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Rapid diagnostic tests for *Plasmodium vivax* malaria in endemic countries (Review)

Agarwal R, Choi L, Johnson S, Takwoingi Y

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[Diagnostic Test Accuracy Review]

Rapid diagnostic tests for *Plasmodium vivax* malaria in endemic countries

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ABSTRACT

Background

Plasmodium vivax (*P vivax*) is a focus of malaria elimination. It is important because *P vivax* and *Plasmodium falciparum* infection are co-endemic in some areas. There are asymptomatic carriers of *P vivax,* and the treatment for *P vivax* and *Plasmodium ovale* malaria differs from that used in other types of malaria. Rapid diagnostic tests (RDTs) will help distinguish *P vivax* from other malaria species to help treatment and elimination. There are RDTs available that detect *P vivax* parasitaemia through the detection of *P vivax*-specific lactate dehydrogenase (LDH) antigens.

Objectives

To assess the diagnostic accuracy of RDTs for detecting *P vivax* malaria infection in people living in malaria-endemic areas who present to ambulatory healthcare facilities with symptoms suggestive of malaria; and to identify which types and brands of commercial tests best detect *P vivax* malaria.

Search methods

We undertook a comprehensive search of the following databases up to 30 July 2019: Cochrane Infectious Diseases Group Specialized Register; Central Register of Controlled Trials (CENTRAL), published in the Cochrane Library; MEDLINE (PubMed); Embase (OVID); Science Citation Index Expanded (SCI-EXPANDED) and Conference Proceedings Citation Index-Science (CPCI-S), both in the Web of Science.

Selection criteria

Studies comparing RDTs with a reference standard (microscopy or polymerase chain reaction (PCR)) in blood samples from patients attending ambulatory health facilities with symptoms suggestive of malaria in *P vivax*-endemic areas.

Data collection and analysis

For each included study, two review authors independently extracted data using a pre-piloted data extraction form. The methodological quality of the studies were assessed using a tailored Quality Assessment of Diagnostic Accuracy Studies-2 (QUADAS-2) tool. We grouped studies according to commercial brand of the RDT and performed meta-analysis when appropriate. The results given by the index tests were based on the antibody affinity (referred to as the strength of the bond between an antibody and an antigen) and avidity (referred to as the strength of the overall bond between a multivalent antibody and multiple antigens). All analyses were stratified by the type of

reference standard. The bivariate model was used to estimate the pooled sensitivity and specificity with 95% confidence intervals (CIs), this model was simplified when studies were few. We assessed the certainty of the evidence using the GRADE approach.

Main results

We included 10 studies that assessed the accuracy of six different RDT brands (CareStart Malaria Pf/Pv Combo test, Falcivax Device Rapid test, Immuno-Rapid Malaria Pf/Pv test, SD Bioline Malaria Ag Pf/Pv test, OnSite Pf/Pv test and Test Malaria Pf/Pv rapid test) for detecting *P vivax* malaria. One study directly compared the accuracy of two RDT brands. Of the 10 studies, six used microscopy, one used PCR, two used both microscopy and PCR separately and one used microscopy corrected by PCR as the reference standard. Four of the studies were conducted in Ethiopia, two in India, and one each in Bangladesh, Brazil, Colombia and Sudan.

The studies often did not report how patients were selected. In the patient selection domain, we judged the risk of bias as unclear for nine studies. We judged all studies to be of unclear applicability concern. In the index test domain, we judged most studies to be at low risk of bias, but we judged nine studies to be of unclear applicability concern. There was poor reporting on lot testing, how the RDTs were stored, and background parasitaemia density (a key variable determining diagnostic accuracy of RDTs). Only half of the included studies were judged to be at low risk of bias in the reference standard domain, Studies often did not report whether the results of the reference standard could classify the target condition or whether investigators knew the results of the RDT when interpreting the results of the reference standard. All 10 studies were judged to be at low risk of bias in the flow and timing domain.

Only two brands were evaluated by more than one study. Four studies evaluated the CareStart Malaria Pf/Pv Combo test against microscopy and two studies evaluated the Falcivax Device Rapid test against microscopy. The pooled sensitivity and specificity were 99% (95% CI 94% to 100%; 251 patients, moderate-certainty evidence) and 99% (95% CI 99% to 100%; 2147 patients, moderate-certainty evidence) for CareStart Malaria Pf/Pv Combo test.

For a prevalence of 20%, about 206 people will have a positive CareStart Malaria Pf/Pv Combo test result and the remaining 794 people will have a negative result. Of the 206 people with positive results, eight will be incorrect (false positives), and of the 794 people with a negative result, two would be incorrect (false negative).

For the Falcivax Device Rapid test, the pooled sensitivity was 77% (95% CI: 53% to 91%, 89 patients, low-certainty evidence) and the pooled specificity was 99% (95% CI: 98% to 100%, 621 patients, moderate-certainty evidence), respectively. For a prevalence of 20%, about 162 people will have a positive Falcivax Device Rapid test result and the remaining 838 people will have a negative result. Of the 162 people with positive results, eight will be incorrect (false positives), and of the 838 people with a negative result, 46 would be incorrect (false negative).

Authors' conclusions

The CareStart Malaria Pf/Pv Combo test was found to be highly sensitive and specific in comparison to microscopy for detecting *P vivax* in ambulatory healthcare in endemic settings, with moderate-certainty evidence. The number of studies included in this review was limited to 10 studies and we were able to estimate the accuracy of 2 out of 6 RDT brands included, the CareStart Malaria Pf/Pv Combo test and the Falcivax Device Rapid test. Thus, the differences in sensitivity and specificity between all the RDT brands could not be assessed. More high-quality studies in endemic field settings are needed to assess and compare the accuracy of RDTs designed to detect *P vivax*.

PLAIN LANGUAGE SUMMARY

Rapid tests for diagnosing malaria caused by *Plasmodium vivax* in people living in areas where malaria is very common

What is the aim of the review?

Malaria infection is caused mainly by two species of malaria parasite: *Plasmodium falciparum* and *Plasmodium vivax*. The aim of this review was to evaluate rapid diagnostic tests (RDTs) to diagnose *P vivax* infection.

Why are rapid tests for *P vivax* malaria important?

For clinical management, knowing which parasite species is causing the malaria is important as the drug treatments differ. For *P vivax* infection, an additional drug is required to eliminate the infection from the liver. For public health control of malaria, we know that *P falciparum* is declining over the previous 15 years, and infections from *P vivax* have therefore increased in importance.

What was studied in this review?

RDTs provide results quickly and are often as a dipstick. We studied RDTs that specifically test for *P vivax* malaria. RDTs are simple to use, point-of-care tests. They are suitable for use in rural settings by primary healthcare workers, using drop of blood on the dipstick that causes colour change and a distinct line that indicates a positive test result. Healthcare workers in rural areas can perform RDTs for *P vivax* without needing a laboratory or special equipment. We wanted to find out which brands of RDTs were the most accurate for diagnosing *P vivax* malaria. We compared the new tests against the standard form of diagnosis with microscopy, and also more recent methods polymerase chain reaction (PCR): a molecular method to identify *P vivax* DNA in blood samples.

What are the main results of the review?

We included 10 studies that looked at the accuracy of six diagnostic test brands for detecting *P vivax* malaria in people with suspected malaria symptoms. The studies were conducted in Ethiopia (four studies), India (two studies) and Bangladesh, Brazil, Colombia, and Sudan (one study each).

Compared with microscopy, the Care Start Malaria Pf/Pv Combo test performed well with 99% sensitivity and specificity (four studies). This means that:

• for every 100 people tested who have *P vivax* malaria, one person will have a negative test result, and might not receive the right treatment soon enough;

• for every 100 people tested who do not have *P vivax* malaria, one will have a positive result, and might receive unnecessary treatment.

Compared with microscopy, the Falcivax Device Rapid test had a sensitivity of 77% and a specificity of 99% (two studies). This means that:

• For every 100 people tested who have *P vivax* malaria, 23 people will have a negative test result; and,

• for every 100 people tested who do not have *P vivax* malaria, one person will have a positive result.

We are moderately confident (certain) in the accuracy results for the Care Start Malaria Pf/Pv Combo test. The results are from a small number of studies (four), so our findings may change when results from further studies become available.

We are less confident in the accuracy results for the Falcivax Device Rapid test, because these came from only two studies. Our findings for this test will probably change when results from further studies become available.

Our results are based on a small number of studies, so we could not reliably assess all six brands of antibody test or compare their accuracy. Most studies included in this review had limitations: it was not clear how people were selected for testing, or how the study results were assessed and checked, which could have affected the results. Some rapid antibody tests were investigated by only one study. Some studies did not report clearly how common *P* malaria was in the area where the study was done.

How up-to-date is this review?

The review authors searched for studies published up to 30 July 2019.

SUMMARY OF FINDINGS

Summary of findings 1. Summary of findings table for RDTs for diagnosing P vivax malaria

Population: people presenting with symptoms of uncomplicated malaria

Prior testing: none

Setting: ambulatory healthcare settings in *P vivax* endemic areas

Index tests: immunochromatography-based rapid diagnostic tests (RDTs) for *P vivax* malaria that meet the WHO malaria RDT performance criteria (WHO 2017b)

Reference standards: conventional microscopy, polymerase chain reaction (PCR)

Target condition: *P vivax* malaria

Importance: accurate and fast diagnosis of *P vivax* from other malaria species allows appropriate treatment to be provided quickly

Study design: retrospective or prospective cohort or cross-sectional

Findings: 10 studies of six different RDT brands (CareStart Malaria Pf/Pv Combo test, Falcivax Device Rapid test, Immuno-Rapid Malaria Pf/Pv test, SD Bioline Malaria Ag Pf/Pv test, OnSite Pf/Pv test and Test Malaria Pf/Pv rapid test) for *P vivax* malaria were included. Only two brands (CareStart Malaria Pf/Pv Combo test and Falcivax Device Rapid test) were evaluated against the same reference standard by more than one study.

Limitations: a small number of studies were included in the analyses and meta-analyses were only possible for two RDT brands. Studies often did not report how patients were selected, the blinding of the RDT results to the reference standard and the storage conditions and lot testing of RDTs.

Outcome	№ of studies	№ of patients	Numbers in a cohort of 1000 patients tested (95% CI)[a]			Certainty of the evidence (GRADE)[b]
			Prevalence of 0.5%	Prevalence of 5%	Prevalence of 20%	
Test (reference standard): CareStart Malaria Pf/Pv Combo test (microscopy), pooled sensitivity (95% CI) = 99% (94% to 100%) and pooled specificity (95% CI) = 99% (99% to 100%), positive likelihood ratio (95% CI) = 141.09 (68.18 to 292.00) and negative likelihood ratio (95% CI) = 0.01 (0.00 to 0.06)						
True positives (patients with *P vivax* malaria)	4	251	5 (5 to 10)	50 (47 to 50)	198 (188 to 200)	⊕⊕⊕⊖ **MODERATE** [1]
False negatives (patients incorrectly classified as not having *P vivax* malaria)			0 (0 to 0)	0 (0 to 3)	2 (0 to 12)	

True negatives (patients without *P vivax* malaria)		2147	985 (980 to 995)	941 (941 to 950)	792 (792 to 800)	⊕⊕⊕⊖ **MODERATE** [1]
False positives (patients incorrectly classified as having *P vivax* malaria)			10 (0 to 10)	9 (0 to 9)	8 (0 to 8)	
Test (reference standard): Falcivax Device Rapid test (microscopy), pooled sensitivity (95% CI) = 77% (53% to 91%) and pooled specificity (95% CI) = 99% (98% to 100%), positive likelihood ratio (95% CI) = 120.31 (43.10 to 335.87) and negative likelihood ratio (95% CI) = 0.23 (0.10 to 0.53)						
True positives (patients with *P vivax* malaria)	2	89	4 (3 to 5)	39 (27 to 46)	154 (106 to 182)	⊕⊕⊖⊖ **LOW** [1,2]
False negatives (patients incorrectly classified as not having *P vivax* malaria)			1 (0 to 2)	11 (4 to 23)	46 (18 to 94)	
True negatives (patients without *P vivax* malaria)		621	985 (975 to 995)	941 (931 to 950)	792 (784 to 800)	⊕⊕⊕⊖ **MODERATE** [1]
False positives (patients incorrectly classified as having *P vivax* malaria)			10 (0 to 20)	9 (0 to 19)	8 (0 to 16)	

[a]Median values were chosen from ranges of prevalence considered to be moderate, low, and very low transmission settings for *P vivax* (WHO 2017c).
[b]Methods are lacking to assess the determinants and extent of publication bias for diagnostic studies. However, in this table, we considered publication bias 'undetected'.
[1]Downgraded for risk of bias by one.
[2]Downgraded for imprecision by two due to wide confidence intervals.
GRADE certainty of the evidence.
High: we are very confident that the true effect lies close to that of the estimate of the effect.
Moderate: we are moderately confident in the effect estimate: the true effect is likely to be close to the estimate of the effect, but there is a possibility that it is substantially different.
Low: our confidence in the effect estimate is limited: the true effect may be substantially different from the estimate of the effect.
Very low: we have very little confidence in the effect estimate: the true effect is likely to be substantially different from the estimate of effect.

BACKGROUND

Target condition being diagnosed

Malaria is a life-threatening disease caused by *Plasmodium* species (*Plasmodium* spp.), transmitted by the bite of a female *Anopheles* mosquito. Currently, there are five established *Plasmodium* spp. that cause malaria in humans. The two most common are *Plasmodium falciparum* (*P falciparum*) and *Plasmodium vivax* (*P vivax*). *P vivax* malaria is a relapsing form, which is rarely fatal but can cause serious anaemia in children (White 2018). There has been an increased focus on *P vivax*, as malaria-endemic settings that also have *P falciparum* have made progress in *P falciparum* control. In the World Health Organization (WHO) regions of the Americas, South-East Asia and Eastern Mediterranean, *P vivax* is the predominant *Plasmodium* spp., and causes 64%, greater than 30%, and greater than 40% of all malaria cases, respectively, in these regions (WHO 2017a). People with malaria caused by *P vivax* can have relapses due to the dormant liver stage hypnozoites. People can carry hypnozoites ranging from a few weeks to more than 12 months before reporting symptoms again (Campo 2015). Primaquine is recommended additionally to standard malaria treatment for *P vivax* and *Plasmodium ovale* (*P ovale*) to clear these liver stage parasites. Due to this, it is important to have diagnostic tests that are highly sensitive and that can specifically detect *P vivax* from other *Plasmodium* spp.

Index test(s)

Rapid diagnostic tests (RDTs) (WHO 2003), detect parasite-specific antigens in a drop of fresh blood through lateral flow immunochromatography (WHO 2006). Generally, RDTs do not require a laboratory, any special equipment, or specialized training. They are easy to use and can give results as a simple positive or negative result within 15 to 20 minutes based on the antibody affinity (referred to as the strength of the bond between an antibody and an antigen) and avidity (referred to as the strength of the overall bond between a multivalent antibody and multiple antigens) (Talman 2007; WHO 2006). Therefore, RDTs are, in general, suitable for remote areas with limited facilities and lack of laboratory expertise. They typically have a shelf life of 24 months and need to be kept dry and away from temperature extremes (greater than 40°C). They may fail to detect malaria where there are low levels of *Plasmodium* parasites (and antigens) in the blood and false positives are possible due to cross reactions with other disease conditions, presence of certain immunological factors, and gametocytaemia (Gatton 2018; Kakkilaya 2003).

There is strong evidence that storage conditions of the RDT affect their performance (Moonasar 2007). The parasite density of the blood sample can also affect the performance of the RDT. The WHO malaria RDT product-testing programme report investigated the effect of parasite density by testing individual products under laboratory conditions using standardized blood samples at low and high parasite densities (200 and 2000 parasites/µL), and reported the 'panel detection score' (WHO 2012). An existing Cochrane Review on non-falciparum RDTs found that parasite density and storage conditions are often poorly reported in field studies (Abba 2014). Moreover, due to the lag period between when the RDT was evaluated by the WHO malaria RDT product testing programme to when the RDT is actually used in the field, manufacturers may have modified the RDT during this period.

Different types of RDT use different types of antibody or combination of antibodies to detect *Plasmodium* antigens. Some antibodies aim to detect a particular species while others are pan-malarial, aiming to detect all types of *Plasmodium* spp. Currently, all commercial RDTs specific for *P vivax* use *P vivax*-specific lactate dehydrogenase (LDH) antigens (WHO 2017b).

