of some high densely arranged nanofiber-reinforced materials makes it difficult for cell permeating into the fiber bundles and leads to abnormal ECM environment, so the material restoration fails to recover a normal tissue mechanical and physical standard finally. In response to this issue, a new method called dynamic cell culture technique can enhance the permeability of the stem cell and the quality of the ECMs [77]. NP cell bears the hydrostatic pressure while the cells between AF and NP enduring the deformation stimulate mostly. The cell geometry is impacted by the micromechanical environment which is affected by the fiber-reinforced technique, and the adapt law of the cell development direction is the reduction of the strain load [78].

The cell growth situation is affected by the material scale. Nanoscaled fibers which can stimulated a variety of

TABLE 2: Typical FRM use in disc implants and some similar tissues.

Fibers	Matrix	Mechanical properties			Fiber scale	Processing	Repaired tissue	literature
		Breaking stress (Mpa)	Breaking strain (%)	Elastic modulus (Mpa)				
Elastin	Hydrogel	0.08~2.08	10.6~247	0.8~3.68	Mircon	3D syringe drops crossed "log-piles"	Cartilage	Agrawal et al., 2013 [33]
PCL	Gelatin				Nano	Electrospun	AF	Beachley and Wen, 2009 [46]
	Dry	5~25		20~120				
	Wet	0.1~0.9		1~10				
Collagen	Elastin-like	1.85~4.08	23~314	5.3~33.1	Mircon	Winding	Abdominal wall	Caves et al., 2011 [47]
Polydioxanone (PDO)	PLA					Electrospun	AF	Cont et al., 2013 [48]
PCL	Hydrogel	Storage modulus 0.03 Mpa	Loss modulus 0.006 Mpa		Nano	Electrospun	NP	Thorvaldsson et al., 2012 [49]
N-vinyl-2-pyrrol-idone (NFC)	Hydrogel	Storage modulus 0.14 Mpa	Loss modulus 0.019 Mpa	0.02~8		Curing	NP	Borges et al., 2010 [50]

interactions at the cellular level may promote greater amounts of specific protein interactions and more efficiently new bone formation [79]. The microstructured calcium phosphate materials concentrate more proteins and also are proved to induce more bone formation. The biocompatibility and bioactivity are also promoted by the nanoscaled materials [80]. The carbon nanotubes can induce cells in soft tissues to form inductive bone by concentrating more proteins including bone-inducing proteins [81, 82], which is also the contribution of the nanoscale structure. Though the nanoscale materials have stimulative effects on cell growth and induced differentiation, the biocompatibility and toxicity still need cautious experiments before *in vivo* attempt [83].

3. The Disc Restoration Objects

The intervertebral disc (IVD) is the mechanical and structural unit of the spine, so the functional restoration is very important and needs some standard indicators for evaluation. There are many mechanical characters of the IVD including nonlinearity, viscoelasticity, anisotropy, heterogeneity, and permeability. Finding limited crucial and reasonable properties is more meaningful. One of the clinical golden standards for IVD examination is the IVD height from MRI and X-ray plain film [84]. As for the experimental purpose, the neutral zone stiffness and the relative length during axial low load can estimate the recovery condition from needle damage and the endplate injury. As for large and severe AF defects and IVD degeneration, the torsion strength and ROM can test the restoration effects [84]. The dynamic modulus and stress relaxation and creep test make a judgment of the viscoelasticity behavior of the repaired disc [53]. For the

implantable tissue replacement methods, shear strength of the interface is very crucial indicator of tissue fusion [85].

The synthesis and maintenance of extracellular matrix (ECM) are necessary for tissue activity and cell reproduction. The collage and GAGs are main substances for disc ECM [86]. Immunohistochemical methods can make qualitative and quantitative measurement for these two ingredients. The DNA transfer and mRNA expression can reflect the activity and propagation of newborn cells [87]. Though the cell-based therapy methods are more mature than whole IVD transplantation, the inadequacy for severe degeneration and disc prolapse stage limited its development and more and more focuses are on the latter. By now, whole IVD tissue engineering method remains under the exploration and experimentation step. Generally, it needs stages from a material to an artificial tissue use *in vivo*. The duration selections for the stages are shown in Table 3. At present, the time span for each period has not obtained a unified time yet.

4. The Application of FRM in Disc Arthroplasty

As was mentioned in the front part, some materials such as coralline and hydroxyapatite could induce osteogenesis. Spinal fusion is widely used in clinical for intervertebral decompression which aims at complete bony fusion. So, these materials have good performance for intervertebral fusion, especially after being reinforced by fibers such as PLA, PEG, or carbon fibers [88–90]. Though fusion is not the ideal choice for disc repair, FRMs used for fusion materials are also a research hotspot at present.

TABLE 3: Time span for experimental validation.

Experimental objects	Plant mode	Culture <i>ex vivo</i> (week)	Culture <i>in vivo</i> (month)
Canine model [51]	Cell-based		3, 6, 9, 12
Polymer scaffolds [52]	Explant	4, 8, 12	
Rodent model [53]	Implant		6
Collagen-gel compound [54]	Explant	3 days	
Alginate composite [55]	Explant	4	



FIGURE 2: The parts and the whole of the disc prosthesis. (a) Two sides of HAPEX endplates, (b) composite hydrogel for IVD substitute, and (c) total IVD substitute prototype [11].

