



OPINION ARTICLE

REVISED Recommendations to enable drug development for inherited neuropathies: Charcot-Marie-Tooth and Giant Axonal Neuropathy [v2; ref status: indexed, <http://f1000r.es/3am>]

Lori Sames¹, Allison Moore^{2,3}, Renee Arnold^{4,5}, Sean Ekins^{1-4,6-9}

¹Hannah's Hope Fund, Rexford, NY, 12148, USA

²BioGAN Therapeutics, Rexford, NY, 12148, USA

³Hereditary Neuropathy Foundation, New York, NY, 10016, USA

⁴Arnold Consultancy & Technology LLC, New York, NY, 10023, USA

⁵Master of Public Health Program, Mount Sinai School of Medicine, New York, NY, 10029, USA

⁶Collaborations in Chemistry, Fuquay Varina, NC27526, USA

⁷Department of Pharmaceutical Sciences, University of Maryland, Baltimore, MD, 21201, USA

⁸Department of Pharmacology, Rutgers-Robert Wood Johnson Medical School, Piscataway, NJ, 08854, USA

⁹Division of Chemical Biology and Medicinal Chemistry, UNC Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC, 27599-7355, USA

v2 First published: 03 Apr 2014, 3:83 (doi: [10.12688/f1000research.3751.1](https://doi.org/10.12688/f1000research.3751.1))
 Latest published: 22 Apr 2014, 3:83 (doi: [10.12688/f1000research.3751.2](https://doi.org/10.12688/f1000research.3751.2))

Abstract

Approximately 1 in 2500 Americans suffer from Charcot-Marie-Tooth (CMT) disease. The underlying disease mechanisms are unique in most forms of CMT, with many point mutations on various genes causing a toxic accumulation of misfolded proteins. Symptoms of the disease often present within the first two decades of life, with CMT1A patients having reduced compound muscle and sensory action potentials, slow nerve conduction velocities, sensory loss, progressive distal weakness, foot and hand deformities, decreased reflexes, bilateral foot drop and about 5% become wheelchair bound. In contrast, the ultra-rare disease Giant Axonal Neuropathy (GAN) is frequently described as a recessively inherited condition that results in progressive nerve death. GAN usually appears in early childhood and progresses slowly as neuronal injury becomes more severe and leads to death in the second or third decade. There are currently no treatments for any of the forms of CMTs or GAN. We suggest that further clinical studies should analyse electrical impedance myography as an outcome measure for CMT. Further, additional quality of life (QoL) assessments for these CMTs are required, and we need to identify GAN biomarkers as well as develop new genetic testing panels for both diseases. We propose that using the [Global Registry of Inherited Neuropathy \(GRIN\)](#) could be useful for many of these studies. Patient advocacy groups and professional organizations (such as the Hereditary Neuropathy Foundation (HNF), Hannah's Hope Fund (HHF), The Neuropathy Association (TNA) and the American Association of Neuromuscular and Electrodiagnostic Medicine (AANEM)) can play a central role in educating clinicians and patients. Undertaking these studies will assist in the correct diagnosis of disease recruiting patients for clinical studies, and will ultimately improve the endpoints for clinical trials. By addressing obstacles that prevent industry investment in various forms of inherited neuropathies, we can envision treatment options for these rare diseases in the near future.

Article Status Summary

Referee Responses

Referees	1	2
v1 published 03 Apr 2014	 report 1	 report 1
v2 published 22 Apr 2014 REVISED	 report	

- 1 **Flávia L. Osório**, University of São Paulo Brazil
- 2 **Jean-Michel Vallat**, CHU Limoges France

Latest Comments

No Comments Yet

Corresponding author: Sean Ekins (ekinssean@yahoo.com)

How to cite this article: Sames L, Moore A, Arnold R and Ekins S. **Recommendations to enable drug development for inherited neuropathies: Charcot-Marie-Tooth and Giant Axonal Neuropathy [v2; ref status: indexed, <http://f1000r.es/3am>]** *F1000Research* 2014, **3**:83 (doi: [10.12688/f1000research.3751.2](https://doi.org/10.12688/f1000research.3751.2))

Copyright: © 2014 Sames L *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution Licence](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Data associated with the article are available under the terms of the [Creative Commons Zero "No rights reserved" data waiver](#) (CC0 1.0 Public domain dedication).

Grant information: Some of this work was funded by the Centers for Disease Control and Prevention (CDC) Grant # U38DD000713-0. *The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

Competing interests: L.S. is founder of Hannah's Hope Fund and CEO BioGan Therapeutics, LLC. A.M. is founder of Hereditary Neuropathy Foundation and president of BioGan Therapeutics. S.E. works for Collaborations in Chemistry, and Consults for Hannah's Hope Fund, the Hereditary Neuropathy Foundation and Arnold Consultancy & Technology, LLC.

First published: 03 Apr 2014, **3**:83 (doi: [10.12688/f1000research.3751.1](https://doi.org/10.12688/f1000research.3751.1))

First indexed: 14 Apr 2014, **3**:83 (doi: [10.12688/f1000research.3751.1](https://doi.org/10.12688/f1000research.3751.1))

REVISED Amendments from Version 1

In response to reviewer 1 we have added the three references to manuscripts or abstracts as suggested. We have made minor changes to the text, for example replacing 'biopsy' with 'sample' and small rephrasing of some sentences pointed to by reviewer 1. In response to reviewer 2 we moved the two figures to the supplemental materials.

See referee reports

Introduction

There are currently no approved treatments for any of the forms of inherited neuropathy (IN), which are each considered rare diseases. Charcot-Marie-Tooth (CMT) Disease 1A is the most common form of IN. The underlying disease mechanisms are unique in the different forms of CMT (*e.g.* CMT1A, CMT2, *etc.*)¹. To date, at least 51 causal genes have been implicated in CMT². CMT Type 1 diseases are demyelinating and Type 2s are axonal neuropathies³. Based on the prevalence of all CMTs being 1:2500 and the recent census figures of over 300 million people in the United States of America (USA), it is likely that more than 75,000 Americans have CMT1A, which is caused by the peripheral myelin protein 22 (PMP22) duplication gene phenotype, which represents 68.7% of all CMT Type 1s in the USA⁴. Symptoms of the disease often present in the first two decades of life⁵, with patients having reduced compound muscle and sensory action potentials, slow nerve conduction velocities, sensory loss, progressive distal weakness, foot and hand deformities, decreased reflexes, bilateral foot drop and about 5% becoming wheelchair bound. Of the axonopathies, CMT2A is the most common form. CMT2A is primarily caused by mutations in the gene encoding mitofusin (*MFN2*)⁶. Mutations or rearrangements in *PMP22*, myelin protein zero (MPZ) and gap junction β 1-protein (GJB1) or *MFN2* account for 90% of molecular diagnoses⁷. CMT2A has been described to severely affect infants and children, with most affected individuals non-ambulatory by age 20, and having profound proprioception loss in addition to weakness⁶.

Other than braces or other orthopedic devices, there are no effective therapies for any of the various forms of CMTs⁸. Unfortunately, CMT1A is also a textbook case of how ground-breaking fundamental discoveries by academic scientists are not being rapidly translated into therapeutics quickly enough. For example, the causal gene duplication defect for the most common form of CMT (PMP22) was identified in 1991^{9,10} and confirmation that the resulting elevated gene dosage is responsible for the pathology of the disease was established in 1993, yet the first high throughput screen (HTS) to search for a small molecule therapeutic was not published until 2012¹¹. This represents a 21-year gap in translation from gene to screen. The tide is now slowly changing and a small pipeline of molecules currently exists for CMT1A. We are aware of at least two companies with preclinical and clinical compounds. Recently, a [press release](#) from Addex Therapeutics reported that the preclinical compound Addex ADX71441 reduces PMP22 expression in a dose-dependent manner comparable to baclofen in a pre-clinical transgenic mouse model of CMT1A disease. This drug is a new molecular entity that will require extensive preclinical testing, thus it is many years (possibly a decade) from US Food and Drug

Administration (FDA) approval. Pharnext is a French pharmaceutical company which has just completed a **Phase II trial** (manuscript in preparation) for a combination therapy (baclofen, naltrexone and sorbitol) for CMT1A^{12,13}. All these components are already FDA-approved, so if the Phase III clinical trial (proposed for 2014, Dr. Daniel Cohen, Pharnext personal communication) is successful, this could be the first treatment to market for any form of IN. In addition, the coming five years will likely be an explosive time in CMT2A research as the long-awaited, very difficult to engineer, CMT2A knock out mouse model is being characterized by Dr. David Pleasure (University of California, Davis). Preliminary data reveal that this model will be useful for better understanding the mechanisms behind the disease (molecular targets) — and will represent a putative model for testing the efficacy and safety of investigational therapeutics.

Recent clinical studies of other agents targeted for the treatment of CMT1A have not been successful, including the use of high-dose ascorbic acid, which exemplified the need for better therapeutic options¹⁴. Further, a 5-year longitudinal study also using ascorbic acid in subjects with CMT1A, failed to identify a physiologically significant biomarker¹⁵ in patients vs controls. Indeed, “*at the end of the study, 30 patients (68%) and 2 controls (8%) stated that their physical condition had deteriorated during the 5-year study period*”¹⁵. Verhamme *et al.* hypothesize that early weakness, summarized as *pes cavus*, lack of reserves, *etc.*, contribute to increased disability in patients, as they aged at the same rate as controls (same decrease in muscle strength and compound Muscle Action Potential [CMAP])¹⁵. The Inherited Neuropathy Consortium Rare Diseases Clinical Research Network (RDCRN) website (as of October 30th 2013) lists the following six studies: 6601: Natural History Evaluation of Charcot Marie Tooth Disease; 6602: Genetics of CMT-Modifiers of CMT1A, New Causes of CMT2; 6603: Development of CMT Peds Scale for Children with CMT; 6606: An Analysis of the Symptomatic Domains Most Relevant to Charcot Marie Tooth Neuropathy (CMT) Patients; 6608: Survey of Current Management of Orthopedic Complications in Charcot Marie Tooth Disease Patients; 6604: Development and Validation of a Disability Severity Index for Charcot Marie Tooth Disease. All but studies 6604 and 6606 are listed as still recruiting. These clinical studies are also listed on [clinicaltrials.gov](#) with “unknown” status, indicating further the suboptimal progression of research studies in these diseases to date.