Clinical pathway

People of any age with malaria typically present to medical care with non-specific symptoms of fever, headache, chills, or rigors. The RDTs are most commonly used at the point of presentation with these symptoms, most often in settings where quality microscopy is not available. Parasitological diagnosis is recommended prior to commencing on any treatment (WHO 2015a).

Prior test(s)

It is unlikely that patients would have had previous testing for their current infection prior to presentation to healthcare centres with symptoms of malaria. One key benefit of RDTs is the ease of use at point of care. For the purpose of this review, we did not address the sensitivity or specificity of *P vivax*-specific RDTs for confirming efficacy of treatment as this is not recommended practice.

Role of index test(s)

Malaria is a common cause of fever in endemic regions. Given the non-specific symptoms patients with malaria often present with, a parasitological test is recommended to make a formal diagnosis (WHO 2015b). Often people of any age or gender presenting to a healthcare clinic with a history of fever in a malaria-endemic region will undergo a malaria test as part of a routine initial work-up. As such, the population receiving the index test would be identified solely on the basis of the clinical history and physical examination. RDTs have a role in malaria diagnosis where there is no access to good quality microscopy services and in outbreak investigation or surveys of parasite prevalence. The pre-test probability of clinical malaria is an important determinant of the RDT performance. In the absence of strong clinical suspicion of malaria, it may not be reliable to use an RDT, because the test results from this device could potentially be misleading or inaccurate. Reliable diagnosis of *P vivax* malaria with RDTs would not only benefit the individual by allowing treatment of the blood stage and latent hypnozoite stage, but also would have benefits at a population level by potentially reducing low-level ongoing transmission due to relapsing disease. Widespread use of accurate RDTs can facilitate greater diagnosis and treatment rates of *P vivax* malaria in areas where there is inadequate access to high-quality microscopy.

True positive results would allow effective treatment of active disease and facilitate prevention of relapse using drugs that target the liver stage hypnozoites such as primaquine or tafenoquine, thus effectively treating individuals and reducing the risk of onward transmission. True negative results facilitate accurate diagnosis by narrowing differential diagnoses of people presenting to care with fever and non-specific symptoms. False positives would potentially lead to over treatment of individuals with primaquine, tafenoquine and either chloroquine or artemisinin combination therapies and would mean that patients are not treated for the actual cause of their symptoms. False negatives would lead to potential relapsing disease and potentially ongoing transmission at the population level.

Alternative test(s)

Microscopic examination of Giemsa-stained thick and thin blood films remains the conventional laboratory method. Microscopic examination has good sensitivity and specificity, and it allows species and stage differentiations and quantification of parasites, all of which are important in assessing disease severity, monitoring response to treatment, and prescribing appropriate therapy. Intensive examination is more likely to reveal parasitaemia so the test is carried out with a fixed number of fields examined. Infections may be missed if slides are not examined carefully (Wongsrichanalai 2007). Very low parasitaemia may be missed even by good quality microscopy; the limit of detection of thick smear microscopy has been estimated at approximately four to 20 asexual parasites per µL, although a threshold of 50 to 100 asexual parasites per µL is more realistic under field conditions (Wongsrichanalai 2007). False positive results are also possible; if blood slides are not prepared carefully, artefacts may be formed, which can be mistaken for *Plasmodium* parasites (Wongsrichanalai 2007).

The polymerase chain reaction (PCR), a molecular method based on DNA amplification, is the most analytically sensitive method of detecting parasites in the blood. Compared to microscopy, PCR is less prone to observer error and more sensitive at low levels of parasitaemia (Han 2017; Snounou 1993). For PCR, the limit of detection may be as low as 0.004 asexual parasites per µL (Hänscheid 2002). This increased ability to detect low level parasitaemia is important as submicroscopic parasitaemiae may have clinical and public health significance and the prevalence of asymptomatic submicroscopic infection is high in some areas (Chen 2016). PCR is currently not widely available due to logistical constraints and the need for specially-trained technicians and a well-equipped laboratory. It is usually used only for research purposes.

Rationale

P vivax is becoming increasingly important, especially in regions targeting malaria elimination. In areas of co-endemicity, *P vivax* malaria is increasing disproportionally compared to *P falciparum* malaria. Moreover, treatment for *P vivax* and *P ovale* malaria differs from treatments for other types of malaria. Therefore, it is important that the RDT correctly distinguish *P vivax* from other species. Geographically, *P vivax* has a much wider infection range compared to other *Plasmodium* spp. This may increase over time due to climate change (Culleton 2012). Historically, autochthonous transmission of *P vivax* also occurred in temperate climates, such as that of England (Dobson 1994). Autochthonous transmission is referred to as the spread of a disease from one individual and received by another individual from the same place. An existing Cochrane Review assessing RDTs for diagnosing uncomplicated non-falciparum malaria was conducted in 2014 (Abba 2014). A subset of this review included RDTs that diagnosed *P vivax*. This review only assesses the diagnostic accuracy of RDTs that specifically detect *P vivax* with *P vivax*-specific LDH) antigens.

OBJECTIVES

To assess the diagnostic accuracy of RDTs for detecting *P vivax* malaria parasitaemia in people living in malaria-endemic areas who present to ambulatory healthcare facilities with symptoms suggestive of malaria, and to identify which types and brands of commercial tests best detect *P vivax* malaria.

METHODS

Criteria for considering studies for this review

Types of studies

We included retrospective or prospective cohort or cross-sectional studies that assessed the accuracy of an RDT, or compared the accuracy of two or more RDTs, in the same study population (i.e. comparative accuracy studies). We excluded case-control studies because they are known to overestimate test accuracy (Whiting 2011). Eligible studies included a consecutive series of patients, or a randomly selected series of patients. If the study did not explicitly state that the sampling was consecutive or random, the study was considered unclear but was still included. We excluded studies if they did not present sufficient data to allow us to extract or deduce the number of true positives, false positives, false negatives, and true negatives (i.e. 2 x 2 table data). We also excluded studies published in predatory journals, which is referred to as journals that accept articles for publication for a fee without providing peer-review or quality checks for plagiarism or ethical approval.

Participants

Studies recruiting people living in *P vivax*-endemic areas attending ambulatory healthcare settings with symptoms of uncomplicated malaria were eligible.

We excluded studies if participants:

- had travelled from non-malarious region to malarious regions, e.g. travellers or displaced populations;
- had been previously treated for their current malaria infection or the test was performed to assess whether treatment was successful, or both;
- had symptoms of severe malaria as defined by the WHO clinical definition (WHO 2014);
- did not have symptoms of malaria as defined by history of fever, headache, or chills/rigors; or
- were recruited through active case finding (for example, door to door surveys).

Index tests

Studies evaluating any immunochromatography-based RDT specifically designed to detect *P vivax* malaria. We only included RDTs that met the WHO malaria RDT performance criteria (WHO 2017b).

Target conditions

Studies aimed at detecting *P vivax* malaria.

Reference standards

Studies that diagnosed *P vivax* malaria using at least one of the following two reference standards:

- Conventional microscopy of thick blood smears and thin blood smears. Presence of asexual parasites of any density is regarded as a positive smear. Once the diagnosis is established – usually by detecting parasites in the thick smear – the laboratory technician can examine the thin smear to determine the malaria species and the parasitaemia, or the percentage of the patient's red blood cells that are infected with malaria parasites. The thin

and thick smears are able to provide all three of these vital pieces of information. Ideally, blood smears would be examined independently and in duplicate with more than 100 high-power fields;

- PCR, including quantitative PCR (qPCR), nested PCR (nPCR), and real-time PCR (rPCR). We also included studies that used loop-mediated isothermal amplification (LAMP). Most PCR-based assays for *P vivax* are only available as laboratory-developed tests, which means they are rarely used clinically outside of research projects where *P vivax* malaria is endemic. They are especially useful for diagnosing asymptomatic people as the assays have high sensitivity. Molecular diagnostics theoretically have a lower limit of detection than both RDTs and microscopy depending on the training of microscopists and quality of samples analysed. Significant variation exists between molecular diagnostics developed including type of input material (DNA, RNA, or whole blood), target gene, (number of) species detected, primer/probe composition and concentration, amplification technique (PCR or isothermal), read-out (gel-electrophoresis, fluorescence detection, lateral flow), and whether it is qualitative or quantitative. However, no important differences have been found in the accuracy of these tests (Roth 2016).

For studies that used both reference standards, we extracted 2 x 2 data for each reference standard and stratified the analyses by reference standard.

Search methods for identification of studies

We attempted to identify all relevant studies regardless of language or publication status (published, unpublished, in press, and in progress).

Electronic searches

We searched the following databases up to 30 July 2019 using the search terms and strategy described in Appendix 1: Cochrane Infectious Diseases Group Specialized Register; Central Register of Controlled Trials (CENTRAL), published in the Cochrane Library (issue 7, 2019); MEDLINE (PubMed, from 1966); Embase (OVID, from 1947); Science Citation Index Expanded (SCI-EXPANDED) and Conference Proceedings Citation Index- Science (CPCI-S), both in the Web of Science, from 1900; LILACS (BIREME).

We also searched the WHO International Clinical Trials Registry Platform (WHO ICTRP; www.who.int/ictrp/en/) and ClinicalTrials.gov (clinicaltrials.gov/ct2/home) for trials in progress, using "vivax malaria", "Plasmodium vivax", and "rapid diagnostic test*" or RDT* as search terms.

Searching other resources

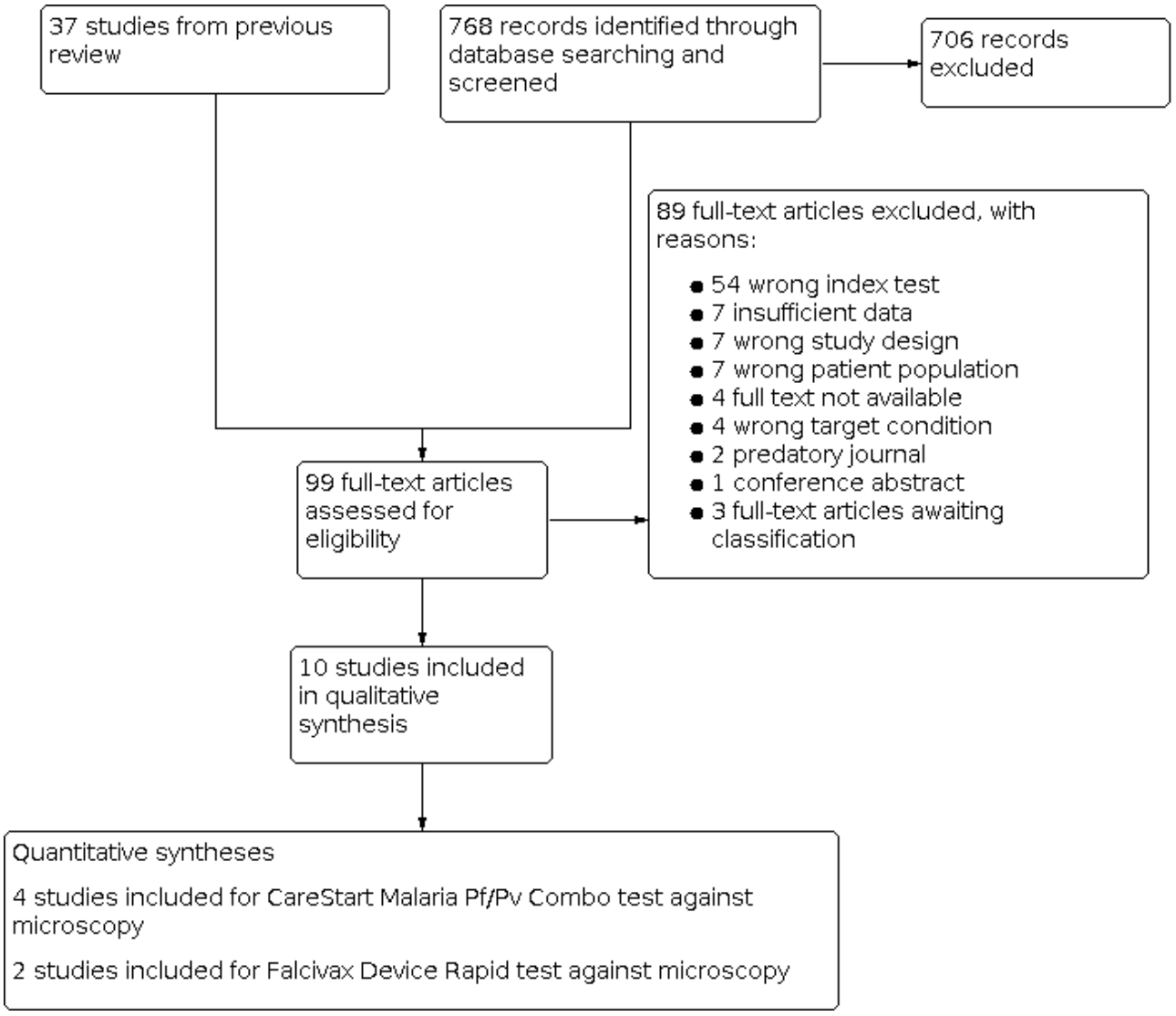
We checked the reference lists of studies identified by the above methods.

Data collection and analysis

Selection of studies

Three review authors (RA, LC and SJ) independently assessed the study eligibility by examining the title and abstract of each article identified by the literature search and excluded obviously irrelevant studies. If a review author considered the abstract to be potentially eligible, we obtained the full-text article. Three review authors independently assessed each full-text article against the predefined inclusion and exclusion criteria, as stated in the 'Criteria for considering studies for this review' section, and resolved any disagreements by discussion. All articles that were excluded after full-text assessment are listed with reasons for exclusion in the 'Characteristics of excluded studies' table. We illustrated the study selection process with a PRISMA flow diagram (Figure 1).

Figure 1. Study flow diagram.



Data extraction and management

Three review authors (RA, LC and SJ) independently extracted data using a pre designed data extraction form.

We extracted the following data.

- Authors, publication year, and journal.
- Study design.
- Study start date.
- Characteristics study participants (age, gender, co morbidities, and pregnancy).
- Study inclusion/exclusion criteria.
- Study setting.
- Malaria species in study setting.
- Malaria prevalence and endemicity in study setting.
- Reference standard.
- Index test (brand name, target antigen, and batch numbers).
- Additional tests (and their results).
- RDT and reference standard setting.
- Lot testing of RDT used.
- Transport and storage conditions of RDTs.
- Training level of person performing index test.
- Training level of person performing reference standard (and if available the WHO certified training level of the microscopist).
- Number of high power fields observed in microscopy.
- Parasite density of microscopy positive cases or PCR.
- Observers or repeats used.
- Number of indeterminate, missing or unavailable test results.
- Number of true positives, false positives, false negatives, and true negatives.
- Type of molecular amplification assay.
- Volume of blood samples.
- Limit of detection for PCR.

We resolved any discrepancies in data extraction by discussion. We contacted the authors of primary studies when we could not resolve any disagreements.

Assessment of methodological quality

We used the revised tool for the Quality Assessment of Diagnostic Accuracy Studies (QUADAS-2) to assess the risk of bias and applicability of included studies (Whiting 2011). We tailored the tool to the context of the review as shown in Appendix 2. Three review authors (RA, LC and SJ) independently assessed methodological quality using the tailored QUADAS-2 tool. We resolved any disagreements through consensus. We used both graphics and text to summarize the results.

Statistical analysis and data synthesis

We stratified all analyses by the type of reference standard used. We plotted estimates of sensitivity and specificity from the included studies in forest plots and in receiver operating characteristic (ROC) space using the software, Review Manager 5 (RevMan 5) (RevMan 2014). We planned to perform meta-analysis using the bivariate model to estimate summary sensitivities and specificities (summary points) (Chu 2006; Macaskill 2010; Takwoingi 2015b). However, due to sparse data or few studies, we simplified the models to univariate random effects logistic regression models to pool sensitivity and specificity separately (Takwoingi 2015a). We performed meta-analyses using the 'meqrlogit' command in Stata (STATA 2015). Due to the limited number of included studies we did not perform meta-analyses to compare the accuracy of different RDT brands as planned. However, we summarized individual study estimates from head-to-head comparisons of brands in a table.

Investigations of heterogeneity

We intended to investigate any heterogeneity from the pooled analyses with pre-specified factors, as stated in our secondary objective. Due to the limited number of studies, we were unable to investigate heterogeneity as planned.