The materials for disc replacement prosthesis need to possess different mechanical properties and composite structures. Flexibility, toughness, and high strength are basic characteristics of the soft biological tissues. For this reason, by employing materials with a single structural arrangement, it is not possible to combine all of these features. So, the structure of natural disc has been reproduced by adopting a biomimetic approach. This led to the development of a fiber-reinforced hydrogel able to match the performances of the natural disc and those of the surrounding tissues [91].

Disc arthroplasty is a kind of disc therapy which uses disc prosthesis (DP) with no biological activities for disc replacement and functional disc repair. The metallic and high polymer-core DP incurs DP sinking and wear and also ADD (adjacent disc degeneration) because of the high stiffness of the DP materials. The ideal artificial disc should be biomimetic which means that the mechanical properties, structure constitution and the motion function are similar with the nature disc tissue. Since, 1990s researchers have begun to use polyethylene and polyurethane fiber-reinforced silicon elastomers to make disc spacers. The artificial disc obtained the same compressive and torsional properties through adjusting the fiber direction, numbers of fiber layers, and the sequence of reinforcement [92]. The compressive modulus increased with the fiber angle and fiber content positively [93].

Hydrogel is always used for disc tissue engineering materials for its close properties to IVD tissue. Using fibers to reinforce hydrogel makes the composite with better stiffness and strength and maintenance of original properties. After fiber have added, the water absorption rate decreased for the new FRMs, is 30 wt% fibers addition with only 25% water absorption [93]. This rate is the same as the collage in disc, and the FRM with 30 wt% fibers could achieve suitable mechanical properties demanded by the normal IVD.

Gloria et al. [11] developed a new disc prosthesis using poly(2-hydroxyethyl methacrylate)/poly(methyl methacrylate) (PHEMA/PMMA) (80/20 w/w) semi-interpenetrating

polymer network (s-IPN) composite hydrogel reinforced with poly(ethylene terephthalate) (PET) fibers as annulus/nucleus substitute and two hydroxyapatite-reinforced polyethylene composites (HAPEXTM) as endplates as was shown in Figure 2. This FRMs' disc performed enough static properties compared to normal disc, with maximum static compressive stiffness 4030 ± 612 N/mm, torsional rigidity 2.8 ± 0.3 Nm/deg, and shear stiffness 205 ± 22.1 N/mm. The ten million times fatigue test indicated that no damage or wear happened during the test, which is much better than some products in the markets. Meanwhile, the hydrogel matrix FRMs' disc has the similar dynamic and viscoelastic properties for the healthy disc to the other prosthesis.

5. The Application of FRM in Disc Tissue Engineering

It is more and more prevalent for disc restoration using tissue engineering method. The cell-based method includes two main directions which are the cell transplantation and the bioactive tissue transplantation. The bioactive tissue material used for disc repair is a research focus at present. The microstructure of the FRMs [94] is proved to promote cell adhesion and growth by supporting a 3D growing space [95]. Some studies have used FRMs for disc repair and various fiber features may lead to different repair effects.

The main cells used for disc regeneration are NP cell, AF cell, and stem cells. The NP cells and AF cells are like the chondrocyte and fibroblast, respectively, in both structure and function. These two kinds of cells are always used for testing materials bioactivity. Some biocompatibility (BC) matrixes after reinforced by the biodegradable nanofibers may improve the brittleness of single fibers and the fiber density become less dense after reinforcing the matrix, making cells easier grow into the fibers. The fibroblasts were cultured in single array and square crossing fibers in FRMs, respectively, and were found to grow along the fibers as a line

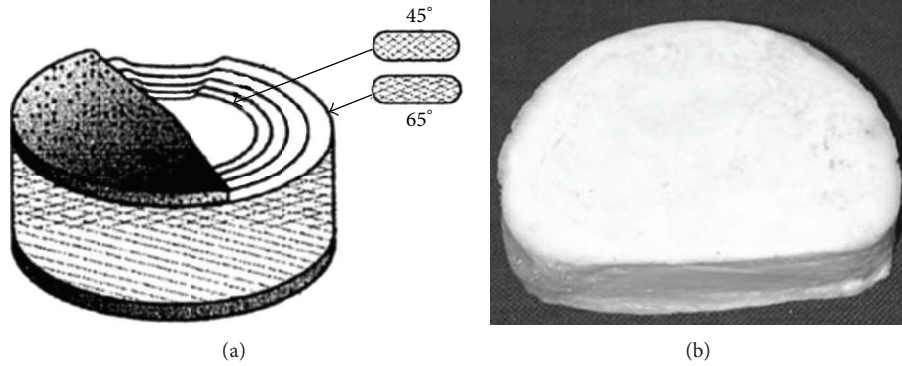


FIGURE 3: (a) A schematic representation of the fiber-reinforced disc substitute with hydroxyapatite reinforcing hydrogel endplates, (b) The total intervertebral disc substitute prototype [12].

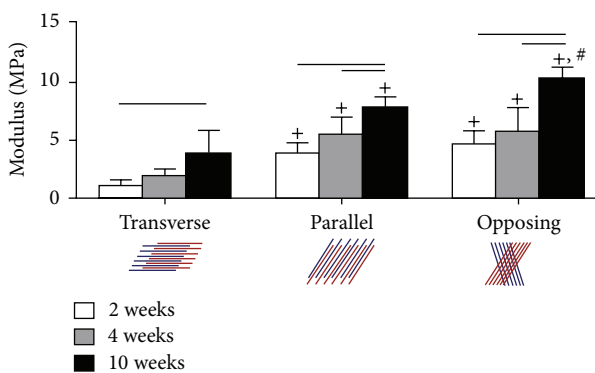


FIGURE 4: Three type of fiber organization between layers and the shear modulus when loaded tensile [13].

in the single array group and distribute in the meshes in the crossing one. The nanoscale FRMs process abilities to induce cell grow directions [96].