In contrast to CMT, which can vary in severity and is relatively prevalent, GAN is the most severe form of IN. GAN is an “ultra-rare” disease with tens of patients globally (with many likely to be undiagnosed¹⁶), but neurologists suspect that some CMT Type 2 patients whose causal gene remains unknown may actually have GAN¹⁷ greatly expanding the patient population. GAN generally appears in early childhood and progresses slowly as the neuronal injury becomes more severe. GAN is a single-gene, autosomal recessive disorder that is fatal by the late teens or early 20s. The *GAN* gene encodes the protein gigaxonin¹⁸. Gigaxonin is an intracellular protein needed for long-term nerve survival. It is a predicted E3 ligase adaptor protein believed to flag substrates for ubiquitination on the proteasome¹⁹. Recent studies also suggest that disturbed cytoskeletal regulation, likely involving the proteasome degradation pathway²⁰, is responsible for the formation of aggregates of

some type 3 and type 4 intermediate filament proteins, which is a morphological characteristic of this disease, and many others.

Based on histopathological analysis of large axonal swellings, “giant axons” are filled with neurofilament (NF) triplet proteins (neurofilament light chain [NF-L], neurofilament medium chain [NF-M] and neurofilament heavy chain [NF-H]) in both the central and peripheral nervous systems of GAN patients²¹. The knockout GAN *-/-* mouse has a similar histological phenotype, with increased levels of NF proteins, α -internexin, peripherin, as well as vimentin, with mRNA transcription levels remaining normal. Additionally, histological analysis of brain sections from the GAN *-/-* mouse has revealed accumulations of NF-H and of α -internexin in the cortex (negative for other NF subunits)²¹.

Using three cellular models, Mahammad *et al.*²⁰ showed that restoring functional gigaxonin in cultured cells had a direct impact on vimentin, peripherin and NF-L. Using GAN patient fibroblasts, they were able to clear the vimentin aggregates by expressing wild type (WT) gigaxonin. Using a rat PC12 cell line stably expressing WT gigaxonin, they were also able to show the loss of peripherin in these cells. Lastly, using a human neuroblastoma cell line, SH-SY-5Y that stably expresses WT gigaxonin, they revealed clearance of NF-L protein by both immunoblotting and immunofluorescence. Since their manuscript²⁰ was accepted for publication, the Goldman and Opal laboratories have obtained evidence that gigaxonin also regulates the turnover of glial fibrillary acidic protein (GFAP) in astrocytes (Dr. Robert Goldman, Northwestern University Feinberg School of Medicine, personal communication).

Likewise, Mussche *et al.* were able to demonstrate proof-of-concept of GAN gene delivery *in vitro* and *in vivo* by clearing IF aggregates. They showed that the restoration of WT gigaxonin with adeno-associated virus vector serotype 2 (AAV2) clears vimentin IF aggregates in the fibroblasts of GAN patients. Using an AAV9 viral vector, (the same vector being used in the GAN gene delivery clinical trials that HHF is sponsoring), GAN knockout mice received an intracisternal injection of an AAV9/GAN vector that allowed the delivery of the GAN gene to the brainstem and spinal cord. The authors observed that “the treated mice showed a nearly complete clearance of peripherin IF accumulations in the brainstem and spinal cord at 3 weeks post-injection”²². These studies demonstrated that gigaxonin gene transfer can reverse the cellular IF aggregate pathology associated with GAN. Based on this and other published studies (described earlier), it is fair to speculate, that vimentin, peripherin, NF-L, NF-M, NF-H, α -internexin and GFAP could be used as putative surrogate biomarkers of disease severity.

As GAN progresses (through effects on the CNS), patients typically become quadriplegics, dependent on a feeding tube and ventilator, and usually die in the second or third decade. The majority of GAN patients that have been identified to date have also had extremely kinky hair. However, there are two confirmed GAN cases with straight hair, further suggesting CMT as a differential diagnosis for GAN²³. Some pathological factors in GAN, like neurofilament light chain (NF-L) and peripherin accumulation, are also hallmarks of many other diseases²⁴ including Alexander disease (AxD), amyotrophic lateral sclerosis (ALS), Alzheimer’s disease,

Parkinson’s disease, dementia with Lewy bodies, neuronal IF inclusion disease (NIFID), diabetic neuropathy, spinal muscular atrophy (SMA), and some forms of CMT (such as CMT 2E)^{20,25}. Thankfully, basic cellular biology research has caught up with clinical research for GAN. Patient cerebrospinal fluid (CSF) samples that have been collected over the past three years can finally be used in pilot studies now that putative surrogate biomarkers have finally been elucidated²⁰.

Over the past five years, HHF, through grassroots efforts, has funded the development and study of a novel GAN gene delivery investigational new biologic (INB) for the treatment of GAN. The gene delivery product is an AAV9 capsid with a self-complementary genome containing the WT-gigaxonin transgene (Dr. Steven Gray, UNC personal communication). This group has achieved broad distribution of the vector throughout the central nervous system (CNS) of both non-human primates and pigs with transduction of the transgene found even deep into the brain parenchyma²². NINDS is sponsoring the Phase I trials that will take place at the National Institute of Health (NIH) Clinical Center in Bethesda, MD, USA. US Recombinant DNA Advisory Committee (RAC) approval was obtained during a public hearing in Washington DC that took place on June 12, 2013. Assuming FDA and (IRB) approvals, a Phase I trial is expected to begin in the second quarter of 2014.

With research reaching such advanced stages we propose that some efforts should be made to fill the various gaps in clinical studies, in order to translate therapeutics to patients more rapidly for the related diseases CMT1A, CMT2A and GAN. An important component will be to train clinicians, identify the diseases and provide information to hereditary neuropathy patients to help them manage their condition. In addition, there need to be continued efforts to bring new patients to the attention of researchers for clinical studies and energize pharmaceutical companies and investors to pursue treatments for hereditary neuropathies.

We propose that in order to translate these preclinical advances to the clinic and result in successful therapeutics there should be 1) alternative outcomes measures for CMT; 2) demonstration that potential treatments are having an effect on the QoL of patients; 3) genetic testing panels to facilitate identification of patients with CMT and GAN for future clinical trials; 4) validation of putative CSF biomarkers for GAN; 5) use of the GRIN database to capture patients for future clinical trials and 6) education of clinicians and patients as to therapeutic options for these diseases. Each of these topics is discussed further below.

Alternative outcome measures: electrical impedance myography

The development of therapies for patients critically depends on the outcome measures used in clinical trials. There are limited efforts to find additional outcome measures for CMT, including *in vivo* confocal microscopy of Meissner corpuscles²⁶, Anterior Tibialis CMAP amplitude²⁷, CMT neuropathy score (second version)²⁸ and patient identified QoL²⁹. We now propose that a promising technique called electrical impedance myography (EIM) could be validated as an outcome measure for CMT1A and CMT2A disease progression, facilitating the execution of therapeutic CMT clinical trials in patients.

Skulpt, Inc. (previously Convergence Medical Devices, Inc.) has developed a handheld device specifically for performing EIM measurements in patients. EIM measures the impedance of skeletal muscle over a frequency range between 1 kHz and 10 MHz. The impedance is measured at each frequency by applying low-intensity electrical current *via* surface electrodes and measuring the resulting voltage signals using a second set of surface electrodes. EIM involves only minimal risk. The estimated testing duration for an individual subject is 15 to 30 minutes, depending on the number of muscles tested. The data obtained with this handheld EIM device have shown marked alterations between healthy individuals and patients. Data from both human subjects and animal disease models, including amyotrophic lateral sclerosis (ALS), Spinal muscular atrophy (SMA), Duchenne muscular dystrophy (DMD) and neurogenic mouse models, show that EIM may be sensitive to a variety of pathological states^{30–34}. We envisage that EIM will help quantify the severity of the disease affecting various muscle groups as well as measure changes in the progression of the disease over time, which are critical aspects to determine if a therapeutic has any value. A recent DMD EIM pilot study enrolled 61 boys with DMD and 31 healthy controls and validated whether EIM allowed detection of diseased muscle tissues *vs* control tissues³⁰. Further, EIM was helpful in quantifying the change in muscle quality over time (Dr. Eugene Williams, Dart Therapeutics, personal communication). This was a single day evaluation study that showed strong inter- and intra-rater reliability. It was also a cross sectional study, which used boys over a wide age range, and allowed to correlate EIM readings with age and measures of disease progression³⁰. While this is not technically showing changes over time (a true “longitudinal study”), it does provide similar information and presents some valuable preliminary data.

Thus, EIM represents a novel, cutting-edge technology that deserves to be evaluated as to its validity in CMT and other INs.