Sensitivity analyses

We did not have sufficient data for sensitivity analyses.

Assessment of the certainty of the evidence

We assessed the certainty of the evidence for comparisons where there were sufficient studies enabling meta-analyses (i.e. quality of evidence or confidence in effect estimates) using the GRADE approach and GRADEpro Guideline Development Tool software (GRADE 2013; GRADEpro GDT 2015). In the context of a systematic review, the ratings of the certainty of the evidence reflect the extent of our confidence that the estimates of test accuracy are correct. As recommended, we rated the certainty of the evidence as either high (not downgraded), moderate (downgraded by one level), low (downgraded by two levels), or very low (downgraded by more than two levels) for four domains: risk of bias, indirectness, inconsistency, and imprecision. For sensitivity and specificity, the certainty of the evidence initially started as high when there were high-quality cross-sectional or cohort studies that enrolled participants with diagnostic uncertainty. If we found a reason for downgrading the certainty of the evidence, we classified the reason as either serious (downgraded by one level) or very serious (downgraded by two levels).

Three review authors (RA, LC and SJ) discussed judgments and reached a consensus. We applied GRADE in the following way.

- Risk of bias: we used QUADAS-2 to assess risk of bias.
- Indirectness: we considered indirectness from the perspective of test accuracy. We used QUADAS-2 to assess applicability concerns and looked for important differences between the populations studied (for example, in the transmission intensity as defined by the WHO World Malaria Report or WHO malaria country profiles for the corresponding year), the setting, and the review question.
- Inconsistency: GRADE recommends downgrading for unexplained inconsistency in sensitivity and specificity estimates.
- Imprecision: we considered the width of the confidence intervals (CIs), and asked ourselves, “would we make a different decision if the lower or upper limit of the 95% confidence interval (CI) represented the truth?” In addition, we calculated absolute numbers of true positives, false negatives, false positives, and true negatives, as well as ranges for these values based on the CIs of the pooled estimates of sensitivity and specificity for various prevalences of *P vivax* malaria; we also made judgements on imprecision using these calculations. We also calculated positive and negative likelihood ratios with their 95% CIs.

Assessment of reporting bias

We did not assess publication bias due to the uncertainty about the determinants of publication bias for diagnostic accuracy studies, and the inadequacy of tests for detecting funnel plot asymmetry (Deeks 2005).

RESULTS

Results of the search

We identified and screened 768 reports through the database searches conducted on 30 July 2019. We excluded 706 of these reports based on their title or abstract alone. We considered the remaining 62 articles for full-text screening, along with the 37 studies included in the non-falciparum malaria review by Abba 2014. Of the 109 articles, we excluded 99 for various reasons as reported in the Characteristics of excluded studies section, shown in Figure 1. We included 10 studies, of which five studies (Alam 2011; Chanie 2011; Mekonnen 2010; Singh 2010; Sharew 2009) were also included in the review by Abba 2014. The 10 studies assessed six different RDT brands (CareStart Malaria Pf/Pv Combo test, Falcivax Device Rapid test, Immuno-Rapid Malaria Pf/Pv test, SD Bioline Malaria Ag Pf/Pv test, OnSite Pf/Pv test and Test Malaria Pf/Pv rapid test). One study directly compared the accuracy of two RDT brands (Falcivax Device Rapid test and OnSite Pf/Pv test) (Alam 2011). The six RDT brands detect *P vivax* as part of a mixed infection with *P vivax*-specific LDH antigens. The tests have two test lines, an HRP-2 line to detect *P falciparum* and an pLDH line to detect *P vivax*. For our analysis we only considered the presence of the pLDH line.

Of the 10 included studies, six used microscopy (Chanie 2011; Costa 2019; Hailu 2014; Mekonnen 2010; Sharew 2009; Singh 2010), one used PCR (Mussa 2019), two used both microscopy and PCR separately (Alam 2011; Saha 2017), and one used microscopy corrected by PCR (Mendoza 2013) as the reference standard. Four of the studies were conducted in Ethiopia (Chanie 2011; Hailu 2014; Mekonnen 2010; Sharew 2009), two in India (Saha 2017; Singh

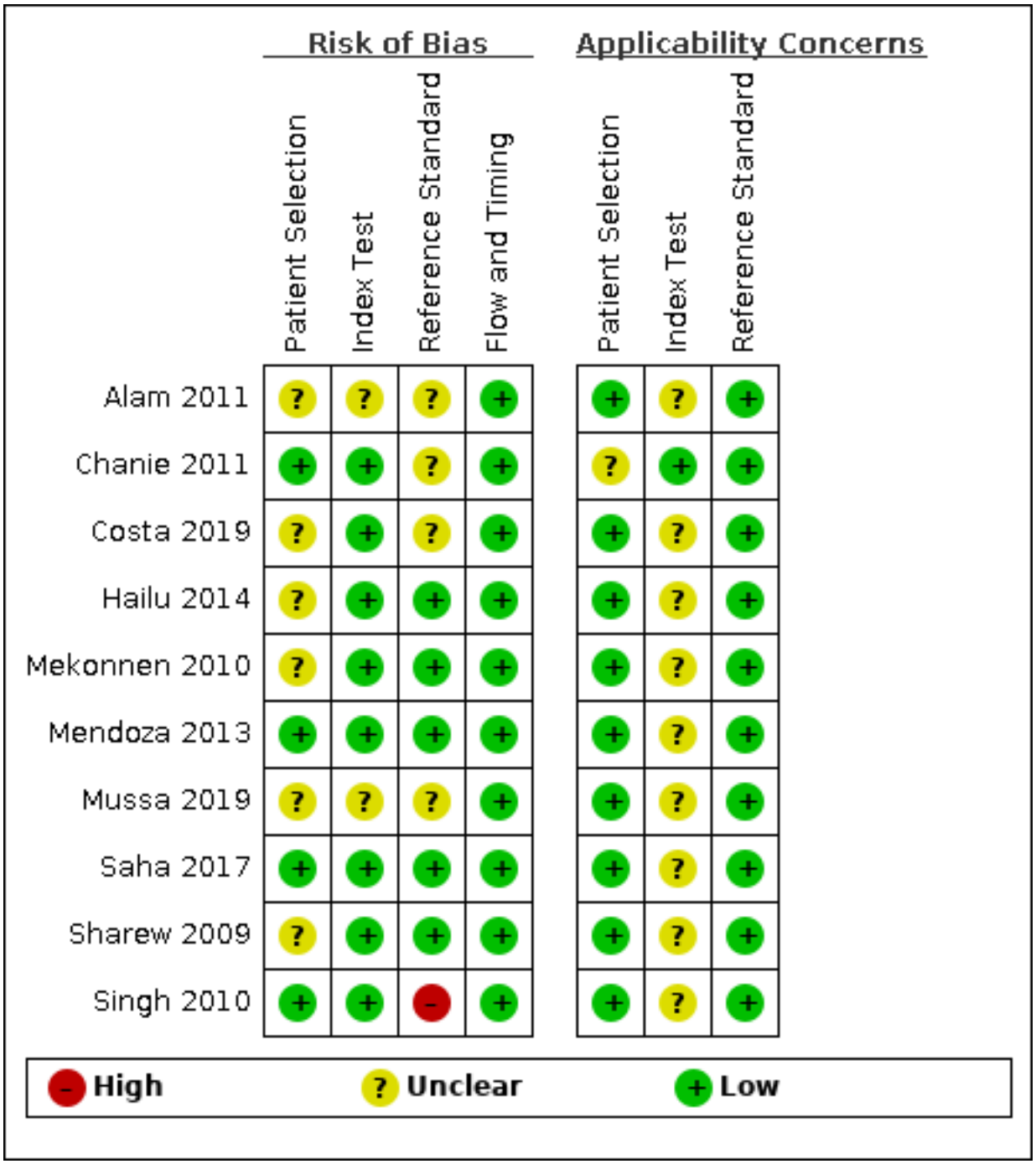
2010), and one each in Bangladesh (Alam 2011), Brazil (Costa 2019), Colombia (Mendoza 2013), and Sudan (Mussa 2019).

There was a lack of detail on how the RDTs were stored and whether RDT lots were quality-controlled prior to testing. Key study characteristics that may affect the performance of RDTs (e.g. training level of person performing the RDT, storage conditions, and parasite density of microscopy-positive cases or PCR) are summarised in Table 1.

Methodological quality of included studies

The results of the risk of bias and applicability assessment are summarised in Figure 2. One study was judged to be at low risk of bias in all four domains of the QUADAS-2 tool (Saha 2017). This study assessed the SD Bioline Malaria Ag Pf/Pv test.

Figure 2. Risk of bias and applicability concerns summary: review authors' judgements about each domain for each included study



Patient selection

Six (60%) studies were at unclear risk of bias in the patient selection domain because the method of participant recruitment (random or consecutive) was unclear (five studies), and/or the exclusion criteria were unclear (five studies). All studies were of low concern regarding applicability as they were all conducted in settings endemic with *P vivax.* However, Saha 2017 and Mussa 2019 did not report the prevalence of *P vivax* malaria. The remaining eight studies reported *P vivax* malaria or malaria in general as prevalent or endemic, but it was unclear to what degree.

Index test

We judged eight (80%) studies to be at low risk of bias in this domain because the results of the RDTs were interpreted without knowledge of the results of the reference standard. We judged the risk of bias for the remaining two studies to be unclear (Alam 2011; Mussa 2019). We judged the applicability of eight studies to be unclear, as poor reporting of the storage conditions or lot testing hampered the assessment. Singh 2010 provided thorough detail of how their RDT was stored, but it was unclear whether these conditions followed the instructions of the manufacturer. Applicability in this study was thus unclear. This study tested the temperature stability of the tests (see Table 1). Chanie 2011 evaluated the CareStart Malaria Pf/Pv Combo test. This was the only study considered to be of low applicability concern, because lot testing was reported.

Reference standard

We judged five studies to be at low risk of bias in the reference standard domain (Mendoza 2013; Saha 2017; Hailu 2014; Mekonnen 2010; Sharew 2009), while we judged one to be at high risk of bias (Singh 2010). We judged the remaining four studies to be at unclear risk of bias in this domain. It was unclear for two studies whether the results of the reference standard were interpreted without knowledge of the RDT results (Alam 2011, Mussa 2019), and it was unclear for two studies whether the results of the reference standard could classify the target condition. Costa 2019 and Chanie 2011 did not provide enough information on the reference standard to deduce if at least two microscopists independently examined the same slides from microscopy. We deemed Singh 2010 to be at high risk of bias because the second microscopist did not verify all of the reference standard results.

Flow and timing

We judged all 10 studies to be at low risk of bias in the flow and timing domain. All studies avoided partial verification, differential verification and incorporation bias, and reasons for any withdrawals were recorded. Nine studies appeared to have no uninterpretable results because the number of participants enrolled matched the number in the analysis. The remaining study reported two invalid RDT results, which were retested with the same test kits by taking fresh blood from the patients (Hailu 2014). However, it was unclear whether the same blood sample was used for the reference standard.

Test comparison

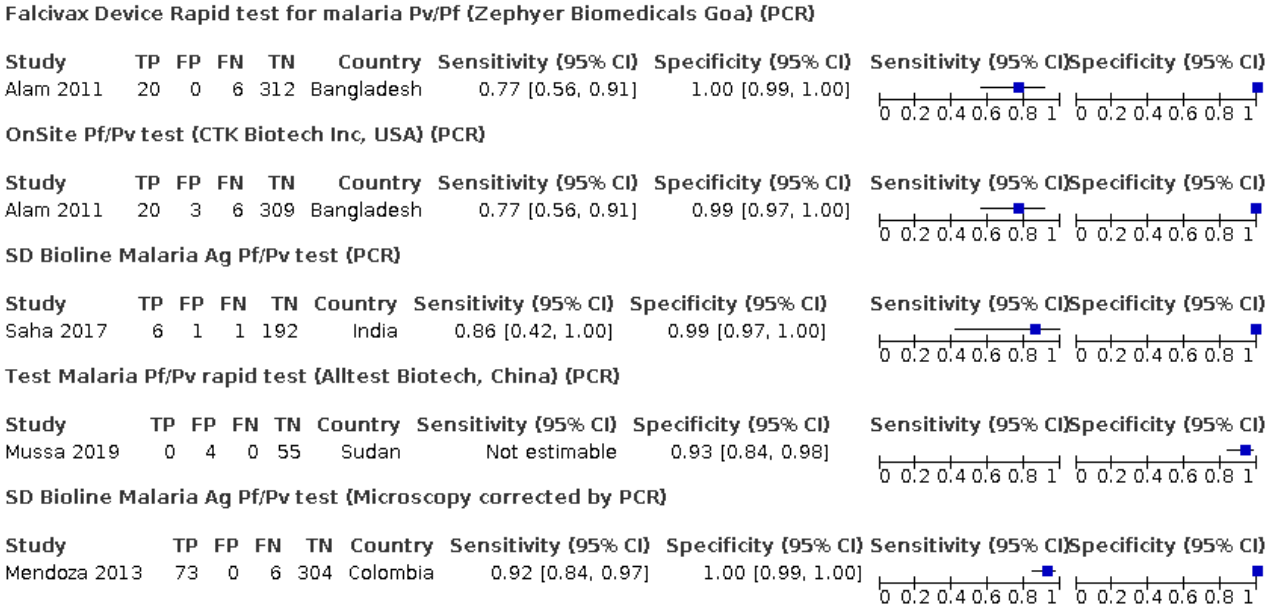
Although the QUADAS-2 tool does not specifically address risk of bias in a test comparison, we additionally considered the potential for such bias in a study that directly compared two RDT brands (OnSite Pf/Pv test and Falcivax Device Rapid test) (Alam 2011). It was unclear whether the results of one RDT brand were interpreted without knowledge of the results of the other brand. The study used both microscopy and PCR as two separate reference standards, but it was unclear whether the conduct and interpretation of the results from these two reference standards were done independently of each other.

Findings

Verified by PCR

Three studies (Alam 2011; Mussa 2019; Saha 2017) evaluated the accuracy of four different brands of RDTs against PCR (Figure 3; Table 2). One of the studies had no cases of *P vivax* malaria, so sensitivity was not estimable (Mussa 2019). The sensitivities of the RDTs ranged between 77% and 86% and the specificities ranged between 93% and 100%.