The mechanical strength of the composite may be enhanced after fiber being reinforced. We could obtain certain properties through increasing fiber content and changing fiber angles. PET fibers' angle changing from 45° to 60° (shown in Figure 3) (angle to the load) which were added into hydrogel makes compressive modulus (CM) increased by 2~3 times. After changing volume fraction of PCL fibers from 0% to 30%, CM increases by 15 times [12]. PGA (polyglycolic acid) is reinforced by PLA fibers; CM increased with volume of PLA fibers linearly, which change by 0%~68% CM increased by 20 times [97]. 3%~17% volume change of elastic fibers was reinforced by collage fibers; the stretch modulus increased by 1 time with the same fiber angles while the angle change for 15 not significant changed the stretch modulus.

Researchers used PCL (polycaprolactone) nanofiber-reinforced hydrogel for NP tissue engineering. The dynamic properties of the materials are not increased with fiber density; on the contrary lower density obtained a closer storage and loss modulus to the real disc. We may draw a conclusion that fiber content got a bigger weighting factor

than the fiber angles on the impact of mechanical properties. Change angles may adjust FRMs in a small range.

The annulus fibrosus is a natural fiber-reinforced structured tissue. Some researches focus on the organization of collagen fibers into planes of alternating alignment and found that it played an important role in annulus fibrosus tissue function. By using MSC-seeded nanofibrous scaffolds and applying the constitutive model to uniaxial tensile stress-strain data for bilayers with three different fiber orientations, they found that fiber orientation of adjacent layers with an opposing style got the biggest strength against the shear between layers under tensile load as shown in Figure 4 [13].

6. Conclusion and Perspectives

Current therapies for disc degeneration and spinal disease mainly focus on the relieving pains instantly instead of functional and physiological repair on the long run. Tissue engineering provides new treatment strategy for disc repair. The application of fiber-reinforced materials in tissue repair has a wide range of use and mature background, considering the excellent mechanical properties, and will make a new direction for disc restoration.

FRMs are on the developing stage for disc repair; some problems and research fields such as the choice of fibers, the interaction of fibers-matrix, and also the processing technology effects for properties coordination deserve more deeply discussion. Meanwhile, the mechanisms of FRMs for cell growth and propagation are still not known yet.

MSC differentiation *in vivo* technique is widely studied for tissue regeneration. Recent research shows that the MSCs' function is related to the substrate stiffness which could be adjusted by fiber-reinforced methods [98, 99]. As the disc cell lives in hypoxia, it has been proved that MSC differentiation shows better cell survival rate and activity under low oxygen environment [100]. By controlling technology and fiber density, FRMs could obtain nanoscale 3D cell growth space and fulfill the oxygen concentration control.

The cartilage endplates of the disc have very important structure and many disc diseases originate from endplate degeneration. As for the special position, the junction of

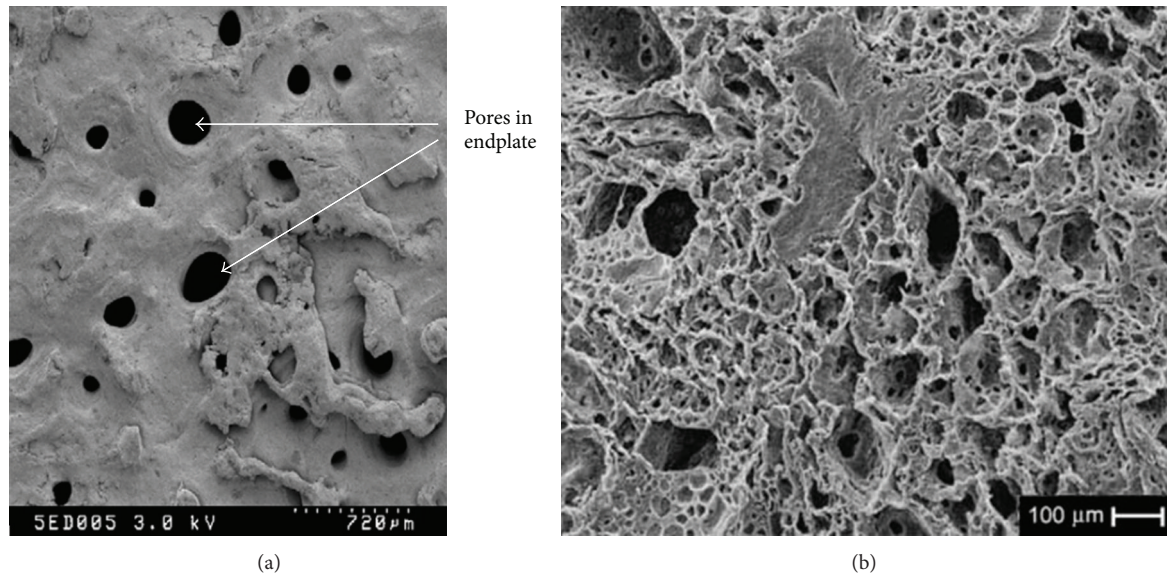


FIGURE 5: The similar scale of the pores in endplate and pores in FRM composite. (a) REM image of a vertebral endplate with 720 μm scale plate [14] and (b) SEM analysis of FRM scaffolds with a 100 μm scale plate [15].

the vertebra, and disc tissue, the endplates are responsible for material exchange. The pores in the endplate (shown in Figure 5(a)) make it possible for mass metabolism between the inside and outside [14]. So the repair of this structure needs not only mechanical function but also diffusion and interface fusion properties. FRMs have a better bone fusion ability and can show better interface strength. At the same time, the microporous (shown in Figure 5(b)) formed by the embedded fibers may ensure the diffusion function [15]. Also, the endplates' tissue replacement needs two different properties in one piece of materials. FRMs' coordinability properties make this possible for a functional endplate.