Demonstration that potential treatments are having an effect on the quality of life of patients

Although CMT is typically a slowly progressive disease, it may have a profound negative impact on patients’ functional, emotional and social abilities, resulting in reduced QoL. A number of studies of the influence of CMT on QoL have been published in the past several years; however, they have been either in small numbers of patients, with non-validated instruments, and/or specific to non-US locations, which reduce their generalizability to US populations^{29,35–37}. These studies have employed generic QoL instruments, such as the EQ-5D^{35,38}, SF-36^{36,37,39}, the Child Health Questionnaire^{40,41}, and qualitative interviews using a semi-structured approach and open-ended questions²⁹. Moreover, different evaluations have produced opposing results. For example, a study of 20 Italian patients with follow-up at two years found worsening strength of distal muscles of upper limbs and proximal muscles of lower limbs, sensory function and walking disability, but no worsening of QoL, as measured by the SF-36³⁶. Due to the small number of patients, it was not possible to stratify according to disease severity, which may have confounded the results. The researchers hypothesized that this may have been due to adaptive mechanisms of the patients because of the slow progression of disease or lack of sensitivity of the QoL instrument to changes specific to CMT. All of these authors³⁶ called

for the development of CMT1A disease-specific QoL instruments (adult and child) as there is currently no disease-specific QoL instrument for CMT. Disease-specific instruments are more responsive to small, but important, changes in health than are generic instruments⁴². A disease-specific instrument for CMT1A would have enhanced validity (having it measure what was intended), reliability (the extent to which it discriminates between individuals in a population in a consistent manner), sensitivity to change (the ability to measure true change regardless of whether it is relevant or meaningful to the patient or clinician) and responsiveness (the ability to detect change that is important to the patient)⁴³.

Johnson and colleagues²⁹ recently completed a study using a comprehensive qualitative interview method to identify symptoms and themes that had the greatest impact on CMT1A-related QoL in 16 adult patients. The themes mentioned most frequently were difficulty with mobility and ambulation, specific-activity impairment and emotional distress, while the most frequently noted individual symptoms in this study included impaired walking, falls and tripping. It is likely given the small number of patients, these did not correlate with CMT Neuropathy Score (CMTNS) as an indication of disease severity.

Padue *et al.* recently described a survey of CMT patients and caregivers and their perspectives and perceptions of efficacy and needs⁴⁴. This cross sectional study used the Rehabilitation Access Questionnaire, the SF36 QoL questionnaire and the family APGAR. This observational survey of 123 patients enrolled through clinical and genetic testing, suggested that patients believe it is important to feel better after physical therapy. There was a discrepancy between the perception of benefit from rehabilitation for the patient *vs* the caregiver.

Barrett and Birdsall⁴⁵ also completed an online qualitative study of adults (N=82) that used a modified van Kaam phenomenological method to explore (until saturation) thoughts, perceptions and feelings, including physical, emotional, and social effects. In addition, the study explored their sense of Barrett’s power as *knowing* participation in change—involving their ability to make informed choices with the intention of freely involving themselves in creating desired changes. The extent of the impact varied with severity, attitude, coping methods, resources, available interventions and treatments. Lack of awareness in the medical community was a predominant theme, as well as extensive description of the physical, emotional, and social impacts. Regarding personal power, searching for ways to educate themselves regarding their disease, to access quality of care, to feel fulfilled despite the severity of symptoms and ways to improve QoL were themes that occupied awareness (A). Regarding choices (C), many patients reflected the ways they were coping with physical limitations and stress, choosing protective and beneficial actions, as well as choosing attitudes taking advantage of available resources, taking charge of health decisions, giving up destructive habits, finding hobbies, altruistic actions, bringing information to physicians, and making necessary career changes. Regarding freedom to act intentionally (F), the range was from most who felt no freedom to those feeling absolutely free in relation to CMT. This was most often related to available and adequate medical care by physicians who “cared”. Regarding involvement in creating

change (I), being able to find the best and affordable medical care and research findings often allowed people to make major changes. Each of these four power dimensions (A, C, F, I) represent areas for different groups (*e.g.* Hereditary Neuropathy Foundation [HNF] or others) to address when planning interventions to guide patients to on how they can be empowered to improve their QoL. In addition, 13 QoL quantitative questions were explored by Barrett and Birdsall⁴⁵ using descriptive statistics.

In summary, variable results have been found in terms of QoL domains affected, with some studies showing that social and emotional aspects were significantly affected by the disease³⁷, and others showing no long-term influence on social and emotional functioning, but instead demonstrating impacts on physical and mental health³⁶. Moreover, the impact of disease progression on QoL is unclear, as some researchers have found paradoxical outcomes of worsening disease with no or only mild deterioration in QoL^{29,36}. This is further confounded by a research study in children with CMT1A, where physical signs and symptoms were found to be of significantly greater value than psychological aspect of QoL, as relayed by their parents⁴⁰. Since there is no clear delineation of disease progression symptoms, relationship to disease severity, longitudinal follow-up, determination of best methods of measuring QoL or disease-specific instrument, future studies should evaluate changes in impact of factors. These factors should include improved medical care, education through the Internet, advocacy support from CMT organizations and other dynamics, on patients' quality of life over the past ten years. Also newer instruments, such as NIH's Patient Reported Outcomes Measurement Information System (PROMIS[®]) and Neuro-QoL, need to be used in comparison to more established tools to determine if any of these tools is/are responsive enough to capture the impact of these diseases on patients' health-related QoL (HRQoL). Neuro-QoL is a new, standardized approach to measuring HRQoL across common neurologic diseases and has been validated in multiple neurological conditions, both in adult and pediatric patients⁴⁶. PROMIS[®] is a tool defined as "*a system of highly reliable, precise measures of patient-reported health status for physical, mental, and social well-being*"⁴⁷. Further psychometric testing within CMT and between CMT and GAN will help evaluate the generalizability of these new tools. Ultimately, there should be separate development and validation of CMT1A (disease-specific) QoL instruments for adults and children for use in future clinical trials.

Develop genetic testing panels for CMT and GAN

INs are widely known as diverse in both their genetic causes and clinical phenotypes. INs are also very frequently underdiagnosed or misdiagnosed⁴⁸. In 2013 a large prospective study found that nearly half of the patients referred to specialist peripheral neuropathy clinics have an unrecognized inherited cause⁴⁹. We are aware that the sequencing company Athena/Quest Diagnostics has sequenced more than 100,000 CMT patients and claims to have the largest variant database in the world. Previous discussions have suggested that published prevalence data are outdated, and that CMT is much more prevalent than previously estimated (Dr. Stephen L. Vincent, Director of Business Development, Quest Diagnostics and Athena Diagnostics, personal communication). It is therefore important that updated prevalence data be communicated to illustrate the large and

underserved patient population in order to motivate the pharmaceutical industry to develop treatments for CMT. There should therefore be coordinated efforts for the genetic diagnosis of this community with initial focus on CMT1A, CMT2A and GAN in order to have well-defined cohorts available for upcoming clinical studies.

To date, 2000 variants have been identified on the causal gene in cystic fibrosis and 18 causal variants have been identified (7). Perhaps CMT should be considered in the same light as the Cystic Fibrosis Foundation/Vertex Pharmaceuticals apparent view of cystic fibrosis, in which each genotype (*e.g.* mutations G551D, Delta F508 *etc.*) is treated as a different disease, requiring separate drugs for patients with different mutations. This ultimately represents an approach to individualized therapy. For INs, a long journey awaits as the field is working to elucidate causal genes in many cases, not to mention the many variants on a given gene that mandate a unique therapeutic approach. This is in stark contrast to the belief and understanding of most neurologists who are content in not investigating the genetic diagnosis of patients because no treatments currently exist. This paradigm must change in order to better understand the variability in the different forms of IN. The genotype must be identified and non-disease causing variants evaluated, as this can lead to the development of therapeutic targets. With the advancements in other personalized medicine approaches such as antisense oligonucleotides, we must endeavor to obtain a thorough understanding of the genetics behind the various forms of IN; otherwise it will be difficult to obtain effective therapies. In our view the field of IN is at least 20 years behind cystic fibrosis, largely because genetic testing has not been a priority. We have coined the phrase "diagnostic complacency" to describe this situation. Just because most forms of IN are not fatal and no treatments exist, it does not mean that CMT and idiopathic neuropathy patients do not deserve to be genetically diagnosed and have the opportunity to contribute to research by sharing their phenotype/genotype relationships.

Athena/Quest Diagnostics currently provides a variety of CMT next-generation sequencing options and plans on adding newly identified genes to these test panels in early 2015 (Dr. Stephen L. Vincent, Quest Diagnostics and Athena Diagnostics, personal communication). In addition, soon they may be able to provide disease-specific exome sequencing. This will allow them to test all known CMT genes in a single test in the same timeframe that it takes to run a single test today. We are not aware of other companies doing sequencing for CMTs and GAN specifically, therefore Athena's/Quest's CMT next-generation sequencing test offerings may provide a rapid result and cost savings for idiopathic CMT subjects in contrast to running multiple single or small test panels over an extended period of time.

The development of new CMT panels that include the GAN gene and other newly-identified CMT Type 2 genes that have a phenotype and electrophysiological findings similar to GAN will be important now that a Phase I trial is planned for GAN.

GAN biomarker study

In order to assist in Phase I and Phase II trials of a first-in-human gene delivery vector delivered to the spinal cord there should be validated putative surrogate biomarkers of GAN disease progression.

CSF has long been exploited as a relatively non-invasive way to sample the CNS⁵⁰. Subjects with GAN will likely be the first disease community in the world to receive a therapeutic gene delivered to the spinal cord, paving the way for other related neurological disorders such as spinal muscular atrophy (SMA) which, according to the [National Organization for Rare Disorders \(NORD\)](#), is the leading genetic cause of death in children.

As it has been reported that gigaxonin regulates GFAP in astrocytes⁵¹, quantifying the levels of GFAP in body fluids may be useful to determine the severity and/or clinical progression of GAN. Normally, low levels of GFAP are seen in body fluids, and will thus require the development of very sensitive detection methods, such as sandwich ELISA assays⁵². CSF GFAP levels have been shown to be elevated in a number of neurological conditions, reflecting a wide range of etiologies (reviewed in⁵³). In several situations GFAP is evaluated relative to other biomarkers linked to neuronal damage, such as neurofilaments^{54,55}. Generally, the levels of GFAP are modest in the slowly progressive conditions such as dementia and multiple sclerosis (MS), and in contrast, higher but transient, in vascular accidents, trauma, and infection. The level of GFAP in CSF may have a prognostic value^{56–58} which is worth exploring. For instance, according to a study on patients with subarachnoid hemorrhage, six days after the event the GFAP level was approximately 8-fold higher in non-survivors compared to survivors⁵⁸. It has been described that GFAP levels in CSF seem to drop in response to treatment in patients with neuromyelitis optica given corticosteroid therapy⁵⁹.