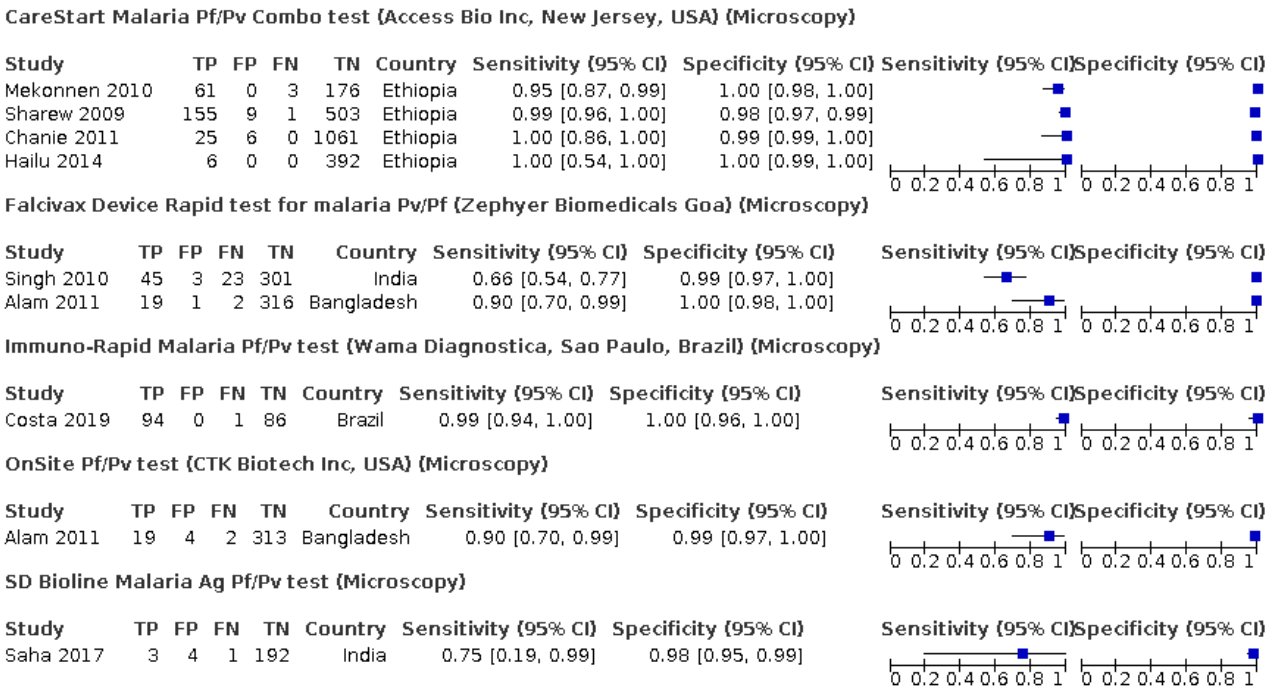
Figure 3. Forest plot of brands of rapid diagnostic tests verified against PCR or microscopy corrected with PCR



Verified by microscopy

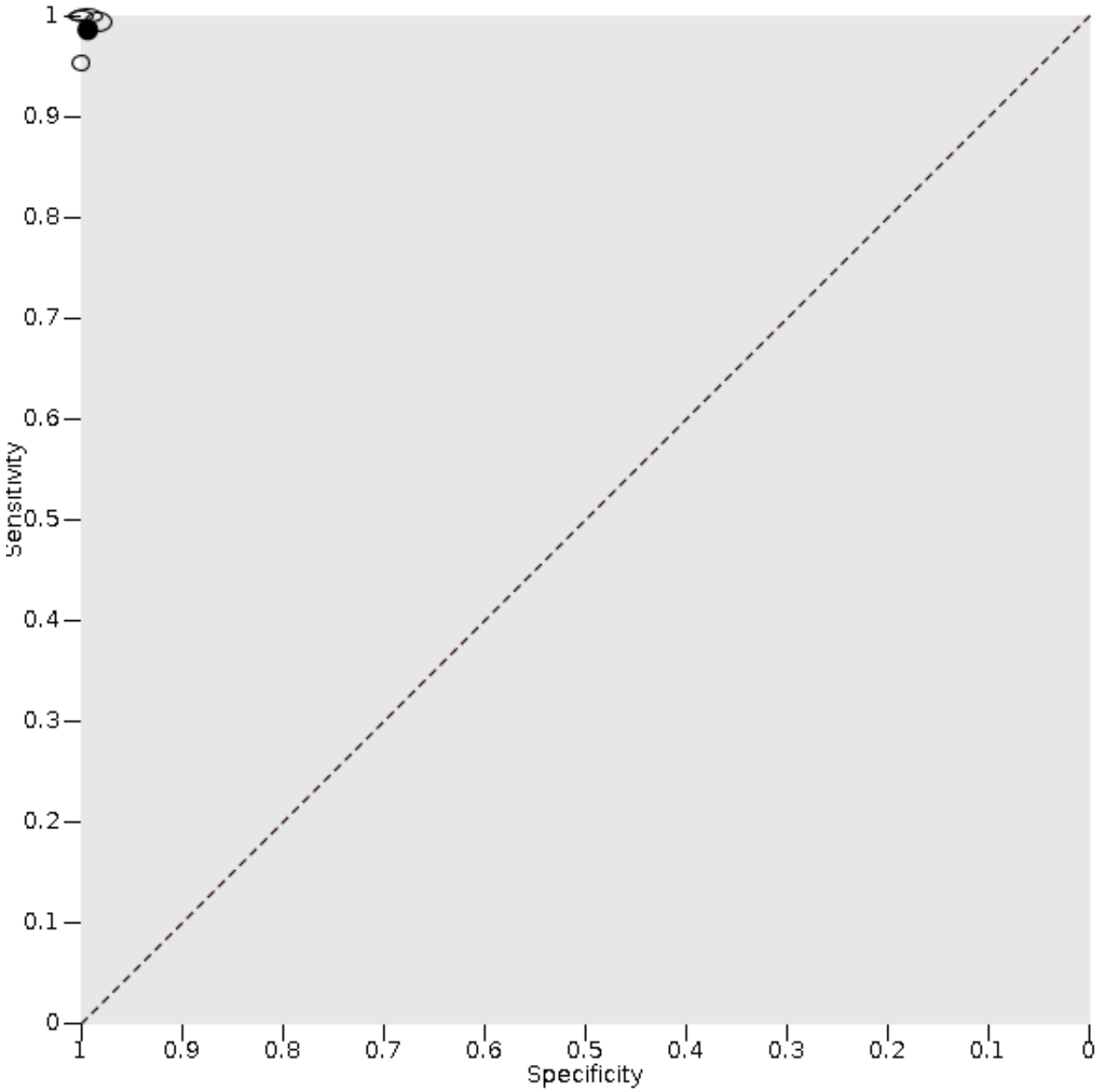
Eight studies conducted in four different countries evaluated the accuracy of RDTs against microscopy (Figure 4). Five different RDT brands were assessed: CareStart Malaria Pf/Pv Combo test (four studies), Falcivax Device Rapid test (two studies), Immuno-Rapid Malaria Pf/Pv test (one study), OnSite Pf/Pv test (one study), and SD Bioline Malaria Ag Pf/Pv test (one study).

Figure 4. Forest plot of brands of rapid diagnostic tests verified against microscopy, within each brand sorted by sensitivity and specificity



In the four CareStart Malaria Pf/Pv Combo test studies (251 *P vivax* malaria cases, 2398 patients), the sensitivity ranged from 95% to 100% and specificity ranged from 98% to 100%. The pooled sensitivity (95% CI) was 99% (94% to 100%) and the pooled specificity (95% CI) was 99% (99% to 100%) (Figure 5). The positive likelihood ratio (95% CI) was 141.09 (68.18 to 292.00) and the negative likelihood ratio (95% CI) was 0.01 (0.00 to 0.06).

Figure 5. Summary ROC plot for CareStart Malaria Pf/Pv Combo test verified against microscopy. The size of each study point was scaled by the sample size of the diseased and non-diseased groups used to estimate the study's sensitivity and specificity respectively, and reflects the precision of sensitivity and specificity in the study relative to other study points.The solid circle (summary point) represents the summary estimate of sensitivity and specificity. The summary point is not surrounded by a 95% confidence region because the bivariate model was simplified to univariate models.



The sensitivities of the Falcivax Device Rapid test from the two studies (89 *P vivax* malaria cases, 710 patients) were 66% (95% CI 54% to 77%) and 90% (95% CI 70% to 99%), and specificities were 99% (95% CI 97% to 100%) and 100% (95% CI 98% to 100%). The pooled sensitivity (95% CI) was 77% (53% to 91%) and the pooled specificity (95% CI) was 99% (98% to 100%). The positive likelihood ratio (95% CI) was 120.31 (43.10 to 335.87) and the negative likelihood ratio (95% CI) was 0.23 (0.10 to 0.53).

The sensitivities of the three remaining RDT brands ranged between 75% and 99% and the specificities ranged between 98% and 100% (Table 2; Figure 4).

Verified by microscopy corrected with PCR

Mendoza 2013 evaluated the accuracy of SD Bioline Malaria Ag Pf/Pv test against microscopy corrected with PCR (Figure 3). When there were discordant results between microscopy and PCR, the result of the PCR was taken, except in those in which the thick drop showed parasitic forms and the PCR was negative. The study reported a sensitivity (95% CI) of 92% (84% to 97%) and a specificity (95% CI) of 100% (99% to 100%).

Comparison between RDT brands

Alam 2011 directly compared the accuracy of Falcivax Device Rapid test and OnSite Pf/Pv test with PCR and microscopy as the reference standards. There was no evidence to suggest a difference in the sensitivity and specificity of the two brands (Table 3). Using microscopy as the reference standard, the absolute difference in sensitivity (95% CI) was 0 percentage points (-17.8 to 17.8 percentage points) and the absolute difference in specificity (95% CI) was 0.9 percentage points (-0.4 to 2.3 percentage points). Using PCR as the reference standard, the differences in sensitivity and specificity were similar.

DISCUSSION

Summary of main results

This systematic review included 10 studies conducted in six different countries (Bangladesh, Brazil, Colombia, Ethiopia, India, and Sudan). The studies assessed six different RDT brands: CareStart Malaria Pf/Pv Combo test (four studies), Falcivax Device Rapid test for malaria Pv/Pf (three studies), Immuno-Rapid Malaria Pf/Pv test (one study), SD Bioline Malaria Ag Pf/Pv test (three study), OnSite Pf/Pv test (two studies), and Test Malaria Pf/Pv rapid test (one study). However, only one study directly compared the accuracy of two brands.

The main findings of the review are summarised in Summary of findings 1, together with illustrations of what the findings mean. We assume median prevalences of ranges that would be classified as moderate, low, and very low transmission areas for *P vivax* (20%, 5%, and 0.5% respectively) in a hypothetical cohort of 1000 people suspected of having *P vivax* malaria (WHO 2017c). The CareStart Malaria Pf/Pv Combo test had a pooled sensitivity (95% CI) and specificity (95% CI) of 99% (94% to 100%) and 99% (99% to 100%) when microscopy was the reference standard. For a prevalence of 20%, about 206 people will have a positive CareStart Malaria Pf/Pv Combo test result and the remaining 794 people will have a negative result. Of the 206 people with positive results, eight will be incorrect (false positives), and of the 794 people with a negative result, two would be incorrect (false negative). The potential consequence of false positive results is unnecessary initiation of treatment and over-treatment of individuals with primaquine and either chloroquine or artemisinin combination therapies, and that patients are not treated for the actual cause of their symptoms. The consequences of false negative results are potential relapsing disease and continued risk of transmission of *P vivax* malaria at population level.

The Falcivax Device Rapid test had a pooled sensitivity and specificity of 77% (53% to 91%) and 99% (98% to 100%) when microscopy was the reference standard. For a prevalence of 20%, about 162 people will have a positive Falcivax Device Rapid test result and the remaining 838 people will have a negative result. Of the 162 people with positive results, eight will be incorrect (false positives), and of the 838 people with a negative result, 46 would be incorrect (false negative). A study that verified the results of the Falcivax Device Rapid test against PCR (Alam 2011), had a similar sensitivity and specificity of 77% (56% to 91%) and 100% (99% to 100%).

Strengths and weaknesses of the review

It is possible that some studies eligible for the inclusion in the review were missed by our search strategy. DTA studies are known to be poorly indexed, thus liable to be missed despite a broad literature search (Whiting 2009). However, our search was systematic, included studies published in all languages, and identified eligible studies from a previous review (Abba 2014). We also corresponded with study authors, when necessary, to obtain additional and unpublished data.

The main limitation of the review was the small number of studies included in the analyses. The meta-analysis of the Falcivax Device Rapid test verified by microscopy included only two studies. Thus, the pooled estimate of sensitivity, and in general from analyses containing a small number of studies, should be interpreted with caution. Comparative accuracy studies are known to be typically scarce (Takwoingi 2013). Only one of the included studies compared the accuracy of two RDT brands, so we were unable to conduct comparative meta-analyses to determine which brands were more sensitive and/or more specific. We intended to investigate any heterogeneity from the pooled analyses with pre-specified factors, as stated in our secondary objective, but this was not possible due to the small number of studies included in the analyses.

For the diagnostic test accuracy of RDTs, there is a lack of a 'perfect reference standard'. PCR is often seen as the gold standard for malaria diagnosis, because it is less prone to observer error and more sensitive at low levels of parasitaemia (Han 2017; Snounou 1993). On the other hand, it is too analytically sensitive to be a gold standard, because it detects subclinical infections (e.g. in patients with partial immunity). Furthermore, PCR sometimes has poor sensitivity for the detection of mixed infections (Shokoples 2009). A small sample of the cases in our review are mixed infections using PCR as the reference standard (Alam 2011; Mendoza 2013), so the analysis may be flawed.

PCR is currently not widely available due to logistical constraints, namely the need for specially-trained technicians and a well-equipped laboratory. It is thus mostly used for research purposes and is less applicable in clinical settings. Thus, microscopy in the correct clinical setting, with well-trained microscopists, remains the acceptable reference standard. This method is less costly than PCR, but infections can be missed if the slides are not examined carefully (Wongsrichanalai 2007). This raises the possibility that in some cases, the RDT results may in fact have been correct and the microscopy results incorrect. Alam 2011 verified RDT results against both microscopy and PCR separately, giving similar results of high specificity but lower sensitivity when verified against PCR. As mentioned previously, microscopy is more prone to observer error

and is less sensitive at low levels of parasitaemia in comparison to PCR.

As reported in the Methodological quality of included studies, there was a high number of 'unclear' evaluations of risk of bias and applicability due to poor reporting of study methods and characteristics. Nine studies (90%) did not provide enough information for us to adequately assess the selection of patients. Eight studies (80%) used an adequate reference standard, which was likely to have classified the target condition, but only four studies (40%) reported that readers of the reference standard were blinded to the results of the RDTs.

Applicability of findings to the review question

Due to the small number of studies included in this review, it is doubtful that the results obtained here can be considered to be generally applicable. Nevertheless, the findings show that the CareStart Malaria Pf/Pv Combo test verified by microscopy appeared to be both highly sensitive (missing 1% of cases) and highly specific (incorrectly classifying 1% of non-cases as positives) in detecting *P vivax* alone or as part of a mixed infection. In contrast, the Falcivax Device Rapid test, verified by microscopy, appeared to be less sensitive (missing 23% of cases), but was similarly highly specific. This result should be interpreted with caution because only two studies were used to obtain the pooled estimates.

Furthermore, the RDTs are heterogeneous in terms of quality. The devices can give ambiguous test results, are prone to drying out in low-humidity climates, resulting in lack of fluid migration. They are often not tested after they have been exposed to field conditions (Maltha 2013). In January 2020, the CareStart Malaria Pf/Pv Combo test produced by Access Bio Inc. was issued a WHO notice of concern due to their manufacturing quality assurance processes, which in turn could impact on patient safety (WHO 2020). Thus, in addition to considering results of test accuracy in published reports, end-users must be attuned to outcomes of periodic monitoring procedures of regulatory authorities and WHO prequalification.

Comparison with previous systematic reviews

An existing Cochrane Review assessing RDTs for diagnosing uncomplicated non-falciparum malaria was conducted in 2014 (Abba 2014). A subset of the review included RDTs that diagnosed *P vivax*. Our review only assessed the diagnostic accuracy of RDTs that specifically detect *P vivax* with *P vivax*-specific LDH antigens, however all the RDTs included in this review are combo tests that are used to detect *P falciparum* as well as *P vivax*. We included 10 studies, of which five studies (Alam 2011; Chanie 2011; Mekonnen 2010; Singh 2010; Sharew 2009) were included in the review by Abba 2014. Four studies were published following the review by Abba 2014 (Costa 2019; Hailu 2014; Mussa 2019; Saha 2017). One study (Mendoza 2013) was excluded by Abba 2014 because non-English language studies were excluded due to resource constraints. We included studies published in all languages.

AUTHORS' CONCLUSIONS

Implications for practice

Differentiating between *Plasmodium* species is particularly important in areas of co-endemicity whereby *P vivax* malaria is increasing proportionally, compared to *P falciparum* malaria. The main analysis included in this review was CareStart Malaria Pf/Pv Combo test against microscopy as the reference standard, and this RDT was found to be both highly sensitive and specific. Owing to concerns regarding methodological quality, these findings should be interpreted with caution. Only two RDT brands were assessed by more than one study in this review, so we could not assess differences in sensitivity and specificity between RDT brands. Studies often did not report on transport, storage conditions and quality control practices for RDTs such as lot testing prior to use, therefore damage to RDTs in transit or during the study period cannot be excluded and may have negatively impacted on test results. Studies also often did not report on the background parasitaemia density. This is an important variable which influences the performance of the RDTs.

Implications for research

More high-quality studies are needed to assess and compare the accuracy of RDTs designed to detect *P vivax*. The studies should clearly report their sampling methods, if exclusion criteria were used and whether the results of index tests and reference standards were blinded from each other. Studies should also report the background parasitaemia density, if and how RDTs were quality assured prior to use, including details of transport, storage conditions, and lot testing.

In the future, the RDTs studied here may no longer be available. The quality of those that remain may be improved by the manufacturers. Thus, this review will require updating.

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REFERENCES

References to studies included in this review

Alam 2011 *{published data only}*

Alam MS, Mohon AN, Mustafa S, Khan WA, Islam N, Karim MJ, et al. Real-time PCR assay and rapid diagnostic tests for the diagnosis of clinically suspected malaria patients in Bangladesh. *Malaria Journal* 2011;**10**:175.