Acknowledgment

This work was supported by the National Natural Science Foundation of China (11372029).

References

- [1] A. I. Kapandji, *The Physiology of the Joints: The Spinal Column Pelvic Girdle and Head*, 6 edition, 2011.
- [2] M. A. Adams, P. Dolan, and D. S. McNally, "The internal mechanical functioning of intervertebral discs and articular cartilage, and its relevance to matrix biology," *Matrix Biology*, vol. 28, no. 7, pp. 384–389, 2009.
- [3] C. C. Guterl, E. Y. See, S. B. Blanquer et al., "Challenges and strategies in the repair of ruptured annulus fibrosus," *European Cells and Materials*, vol. 25, pp. 1–21, 2013.
- [4] M. A. Adams and P. Dolan, "Spine biomechanics," *Journal of Biomechanics*, vol. 38, no. 10, pp. 1972–1983, 2005.
- [5] M. A. Adams, D. S. McNally, and P. Dolan, "Stress' distributions inside intervertebral discs: the effects of age and degeneration," *Journal of Bone and Joint Surgery B*, vol. 78, no. 6, pp. 965–972, 1996.
- [6] E. R. Acaroglu, J. C. Iatridis, L. A. Setton, R. J. Foster, V. C. Mow, and M. Weidenbaum, "Degeneration and aging affect the tensile behavior of human lumbar annulus fibrosus," *Spine*, vol. 20, no. 24, pp. 2690–2701, 1995.
- [7] J. C. Iatridis, S. Kumar, R. J. Foster, M. Weidenbaum, and V. C. Mow, "Shear mechanical properties of human lumbar annulus fibrosus," *Journal of Orthopaedic Research*, vol. 17, no. 5, pp. 732–737, 1999.
- [8] D. C. Wilson, R. J. Connolly, Q. Zhu et al., "An ex vivo biomechanical comparison of a novel vertebral compression fracture treatment system to kyphoplasty," *Clinical Biomechanics*, vol. 27, no. 4, pp. 346–353, 2012.
- [9] R. Lindsay, S. L. Silverman, C. Cooper et al., "Risk of new vertebral fracture in the year following a fracture," *Journal of the American Medical Association*, vol. 285, no. 3, pp. 320–323, 2001.
- [10] A. H. Milani, A. J. Freemont, J. A. Hoyland, D. J. Adlam, and B. R. Saunders, "Injectable doubly cross-linked microgels for improving the mechanical properties of degenerated intervertebral discs," *Biomacromolecules*, vol. 13, no. 9, pp. 2793–2801, 2012.
- [11] A. Gloria, R. De Santis, L. Ambrosio, F. Causa, and K. E. Tanner, "A multi-component fiber-reinforced PHEMA-based hydrogel/HAPEXTM device for customized intervertebral disc prosthesis," *Journal of Biomaterials Applications*, vol. 25, no. 8, pp. 795–810, 2011.
- [12] A. Gloria, F. Causa, R. de Santis, P. A. Netti, and L. Ambrosio, "Dynamic-mechanical properties of a novel composite intervertebral disc prosthesis," *Journal of Materials Science*, vol. 18, no. 11, pp. 2159–2165, 2007.
- [13] N. L. Nerurkar, R. L. S. Mauck, and D. M. Elliott, "Modeling interlamellar interactions in angle-ply biologic laminates for annulus fibrosus tissue engineering," *Biomechanics and Modeling in Mechanobiology*, vol. 10, no. 6, pp. 973–984, 2011.
- [14] G. Huber, M. M. Morlock, and K. Ito, "Consistent hydration of intervertebral discs during in vitro testing," *Medical Engineering and Physics*, vol. 29, no. 7, pp. 808–813, 2007.