Alexander disease (closely related to GAN) is caused by dominantly-acting gain of functional mutations in GFAP. These mutations lead to increased levels of expression, measured either by mRNA or protein. GFAP has been measured in the CSF from three Alexander disease patients⁶⁰, and was elevated in them all. Since mutations in GFAP are the root cause of Alexander disease, and increased levels of GFAP within astrocytes are thought to be central to the pathogenesis⁶¹, GFAP measurements might prove especially valuable for assessing either severity or progression of this particular condition. For instance, the highest levels of GFAP in this study were found in a patient who, although intermediate in terms of age of onset, was rapidly deteriorating at the time of biopsy. Recent studies in the mouse model of Alexander disease also showed a positive correlation between the levels of protein present in brain parenchyma and the levels of protein in CSF, with an increased expression correlating with the severity of phenotype⁶².

It is perhaps not surprising that Alexander disease and GAN show interesting similarities. The hallmark pathological feature in Alexander disease is the presence of stress protein aggregates (known as “Rosenthal fibers”) within the cytoplasm of astrocytes, along with variable degrees of leukodystrophy. Indeed, the presence of Rosenthal fibers has also been described in several patients with GAN^{63–65}. It is intriguing to speculate therefore that loss of gigaxonin affects GFAP, causing accumulation and aggregation of this protein, in a manner similar to its effects on other IFs, and may even produce Alexander disease–like pathology. A recent authoritative review on the MRI features of leukodystrophies commented on the similarities between GAN and Alexander disease but was limited to focusing on lesions in the white matter⁶⁶. We could predict that

GFAP levels will be increased in the CSF of GAN patients and will likely rise with the progression of the disease. We further hypothesize that GFAP levels will stabilize and potentially fall in response to treatment, thus serving as a useful biomarker for the GAN gene delivery clinical trial. It will be interesting to see if the abundance of GFAP, NF-H, NF-M, NF-L, vimentin, α -internexin and peripherin (all proteins implicated in GAN morphology) in CSF, changes over time and correlates with disease severity in the future trial. It is also intriguing to think that correlating NF-L abundance in CSF to disease progression in GAN may also translate to CMT2E, which is caused by dominant point mutations in NF-L. Most intriguing is the prospect that the markers for this devastating disorder in the pilot study could potentially be valuable surrogate endpoints demonstrating efficacy of the first-in-human AAV9 GAN gene delivery trials.

Global Registry for Inherited Neuropathy (GRIN)

A patient registry is a place to store detailed information about patients with a specific disease or syndrome. In general, collection, analysis and dissemination of data on disease progression and patient responses to long-term disease management strategies represents a valuable way to improve understanding of the disease and keep medical professionals up to date on the latest advances. The current Inherited Neuropathy Consortium (INC) has a very basic contact registry only for CMT1A, CMT1B, CMT2A, CMTX, CMT4, other known CMT peripheral neuropathy and other unknown CMT peripheral neuropathy. In contrast, the HNF & HHF launched the [Global Registry for Inherited Neuropathies \(GRIN, Supplemental Figure 1\)](#) – in 2013 with the primary objective of identifying patients with INs to gain a better understanding of disease phenotypes, which vary among all the different types and sub-types of CMT and related INs. This registry is far more extensive ([Supplemental Figure 2–Supplemental Figure 10](#)) than anything else available for INs such as basic contact registries, as it covers all INs and is managed by [Patient Crossroads](#), which is a professional organization that has developed registries for more than 250 diseases. These registries are used by participants in more than 75 countries. GRIN can be used to connect patients, healthcare providers, researchers, clinical investigators, regulators, policy-makers industry and beyond.

The GRIN collects data on patients who have either been clinically diagnosed by a doctor or genetically diagnosed with a form of IN ([Supplemental Figure 1](#)). Establishing the registry addresses three critical needs. First, the scientists who study various forms of IN need accurate information to understand the specific forms of neuropathy; second, the scientists who are conducting preclinical studies may need tissue samples from registrants who have already agreed to be contacted by GRIN if samples are needed to advance research for their form of IN. Additionally, clinicians, pharmaceutical and biotechnology companies ready to begin trials would need the GRIN administrator to distribute and advertise the study and clinical-trials.gov study links. The Patient Crossroads platform on which the database is built provides robust query functionality and allows researchers to narrow in on specific phenotypes and genotypes. If a patient looks like a good match for research, the scientist will contact HNF. The foundation or its agent will then contact the IN family. Scientists will not contact the families directly and will only be given de-identified patient data.

Although there is no direct benefit to any patients participating in the registry, by collecting information on patients in the registry, scientists can:

- Study why individuals have different symptoms.
- Learn about phenotype/genotype relationships (how a particular mutation type can lead to different or unique symptoms).
- Learn how future treatments work or don't work for a given patient population.
- Obtain tissue samples from registrants who agreed to be contacted for such purpose.
- Help medical professionals improve how they treat patients with IN, for example, pain management.
- Speed up research in INs by collecting information that scientists can use.
- Let patients (or their families) know when they may be eligible for clinical research studies (clinical trials).
- Let pharmaceutical companies know the number of patients with a particular form of IN who would be willing to consider volunteering for experimental trials.

This longitudinal database of patient-contributed data includes the ability for patients to create an account that allows them to login and update information, complete questionnaires or provide patient-reported outcome measures and track results, compare their responses to the de-identified responses of other registry participants, and upload medical records for use in identifying confirmed diagnosis of hereditary neuropathy patients. GRIN also provides the ability to monitor registration metrics, respond to registrant inquiries, add researcher-specific questions, and export de-identified data in CSV format that is widely accepted by statistical and scientific software.

GRIN currently collects data on meaningful characteristics such as *pes cavus*, absent-to-poor reflexes, toe walking, mobility devices on various levels of disability deficits, *etc.*, management, and outcomes of disease progression with and without palliative treatments. Another critical component of the registry is to provide a better understanding of disease progression in the general population of both diagnosed and undiagnosed patients. Environmental factors and **contraindicated drugs** have been confirmed to exacerbate CMT and/or present onset of symptoms in asymptomatic patients⁶⁷. This obviously can alter disease progression and may be relevant to the natural history and clinical outcomes of treatments.

The registry has several objectives:

1. Offer additional data to enable the pharmaceutical industry to judge the risk – reward of the patient population for pursuing future clinical trials.
2. Obtain a better understanding of natural history from the perspective of the patient. Currently the patient, caregiver or parent of a minor, answers the questions, and when tests (needle electromyography, nerve conduction velocities or genetic tests) are available, patients and/or physicians upload the results to the registry or contact HNF directly.

3. Evaluate safety, efficacy, and influence of therapy on patients' QoL, which is not currently a feature of any IN registry.
4. Record patient experience in a period prior to baseline as a control for future clinical trials.
5. Help to establish relationship of genotype – phenotype on disease progression.
6. Help to define data that are clinically meaningful.

GRIN includes a comprehensive series of questions covering many areas and samples of these are now provided for: diagnosis/assessments, genetic test results, general health, neurology, orthopedic, alternative/complimentary therapies, skin/hair, clinical trials, and research/bio-repository (**Supplemental Figure 1–Supplemental Figure 10**). GRIN will help close the data gap to enable a more accurate representation of the patients' experience. The registry contains well thought out questions and was beta tested by patients, caregivers and medical professionals. GRIN is also IRB-approved through the Chesapeake IRB. HNF's diverse team of medical advisors, industry leaders, and the data used from focus groups conducted and analyzed through the National CMT Resource Center in 2010 helped to develop the purpose, objective, feasibility and the robust registry. GRIN is being marketed globally through aggressive pay-per-click campaigns (via a Google grant) to actively search out patients with CMT and other neuropathies. These marketing efforts ask them to join the registry and encourage them to be seen at one of the currently-funded RDCRN CMT Centers, Muscular Dystrophy Association Centers or by medical professionals trained to recognize, diagnose and treat CMT where patients are not economically able or are too severely affected to travel.

Training and advocacy

If we are to develop new technologies and drugs for IN then we need to continually train clinicians to be aware of the state of the art knowledge of the diseases such as **the growing list of known neurotoxins that must be avoided**. At the same time we need to educate patients about their disease and what is in the research and clinical pipeline. The training of future physician-scientists is therefore critical to accelerating research in the hereditary neuropathies field. There is also the responsibility of foundations like HNF to educate the entire medical community to be aware of hereditary neuropathies. This would lead to faster and more accurate patient diagnosis that could have an impact on medical expenses and patient QoL.