Chanie 2011 *{published data only}*

Chanie M, Erko B, Animut A, Legesse M. Performance of CareStart™ Malaria Pf/Pv Combo test for the diagnosis of Plasmodium falciparum and Plasmodium vivax infections in the afar region, North East Ethiopia. *Ethiopian Journal of Health Development* 2011;**25**(3):206-11.

Costa 2019 *{published data only}*

Costa MRF, Barcelos ALR, Camargo MA, Melo GC, Almeida AC, Costa AG, et al. Performance of an immuno-rapid malaria Pf/Pv rapid diagnostic test for malaria diagnosis in the Western Brazilian Amazon. *Revista da Sociedade Brasileira de Medicina Tropical* 2019;**52**:e-20170450.

Hailu 2014 *{published data only}*

Hailu T, Kebede T. Assessing the performance of CareStart Malaria Pf/Pv Combo test against thick blood film in the diagnosis of malaria in northwest Ethiopia. *The American Journal of Tropical Medicine and Hygiene* 2014;**90**(6):1109-12.

Mekonnen 2010 *{published data only}*

Mekonnen Z, Ali S, Belay G, Suleman S, Chatterjee S. Evaluation of the performance of CareStart™ Malaria Pf/Pv Combo rapid diagnostic test for the diagnosis of malaria in Jimma, Southwestern Ethiopia. *Acta Tropica* 2010;**113**(3):285-8.

Mendoza 2013 *{published data only}*

Mendoza NM, Cucunubá ZM, Aponte S, González NE, Bernal SD. Field evaluation for diagnostic accuracy of the rapid test SD Bioline Malaria Antigen Pf/Pv® in Colombia [Evaluación de campo de la precisión de la prueba de diagnóstico rápido SD Bioline Malaria Antigen Pf/Pv® en Colombia]. *Biomedica* 2013;**33**(4):587-97.

Mussa 2019 *{published data only}*

Mussa A, Talib M, Mohamed Z, Hajissa K. Genetic diversity of Plasmodium falciparum histidine-rich protein 2 (*Pf*HRP2) and its effect on the performance of *Pf*HRP2-based rapid diagnostic tests. *BMC Research Notes* 2019;**12**:334.

Saha 2017 *{published data only}*

Saha S, Narang R, Deshmukh P, Pote K, Anvikar A, Narang P. Diagnostic efficacy of microscopy, rapid diagnostic test and polymerase chain reaction for malaria using bayesian latent class analysis. *Indian Journal of Medical Microbiology* 2017;**35**(3):376-80.

Sharew 2009 *{published data only}*

Sharew B, Legesse M, Animut A, Jima D, Medhin G, Erko B. Evaluation of the performance of CareStart™ Malaria Pf/Pv Combo and Paracheck Pf tests for the diagnosis of malaria in Wondo Genet, Southern Ethiopia. *Acta Tropica* 2009;**111**(3):321-4.

Singh 2010 *{published data only}*

Singh N, Shukla MM, Shukla MK, Mehra RK, Sharma S, Bharti PK, et al. Field and laboratory comparative evaluation of rapid malaria diagnostic tests versus traditional and molecular techniques in India. *Malaria Journal* 2010;**9**:191.

References to studies excluded from this review

Abdelraheem 2016 *{published data only}*

Abdelraheem MH, Albsheer MM, Mohamed HS, Amin M, Mahdi Abdel Hamid M. Transmission of Plasmodium vivax in Duffy-negative individuals in Central Sudan. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2016;**110**(4):258-60.

Adams 2015 *{published data only}*

Adams M, Joshi SN, Mbambo G, Mu AZ, Roemmich SM, Shrestha B, et al. An ultrasensitive reverse transcription polymerase chain reaction assay to detect asymptomatic low-density Plasmodium falciparum and Plasmodium vivax infections in small volume blood samples. *Malaria Journal* 2015;**14**:520.

Adnan 2017 *{published data only}*

Adnan STA, Warraich RA, Khan MA, Tabassum S. Community based evaluation of malaria rapid diagnostic test (ICT) comparing with conventional method of microscopy. *Medical Forum Monthly* 2017;**28**(7):60-63.

Ageep 2013 *{published data only}*

Ageep AK. Diagnosis of malaria in Red Sea State, Sudan. *Annals of Tropical Medicine and Public Health* 2013;**6**(2):232-5.

Andrade 2010 *{published data only}*

Andrade BB, Reis-Filho A, Barros AM, Souza-Neto SM, Nogueira LL, Fukatano KF, et al. Towards a precise test for malaria diagnosis in the Brazilian Amazon: Comparison among field microscopy, a rapid diagnostic test, nested PCR, and a computational expert system based on artificial neural networks. *Malaria Journal* 2010;**9**:117.

Arvind 2015 *{published data only}*

Kumar A. Evaluation of parasite LDH detection strip test (OptiMAL) for rapid diagnosis of malaria in children. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 2015;**6**(1):557-61.

Ashton 2010 *{published data only}*

Ashton RA, Kefyalew T, Tesfaye G, Counihan H, Yadeta D, Cundill B, et al. Performance of three multi-species rapid diagnostic tests for diagnosis of Plasmodium falciparum and Plasmodium vivax malaria in Oromia Regional State, Ethiopia. *Malaria Journal* 2010;**9**:297.

Ayorinde 2016 *{published data only}*

Ayorinde AF, Oyeyiga AM, Nosegbe NO, Folarin OA. A survey of malaria and some arboviral infections among suspected febrile patients visiting a health centre in Simawa, Ogun State, Nigeria. *Journal of Infection and Public Health* 2016;**9**(1):52-9.

Ba 2017 *{published data only}*

Ba H, Ahouidi AD, Duffy CW, Deh YB, Diedhiou C, Tandia A, et al. Evaluation of malaria rapid diagnostic test Optimal-IT® pLDH along the Plasmodium falciparum distribution limit in Mauritania [Evaluation du test de diagnostic rapide du paludisme OptiMal-IT® pLDH à la limite de la distribution de Plasmodium falciparum en Mauritanie]. *Bulletin de la Societe de pathologie exotique* 2017;**110**(1):31-7.

Bahk 2018 *{published data only}*

Bahk YY, Park SH, Lee W, Jin K, Ahn SK, Na BK, et al. Comparative Assessment of Diagnostic Performances of Two Commercial Rapid Diagnostic Test Kits for Detection of Plasmodium spp. in Ugandan Patients with Malaria. *Korean Journal of Parasitology* 2018;**56**(5):447-52.

Barber 2013 *{published data only}*

Barber BE, William T, Grigg MJ, Piera K, Yeo TW, Anstey NM. Evaluation of the sensitivity of a pLDH-based and an aldolase-based rapid diagnostic test for diagnosis of uncomplicated and severe malaria caused by PCR-confirmed Plasmodium knowlesi, Plasmodium falciparum, and Plasmodium vivax. *Journal of Clinical Microbiology* 2013;**51**(4):1118–23.

Bell 2001 *{published data only}*

Bell D, Go R, Miguel C, Walker J, Cacal L, Saul A. Diagnosis of malaria in a remote area of the Philippines: Comparison of techniques and their acceptance by health workers and the community. *Bulletin of the World Health Organization* 2001;**79**(10):933-41.

Bendezu 2010 *{published data only}*

Bendezu J, Rosas A, Grande T, Rodriguez H, Llanos-Cuentas A, Escobedo J, et al. Field evaluation of a rapid diagnostic test (Parascreen™) for malaria diagnosis in the Peruvian Amazon. *Malaria Journal* 2010;**9**:154.

Berhane 2017 *{published data only}*

Berhane A, Russom M, Bahta I, Hagos F, Ghirmai M, Uqubay S. Rapid diagnostic tests failing to detect Plasmodium falciparum infections in Eritrea: An investigation of reported false negative RDT results. *Malaria Journal* 2017;**16**(1):105.

Berzosa 2018 *{published data only}*

Berzosa P, de Lucio A, Romay-Barja M, Herrador Z, González V, García L, et al. Comparison of three diagnostic methods (microscopy, RDT, and PCR) for the detection of malaria parasites in representative samples from Equatorial Guinea. *Malaria Journal* 2018;**17**(1):333.

Bharti 2008 *{published data only}*

Bharti PK, Silawat N, Singh PP, Singh MP, Shukla M, Chand G, et al. The usefulness of a new rapid diagnostic test, the First Response Malaria Combo (pLDH/HRP2) Card Test, for malaria diagnosis in the forested belt of Central India. *Malaria Journal* 2008;**7**:126.

Bharti 2013 *{published data only}*

Bharti PK, Chand SK, Singh MP, Mishra S, Shukla MM, Singh R, et al. Emergence of a new focus of Plasmodium malariae in forest villages of District Balaghat, Central India: Implications for the diagnosis of malaria and its control. *Tropical Medicine and International Health* 2013;**18**(1):12-7.

Bhide 2014 *{published data only}*

Bhide M, Parekh V. Automated screening for malaria parasites on Mindray BC-6800 Hematology Analyzer: A follow-up report. *International Journal of Laboratory Hematology* 2014;**36**:80-1.

Birhanie 2016 *{published data only}*

Birhanie M. Comparison of Partec Rapid Malaria Test with Conventional Light Microscopy for Diagnosis of Malaria in Northwest Ethiopia. *Journal of Parasitology Research* 2016;**2016**:3479457.

Bisoffi 2014 *{published data only}*

Bisoffi Z, Gobbi F, Van den Ende J. Rapid diagnostic tests for malaria. *BMJ* 2014;**348**:g3846.

Britton 2016 *{published data only}*

Britton S, Cheng Q, Grigg MJ, Poole CB, Pasay C, William T, et al. Sensitive Detection of Plasmodium vivax Using a High-Throughput, Colourimetric Loop Mediated Isothermal Amplification (HtLAMP) Platform: A Potential Novel Tool for Malaria Elimination. *PLoS Neglected Tropical Diseases* 2016;**10**(2):e0004443.

Chayani 2004 *{published data only}*

Chayani N, Das B, Sur M, Bajoria S. Comparison of parasite lactate dehydrogenase based immunochromatographic antigen detection assay (optimal) with microscopy for detection of malaria parasites. *Indian Journal of Medical Microbiology* 2004;**22**(2):104-6.

Cho 2016 *{published data only}*

Cho SJ, Lee J, Lee HJ, Jo HY, Sinniah M, Kim HY, et al. A Novel Malaria Pf/Pv Ab Rapid Diagnostic Test Using a Differential Diagnostic Marker Identified by Network Biology. *International Journal of Biological Sciences* 2016;**12**(7):824-35.

Dahesh 2015 *{published data only}*

Dahesh SM, Mostafa HI. Reevaluation of malaria parasites in El-Fayoum Governorate, Egypt using Rapid Diagnostic Tests (RDTs). *Journal of the Egyptian Society of Parasitology* 2015;**45**(3):617-28.

Dash 2013 *{published data only}*

Dash M. Comparison of rapid immunochromatographic assays (ICT malaria P.f./P.v. test and OptiMAL test) with microscopy for detection of malaria parasites. *International Journal of Pharma and Bio Sciences* 2013;**4**(4):322-7.

Deida 2019 *{published data only}*

Deida J, Tahar R, Khalef YO, Lekweiry KM, Hmeyade A, Khairy MLO, et al. Oasis Malaria, Northern Mauritania. *Emerging Infectious Diseases* 2019;**25**(2):273-80.

DeKoninck 2017 *{published data only}*

De Koninck AS, Cnops L, Hofmans M, Jacobs J, Van den Bossche D, Philippé J. Diagnostic performance of the loop-mediated isothermal amplification (LAMP) based illumigene® malaria assay in a non-endemic region. *Malaria Journal* 2017;**16**(1):418.

Dev 2004 *{published data only}*

Dev V. Relative utility of dipsticks for diagnosis of malaria in mesoendemic area for Plasmodium falciparum and P. vivax in Northeastern India. *Vector Borne and Zoonotic Diseases* 2004;**4**(2):123-30.

Dinzouna-Boutamba 2014 *{published data only}*

Dinzouna-Boutamba SD, Yang HW, Joo SY, Jeong S, Na BK, Inoue N, et al. The development of loop-mediated isothermal amplification targeting alpha-tubulin DNA for the rapid detection of Plasmodium vivax. *Malaria Journal* 2014;**13**:248.

Dzakah 2014 *{published data only}*

Dzakah EE, Kang K, Ni C, Tang S, Wang J, Wang J. Comparative performance of aldolase and lactate dehydrogenase rapid diagnostic tests in Plasmodium vivax detection. *Malaria Journal* 2014;**13**:272.

Ehtesham 2015 *{published data only}*

Ehtesham R, Fazaeli A, Raeisi A, Keshavarz H, Heidari A. Detection of mixed-species infections of Plasmodium falciparum and Plasmodium vivax by nested PCR and rapid diagnostic tests in Southeastern Iran. *American Journal of Tropical Medicine and Hygiene* 2015;**93**(1):181-5.

Eibach 2013 *{published data only}*

Eibach D, Traore B, Bouchrik M, Coulibaly B, Coulibaly N, Siby F, et al. Evaluation of the malaria rapid diagnostic test VIKIA Malaria Ag Pf/Pan™ in endemic and non-endemic settings. *Malaria Journal* 2013;**12**:188.

Elahi 2013 *{published data only}*

Elahi R, Mohon AN, Khan WA, Haque R, Alam MS. Performance of a HRP-2/pLDH based rapid diagnostic test at the Bangladesh-India-Myanmar border area for diagnosis of clinical malaria. *Malaria Journal* 2013;**12**:378.

Endeshaw 2012 *{published data only}*

Endeshaw T, Graves PM, Ayele B, Mosher AW, Gebre T, Ayalew F, et al. Performance of local light microscopy and the ParaScreen Pan/Pf rapid diagnostic test to detect malaria in health centers in Northwest Ethiopia. *PLoS One* 2012;**7**(4):e33014.

Falade 2016 *{published data only}*

Falade CO, Ajayi IO, Nsungwa-Sabiiti J, Siribie M, Diarra A, Serme L, et al. Malaria Rapid Diagnostic Tests and Malaria Microscopy for Guiding Malaria Treatment of Uncomplicated Fevers in Nigeria and Prereferral Cases in 3 African Countries. *Clinical Infectious Diseases* 2016;**63**:S290-s297.

Fernando 2004 *{published data only}*

Fernando SD, Karunaweera ND, Fernando WP, Attanayake N, Wickremasinghe AR. A cost analysis of the use of the rapid, whole-blood, immunochromatographic P.f/P.v assay for the diagnosis of Plasmodium vivax malaria in rural areas of Sri Lanka. *Annals of Tropical Medicine and Parasitology* 2004;**98**(1):5-13.

Fernando 2004a *{published data only}*

Fernando SD, Karunaweera ND, Fernando WP. Evaluation of a rapid whole blood immunochromatographic assay for the diagnosis of Plasmodium falciparum and Plasmodium vivax malaria. *Ceylon Medical Journal* 2004;**49**(1):7-11.

Foster 2014 *{published data only}*

Foster D, Cox-Singh J, Mohamad DS, Krishna S, Chin PP, Singh B. Evaluation of three rapid diagnostic tests for the detection of human infections with Plasmodium knowlesi. *Malaria Journal* 2014;**13**:60.

Fransisca 2015 *{published data only}*

Fransisca L, Kusnanto JH, Satoto TB, Sebayang B, Supriyanto, Andriyan E, et al. Comparison of rapid diagnostic test Plasmotec Malaria-3, microscopy, and quantitative real-time PCR for diagnoses of Plasmodium falciparum and Plasmodium vivax infections in Mimika Regency, Papua, Indonesia. *Malaria Journal* 2015;**14**:103.

Gabrielli 2016 *{published data only}*

Gabrielli S, Bellina L, Milardi GL, Katende BK, Totino V, Fullin V, et al. Malaria in children of Tshimbulu (Western Kasai, Democratic Republic of the Congo): Epidemiological data and accuracy of diagnostic assays applied in a limited resource setting. *Malaria Journal* 2016;**15**:81.

Ghai 2016 *{published data only}*

Ghai RR, Thurber MI, El Bakry A, Chapman CA, Goldberg TL. Multi-method assessment of patients with febrile illness reveals over-diagnosis of malaria in rural Uganda. *Malaria Journal* 2016;**15**:460.