- [15] V. Guarino, F. Causa, P. Taddei et al., "Polylactic acid fibre-reinforced polycaprolactone scaffolds for bone tissue engineering," *Biomaterials*, vol. 29, no. 27, pp. 3662–3670, 2008.
- [16] L. H. Leung, A. DiRosa, and H. E. Naguib, "Physical and mechanical properties of poly(ϵ -caprolactone) hydroxyapatite composites for bone tissue engineering applications," in *Proceedings of the ASME International Mechanical Engineering Congress and Exposition (IMECE '09)*, pp. 17–23, New York, NY, USA, November 2009.
- [17] L. A. Vasconcellos and L. A. dos Santos, "Calcium phosphate cement scaffolds with PLGA fibers," *Materials Science and Engineering C*, vol. 33, no. 3, pp. 1032–1040, 2013.
- [18] L. F. Charles, E. R. Kramer, M. T. Shaw, J. R. Olson, and M. Wei, "Self-reinforced composites of hydroxyapatite-coated PLLA fibers: fabrication and mechanical characterization," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 17, pp. 269–277, 2013.
- [19] R. Kruger and J. Groll, "Fiber reinforced calcium phosphate cements: on the way to degradable load bearing bone substitutes?" *Biomaterials*, vol. 33, no. 25, pp. 5887–5900, 2012.
- [20] X. M. Li and Q. L. Feng, "Dynamic rheological behaviors of the bone scaffold reinforced by chitin fibres," *Materials Science Forum*, vol. 475–479, pp. 2387–2390, 2005.
- [21] X. Li, Q. Feng, and F. Cui, "In vitro degradation of porous nano-hydroxyapatite/collagen/PLLA scaffold reinforced by chitin fibres," *Materials Science and Engineering C*, vol. 26, no. 4, pp. 716–720, 2006.
- [22] X. Li, Y. Yang, Y. Fan, Q. Feng, F. Z. Cui, and F. Watari, "Bio-composites reinforced by fibers or tubes as scaffolds for tissue engineering or regenerative medicine," *Journal of Biomedical Materials Research A*, 2013.
- [23] X. Li, Q. Feng, X. Liu, W. Dong, and F. Cui, "Collagen-based implants reinforced by chitin fibres in a goat shank bone defect model," *Biomaterials*, vol. 27, no. 9, pp. 1917–1923, 2006.
- [24] X. Liu, X. Li, Y. Fan et al., "Repairing goat tibia segmental bone defect using scaffold cultured with mesenchymal stem cells," *Journal of Biomedical Materials Research B*, vol. 94, no. 1, pp. 44–52, 2010.
- [25] X. Li, X. Liu, G. Zhang et al., "Repairing 25 mm bone defect using fibres reinforced scaffolds as well as autograft bone," *Bone*, vol. 43, Supplement 1, S94 pages, 2008.
- [26] X. Niu, Y. Fan, X. Liu et al., "Repair of bone defect in femoral condyle using microencapsulated chitosan, nanohydroxyapatite/collagen and poly(L-lactide)-based microsphere-scaffold delivery system," *Artificial Organs*, vol. 35, no. 7, pp. E119–E128, 2011.
- [27] X. Li, X. Liu, W. Dong et al., "In vitro evaluation of porous poly(L-lactic acid) scaffold reinforced by chitin fibers," *Journal of Biomedical Materials Research B*, vol. 90, no. 2, pp. 503–509, 2009.
- [28] Y. B. Kim and G. Kim, "Rapid-prototyped collagen scaffolds reinforced with PCL/ β -TCP nanofibres to obtain high cell seeding efficiency and enhanced mechanical properties for bone tissue regeneration," *Journal of Materials Chemistry*, vol. 22, no. 33, pp. 16880–16889, 2012.
- [29] X. Li, H. Gao, M. Uo et al., "Effect of carbon nanotubes on cellular functions in vitro," *Journal of Biomedical Materials Research A*, vol. 91, no. 1, pp. 132–139, 2009.
- [30] X. Li, H. Gao, M. Uo et al., "Maturation of osteoblast-like SaoS_2 induced by carbon nanotubes," *Biomedical Materials*, vol. 4, no. 1, Article ID 015005, 2009.