Close affiliations with other patient advocacy organizations focused on INs is also important to educate all members of the community. As indicated throughout this document, key advocates include: HNF, HNF, The Neuropathy Association, Neuropathy Action Foundation, and NORD. To date HNF has actively supported the efforts of the RDCRN Inherited Neuropathies Consortium by including new studies for recruitment in monthly newsletters to over 7,000 members and aggressive social media outreach. Platforms used include Facebook, Twitter and the **CMT Inspire Community**. With well over 1,000 patients, the Inspire CMT community has dramatic branding and is a highly targeted and engaged population with valuable content for patients and clinicians. Inspire offers access to members for clinical studies that include market research and awareness campaigns where each member receives daily or

weekly e-mails from Inspire — all aimed at getting the community to take action and move research forward. The mission of the Neuropathy Association is to bring help, hope, and healing to the estimated over 20 million people living with over 100 various forms of neuropathy. A significant segment of this population has an idiopathic neuropathy. Experts in the field have found that there is a high likelihood that many of these are actually unrecognized inherited neuropathies. The Neuropathy Association's extensive network of patients, through their membership base (over 50,000), extensive online and social media networks (35,000–45,000 unique website visitors per month and 12,000 followers on Facebook) and the 15 designated Centers of Excellence at prominent academic and medical institutions across the USA, are critical to engaging clinicians – and their patients – to participate in future clinical trials. The Neuropathy Association and the HNF recently hosted a monthly Facebook Chat (July 2013), focusing on hereditary neuropathies, with the goal of giving participants a chance to hear from — and ask questions to — health experts in the field, the Association and Foundation staff, collaborating patient advocacy groups, as well as each other. These examples show that by actively engaging the advocacy groups and their networks through social media we can reach patients which, in previous years, would have been impossible without extensive media expenditures.

The Neuropathy Association has awarded grants for scientific research on hereditary neuropathies and the supported research projects have yielded significant results. For example, Landsverk *et al.* identified the gene causing a hereditary neuropathy (hereditary neuralgic amyotrophy [HNA]) that attacks the muscles responsible for controlling the movement of shoulders and arms⁶⁸. In addition, Florian Thomas (St. Louis University, St. Louis, USA) and his international team have uncovered three different genetic mutations specific to dominant intermediate CMT neuropathy type C, a form of CMT disease⁶⁹. Organizations like The American Association of Neuromuscular and Electrodiagnostic Medicine (AANEM) could be funded to support awarding of fellowships that focus on rare neuromuscular disease research. This could have a significant impact on training and education for the IN clinical community.

Additionally, a regular Hereditary Neuropathies Workshop could serve as a forum to promote awareness of IN, encourage patient recruitment for future clinical trials and GRIN, and provide a means for encouraging more participation by other researchers and companies in the field, as well as provide a learning platform to benefit new researchers; this would encourage the researchers' development in this area and promote their interest in pursuing additional research projects in the field of hereditary neuropathies.

Conclusion

In conclusion, we have recognized the following gaps in translation and obstacles to future clinical studies for IN *via* lack of: a validated QoL instrument that is sensitive enough to capture disease impact; validated biomarkers that correlate with severity of disease progression; a large, robust, clinical patient registry that accurately captures severity and range of symptoms, along with genotypes; physician

education and awareness about IN; accurate genetic diagnoses; patient and physician education regarding the need for an early diagnosis; prevalence data; data regarding patient willingness to consider volunteering for experimental trials for various forms of IN. We have suggested some recommendations that could perhaps remedy this situation—positively impacting patients with CMT, GAN and other INs.

Author contributions

All authors conceived and contributed to the literature review, and the writing of the article. All authors read and approved the final manuscript.

Competing interests

L.S. is founder of Hannah's Hope Fund and CEO BioGan Therapeutics, LLC. A.M. is founder of Hereditary Neuropathy Foundation and president of BioGan Therapeutics. S.E. works for Collaborations in Chemistry, and Consults for Hannah's Hope Fund, the Hereditary Neuropathy Foundation and Arnold Consultancy & Technology, LLC.

Grant information

Some of this work was funded by the Centers for Disease Control and Prevention (CDC) Grant # U38DD000713-0.

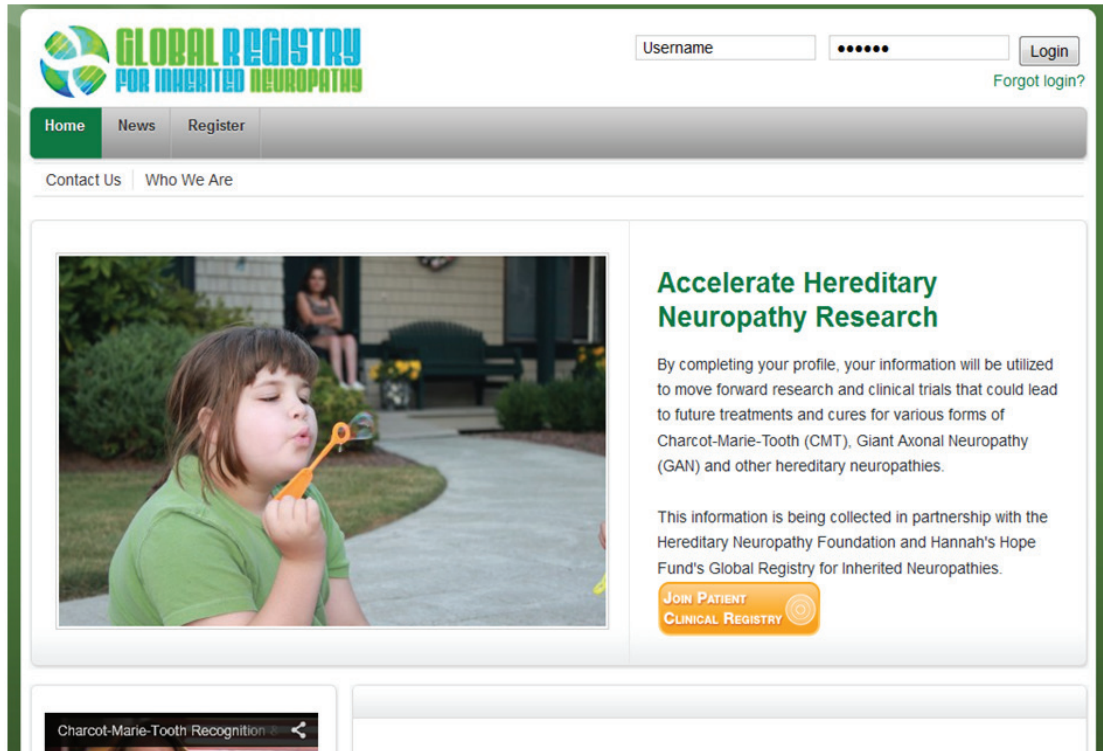
The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgments

We gratefully acknowledge all of the scientists, patient advocacy groups and other organizations we collaborate with (some of whom have kindly provided unpublished information) and the individuals and companies who have so generously supported our foundations. These include: Thomas H. Brannagan, III MD (Columbia University); Adrienne Rhodes (Blue Fountain Media); Steven Gray, PhD (University of North Carolina); Lisa M. Phelps (NORD); Victoria H. Lawson, MD (Ohio State University); Florian P. Thomas, MD (Saint Louis University); Pragna Patel, PhD (University of Southern California); Jose Bohorquez, PhD (Skulpt, Inc); Kyle Brown (PatientCrossroads); Carsten G. Bönnemann, MD (NIH); Daniel Cohen, MD, PhD (Pharnext); Shirlyn A. Adkins, JD, (AANEM); Stanley S. Young (NISS); Xavier Badia MD, MPH, PhD (University of Barcelona); Dominick V. Spatafora (Neuropathy Action Foundation); Michael Serada MD, (Max-Planck Institute of Experimental Medicine); Stephen L. Vincent, PhD, (Quest Diagnostics and Athena Diagnostics); Leslie Hinyard, PhD, MSW (Saint Louis University); Eugene Williams (Dart Therapeutics), Mickaël Guedj, PhD (Pharnext); Elizabeth A.M. Barrett (Hunter College); Natacha T. Pires, MBBS (The Neuropathy Association); Albee Messing PhD, (University of Wisconsin-Madison), Gerry Shaw PhD, (University of Florida); David Pleasure MD, (University of California-Davis); Tina M. Tockarszewsky (The Neuropathy Association); Yonglong Zhou PhD, (Arnold Consultancy & Technology); Katrina A. Gwinn MD, (NIH); Robin Conwit MD, (NIH); and Courtney Hollett (The Hereditary Neuropathy Association).

Supplementary materials

Figures relating to the Global Registry for Inherited Neuropathies.



Supplemental Figure 1. Website landing page for Global Registry for Inherited Neuropathy (GRIN).

Genetic Test Results

If you have had DNA/genetic testing, was a genetic diagnosis made? (i.e., A disease causing mutation was found?)

- A non-disease causing mutation was found
- Yes
- No
- Unsure

What are the results of the genetic test? (CMT = Charcot-Marie-Tooth, HMSN = Hereditary Motor and Sensory Neuropathy)

Please upload copies of NCV reports, muscle biopsy report, DNA testing and MRI report. If you are unable to upload please fax to 518-357-3734. Directions to upload results are provided at the end of the survey.

- No mutation was identified. (My disease causing gene is unknown.)
- Not able to interpret the genetic report (Upload results or Fax report)
- My genetic diagnosis is not listed (Upload results or Fax report)
- Andermann (KCC3 gene, Corpus Callosum)
- α -Methylacyl-CoA racemase (AMACR)
- CMT2A (Lamin A/C gene, autosomal recessive)
- CMT2B (MED25 gene, autosomal recessive)
- CMT2 (LRSAM1 gene, recessive)
- CMT2 (NEFL gene, autosomal recessive, severe & early onset)

Supplemental Figure 2. Genetic tests.

General Health

At what age did the participant first show signs of puberty? (In boys, this refers to body odor or an increase in testicle size, most commonly. In girls, this refers to body odor or breast development, most commonly.)

Please select ... ▼

For female participants, what is the age of onset of menses (age at first period)?

Please select ... ▼

Has a medical professional ever told you that you have a hormonal imbalance?

- Yes
- No
- Unsure

Has the participant been diagnosed with an autoimmune disorder?

- Yes
- No
- Unsure

Does the participant have an allergy or sensitivity to any of the following? (Select all that apply.)

Supplemental Figure 3. General health.

Neurology

Has the participant ever had, or currently has any of the following symptoms: (Select all that apply.)