Gupta 2018 *{published data only}*

Gupta P Gupta P, Rao S, Singh N, Kalita D. Comparison between microscopy and rapid diagnostic tests in diagnosis of malaria at a tertiary care medical institution in Uttarakhand (A 3-year study). *Asian Journal of Pharmaceutical and Clinical Research* 2018;**11**(2):94-6.

Harani 2006 *{published data only}*

Harani MS, Beg MA, Khaleeq L, Adil SN, Kakepoto GN, Khurshid M. Role of ICT malaria immunochromatographic test for rapid diagnosis of malaria. *Journal of the Pakistan Medical Association* 2006;**56**(4):167-71.

Hawash 2019 *{published data only}*

Hawash Y, Ismail K, Alsharif K, Alsanie W. Malaria prevalence in a low transmission area, Jazan District of Southwestern Saudi Arabia. *Korean Journal of Parasitology* 2019;**57**(3):233–42.

Jabeen 2016 *{published data only}*

Jabeen S, Farrukh U, Hameed SA, Kanwal S, Qayyum M. An investigation on the prevalence and efficiency of immunochromatographic testing in suspected malarial patients of Rawalpindi and Islamabad, Pakistan. *Turkish Journal of Medical Sciences* 2016;**46**(5):1329-34.

Jahan 2019 *{published data only}*

Jahan F, Khan NH, Wahid S, Ullah Z, Kausar A, Ali N. Malaria epidemiology and comparative reliability of diagnostic tools in Bannu; an endemic malaria focus in south of Khyber Pakhtunkhwa, Pakistan. *Pathogens and Global health* 2019;**113**(2):75-85.

Joseph 2018 *{published data only}*

Joseph N, Uchila AK. Validation of malaria antigen detecting rapid diagnostic test kit: A study from highly endemic area in Coastal India. *Journal of Clinical and Diagnostic Research* 2018;**12**(9):16-20.

Karimov 2013 *{published data only}*

Karimov SS, Saiburkhonov DS. Evaluation of the efficiency of rapid tests in identifying malaria patients and parasite carriers in the Republic of Tajikistan. *Meditsinskaia parazitologiia i parazitarnye bolezni* 2013;**(1)**:44-5.

Kim 2013 *{published data only}*

Kim JY, Ji SY, Goo YK, Na BK, Pyo HJ, Lee HN, et al. Comparison of rapid diagnostic tests for the detection of Plasmodium vivax malaria in South Korea. *PLoS One* 2013;**8**(5):e64353.

Kolaczinski 2004 *{published data only}*

Klolaczinski J, Mohammed N, Ali I, Ali M, Khan N, Ezard N, et al. Comparison of the OptiMAL rapid antigen test with field microscopy for the detection of Plasmodium vivax and P. falciparum: Considerations for the application of the rapid test in Afghanistan. *Annals of Tropical Medicine and Parasitology* 2004;**98**(1):15-20.

Kosack 2013 *{published data only}*

Kosack CS, Naing WT, Piriou E, Shanks L. Routine parallel diagnosis of malaria using microscopy and the malaria rapid diagnostic test SD 05FK60: The experience of Médecins Sans Frontières in Myanmar. *Malaria Journal* 2013;**12**:167.

Kumari 2014 *{published data only}*

Kumari PR. A study of evaluation of rapid diagnostic techniques of Malaria in Urban Slums of Vijayawada, Krishna District, Andhra Pradesh, India. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 2014;**5**(4):1428-49.

Liu 2013 *{published data only}*

Liu H, Li XR, Li CF, Li XL, Wang HY, Nie RH. Field evaluation of SD(BIOLINE) malaria antigen Plasmodium falciparum/ Plasmodium vivax rapid test kit. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 2013;**31**(2):160-1.

Mallepaddi 2019 *{published data only}*

Mallepaddi PC, Maity SN, Poonati R, Pyadala N, Polavarapu R, Mangamuri UK, et al. Selecting better diagnostic kits for diagnosis of malarial parasites at point of care. *3 Biotech* 2019;**9**(1):36.

Metzger 2011 *{published data only}*

Metzger WG, Vivas-Martínez S, Giron A, Vaccari E, Campos E, Rodríguez I, et al. Assessment of routine malaria diagnosis in the Venezuelan Amazon. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2011;**105**(5):262-8.

Moges 2012 *{published data only}*

Moges B, Amare B, Belyhun Y, Tekeste Z, Gizachew M, Workineh M, et al. Comparison of CareStart HRP2/pLDH Combo rapid malaria test with light microscopy in North-West Ethiopia. *Malaria Journal* 2012;**11**:234.

Mohon 2012 *{published data only}*

Mohon AN, Elahi R, Podder MP, Mohiuddin K, Hossain MS, Khan WA, et al. Evaluation of the OnSite (Pf/Pan) rapid diagnostic test for diagnosis of clinical malaria. *Malaria Journal* 2012;**11**:415.

Olasehinde 2018 *{published data only}*

Olasehinde GI, Oyeka UC, Oniha MI, Onile-Ere OA, Ayepola OO, Ajayi AA, et al. Data set on rapid diagnostic tests (RDTs) and microscopy for diagnosing Plasmodium falciparum and Plasmodium vivax. *Data Brief* 2018;**20**:503-9.

Pakalapati 2013 *{published data only}*

Pakalapati D, Garg S, Middha S, Kochar A, Subudhi AK, Arunachalam BP, et al. Comparative evaluation of microscopy, OptiMAL(®) and 18S rRNA gene based multiplex PCR for detection of Plasmodium falciparum & Plasmodium vivax from field isolates of Bikaner, India. *Asian Pacific Journal of Tropical Medicine* 2013;**6**(5):346-51.

Pattanasin 2003 *{published data only}*

Pattanasin S, Proux S, Chompasuk D, Luwiradaj K, Jacquier P, Looareesuwan S, et al. Evaluation of a new Plasmodium lactate dehydrogenase assay (OptiMAL-IT) for the detection of malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2003;**97**(6):672-4.

Puri 2013 *{published data only}*

Puri B, Mehta P, Ingole NA, Prasad P, Mathure T. Laboratory tests for malaria: A diagnostic conundrum? *South African Medical Journal* 2013;**103**(9):625-7.

Rakotonirina 2008 *{published data only}*

Rakotonirina H, Barnadas C, Raherijafy R, Andrianantenaina H, Ratsimbasoa A, Randrianasolo L, et al. Accuracy and reliability of malaria diagnostic techniques for guiding febrile outpatient treatment in malaria-endemic countries. *American Journal of Tropical Medicine and Hygiene* 2008;**78**(2):217-21.

Ranjan 2016 *{published data only}*

Ranjan P, Ghoshal U. Utility of nested polymerase chain reaction over the microscopy and immuno-chromatographic test in the detection of Plasmodium species and their clinical spectrum. *Parasitology Research* 2016;**115**(9):3375-85.

Ratsimbasoa 2007 *{published data only}*

Ratsimbasoa A, Randriamanantena A, Raherinjafy R, Rasoarilalao N, Ménard D. Which malaria rapid test for Madagascar? Field and laboratory evaluation of three test and expert microscopy of samples from suspected malaria patients in Madagascar. *American Journal of Tropical Medicine and Hygiene* 2007;**76**(3):481-5.

Ratsimbasoa 2008 *{published data only}*

Ratsimbasoa A, Fanazava L, Radrianjafy R, Ramilijaona J, Rafanomezantsoa H, Ménard D. Evaluation of two new immunochromatographic assays for diagnosis of malaria. *American Journal of Tropical Medicine and Hygiene* 2008;**79**(5):670-2.

Samane 2010 *{published data only}*

Samane AK, Nahid HZ, Saaed S, Khazen H, Ali H, Ahmad R, et al. Comparison of microscopy and RDTs techniques for laboratory detection of malaria. *African Journal of Biotechnology* 2010;**9**(10):1514-6.

Selimuzzaman 2010 *{published data only}*

Selimuzzaman SM, Islam SJ, Nahar Z, Das R, Rahman MA, Rahman MA. Malariagen malaria Pf/Pv antigen rapid test: A simple and effective tool for diagnosis of malaria in the far-flung hilly areas of Bangladesh. *Mymensingh Medical Journal* 2010;**19**(1):94-9.

Shaikh 2013 *{published data only}*

Shaikh S, Memon S, Memon H, Ahmed I. Role of rapid diagnostic tests for guiding outpatient treatment of febrile illness in Liaquat University Hospital. *Pakistan Journal of Medical Sciences* 2013;**29**(5):1167-72.

Sharma 2014 *{published data only}*

Sharma J. Prevalence of malaria cases in tea garden areas of Lakhimpur District, Assam. *International Journal of Pharmacy and Pharmaceutical Sciences* 2014;**6**(8):571-3.

Singh 2000a *{published data only}*

Singh N, Saxena A, Valecha N. Field evaluation of the ICT malaria P.f/P.v immunochromatographic test for diagnosis of Plasmodium falciparum and P. vivax infection in forest villages in Chhindwara, Central India. *Tropical Medicine and International Health* 2000;**5**(11):765-70.

Singh 2003 *{published data only}*

Singh N, Valecha N, Nagpal AC, Mishra SS, Varma HS, Subbaro SK. The hospital- and field-based performances of the OptiMAL test, for malaria diagnosis and treatment monitoring in Central India. *Annals of Tropical Medicine and Parasitology* 2003;**97**(1):5-13.

Singh 2013 *{published data only}*

Singh N, Bharti PK, Singh MP, Mishra S, Shukla MM, Sharma RK, et al. Comparative evaluation of bivalent malaria rapid diagnostic tests versus traditional methods in field with special reference to heat stability testing in Central India. *PLoS One* 2013;**8**(3):e58080.

Siwal 2018 *{published data only}*

Siwal N, Singh US, Dash M, Kar S, Rani S, Rawal C, et al. Malaria diagnosis by PCR revealed differential distribution of mono and mixed species infections by Plasmodium falciparum and P. vivax in India. *PLoS One* 2018;**13**(3):e0193046.

Stijnberg 2013 *{published data only}*

Stijnberg D, Cairo H. Rapid diagnostic test (RDT) performance of the malaria gold mining program in Suriname: Comparing the performance of two RDT's. In: 62nd Annual Meeting of the American Society of Tropical Medicine and Hygiene; 2013 13-17 November; Washington, DC. Vol. 89(5 Suppl 1). Washington, DC, USA: The American Society of Tropical Medicine and Hygiene, 2013:384.

Strom 2014 *{published data only}*

Strøm GE, Moyo S, Fataki M, Langeland N, Blomberg B. PCR targeting Plasmodium mitochondrial genome of DNA extracted from dried blood on filter paper compared to whole blood. *Malaria Journal* 2014;**13**:137.

Thongdee 2014 *{published data only}*

Thongdee P, Chaijaroenkul W, Kuesap J, Na-Bangchang K. Nested-PCR and a new ELISA-based NovaLisa test kit for malaria diagnosis in an endemic area of Thailand. *Korean Journal of Parasitology* 2014;**52**(4):377-81.

Tjitra 1999 *{published data only}*

Tjitra E, Suprianto S, Dyer M, Currie BJ, Anstey NM. Field evaluation of the ICT malaria P.f/P.v immunochromatographic test in detection of Plasmodium falciparum and Plasmodium vivax in patients with a presumptive clinical diagnosis of malaria in Eastern Indonesia. *Journal of Clinical Microbiology* 1999;**37**(8):2412-7.

Trouvay 2013 *{published data only}*

Trouvay M, Palazon G, Berger F, Volney B, Blanchet D, Faway E, et al. High performance of histidine-rich protein 2 based rapid diagnostic tests in French Guiana are explained by the absence of pfhrp2 gene deletion in P. falciparum. *PLoS One* 2013;**8**(9):e74269.

Valecha 2003 *{published data only}*

Valecha N, Singh N, Yadav RS, Dev V, Aggarwal A, Subbarao SK. Field evaluation of OptiMAL48 rapid malaria diagnostic test in India. *Acta Parasitologica* 2003;**48**(3):229-32.

van den Broek 2006 *{published data only}*

van den Broek I, Hill O, Gordillo F, Angarita B, Hamade P, Counihan H, et al. Evaluation of three rapid tests for diagnosis of P. falciparum and P. vivax malaria in Colombia. *American Journal of Tropical Medicine and Hygiene* 2006;**75**(6):1209-15.

Vohra 2014 *{published data only}*

Vohra P, Mengi S, Bunger R, Pathania D, Singh VA. Comparison of peripheral blood film stained by Giemsa stain, acridine orange staining and rapid diagnostic tests for detection of P. vivax and P. falciparum in clinically suspected cases of malaria. *International Journal of Pharma Medicine and Biological Sciences* 2014;**3**(3):108-112.

Wang 2014 *{published data only}*

Wang ZY, Jiang L, Zhang YG, Zhang XP, Cai L. Comparison of two rapid diagnostic tests in detection of malaria parasites. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 2014;**32**(1):50-3.

Wongsrichanalai 2003 *{published data only}*

Wongsrichanalai G, Arevalo I, Laoboonchai A, Yingyuen K, Miller RS, Magill AJ, et al. Rapid diagnostic devices for malaria: Field evaluation of a new prototype immunochromatographic assay for the detection of Plasmodium falciparum and non-falciparum Plasmodium. *American Journal of Tropical Medicine and Hygiene* 2003;**69**(1):26-30.

Woyessa 2013 *{published data only}*

Woyessa A, Deressa W, Ali A, Lindtjørn B. Evaluation of CareStart™ Malaria Pf/Pv Combo test for Plasmodium falciparum and Plasmodium vivax malaria diagnosis in Butajira area, South-Central Ethiopia. *Malaria Journal* 2013;**12**:218.

Xiaodong 2013 *{published data only}*

Xiaodong S, Tambo E, Chun W, Zhibin C, Yan D, Jian W, et al. Diagnostic performance of CareStart™ malaria HRP2/pLDH (Pf/pan) Combo test versus standard microscopy on falciparum and vivax malaria between China-Myanmar endemic borders. *Malaria Journal* 2013;**12**:6.

Yan 2013 *{published data only}*

Yan J, Li N, Wei X, Li P, Zhao Z, Wang L, et al. Performance of two rapid diagnostic tests for malaria diagnosis at the China-Myanmar border area. *Malaria Journal* 2013;**12**:73.

References to studies awaiting assessment

Boni 2015 *{published data only}*

Boni MF, Lover AA, Ngo TD, Nguyen TX, Sutamihardja A, Tran DT, et al. Potential impact of rapid diagnostic test diagnostic error on increased malaria transmission of emerging untreatable malaria. In: 64th Annual Meeting of the American Society of Tropical Medicine and Hygiene; 2015 October 25-29; Philadelphia (PA). Vol. 93(4 Suppl). Philadelphia (PA), USA: The American Society of Tropical Medicine and Hygiene, 2015:371.

Cheng 2013 *{published data only}*

Cheng Z, Sun X, Yang Y, Wang H, Zheng Z. A novel, sensitive assay for high-throughput molecular detection of Plasmodia for active screening of malaria for elimination. *Journal of Clinical Microbiology* 2013;**51**(1):125-30.

Reda 2016 *{published data only}*

Reda AG. Evaluation of the performance of SD bioline malaria AG PF/PV and carestartTM malaria PF/PV combo tests for the diagnosis of malaria in two malarious areas in central Ethiopia. In: 65th Annual Meeting of the American Society of Tropical Medicine and Hygiene; 2016 November 13-17; Atlanta (GA). Vol. 95(5 Suppl). Atlanta (GA), USA: The American Society of Tropical Medicine and Hygiene, 2016:82-83.

Additional references

Abba 2014

Abba K, Kirkham AJ, Olliaro PL, Deeks JJ, Donegan S, Garner P, et al. Rapid diagnostic tests for diagnosing uncomplicated non-falciparum or Plasmodium vivax malaria in endemic countries. *Cochrane Database of Systematic Reviews* 2014, Issue 12. Art. No: CD011431. [DOI: 10.1002/14651858.CD011431]

Campo 2015

Campo B, Vandal O, Wesche DL, Burrows JN. Killing the hypnozoite – drug discovery approaches to prevent relapse in Plasmodium vivax. *Pathogens and Global Health* 2015;**109**(3):107-22.

Chen 2016

Chen I, Clarke SE, Gosling R, Hamainza B, Killeen G, Magill A, et al. "Asymptomatic" malaria: A chronic and debilitating infection that should be treated. *PLoS Medicine* 2016;**13**(1):e1001942.