- [31] S. C. W. Chan and B. Gantenbein-Ritter, "Intervertebral disc regeneration or repair with biomaterials and stem cell therapy: feasible or fiction?" *Swiss Medical Weekly*, vol. 142, Article ID w13598, 2012.
- [32] S. Reitmaier, U. Wolfram, A. Ignatius et al., "Hydrogels for nucleus replacement: facing the biomechanical challenge," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 14, pp. 67–77, 2012.
- [33] A. Agrawal, N. Rahbar, and P. D. Calvert, "Strong fiber-reinforced hydrogel," *Acta Biomaterialia*, vol. 9, no. 2, pp. 5313–5318, 2013.
- [34] A. T. Reza and S. B. Nicoll, "Characterization of novel photocrosslinked carboxymethylcellulose hydrogels for encapsulation of nucleus pulposus cells," *Acta Biomaterialia*, vol. 6, no. 1, pp. 179–186, 2010.
- [35] E. C. Collin, S. Grad, D. I. Zeugolis et al., "An injectable vehicle for nucleus pulposus cell-based therapy," *Biomaterials*, vol. 32, no. 11, pp. 2862–2870, 2011.
- [36] B. Huang, C.-Q. Li, Y. Zhou, G. Luo, and C.-Z. Zhang, "Collagen II/hyaluronan/chondroitin-6-sulfate tri-copolymer scaffold for nucleus pulposus tissue engineering," *Journal of Biomedical Materials Research B*, vol. 92, no. 2, pp. 322–331, 2010.
- [37] A. D. Diwan, H. K. Parvataneni, S. N. Khan, H. S. Sandhu, F. P. Girardi, and F. P. Cammisa Jr., "Current concepts in intervertebral disk restoration," *Orthopedic Clinics of North America*, vol. 31, no. 3, pp. 453–464, 2000.
- [38] G. Lewis, "Nucleus pulposus replacement and regeneration/repair technologies: present status and future prospects," *Journal of Biomedical Materials Research B*, vol. 100, no. 6, pp. 1702–1720, 2012.
- [39] B. R. Whatley and X. Wen, "Intervertebral disc (IVD): structure, degeneration, repair and regeneration," *Materials Science and Engineering C*, vol. 32, no. 2, pp. 61–77, 2012.
- [40] W. Fu and Z. Xiang, "Regeneration strategies of intervertebral disc," *Chinese Journal of Reparative and Reconstructive Surgery*, vol. 27, no. 2, pp. 227–232, 2013.
- [41] R. Kruger and J. Groll, "Fiber reinforced calcium phosphate cements: on the way to degradable load bearing bone substitutes?" *Biomaterials*, vol. 33, no. 25, pp. 5887–5900, 2012.
- [42] D. T.-J. Barone, J.-M. Raquez, and P. Dubois, "Bone-guided regeneration: from inert biomaterials to bioactive polymer (nano)composites," *Polymers for Advanced Technologies*, vol. 22, no. 5, pp. 463–475, 2011.
- [43] T. Eliades, "Orthodontic materials research and applications. Part 2: current status and projected future developments in materials and biocompatibility," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 131, no. 2, pp. 253–262, 2007.
- [44] A. Parashar and P. Mertiny, "Adhesively bonded composite tubular joints: review," *International Journal of Adhesion and Adhesives*, vol. 38, pp. 58–68, 2012.
- [45] K. G. Bomshtein, V. I. Danilov, and V. N. Pravetskii, "Statics and dynamics of intervertebral discs," *Mechanics of Composite Materials*, vol. 15, no. 4, pp. 419–423, 1980.
- [46] V. Beachley and X. Wen, "Fabrication of nanofiber reinforced protein structures for tissue engineering," *Materials Science and Engineering C*, vol. 29, no. 8, pp. 2448–2453, 2009.
- [47] J. M. Caves, W. Cui, J. Wen, V. A. Kumar, C. A. Haller, and E. L. Chaikof, "Elastin-like protein matrix reinforced with collagen microfibers for soft tissue repair," *Biomaterials*, vol. 32, no. 23, pp. 5371–5379, 2011.