- Arm or leg weakness
- Clumsiness
- Coordination/balancing problems
- Developmental delay (fine or gross motor)
- Difficulty swallowing (choking on food and/or liquids)
- Difficulty with physical activities
- Dizziness (vertigo)
- Extreme fatigue
- Flacid/loose joints that hyper-extend
- Gait disturbance
- Hearing problems
- Low muscle tone (hypotonia)
- Muscle atrophy
- Neuropathic pain
- Numbness
- Nystagmus (involuntary rapid eye movements)
- Poor circulation

Supplemental Figure 4. Neurology.

Orthopedic

Has the participant had any of the following hand or foot conditions? (Select all that apply.)

- Absent or poor reflexes
- Claw hands (curled fingers)
- Contracture of fingers or toes
- Cramps/spasms
- Drop foot
- Flat feet (collapsed arches)
- Fractures
- Hammer and/or curled toes
- Hyperextended joints (knees or elbows)
- Inversion
- Leg length discrepancy
- Pes Cavus (high arches)
- Pronation
- Toe walking
- Weak ankles
- None
- Unsure

Supplemental Figure 5. Orthopedic.

Alternative/Complimentary Therapies

Which of the following therapies are currently used? (Select all that apply.)

- Acupuncture
- Ayurveda
- Biofeedback
- Chiropractor
- Exercise
- Guided imagery
- Homeopathy (supplements)
- Massage
- Meditation
- Psychotherapy
- Relaxation Techniques
- Support groups
- Tai Chi/Chiquong
- None
- Unsure
- Other :

Supplemental Figure 6. Alternative/complimentary therapies.

Skin/Hair

Does the participant have kinky (extremely curly) hair, unlike parent's hair?

- Yes
- No
- Unsure

Is the participant's hair dry/dull (lacks luster)?

- Yes
- No
- Unsure

Does the participant have dry skin and/or eczema?

- Yes
- No
- Unsure

Next >>

Supplemental Figure 7. Skin/hair.

Clinical Trials

Is the participant CURRENTLY participating in a clinical trial or study (treatment trial or observational/non-treatment study)?

- Yes
- No
- Unsure

Provide the name, city, state/province, and country of the clinical trial or study where the participant is CURRENTLY participating.

Has the participant participated in a clinical trial that is no longer active?

- Yes
- No
- Unsure

Supplemental Figure 8. Clinical trials.

Research/Biorepository

Was cord blood saved and stored by a cord blood bank?

- Yes
- No
- Unsure

Has the participant previously donated a biospecimen (such as blood or a tissue sample) for research?

- Yes
- No
- Unsure

Provide the name, city, state/province and country of the facility where the participant donated the biospecimen.

If a FDA approved drug therapy existed for your form of hereditary neuropathy, would you take the drug if prescribed, or are your symptoms too mild to consider drug intervention?

Supplemental Figure 9. Research/biorepository.

Registry

Which of the following explains the reason why the participant chose to join this registry? (Select all that apply).

- Want to make a difference for future and current survivors of my diagnosis
- Think emotional and social support are an important part of a complete diagnosis treatment plan
- Hope results will help me and other survivors of my diagnosis live better with the disease
- Want to give back to the diagnosis community
- Because my doctor suggested it
- Because the foundation suggested it
- For participation in clinical trials
- Participate in basic research (provide tissue sample)
- All of the above
- Unsure
- Other :

How did the participant first hear about this registry? (Select all that apply.)

- Advocacy foundation
- Billboard/airport ad
- Facebook
- Friend/family

Supplemental Figure 10. Registry.



The screenshot shows the website for the Global Registry for Inherited Neuropathy. At the top left is the logo, which consists of a stylized globe icon and the text 'GLOBAL REGISTRY FOR INHERITED NEUROPATHY'. To the right of the logo is a 'Logout' button. Below the logo is a navigation menu with the following items: Home, News, Register, Registry Data, and My Profile. The main content area has a heading 'Survey Submitted' and a sub-heading 'Follow the steps below to complete your registration:'. Below this, it says 'Click on My Profile from the main menu to:' followed by a numbered list of three steps: 1. Add the participant's doctors in the "Health Care Providers" tab; 2. Upload test results directly from your computer in the "Attachments" tab following the instructions you will find there. 3. Add a family member. At the bottom of the list, there is a note: 'Note: You can log back into the Registry at any time to complete unanswered questions or update your profile. We encourage you to update your profile anytime there is a change in the participant's health status, medications, treatments, testing, or doctors. Periodically we will send you an email reminder to update your profile.'

Supplemental Figure 11. Survey submission acknowledgment.

Progress 0%

Diagnosis/Assessments

How old was the participant when the hereditary neuropathy symptoms first started?

Please select... ▼

What led to the initial medical evaluation? (Select all that apply.)

- Diagnosis of a family member
- Symptoms noticed by patient/family
- Symptoms noticed by medical provider
- Symptoms noticed by other professional (teacher, daycare, coach, etc.)
- Noticed from unrelated imaging study (MRI, CT, X-ray, ultrasound, etc.)
- Noticed from unrelated lab test (blood test, urine test, etc.)
- Noticed from an unrelated tissue test (biopsy, surgery, operation, etc.)
- None of the above
- Unsure
- Other :

Has the participant been diagnosed by a doctor as having either of the following? (Other specific types of neuropathies are listed in another section of this questionnaire.)

- Charcot-Marie-Tooth Type 1 (demyelinating)
- Charcot-Marie-Tooth Type 2 (axonal)
- No
- Unsure
- Other :

At what age was the diagnosis made?

Please select... ▼

Which type of health care provider FIRST diagnosed the hereditary neuropathy?

Please select... ▼

- Other :

Have you been diagnosed with Diabetic Neuropathy?

- Yes
- No
- Unsure

Supplemental Figure 12. Diagnosis page in Global Registry for Inherited Neuropathy (GRIN).