Chu 2006

Chu H, Cole SR. Bivariate meta-analysis of sensitivity and specificity with sparse data: A generalized linear mixed model approach. *Journal of Clinical Epidemiology* 2006;**59**(12):1331-2; author reply 1332-3. [PMID: 17098577]

Culleton 2012

Culleton R, Carter R. African Plasmodium vivax: Distribution and origins. *International Journal for Parasitology* 2012;**42**(12):1091-7.

Deeks 2005

Deeks JJ, Macaskill P, Irwig L. The performance of tests of publication bias and other sample size effects in systematic reviews of diagnostic test accuracy was assessed. *Journal of Clinical Epidemiology* 2005;**58**(9):882-93.

Dobson 1994

Dobson MJ. Malaria in England: A geographical and historical perspective. *Parassitologia* 1994;**36**(1-2):35-60.

Gatton 2018

Gatton ML, Ciketic S, Barnwell JW, Cheng Q, Chiodini PL, Incardona S, et al. An assessment of false positive rates for malaria rapid diagnostic tests caused by non-Plasmodium infectious agents and immunological factors. *PLoS One* 2018;**13**(5):e0197395.

GRADE 2013

Schünemann H, Brożek J, Oxman A, editors. GRADE handbook for grading quality of evidence and strength of recommendations. Updated October 2013. Available from www.gdt.guidelinedevelopment.org/app/handbook/handbook.html (accessed 5 December 2017).

GRADEpro GDT 2015 [Computer program]

McMaster University (developed by Evidence Prime) GRADEpro GDT. Version accessed 5 December 2017. Hamilton (ON): McMaster University (developed by Evidence Prime), 2015. Available at gradepro.org.

Han 2017

Han TZ, Han KT, Aye KH, Hlaing T, Thant KZ, Vythilingam I. Comparison of microscopy and PCR for the detection of human Plasmodium species and Plasmodium knowlesi in Southern Myanmar. *Asian Pacific Journal of Tropical Biomedicine* 2017;**7**(8):680-5.

Hänscheid 2002

Hänscheid T, Grobusch MP. How useful is PCR in the diagnosis of malaria? *Trends in Parasitology* 2002;**18**(9):395-8.

Kakkilaya 2003

Kakkilaya BS. Rapid diagnosis of malaria. *Laboratory Medicine* 2003;**34**(8):602-8.

Macaskill 2010

Macaskill P, Gatsonis C, Deeks JJ, Harbord RM, Takwoingi Y. Chapter 10: Analysing and presenting results. In: Deeks JJ, Bossuyt PM, Gatsonis C, editor(s). Cochrane Handbook for Systematic Reviews of Diagnostic Test Accuracy Version 1.0. The Cochrane Collaboration 2010. Available from: methods.cochrane.org/sdt/handbook-dta-reviews.

Maltha 2013

Maltha J, Gillet P, Jacobs J. Malaria rapid diagnostic tests in endemic settings. *Clinical Microbiology and Infection* 2013;**19**(5):399-407.

Moonasar 2007

Moonasar D, Goga AE, Frean J, Kruger P, Chandramohan D. An exploratory study of factors that affect the performance and usage of rapid diagnostic tests for malaria in the Limpopo Province, South Africa. *Malaria Journal* 2007;**6**:74.

RevMan 2014 [Computer program]

Nordic Cochrane Centre, The Cochrane Collaboration Review Manager 5 (RevMan 5). Version 5.3. Copenhagen: Nordic Cochrane Centre, The Cochrane Collaboration, 2014.

Roth 2016

Roth J, Korevaar D, Leeflang MM, Mens P. Molecular malaria diagnostics: A systematic review and meta-analysis. *Critical Reviews in Clinical Laboratory Sciences* 2016;**53**(2):87-105.

Shokoples 2009

Shokoples SE, Ndao M, Kowalewska-Grochowska K, Yanow SK. Multiplexed real-time PCR assay for discrimination of Plasmodium species with improved sensitivity for mixed infections. *Journal of Clinical Microbiology* 2009 Apr 1;**47**(4):975-80.

Snounou 1993

Snounou G, Viriyakosol S, Zhu XP, Jarra W, Pinheiro L, do Rosario VE, et al. High sensitivity of detection of human malaria parasites by the use of nested polymerase chain reaction. *Molecular and Biochemical Parasitology* 1993;**61**(2):315-20.

STATA 2015 [Computer program]

Stata statistical software. Version 15. College Station, TX: StataCorp LLC, 2017.

Takwoingi 2013

Takwoingi Y, Leeflang MM, Deeks JJ. Empirical evidence of the importance of comparative studies of diagnostic test accuracy. *Annals of Internal Medicine* 2013;**158**(7):544-54. [MEDLINE: 23546566]

Takwoingi 2015a

Takwoingi Y, Guo B, Riley RD, Deeks JJ. Performance of methods for meta-analysis of diagnostic test accuracy with few studies or sparse data. *Statistical Methods in Medical Research* 2015;**26**(4):1896-911.

Takwoingi 2015b

Takwoingi Y, Riley DD, Deeks JJ. Meta-analysis of diagnostic accuracy studies in mental health. *Evidence-Based Mental Health* 2015;**18**(4):103-9. [DOI: 10.1136/eb-2015-102228]

Talman 2007

Talman AM, Duval L, Legrand E, Hubert V, Yen S, Bell D, et al. Evaluation of the intra and inter-specific genetic variability of Plasmodium lactate dehydrogenase. *Malaria Journal* 2007;**6**:140.

White 2018

White NJ. Anaemia and malaria. *Malaria Journal* 2018;**17**:371.

Whiting 2009

Whiting P, Westwood M, Burke M, Sterne J, Glanville J. Systematic reviews of test accuracy should search a range of databases to identify primary studies. *Journal of Clinical Epidemiology* 2009;**61**(4):357–64.

Whiting 2011

Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Annals of Internal Medicine* 2011;**155**(8):529-36.

WHO 2003

World Health Organization. Malaria rapid diagnosis:making it work: World Health Organization. Meeting report 20-23 January, Manila. apps.who.int/iris/bitstream/handle/10665/208030/RS_2003_GE_05_PHL_eng.pdf?sequence=1&isAllowed=y (accessed 30 Oct 2018).

WHO 2006

World Health Organization. Towards quality testing of malaria rapid diagnostic tests: evidence and methods. Proceedings of the WHO Informal Consultation on development and methods for testing malaria rapid diagnostic tests 28 February - 2 March 2006. Available at www.portal.pmnch.org/malaria/publications/atoz/929061238X/en/ (accessed 30 Oct 2018).

WHO 2012

World Health Organization. Malaria rapid diagnostic test performance: results of WHO product testing of malaria RDTs: Round 4 (2012). December 2012. Available at www.who.int/malaria/publications/rapid_diagnostic/en/ (accessed 30 Oct 2018).

WHO 2014

World Health Organization, Medicines for Malaria Venture, Roll Back Malaria, and the Wellcome Trust. Severe malaria. *Tropical Medicine & International Health* 2014;**19**(s1):7-131.

WHO 2015a

World Health Organization. Guidelines for the treatment of malaria. Third edition. April 2015. http://apps.who.int/iris/bitstream/handle/10665/162441/9789241549127_eng.pdf;jsessionid=9F90A5F3F058C3DE058158B4003EDA38?sequence=1 (accessed 26 July 2018).

WHO 2015b

World Health Organization. The role of RDTs in malaria control. www.who.int/malaria/areas/diagnosis/rapid-diagnostic-tests/role_in_malaria_control/en/ (accessed 4 December 2017).

WHO 2017a

World Health Organization. World Malaria Report 2017. apps.who.int/iris/bitstream/handle/10665/259492/9789241565523-eng.pdf;jsessionid=85F16FF476D140F1E6DEF84CF03783DC?sequence=1 (accessed 9 April 2018).

WHO 2017b

World Health Organization. WHO-FIND malaria RDT evaluation programme. Last update 10 July 2017. www.who.int/malaria/areas/diagnosis/rapid-diagnostic-tests/rdt-evaluation-programme/en/ (accessed 4 December 2017).

WHO 2017c

World Health Organization. A framework for malaria elimination. Last update March 2017. www.who.int/malaria/publications/atoz/9789241511988/en/ (accessed 29 April 2020).

WHO 2020

World Health Organization. WHO information notice for users. Last updated January 2020. www.who.int/diagnostics_laboratory/procurement/accessbioinc_20012020_noticeforusers.pdf?ua=1 (accessed 30 July 2020).

Wongsrichanalai 2007

Wongsrichanalai C, Barcus MJ, Muth S, Sutamihardja A, Wernsdorfer WH. A review of malaria diagnostic tools: Microscopy and rapid diagnostic test (RDT). *American Journal of Tropical Medicine and Hygiene* 2007;**77**(6 Suppl):119-27. [PMID: 18165483]

CHARACTERISTICS OF STUDIES

Characteristics of included studies *[ordered by study ID]*

Alam 2011

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** did not state consecutive or random sampling **Study period:** May 2009 to August 2010 **Population:** 338 febrile patients referred for microscopy to diagnose malaria diagnosis at a health facility **Inclusion and exclusion criteria:** not reported
Patient characteristics and setting	**Sex:** 49.7% male, 50.3% female **Age:** median = 14 years, range 18 months to 82 years **Setting:** Matiranga Upazila Health Complex (UHC), in Matiranga Upazila (sub-district) of Khagrachari district, south-eastern part of Bangladesh **Malaria transmission:** perennial transmission of malaria with 2 peaks in pre-monsoon (March to May) and post-monsoon (September to November) periods
Index tests	**RDT brand(s):** OnSite Pf/Pv test (CTK Biotech Inc, USA) and Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** unclear, reported manufacturer's instructions were followed for use

Alam 2011 *(Continued)*

	Blinding: not reported
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** PCR and microscopy **Microscopy details:** • 200 high powered fields • Two microscopists independently examined each microscopic slide; one of which was employed by the study and the other was posted at Matiranga UHC. • Slide considered positive only when the two microscopists were in agreement. Discrepancies were resolved by a third microscopist. **PCR details:** • Did not report who performed PCR. • Detection limit of 5-10 parasites/µL. **Blinding:** not reported
Flow and timing	**Appropriate interval between index test and reference standard:** one blood sample taken from each patient. **Invalid test results:** None reported.
Comparative	
Notes	

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Unclear		
Could the selection of patients have introduced bias?		Unclear risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		

Alam 2011 *(Continued)*

Could the conduct or interpretation of the index test have introduced bias?		Unclear risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Unclear risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Chanie 2011

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** consecutive **Study period:** December 2009 and January 2010 **Population:** 1092 febrile patients who had clinical symptoms of malaria and visited the outpatient department (OPD) of three health facilities. **Inclusion and exclusion criteria:** no exclusion criteria, unless the patient or the guardians of children less than 18 years old did not consent to participate.
Patient characteristics and setting	**Sex:** 51.4% male, 48.6% female **Age:** median = 22.3 years, SD: 12.8 **Setting:** 75 (61.81%), 238 (21.8%) and 179 (16.4%) patients were respectively from Melkawerer Health Centre, Gewane Health Centre and Dubti Hospital, in the Afar Region, Northeast Ethiopia

Chanie 2011 *(Continued)*

	Malaria transmission: The study reported the transmission of malaria as unstable, with some areas of perennial transmission. **Other patient characteristics:** 12.5% of the patients had anti-malarial therapy in the preceding month.
Index tests	**RDT brand(s):** CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ) **Batch number:** not reported **Lot testing:** Lot No H38 IV and Lot No H28 IV **Storage conditions:** Test kits were kept at the local temperature of the region without any controlling system of the storage temperature during data collection. The quality of the package desiccant was checked before use. **Blinding:** Microscopy and RDT were performed independently by malaria technicians, results were recorded separately.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy **Microscopy details:** • minimum of 100 high powered fields • microscopy performed by experienced malaria technicians. • The study did not explicitly state the number of observers or repeats. • 20% of the positive and 10% of the negative slides and discordant results between CareStartTM Malaria Pf/Pv Combo test and those of microscopy were examined by another well experienced technician. **Blinding:** Microscopy and RDT was performed independently by malaria technicians, results were recorded separately.
Flow and timing	**Appropriate interval between index test and reference standard:** one blood sample taken from each patient. **Invalid test results:** None reported.
Comparative	
Notes	

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
Could the selection of patients have introduced bias?		Low risk	

Chanie 2011 *(Continued)*

Are there concerns that the included patients and setting do not match the review question?			Unclear
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Low concern
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Unclear		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Unclear risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Costa 2019

Study characteristics	
Patient Sampling	**Study design:** prospective cross-sectional study **Recruitment:** consecutive **Study period:** November 2016 and April 2017

Costa 2019 *(Continued)*

	Population: 181 febrile patients were recruited based on the hospital's regular admissions **Inclusion and exclusion criteria:** not reported
Patient characteristics and setting	**Sex:** 64.1% male, 35.9% female **Age:** median = 41.7 years, SD: 14.4 **Setting:** Tertiary health unit at Fundação de Medicina Tropical Dr. Heitor Vieira Dourado in Western Brazilian Amazon **Malaria transmission:** study reported *P vivax* as "the most prevalent species in Brazil and extra-Afican areas", but did not specify to what degree. **Other patient characteristics:** 93.3% of the patients had up to three previous episodes of malaria.
Index tests	**RDT brand(s):** Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** reported manufacturer's instructions were followed (2ºC-30ºC until the expiry date) **Blinding:** The hospital laboratory staff who performed the RDT and thick blood smear analysis were blinded.
Target condition and reference standard(s)	**Target condition(s):** *P vivax* and *P falciparum* **Reference standard(s):** microscopy **Microscopy details:** • number of high powered fields not reported • Experienced microscopists examined the slides, however the study did not explicitly state the number of observers or repeat. **Blinding:** The hospital laboratory staff who performed the RDT and thick blood smear analysis were blinded.
Flow and timing	**Appropriate interval between index test and reference standard:** RDT and microscopy were performed on admission. **Invalid test results:** None reported
Comparative	
Notes	

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			

Costa 2019 *(Continued)*

Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Unclear		
Could the selection of patients have introduced bias?		Unclear risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Unclear		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Unclear risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Hailu 2014

Study characteristics

Hailu 2014 *(Continued)*

Patient Sampling	**Study design:** cross-sectional study **Recruitment:** did not state consecutive or random sampling **Study period:** patients recruited in December 2011 **Population:** 398 febrile patients who visited the outpatient department of a health centre **Inclusion and exclusion criteria:** Patients with acute febrile illnesses (body temperature > 37.5 degrees C) or a history of fever during the last 2 weeks at the date of data collection were included. Patients who took antimalarials within the last 2 weeks before the data collection date or refused participation were excluded.
Patient characteristics and setting	**Sex:** 44.2% male, 55.8% female **Age range:** 1 and 70 years **Setting:** Felegeselam Health Center in Pawe Special Woredam, Northwest Ethiopia **Malaria transmission:** malaria transmission takes place throughout the year and that *P falciparum* and *P vivax* are co-endemic.
Index tests	**RDT brand(s):** CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** RDT was stored based at room temperature and the quality of package and expiration date was checked before use. **Blinding:** The results of the RDT were determined earlier than microscopic results, with strict blinding to microscopic examination.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy **Microscopy details:** • number of high powered fields for microscopy not reported • Two experienced malaria technologists conducted the microscopic examination independently and blindly. • Results of their observation were recorded for later comparison and all discordant results were repeated and rechecked by the principal investigator who was also experienced. **Blinding:** RDT with strict blinding to microscopic examination.
Flow and timing	**Appropriate interval between index test and reference standard:** Blood sample was collected from each patient. Two out of 398 patients with invalid test results from RDT were retested by taking fresh blood. Unclear whether the same blood sample was used for microscopy for these patients. **Invalid test results:** Two patients with invalid test results.
Comparative	
Notes	
Methodological quality	

Hailu 2014 *(Continued)*

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
Could the selection of patients have introduced bias?		Unclear risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Low risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		

Hailu 2014 *(Continued)*

Were all patients included in the analysis?	Yes
Could the patient flow have introduced bias?	Low risk

Mekonnen 2010

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** did not state consecutive or random sampling **Study period:** October 2007 and December 2008 **Population:** 240 febrile patients who were clinically suspected of malaria and visited the outpatient department of a health cent er. **Inclusion and exclusion criteria:** not reported.
Patient characteristics and setting	**Sex:** 57.5% male, 42.5% female **Age:** mean = 25 years, range 1 and 60 years **Setting:** Serbo health cent er in Jimma zone, southern Ethiopia **Malaria transmission:** Study reported that *P falciparum* and *P vivax* were both prevalent.
Index tests	**RDT brand(s):** CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** manufacturer's instruction was followed and the quality of package desiccant was checked before use. **Blinding:** The RDT and microscopy were performed by three experienced malaria technicians independently. The results of the RDT were determined before microscopic results with strict blinding to microscopic examination.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy **Microscopy details:** • At least 300 high powered fields • Three experienced technicians examined the slides independently. Results of their observation were recorded for later comparison and discordant results between microscopy and RDT were repeated. **Blinding:** RDT with strict blinding to microscopic examination
Flow and timing	**Appropriate interval between index test and reference standard:** Blood sample was collected from each patient. **Invalid test results:** None reported.