- [48] L. Cont, D. Grant, C. Scotchford, M. Todea, and C. Popa, "Composite PLA scaffolds reinforced with PDO fibers for tissue engineering," *Journal of Biomaterials Applications*, vol. 27, no. 6, pp. 707–716, 2013.
- [49] A. Thorvaldsson, J. Silva-Correia, J. M. Oliveira et al., "Development of nanofiber-reinforced hydrogel scaffolds for nucleus pulposus regeneration by a combination of electrospinning and spraying technique," *Journal of Applied Polymer Science*, vol. 128, no. 2, pp. 1158–1163, 2013.
- [50] A. C. Borges, P.-E. Bourban, D. P. Pioletti, and J.-A. E. Månson, "Curing kinetics and mechanical properties of a composite hydrogel for the replacement of the nucleus pulposus," *Composites Science and Technology*, vol. 70, no. 13, pp. 1847–1853, 2010.
- [51] T. Ganey, J. Libera, V. Moos et al., "Disc chondrocyte transplantation in a Canine model: a treatment for degenerated or damaged intervertebral disc," *Spine*, vol. 28, no. 23, pp. 2609–2620, 2003.
- [52] J. M. Moran, D. Pazzano, and L. J. Bonassar, "Characterization of poly(lactic acid)-poly(glycolic acid) composites for cartilage tissue engineering," *Tissue Engineering*, vol. 9, no. 1, pp. 63–70, 2003.
- [53] R. D. Bowles, H. H. Gebhard, R. Härtl, and L. J. Bonassar, "Tissue-engineered intervertebral discs produce new matrix, maintain disc height, and restore biomechanical function to the rodent spine," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 32, pp. 13106–13111, 2011.
- [54] R. D. Bowles, R. M. Williams, W. R. Zipfel, and L. J. Bonassar, "Self-assembly of aligned tissue-engineered annulus fibrosus and intervertebral disc composite via collagen gel contraction," *Tissue Engineering A*, vol. 16, no. 4, pp. 1339–1348, 2010.
- [55] J. L. Bron, L. A. Vonk, T. H. Smit, and G. H. Koenderink, "Engineering alginate for intervertebral disc repair," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 4, no. 7, pp. 1196–1205, 2011.
- [56] I. Kramer, "Biomechanical changes in the lumbar section of movements," *Die Wirbelsäule in Forschung und Praxis*, vol. 58, p. 109, 1973.
- [57] T. Steffen, H. G. Baramki, R. Rubin, J. Antoniou, and M. Aebi, "Lumbar intradiscal pressure measured in the anterior and posterolateral annular regions during asymmetrical loading," *Clinical Biomechanics*, vol. 13, no. 7, pp. 495–505, 1998.
- [58] A. Bisschop, J. H. van Dieën, I. Kingma et al., "Torsion biomechanics of the spine following lumbar laminectomy: a human cadaver study," *European Spine Journal*, vol. 22, no. 8, pp. 1785–1793, 2013.
- [59] E. Karadogan and R. L. Williams, "Three-dimensional static modeling of the Lumbar spine," *Transactions of the ASME-K-Journal of Biomechanical Engineering*, vol. 134, no. 8, Article ID 084504, 2012.
- [60] A. L. Freeman, G. R. Buttermann, B. P. Beaubien, and W. E. Rochefort, "Compressive properties of fibrous repair tissue compared to nucleus and annulus," *Journal of Biomechanics*, vol. 46, no. 10, pp. 1714–1721, 2013.
- [61] Y. Schroeder, D. M. Elliott, W. Wilson, F. P. T. Baaijens, and J. M. Huyghe, "Experimental and model determination of human intervertebral disc osmoviscoelasticity," *Journal of Orthopaedic Research*, vol. 26, no. 8, pp. 1141–1146, 2008.
- [62] W. D. Merryman, K. Loveless, and M. Kasra, "Disc nucleus cellular response to dynamic pressures at critical frequencies: a pig model," in *Proceedings of the ASME International Mechanical Engineering Congress*, pp. 261–262, New York, NY, USA, November 2003.
- [63] W. Johannessen and D. M. Elliott, "Effects of degeneration on the biphasic material properties of human nucleus pulposus in confined compression," *Spine*, vol. 30, no. 24, pp. E724–E729, 2005.
- [64] G. D. O'Connell, E. J. Vresilovic, and D. M. Elliott, "Human intervertebral disc internal strain in compression: the effect of disc region, loading position, and degeneration," *Journal of Orthopaedic Research*, vol. 29, no. 4, pp. 547–555, 2011.
- [65] A. Tsantrizos, K. Ito, M. Aebi, and T. Steffen, "Internal strains in healthy and degenerated lumbar intervertebral discs," *Spine*, vol. 30, no. 19, pp. 2129–2137, 2005.
- [66] J. M. Cloyd, N. R. Malhotra, L. Weng, W. Chen, R. L. Mauck, and D. M. Elliott, "Material properties in unconfined compression of human nucleus pulposus, injectable hyaluronic acid-based hydrogels and tissue engineering scaffolds," *European Spine Journal*, vol. 16, no. 11, pp. 1892–1898, 2007.
- [67] T. P. Driscoll, R. H. Nakasone, S. E. Szczyzny, D. M. Elliott, and R. L. Mauck, "Biaxial mechanics and inter-lamellar shearing of stem-cell seeded electrospun angle-ply laminates for annulus fibrosus tissue engineering," *Journal of Orthopaedic Research*, vol. 31, no. 6, pp. 864–870, 2013.
- [68] X. Li, Q. Feng, Y. Jiao, and F. Cui, "Collagen-based scaffolds reinforced by chitosan fibres for bone tissue engineering," *Polymer International*, vol. 54, no. 7, pp. 1034–1040, 2005.
- [69] A. Joshi, S. Mehta, E. Vresilovic, A. Karduna, and M. Marco-longo, "Nucleus implant parameters significantly change the compressive stiffness of the human lumbar intervertebral disc," *Journal of Biomechanical Engineering*, vol. 127, no. 3, pp. 536–540, 2005.
- [70] P. R. van den Broek, J. M. Huyghe, and K. Ito, "Biomechanical behavior of a biomimetic artificial intervertebral disc," *Spine*, vol. 37, no. 6, pp. E367–E373, 2012.
- [71] R. D. Bowles, H. H. Gebhard, R. Härtl, and L. J. Bonassar, "Tissue-engineered intervertebral discs produce new matrix, maintain disc height, and restore biomechanical function to the rodent spine," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 32, pp. 13106–13111, 2011.
- [72] D. M. K. Aladin, K. M. C. Cheung, A. H. W. Ngan et al., "Nanostructure of collagen fibrils in human nucleus pulposus and its correlation with macroscale tissue mechanics," *Journal of Orthopaedic Research*, vol. 28, no. 4, pp. 497–502, 2010.
- [73] T. D. Nguyen, R. E. Jones, and B. L. Boyce, "Modeling the anisotropic finite-deformation viscoelastic behavior of soft fiber-reinforced composites," *International Journal of Solids and Structures*, vol. 44, no. 25–26, pp. 8366–8389, 2007.
- [74] G. Limbert and M. Taylor, "On the constitutive modeling of biological soft connective tissues. A general theoretical framework and explicit forms of the tensors of elasticity for strongly anisotropic continuum fiber-reinforced composites at finite strain," *International Journal of Solids and Structures*, vol. 39, no. 8, pp. 2343–2358, 2002.
- [75] A. E. Ehret, M. Itskov, and G. W. Weinhold, "A micromechanically motivated model for the viscoelastic behaviour of soft biological tissues at large strains," *Nuovo Cimento della Societa Italiana di Fisica C*, vol. 32, no. 1, pp. 73–80, 2009.
- [76] K. L. Goh, J. R. Meakin, R. M. Aspden, and D. W. L. Hukins, "Stress transfer in collagen fibrils reinforcing connective tissues: effects of collagen fibril slenderness and relative stiffness," *Journal of Theoretical Biology*, vol. 245, no. 2, pp. 305–311, 2007.