References

1. Lee SM, Chin LS, Li L: **Protein misfolding and clearance in demyelinating peripheral neuropathies: Therapeutic implications.** *Commun Integr Biol.* 2012; 5(1): 107–10.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
2. Shy ME: **Lessons from London.** *J Neurol Neurosurg Psychiatry.* 2012; 83(8): 767–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
3. Bort S, Sevilla T, Vilchez JJ, et al.: **[The diagnosis and prevalence of locus CMT1A duplication in Charcot-Marie-Tooth disease type 1].** *Med Clin (Barc).* 1995; 104(17): 648–52.
[PubMed Abstract](#)
4. Naef R, Suter U: **Many facets of the peripheral myelin protein PMP22 in myelination and disease.** *Microsc Res Tech.* 1998; 41(5): 359–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
5. Kaya F, Belin S, Bourgeois P, et al.: **Ascorbic acid inhibits PMP22 expression by reducing cAMP levels.** *Neuromuscul Disord.* 2007; 17(3): 248–53.
[PubMed Abstract](#) | [Publisher Full Text](#)
6. Feely SM, Laura M, Siskind CE, et al.: **MFN2 mutations cause severe phenotypes in most patients with CMT2A.** *Neurology.* 2011; 76(20): 1690–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Murphy SM, Laura M, Fawcett K, et al.: **Charcot-Marie-Tooth disease: frequency of genetic subtypes and guidelines for genetic testing.** *J Neurol Neurosurg Psychiatry.* 2012; 83(7): 706–10.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. Patel PI, Pleasure D: **Whither hope for pharmacological treatment of Charcot-Marie-Tooth disease type 1A?** *JAMA Neurol.* 2013; 70(8): 969–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. Raeymaekers P, Timmerman V, Nelis E, et al.: **Duplication in chromosome 17p11.2 in Charcot-Marie-Tooth neuropathy type 1a (CMT 1a). The HMSN Collaborative Research Group.** *Neuromuscul Disord.* 1991; 1(2): 93–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
10. Lupinski JR, Garcia CA: **Molecular genetics and neuropathy of Charcot-Marie-Tooth disease type 1A.** *Brain Pathol.* 1992; 2(4): 337–349.
[Reference Source](#)
11. Jang SW, Lopez-Anido C, MacArthur R, et al.: **Identification of drug modulators targeting gene-dosage disease CMT1A.** *ACS Chem Biol.* 2012; 7(7): 1205–1213.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Ainsworth C: **Networking for new drugs.** *Nat Med.* 2011; 17(10): 1166–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
13. Attarian S, et al.: **A phase II randomized, placebo-controlled multicenter clinical trial of three doses of PXT3003 in 80 adult patients with CMT1A treated for 1 year.** *J Peripher Nerv Syst.* 2013; 8: 197–198.
14. Lewis RA, McDermott MP, Herrmann DN, et al.: **High-dosage ascorbic acid treatment in Charcot-Marie-Tooth disease type 1A: results of a randomized, double-masked, controlled trial.** *JAMA Neurol.* 2013; 70(8): 981–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Verhamme C, de Haan RJ, Vermeulen M, et al.: **Oral high dose ascorbic acid treatment for one year in young CMT1A patients: a randomised, double-blind, placebo-controlled phase II trial.** *BMC Med.* 2009; 7: 70.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Israni A, Chakrabarty B, Gulati S, et al.: **Giant axonal neuropathy: a clinicoradiopathologic diagnosis.** *Neurology.* 2013; 82(9): 816–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
17. Yiu EM, Ryan MM: **Genetic axonal neuropathies and neuronopathies of pre-natal and infantile onset.** *J Peripher Nerv Syst.* 2012; 17(3): 285–300.
[PubMed Abstract](#) | [Publisher Full Text](#)
18. Bomont P, Cavalier L, Blondeau F, et al.: **The gene encoding gigaxonin, a new member of the cytoskeletal BTB/kelch repeat family, is mutated in giant axonal neuropathy.** *Nat Genet.* 2000; 26(3): 370–4.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Cleveland DW, Yamanaka K, Bomont P: **Gigaxonin controls vimentin organization through a tubulin chaperone-independent pathway.** *Hum Mol Genet.* 2009; 18(8): 1384–94.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
20. Mahammad S, Murthy SN, Didonna A, et al.: **Giant axonal neuropathy-associated gigaxonin mutations impair intermediate filament protein degradation.** *J Clin Invest.* 2013; 123(5): 1964–1975.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Dequen F, Bomont P, Gowing G, et al.: **Modest loss of peripheral axons, muscle atrophy and formation of brain inclusions in mice with targeted deletion of gigaxonin exon 1.** *J Neurochem.* 2008; 107(1): 253–64.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Müssche S, Devreese B, Nagabhushan Kalburgi S, et al.: **Restoration of cytoskeleton homeostasis after gigaxonin gene transfer for giant axonal neuropathy.** *Hum Gene Ther.* 2013; 24(2): 209–19.
[PubMed Abstract](#) | [Publisher Full Text](#)
23. Roth LA, Johnson-Kerner BL, Marra JD, et al.: **The absence of curly hair is associated with a milder phenotype in Giant Axonal Neuropathy.** *Neuromuscul Disord.* 2014; 24(1): 48–55.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Opal P, Goldman RD: **Explaining intermediate filament accumulation in giant axonal neuropathy.** *Rare Diseases.* 2013; 1: e25378.
[Publisher Full Text](#)
25. Fabrizi GM, Cavallaro T, Angiari C, et al.: **Charcot-Marie-Tooth disease type 2E, a disorder of the cytoskeleton.** *Brain.* 2007; 130(Pt 2): 394–403.
[PubMed Abstract](#) | [Publisher Full Text](#)
26. Almodovar JL, Ferguson M, McDermott MP, et al.: **In vivo confocal microscopy of Meissner corpuscles as a novel sensory measure in CMT1A.** *J Peripher Nerv Syst.* 2011; 16(3): 169–74.
[PubMed Abstract](#) | [Publisher Full Text](#)
27. Komyathy K, Neal S, Feely S, et al.: **Anterior tibialis CMAP amplitude correlations with impairment in CMT1A.** *Muscle Nerve.* 2013; 47(4): 493–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
28. Murphy SM, Herrmann DN, McDermott MP, et al.: **Reliability of the CMT neuropathy score (second version) in Charcot-Marie-Tooth disease.** *J Peripher Nerv Syst.* 2011; 16(3): 191–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
29. Johnson NE, Heatwole CR, Ferguson M, et al.: **Patient identification of the symptomatic impact of charcot-marie-tooth disease type 1A.** *J Clin Neuromuscul Dis.* 2013; 15(1): 19–23.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. Rutkove SB, Darras BT: **Electrical impedance myography for the assessment of children with muscular dystrophy: a preliminary study.** *J Phys Conf Ser.* 2013; 434(1): pii: 012069.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
31. Rutkove SB, Caress JB, Cartwright MS, et al.: **Electrical impedance myography as a biomarker to assess ALS progression.** *Amyotroph Lateral Scler.* 2012; 13(5): 439–45.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
32. Rutkove SB, Gregas MC, Darras BT: **Electrical impedance myography in spinal muscular atrophy: a longitudinal study.** *Muscle Nerve.* 2012; 45(5): 642–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
33. Rutkove SB, Shefner JM, Gregas M, et al.: **Characterizing spinal muscular atrophy with electrical impedance myography.** *Muscle Nerve.* 2010; 42(6): 915–21.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. Rutkove SB, Caress JB, Cartwright MS, et al.: **Electrical impedance myography correlates with standard measures of ALS severity.** *Muscle Nerve.* 2013; 49(3): 441–3.
[PubMed Abstract](#) | [Publisher Full Text](#)
35. Calvert M, Pall H, Hoppitt T, et al.: **Health-related quality of life and supportive care in patients with rare long-term neurological conditions.** *Qual Life Res.* 2013; 22(6): 1231–1238.
[PubMed Abstract](#) | [Publisher Full Text](#)
36. Padua L, Pareyson D, Aprile I, et al.: **Natural history of Charcot-Marie-Tooth 2: 2-year follow-up of muscle strength, walking ability and quality of life.** *Neurol Sci.* 2010; 31(2): 175–178.
[PubMed Abstract](#) | [Publisher Full Text](#)
37. Taniguchi JB, Elui VM, Osório FL, et al.: **Quality of life in patients with Charcot-Marie-Tooth disease type 1A.** *Arq Neuropsiquiatr.* 2013; 71(6): 392–396.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Ajay A, Walters WP, Murcko MA: **Can we learn to distinguish between “drug-like” and “non-drug-like” molecules?** *J Med Chem.* 1998; 41(18): 3314–3324.
[PubMed Abstract](#) | [Publisher Full Text](#)
39. Ware JE Jr, Sherbourne CD: **The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection.** *Med Care.* 1992; 30(6): 473–483.
[PubMed Abstract](#) | [Publisher Full Text](#)
40. Burns J, Ramchandren S, Ryan MM, et al.: **Determinants of reduced health-related quality of life in pediatric inherited neuropathies.** *Neurology.* 2010; 75(8): 726–731.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
41. Landgraf JL, Abetz L, Ware J: **The Child Health Questionnaire (CHQ): A User's Manual.** 1999, HealthAct: Boston, MA.
[Reference Source](#)
42. Wiebe S, Guyatt G, Weaver B, et al.: **Comparative responsiveness of generic and specific quality-of-life instruments.** *J Clin Epidemiol.* 2003; 56(1): 52–60.
[PubMed Abstract](#) | [Publisher Full Text](#)
43. Bryant D, Guyatt G, Arnold R: **Patient-reported outcome measures.** In *Pharmacoeconomics: from theory to practice*, R. Arnold, Editor, CRC Press (Taylor and Francis Group, LLC): Boca Raton, 2010; 149–162.
[Publisher Full Text](#)
44. Padua L, Pazzaglia C, Schenone A, et al.: **Rehabilitation for Charcot Marie tooth: a survey study of patients and familiar/caregiver perspective and perception of efficacy and needs.** *Eur J Phys Rehabil Med.* 2014; 50(1): 25–30.
[PubMed Abstract](#)
45. Barrett E, Birdsall C: **What It's Like to Live with CMT? Manuscript in preparation.**
[Reference Source](#)
46. Andersson T, Miners JO, Veronese ME, et al.: **Diazepam metabolism by human liver microsomes is mediated by both S-mephenytoin hydroxylase and CYP3A isoforms.** *Br J Clin Pharmacol.* 1994; 38(2): 131–137.
[PubMed Abstract](#) | [Free Full Text](#)

47. Anthony LB, Bennett RE, Deegan PM, *et al.*: **Cytochrome P-450 in vitro activation of cyclophosphamide (CPA) and ifosfamide (IFOS).** *Clin Pharmacol Ther.* 1996; **59**: 1996. [PubMed Abstract](#) | [Publisher Full Text](#)
48. Dyck PJ, Oviatt KF, Lambert EH: **Intensive evaluation of referred unclassified neuropathies yields improved diagnosis.** *Ann Neurol.* 1981; **10**(3): 222–6. [PubMed Abstract](#) | [Publisher Full Text](#)
49. Klein CJ, Duan X, Shy ME: **Inherited neuropathies: clinical overview and update.** *Muscle Nerve.* 2013; **48**(4): 604–22. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
50. Conly JM, Ronald AR: **Cerebrospinal fluid as a diagnostic body fluid.** *Am J Med.* 1983; **75**(1B): 102–108. [PubMed Abstract](#) | [Publisher Full Text](#)
51. Liem RK, Messing A: **Dysfunctions of neuronal and glial intermediate filaments in disease.** *J Clin Invest.* 2009; **119**(7): 1814–1824. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
52. Albrechtsen M, Sorensen PS, Gjerris F, *et al.*: **High cerebrospinal fluid concentration of glial fibrillary acidic protein (GFAP) in patients with normal pressure hydrocephalus.** *J Neurol Sci.* 1985; **70**(3): 269–274. [PubMed Abstract](#) | [Publisher Full Text](#)
53. Liem RK, Messing A: **Dysfunctions of neuronal and glial intermediate filaments in disease.** *J Clin Invest.* 2009; **119**(7): 1814–1824. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
54. Blennow M, Sävmann K, Ilves P, *et al.*: **Brain-specific proteins in the cerebrospinal fluid of severely asphyxiated newborn infants.** *Acta Paediatr.* 2001; **90**(10): 1171–1175. [PubMed Abstract](#) | [Publisher Full Text](#)
55. Tullberg M, Rosengren L, Blomsterwall E, *et al.*: **CSF neurofilament and glial fibrillary acidic protein in normal pressure hydrocephalus.** *Neurology.* 1998; **50**(4): 1122–1127. [PubMed Abstract](#) | [Publisher Full Text](#)
56. Blennow M, Rosengren L, Jonsson S, *et al.*: **Glial fibrillary acidic protein is increased in the cerebrospinal fluid of preterm infants with abnormal neurological findings.** *Acta Paediatr.* 1996; **85**(4): 485–489. [PubMed Abstract](#) | [Publisher Full Text](#)
57. Norgren N, Sundström P, Svenningsson A, *et al.*: **Neurofilament and glial fibrillary acidic protein in multiple sclerosis.** *Neurology.* 2004; **63**(9): 1586–1590. [PubMed Abstract](#) | [Publisher Full Text](#)
58. Petzold A, Keir G, Kerr M, *et al.*: **Early identification of secondary brain damage in subarachnoid hemorrhage: a role for glial fibrillary acidic protein.** *J Neurotrauma.* 2006; **23**(7): 1179–1184. [PubMed Abstract](#) | [Publisher Full Text](#)
59. Misu T, Takano R, Fujihara K, *et al.*: **Marked increase in cerebrospinal fluid glial fibrillary acidic protein in neuromyelitis optica: an astrocytic damage marker.** *J Neurol Neurosurg Psychiatry.* 2009; **80**(5): 575–577. [PubMed Abstract](#) | [Publisher Full Text](#)
60. Kyllerman M, Rosengren L, Wiklund LM, *et al.*: **Increased levels of GFAP in the cerebrospinal fluid in three subtypes of genetically confirmed Alexander disease.** *Neuropediatrics.* 2005; **36**(5): 319–323. [PubMed Abstract](#) | [Publisher Full Text](#)
61. Messing A, Brenner M, Feany MB, *et al.*: **Alexander disease.** *J Neurosci.* 2012; **32**(15): 5017–5023. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
62. Jany PL, Hagemann TL, Messing A: **GFAP expression as an indicator of disease severity in mouse models of Alexander disease.** *ASN Neuro.* 2013; **5**(1): e00109. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
63. Kretzschmar HA, Berg BO, Davis RL: **Giant axonal neuropathy. A neuropathological study.** *Acta Neuropathol.* 1987; **73**(2): 138–144. [PubMed Abstract](#) | [Publisher Full Text](#)
64. Kumar K, Barre P, Nigro M, *et al.*: **Giant axonal neuropathy: clinical, electrophysiologic, and neuropathologic features in two siblings.** *J Child Neurol.* 1990; **5**(3): 229–234. [PubMed Abstract](#) | [Publisher Full Text](#)
65. Thomas C, Love S, Powell HC, *et al.*: **Giant axonal neuropathy: correlation of clinical findings with postmortem neuropathology.** *Ann Neurol.* 1987; **22**(1): 79–84. [PubMed Abstract](#) | [Publisher Full Text](#)
66. van der Knaap MS, Valk J: **Magnetic resonance of myelination and myelin disorders.** Berlin: Springer Vol. Third, 2005. [Publisher Full Text](#)
67. McDonald CM: **Clinical approach to the diagnostic evaluation of hereditary and acquired neuromuscular diseases.** *Phys Med Rehabil Clin N Am.* 2012; **23**(3): 495–563. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
68. Landsverk ML, Ruzzo EK, Mefford HC, *et al.*: **Duplication within the SEPT9 gene associated with a founder effect in North American families with hereditary neuralgic amyotrophy.** *Hum Mol Genet.* 2009; **18**(7): 1200–1208. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
69. Jordanova A, Thomas FP, Guergueltcheva V, *et al.*: **Dominant intermediate Charcot-Marie-Tooth type C maps to chromosome 1p34-p35.** *Am J Hum Genet.* 2003; **73**(6): 1423–1430. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Current Referee Status:

Referee Responses for Version 2



Jean-Michel Vallat

Centre de référence neuropathies périphériques rares, CHU Limoges, Limoges, France

Approved: 30 April 2014

Referee Report: 30 April 2014

doi:[10.5256/f1000research.4270.r4588](https://doi.org/10.5256/f1000research.4270.r4588)

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.



Flávia L. Osório

Department of Neurosciences and Behavior, University of São Paulo, São Paulo, Brazil

Approved: 23 April 2014

Referee Report: 23 April 2014

doi:[10.5256/f1000research.4270.r4538](https://doi.org/10.5256/f1000research.4270.r4538)

I approve the indexation of this revised article.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

Referee Responses for Version 1



Jean-Michel Vallat

Centre de référence neuropathies périphériques rares, CHU Limoges, Limoges, France

Approved: 14 April 2014

Referee Report: 14 April 2014

doi:[10.5256/f1000research.4019.r4493](https://doi.org/10.5256/f1000research.4019.r4493)

This is a quite interesting paper which stresses that genetic treatments for neuropathies should be further investigated by neuroscientists, MD's who have such patients in their charge, and pharmaceutical companies. On the other hand, we all agree that discoveries in molecular biology should induce increasingly personalized medicine.

Nevertheless, some points have to be explained or clarified:

- The general classification of ‘inherited neuropathies’ includes a great number of disorders. The title of this paper has to be explained in the introduction: it is clearly understandable why the authors have chosen CMT, as it is the most frequent genetic neuropathy. Now it should be discussed why they also included GAN; maybe because it is one of the rarest hereditary neuropathies? But many patients with other inherited neuropathies - such as: porphyria, Chédiak-Higashi syndrome, storage disorders with neuropathy etc. - could argue that they have been forgotten. It could be useful to present a simplified classification of inherited neuropathies in the introduction (perhaps in the form of a table?).
- The frequency of GAN is discussed and I agree is clearly underestimated: see a recent paper in *Neurology*¹ (which could be mentioned). It states: “*Giant axonal neuropathy is a rare (worldwide 50 families reported) autosomal recessive disorder*”. Actually, in our neuropathology laboratory we have about 13 GAN nerve biopsies, all from Algeria.
- I am not sure that GAN is “*the most severe form of IN*” - even though, of course, it is very severe - leukodystrophies and storage disorders induce very severe encephalopathy.
- It should be explained that many symptoms of GAN are mostly explained by CNS involvement (impaired cognition, cerebellar dysfunction) - which is not at all the case in most CMT - so an efficient drug for this disease should also be able to cross the blood brain barrier.
- I disagree with this sentence: “*CSF has long been exploited as a relatively non-invasive way to biopsy the CNS*”. It is true that CSF data can be very important in CNS disorders, but in no case can it be treated as a biopsy!
- Actually the preliminary results of the Pharnext phase II trial have been published.²
- I agree that, at the present time, QoL should be the primary objective in therapeutic trials in CMT. I am not sure that Neuro-Qol would be a good tool as it takes into account pain and sensory signs which are not usually the main symptoms of CMT. Anyway, it will take a long time to validate such scales (PROMIS also) in CMT.
- It is correct that EIM may be a very interesting tool to follow CMT progression; so, it has to be validated - which will take some time. The possibility of utilizing this test in the coming CMT therapeutic trials could be discussed. Rutkove *et al.*³ have recently shown that EIM data correlates with standard measures of ALS severity.

References

1. Israni A, Chakrabarty B, Gulati S, Mohamed A, Kumar A, Sable MN, Suri V: Giant axonal neuropathy: A clinicoradiopathologic diagnosis. *Neurology*. 2014; **82** (9): 816-817 [PubMed Abstract](#) | [Free Full Text](#) | [Publisher Full Text](#)
2. Attarian S, Dubourg O, Funalot B, Gonnaud P-M, Lacour A, Magy L, Micallef J, Péréon Y, Pouget J, Stojkovic T, Vallat J-M, Nave K-A, Bertrand V, Cholet N, Graudens E, Guerassimenko O, Milet A, Mandel J, Scart Gres C, Vial E, Guedj M, Nabirotkin S, Chumakov I, Cohen D: A phase II randomized, placebo-controlled multicenter clinical trial of three doses of PXT3003 in 80 adult patients with CMT1A treated for 1 year. *J Peripher Nerv Syst*. 2013; **8** (3): 197-198 [PubMed Abstract](#) | [Publisher Full Text](#)
3. Rutkove SB, Caress JB, Cartwright MS, Burns TM, Warder J, David WS, Goyal N, Maragakis NJ,

Benatar M, Sharma KR, Narayanaswami P, Raynor EM, Watson ML, Shefner JM: Electrical impedance myography correlates with standard measures of ALS severity. *Muscle Nerve*. 2014; **49** (3): 441-443
[PubMed Abstract](#) | [Publisher Full Text](#)

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

1 Comment

Author Response

Sean Ekins, Collaborations in Chemistry, USA

Posted: 16 Apr 2014

We thank the reviewer for their positive feedback. We have made most of the suggested changes. Each of the following bulletpoints refers to their comments.

- We do not think the title needs any explanation as it just focuses on CMT and GAN as examples of inherited neuropathies (INs). These two diseases also represent the extremes. Yes we acknowledge there are other INs but two of the authors' families suffer from these diseases hence our interest in them specifically.
- We have added the reference for GAN as suggested.
- We have edited out "most" to leave "severe".
- We have added "As GAN progresses (through effects on the CNS),..." in the text. We agree that ability to cross the BBB will be important for a GAN small molecule but we do not describe small molecule work on GAN, only gene therapy which is close to human trials. Therefore we found no appropriate place to add this in the text. We hope these comments will suffice.
- We replaced "biopsy" with "sample".
- We have added this Pharnext reference to a meeting abstract, but the full paper is still forthcoming.
- Thank you for agreeing that QoL is important for CMT trials. As neither Neuro QoL or PROMIS have been used to our knowledge with CMT, we feel they should at least be tried.
- Thank you for agreeing that EIM may be a very interesting tool to follow CMT progression. We have added a further reference on ALS and EIM as you suggested.

Competing Interests: No competing interests were disclosed.



Flávia L. Osório

Department of Neurosciences and Behavior, University of São Paulo, São Paulo, Brazil

Approved: 07 April 2014

Referee Report: 07 April 2014

doi:[10.5256/f1000research.4019.r4344](https://doi.org/10.5256/f1000research.4019.r4344)

In the present article the authors review the literature on inherited neuropathies - especially Charcot-Marie-Tooth and Giant Axonal Neuropathy - based on recent clinical evidence. The authors then offer several suggestions to overcome gaps in this area of knowledge, with criticism and argumentation.

I approve this article for indexation. My only suggestion is that the authors remove figures 1 and 2 from the main body of the article and include them in the Supplementary materials instead.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

1 Comment

Author Response

Sean Ekins, Collaborations in Chemistry, USA

Posted: 16 Apr 2014

We thank the reviewer for their positive feedback. We have made the suggested changes and removed the two figures to the supplemental materials.

Competing Interests: No competing interests were disclosed.