- [77] N. L. Nerurkar, S. Sen, B. M. Baker, D. M. Elliott, and R. L. Mauck, "Dynamic culture enhances stem cell infiltration and modulates extracellular matrix production on aligned electrospun nanofibrous scaffolds," *Acta Biomaterialia*, vol. 7, no. 2, pp. 485–491, 2011.
- [78] A. E. Baer, T. A. Laursen, F. Guilak, and L. A. Setton, "The micromechanical environment of intervertebral disc cells determined by a finite deformation, anisotropic, and biphasic finite element model," *Journal of Biomechanical Engineering*, vol. 125, no. 1, pp. 1–11, 2003.
- [79] X. Li, L. Wang, Y. Fan, Q. Feng, F. Z. Cui, and F. Watari, "Nanostructured scaffolds for bone tissue engineering," *Journal of Biomedical Materials Research A*, vol. 101, no. 8, pp. 2424–2435, 2013.
- [80] X. Li, R. Cui, W. Liu et al., "The use of nano-scaled fibers or tubes to improve biocompatibility and bioactivity of biomedical materials," *Journal of Nanomaterials*, vol. 2013, Article ID 728130, 16 pages, 2013.
- [81] X. Li, Y. Fan, and F. Watari, "Current investigations into carbon nanotubes for biomedical application," *Biomedical Materials*, vol. 5, no. 2, Article ID 022001, 2010.
- [82] X. Li, H. Liu, X. Niu et al., "The use of carbon nanotubes to induce osteogenic differentiation of human adipose-derived MSCs in vitro and ectopic bone formation in vivo," *Biomaterials*, vol. 33, no. 19, pp. 4818–4827, 2012.
- [83] X. Li, L. Wang, Y. Fan, Q. Feng, and F. Z. Cui, "Biocompatibility and toxicity of nanoparticles and nanotubes," *Journal of Nanomaterials*, vol. 2012, Article ID 548389, 19 pages, 2012.
- [84] J. C. Iatridis, S. B. Nicoll, A. J. Michalek, B. A. Walter, and M. S. Gupta, "Role of biomechanics in intervertebral disc degeneration and regenerative therapies: what needs repairing in the disc and what are promising biomaterials for its repair?" *Spine Journal*, vol. 13, no. 3, pp. 243–262, 2013.
- [85] D. J. Hamilton, C. A. Séguin, J. Wang, R. M. Pilliar, and R. A. Kandel, "Formation of a nucleus pulposus-cartilage endplate construct in vitro," *Biomaterials*, vol. 27, no. 3, pp. 397–405, 2006.
- [86] A. D. Diwan, H. K. Parvataneni, S. N. Khan, H. S. Sandhu, F. P. Girardi, and F. P. Cammisa Jr., "Current concepts in intervertebral disk restoration," *Orthopedic Clinics of North America*, vol. 31, no. 3, pp. 453–464, 2000.
- [87] B. M. Holzapfel, J. C. Reichert, J. T. Schantz et al., "How smart do biomaterials need to be? A translational science and clinical point of view," *Advanced Drug Delivery Reviews*, vol. 65, no. 4, pp. 581–603, 2013.
- [88] I. Ahmed, I. A. Jones, A. J. Parsons et al., "Composites for bone repair: phosphate glass fibre reinforced PLA with varying fibre architecture," *Journal of Materials Science*, vol. 22, no. 8, pp. 1825–1834, 2011.
- [89] C. V. Rahman, G. Kuhn, L. J. White et al., "PLGA/PEG-hydrogel composite scaffolds with controllable mechanical properties," *Journal of Biomedical Materials Research B*, vol. 101, no. 4, pp. 648–655, 2013.
- [90] D. Mikociak, S. Blazewicz, and J. Michalowski, "Biological and mechanical properties of nanohydroxyapatite-containing carbon/carbon composites," *International Journal of Applied Ceramic Technology*, vol. 9, no. 3, pp. 468–478, 2012.
- [91] A. Gloria, D. Ronca, T. Russo et al., "Technical features and criteria in designing fiber-reinforced composite materials: from the aerospace and aeronautical field to biomedical applications," *Journal of Applied Biomaterials and Biomechanics*, vol. 9, no. 2, pp. 151–163, 2011.
- [92] N. A. Langrana, J. R. Parsons, C. K. Lee, M. Vuono-Hawkins, S. W. Yang, and H. Alexander, "Materials and design concepts for an intervertebral disc spacer. I: fiber-reinforced composite design," *Journal of Applied Biomaterials*, vol. 5, no. 2, pp. 125–132, 1994.
- [93] L. Ambrosio, P. A. Netti, S. Iannace, S. J. Huang, and L. Nicolais, "Composite hydrogels for intervertebral disc prostheses," *Journal of Materials Science*, vol. 7, no. 5, pp. 251–254, 1996.
- [94] X. Li, C. A. van Blitterswijk, Q. Feng, F. Cui, and F. Watari, "The effect of calcium phosphate microstructure on bone-related cells in vitro," *Biomaterials*, vol. 29, no. 23, pp. 3306–3316, 2008.
- [95] K. D. Hudson, M. Alimi, P. Grunert, R. Hartl, and L. J. Bonassar, "Recent advances in biological therapies for disc degeneration: tissue engineering of the annulus fibrosus, nucleus pulposus and whole intervertebral discs," *Current Opinion in Biotechnology*, vol. 24, no. 5, pp. 872–879, 2013.
- [96] V. Beachley and X. Wen, "Fabrication of nanofiber reinforced protein structures for tissue engineering," *Materials Science and Engineering C*, vol. 29, no. 8, pp. 2448–2453, 2009.
- [97] J. M. Moran, D. Pazzano, and L. J. Bonassar, "Characterization of polylactic acid-polyglycolic acid composites for cartilage tissue engineering," *Tissue Engineering*, vol. 9, no. 1, pp. 63–70, 2003.
- [98] X. M. Li, Y. Huang, L. S. Zheng et al., "Effect of substrate stiffness on the functions of rat bone marrow and adipose tissue derived mesenchymal stem cells in vitro," *Journal of Biomedical Materials Research A*, 2013.
- [99] X. Li, H. Liu, X. Niu et al., "Osteogenic differentiation of human adipose-derived stem cells induced by osteoinductive calcium phosphate ceramics," *Journal of Biomedical Materials Research B*, vol. 97, no. 1, pp. 10–19, 2011.
- [100] P. Malladi, Y. Xu, M. Chiou, A. J. Giaccia, and M. T. Longaker, "Effect of reduced oxygen tension on chondrogenesis and osteogenesis in adipose-derived mesenchymal cells," *The American Journal of Physiology*, vol. 290, no. 4, pp. C1139–C1146, 2006.