Mekonnen 2010 *(Continued)*

Comparative
Notes

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Unclear		
Could the selection of patients have introduced bias?		Unclear risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Low risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		

Mekonnen 2010 *(Continued)*

Did all patients receive the same reference standard?	Yes	
Were all patients included in the analysis?	Yes	
Could the patient flow have introduced bias?		Low risk

Mendoza 2013

Study characteristics	
Patient Sampling	**Study design:** retrospective cross-sectional study **Recruitment:** consecutive **Study period:** November 16 to December 2, 2010 in Córdoba and from June 14 to 25, 2011 in Chocó, Colomba **Population:** 383 patients who attended one of three clinics with microscopy for diagnosis of malaria. **Inclusion and exclusion criteria:** patient was considered as a probable case of malaria and at least 6 years of age (with consent), were included. Probable case defined as patients presenting with current or recent fever within 72 hours, who came from an endemic area in the last 15 days and who may or may not have an epidemiological relationship with diagnosed cases. The study excluded patients who did not consent to participation, lack of diligence of the clinical epidemiological record or who presented with symptoms of complicated malaria.
Patient characteristics and setting	**Sex:** 52.5% male, 47.5% female **Age range:** 6 and 92 years **Setting:** 233 patients came from Córdoba, of which 121 were from Tierralta and 112 from Puerto Libertador. The remaining 150 patients were recruited in the department of Chocó, in the municipality of Quibdo. **Malaria transmission:** The study reported Córdoba had the highest prevalence of *P vivax*, unclear for Chocó. **Other patient characteristics:** 7.8% of the patients received treatment for malaria in the previous month of recruitment.
Index tests	**RDT brand(s):** SD Bioline Malaria Ag Pf/Pv test (Standard Diagnostics Inc) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** reported manufacturer's instructions were followed (1°C-40°C) **Blinding:** The results of the RDT were determined and kept separate so it does not interfere with the reference standard results.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy corrected by PCR **Microscopy details:** • Number of high powered fields not reported.

Mendoza 2013 *(Continued)*

	• Blood films were examined by two experienced readers independently and blindly. • Discordant results were checked by a third reader. **PCR details:** • Did not report who performed PCR. • Detection limit not reported. • When there were discordant results between microscopy and PCR, the result of the PCR was taken, except in those in which the thick drop showed parasitic forms and the PCR was negative. **Blinding:** The results of the RDT were determined and kept seperate so it does not interfere with the reference standard results.
Flow and timing	**Appropriate interval between index test and reference standard:** Multiple blood samples were taken at the same time for each patient. **Invalid test results:** None reported.
Comparative	
Notes	

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
Could the selection of patients have introduced bias?		Low risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	

Mendoza 2013 *(Continued)*

Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Low risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Mussa 2019

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** did not state consecutive or random sampling **Study period:** Not reported **Population:** 59 suspected patients with *P falciparum* infection from different clinical centers were recruited. **Inclusion and exclusion criteria:** not reported
Patient characteristics and setting	**Sex:** 45.8% male, 54.2% female **Age:** Not reported **Setting:** different clinics in Omdurman, Sudan **Malaria transmission:** Not reported

Mussa 2019 *(Continued)*

Index tests	**RDT brand(s):** Test Malaria Pf/Pv rapid test cassette (Alltest Biotech, China) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** Unclear, reported manufacturer's instructions were followed for use **Blinding:** not reported
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* **Reference standard(s):** PCR **PCR details:** • Did not report who performed PCR. • Did not report detection limit. **Blinding:** not reported
Flow and timing	**Appropriate interval between index test and reference standard:** one blood sample taken from each patient. **Invalid test results:** None reported.
Comparative	
Notes	Contacted author specifically for *P vivax* results.

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Unclear		
Could the selection of patients have introduced bias?		Unclear risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Unclear risk	

Mussa 2019 *(Continued)*

Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Unclear risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Saha 2017

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** consecutive **Study period:** Not reported **Population:** 200 febrile patients in whom clinicians suspected malaria and raised the investigations. **Inclusion and exclusion criteria:** Patients having a fever with chills and rigor in the absence of any obvious cause such as upper respiratory tract infection. All patients diagnosed and/or treated with antimalarial drugs within the past six months were excluded.
Patient characteristics and setting	**Sex:** 56.0% male, 44.0% female **Age:** mean = 34.6 years, <10 years: 2.5%, 11-20 years: 20.5%, 21-60 years: 68.5%, >61 years: 8.5% **Setting:** tertiary care hospital setting at the outpatient department of Kasturba Hospital, Sewagram, Wardha in Central India **Malaria transmission:** Not reported
Index tests	**RDT brand(s):** SD Bioline Malaria Ag Pf/Pv test (Standard Diagnostics Inc)

Saha 2017 *(Continued)*

	Batch number: not reported **Lot testing:** not reported **Storage conditions:** Unclear, although reported manufacturer's instructions were followed for use. **Blinding:** The RDT, microscopy and PCR were performed by different technicians and results of all three tests were kept blind.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** PCR and microscopy **Microscopy details:** • Number of high powered fields not reported • Two microscopists having >15years of experience independently examined the slides. • If there was discordance, this was resolved by a third reader (microbiologists). **PCR details:** • Different techinicans conducted the microscopy and PCR. • Limit detection not reported. **Blinding:** The RDT, microscopy and PCR were performed by different technicians and results of all three tests were kept blind.
Flow and timing	**Appropriate interval between index test and reference standard:** one blood sample taken from each patient. **Invalid test results:** None reported.
Comparative	
Notes	Contacted author specifically for *P vivax* results.

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
Could the selection of patients have introduced bias?		Low risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			

Saha 2017 *(Continued)*

Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Low risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Sharew 2009

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** did not state consecutive or random sampling **Study period:** November and December 2008 **Population:** 668 febrile patients who were clinically suspected of malaria and visited the outpatient department of two health centers. **Inclusion and exclusion criteria:** not reported.
Patient characteristics and setting	**Sex:** 54.0% male, 46.0% female **Age range:** 6 months and 75 years

Sharew 2009 *(Continued)*

	Setting: Bussa and Kella health centers in Wondo Genet area, southern Ethiopa **Malaria transmission:** Study reported that *P falciparum* and *P vivax* were both prevalent.
Index tests	**RDT brand(s):** CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** manufacturer's instruction was followed and the quality of package desiccant was checked before use. **Blinding:** The RDT and microscopy were performed by two experienced malaria technicians independently. The results of the RDT were determined before microscopic results with strict blinding to microscopic examination.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy **Microscopy details:** • At least 100 high powered fields • Two experienced technicians examined the slides independently, which was checked by the team leader who is also experienced. Discordant results between microscopy and RDT were repeated. **Blinding:** RDT with strict blinding to microscopic examination
Flow and timing	**Appropriate interval between index test and reference standard:** Blood sample was collected from each patient. **Invalid test results:** None reported.
Comparative	
Notes	

Methodological quality

Item	**Authors' judgement**	**Risk of bias**	**Applicability concerns**
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Unclear		
Could the selection of patients have introduced bias?		Unclear risk	

Sharew 2009 *(Continued)*

Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		Low risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Singh 2010

Study characteristics	
Patient Sampling	**Study design:** cross-sectional study **Recruitment:** consecutive **Study period:** August and December 2009 **Population:** 372 febrile patients with clinical suspicion of malaria who visited field clinics

Singh 2010 *(Continued)*

	Inclusion and exclusion criteria: excluded pregnant women and patients who took antimalarials.
Patient characteristics and setting	**Sex:** Not reported **Age:** mean = 15 years, SD: 14.1 **Setting:** Bajag Primary Health Centre (PHC) of district Dindori and Satanwada PHC of district Shivpuri, India **Malaria transmission:** study reported that both *P falciparum* and *P vivax* as co-endemic in the study area.
Index tests	**RDT brand(s):** Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa) **Batch number:** not reported **Lot testing:** not reported **Storage conditions:** "For testing temperature stability of the tests, RDTs were stored at 25°C on receipt in the study sites, then allocated to separate groups for storage at 35°C & 45°C for 90 days, at 60°C for 48 hours, and at -10°C for 60 minutes before testing [21]. At the start of the study, the incubators were stabilized at the required temperature for three days before the RDTs to be tested were placed inside. RDTs were removed from storage to reach room temperature for 2 hours before testing and comparisons were made with control RDTs kept at 25°C until use." **Blinding:** Microscopy examination was conducted without reference to the results of RDTs.
Target condition and reference standard(s)	**Target condition(s):** *P falciparum* and *P vivax* **Reference standard(s):** microscopy **Microscopy details:** • 100 high powered fields • Only one experienced microscopist conducted the examination in the laboratory. • Any discrepancies between the reference standard or index test were re-examined by another expert technician who was blinded to the results of microscopy and RDT. **Blinding:** microscopy conducted without reference to the results of RDTs.
Flow and timing	**Appropriate interval between index test and reference standard:** Multiple samples were taken at the same time. **Invalid test results:** None reported.
Comparative	
Notes	

Methodological quality

Item	Authors' judgement	Risk of bias	Applicability concerns
DOMAIN 1: Patient Selection			

Singh 2010 *(Continued)*

Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
Could the selection of patients have introduced bias?		Low risk	
Are there concerns that the included patients and setting do not match the review question?			Low concern
DOMAIN 2: Index Test (All tests)			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
Could the conduct or interpretation of the index test have introduced bias?		Low risk	
Are there concerns that the index test, its conduct, or interpretation differ from the review question?			Unclear
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	No		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
Could the reference standard, its conduct, or its interpretation have introduced bias?		High risk	
Are there concerns that the target condition as defined by the reference standard does not match the question?			Low concern
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
Could the patient flow have introduced bias?		Low risk	

Characteristics of excluded studies *[ordered by study ID]*

Study	Reason for exclusion
Abdelraheem 2016	Insufficient data
Adams 2015	Wrong patient population
Adnan 2017	Insufficient data
Ageep 2013	Wrong target condition
Andrade 2010	Wrong index test
Arvind 2015	Predatory journal
Ashton 2010	Wrong index test
Ayorinde 2016	Wrong target condition
Ba 2017	Wrong index test
Bahk 2018	Wrong index test
Barber 2013	Wrong index test
Bell 2001	Wrong index test
Bendezu 2010	Wrong index test
Berhane 2017	Wrong study design
Berzosa 2018	Wrong index test
Bharti 2008	Wrong index test
Bharti 2013	Wrong study design
Bhide 2014	Wrong index test
Birhanie 2016	Wrong index test
Bisoffi 2014	Wrong study design
Britton 2016	Wrong index test
Chayani 2004	Wrong index test
Cho 2016	Wrong index test
Dahesh 2015	Full text not available
Dash 2013	Insufficient data
Deida 2019	Wrong index test

Study	Reason for exclusion
DeKoninck 2017	Wrong patient population
Dev 2004	Wrong index test
Dinzouna-Boutamba 2014	Wrong index test
Dzakah 2014	Wrong patient population
Ehtesham 2015	Wrong patient population
Eibach 2013	Wrong index test
Elahi 2013	Wrong index test
Endeshaw 2012	Wrong index test
Falade 2016	Wrong target condition
Fernando 2004	Wrong index test
Fernando 2004a	Wrong index test
Foster 2014	Wrong study design
Fransisca 2015	Wrong index test
Gabrielli 2016	Wrong patient population
Ghai 2016	Wrong index test
Gupta 2018	Wrong index test
Harani 2006	Wrong index test
Hawash 2019	Wrong patient population
Jabeen 2016	Insufficient data
Jahan 2019	Wrong index test
Joseph 2018	Wrong index test
Karimov 2013	Full text not available
Kim 2013	Wrong index test
Kolaczinski 2004	Wrong index test
Kosack 2013	Wrong index test
Kumari 2014	Predatory journal
Liu 2013	Full text not available
Mallepaddi 2019	Wrong index test

Study	Reason for exclusion
Metzger 2011	Wrong index test
Moges 2012	Wrong index test
Mohon 2012	Wrong index test
Olasehinde 2018	Wrong index test
Pakalapati 2013	Wrong index test
Pattanasin 2003	Wrong index test
Puri 2013	Wrong index test
Rakotonirina 2008	Wrong index test
Ranjan 2016	Insufficient data
Ratsimbasoa 2007	Wrong index test
Ratsimbasoa 2008	Wrong index test
Samane 2010	Wrong index test
Selimuzzaman 2010	Wrong index test
Shaikh 2013	Insufficient data
Sharma 2014	Wrong study design
Singh 2000a	Wrong index test
Singh 2003	Wrong index test
Singh 2013	Wrong study design
Siwal 2018	Wrong patient population
Stijnberg 2013	Conference abstract
Strom 2014	Wrong target condition
Thongdee 2014	Wrong index test
Tjitra 1999	Wrong index test
Trouvay 2013	Wrong index test
Valecha 2003	Wrong index test
van den Broek 2006	Wrong index test
Vohra 2014	Insufficient data
Wang 2014	Full text not available

Study	Reason for exclusion
Wongsrichanalai 2003	Wrong index test
Woyessa 2013	Wrong study design
Xiaodong 2013	Wrong index test
Yan 2013	Wrong index test

Characteristics of studies awaiting classification *[ordered by study ID]*

Boni 2015

Patient Sampling	Patients were recruited from two provinces in Central Vietnam between January and August 2015. The sampling method and inclusion/exclusion criteria were not reported in the abstract.
Patient characteristics and setting	The prevalence, number of patients recruited and characteristics of patients were not reported in the abstract.
Index tests	RDT brand was not reported in abstract. No information on blinding, batch number of RDT, lot testing or storage conditions in the abstract.
Target condition and reference standard(s)	The target conditions were *P falciparum* and *P vivax*. The reference standard was microscopy examined by at least two expert microscopists. No information on number of high powered field or blinding in the abstract.
Flow and timing	Unclear whether the index test and reference standard were performed at the same time and if blood sample was taken at the same time for the tests.
Comparative	
Notes	Unable to deduce the number of true positives, false positives, false negatives and true negatives for *P vivax*. Contacted authors for more details on methodology and results.

Cheng 2013

Patient Sampling	This was a cross-sectional study, patients were recruited in 2008 and in 2011. The study did not explicitly state consecutive or random sampling. Inclusion and exclusion criteria were not reported. Unclear how the febrile patients were recruited, i.e. whether they presented themselves to a health centre.
Patient characteristics and setting	202 febrile patients (49 patients in 2008 and 153 in 2011) with fever of unknown origin in Kachine Myanmar and in Yunnan, China were recruited. 13 healthy patients were also recruited in Beijing, China, however they were not used in the analysis of the RDT. The study did not describe the characteristics of patients recruited. The study reported malaria as endemic in study area.
Index tests	CareStart Malaria HRP2/pLDH combo test (Access Bio Inc., Somerset, NJ). In the study there was no information on batch number, lot testing or storage conditions, however the study stated that the RDT was done according to manufacturer's protocol. The study did not mention blinding.
Target condition and reference standard(s)	The target conditions were *P falciparum* and *P vivax*. The reference standards were PCR and microscopy. The study reported a minimum of 100 high powered

Cheng 2013 *(Continued)*

	fields for microscopy. Two professional microscopists conducted the microscopic examination independently. Unclear how discrepancies for microscopy results between the two microscopists were handled, if any. With PCR, unable to deduce the number of true positives, false positives, false negatives and true negatives.
Flow and timing	The tests were performed at different times, but the blood sample was taken at the same time. All samples were frozen and stored at -80 ºC but 2008 samples suffered freeze–and–thaw cycles. Only the 2011 samples were tested using the RDT but this was stated as 143 patients in the results rather than 153.
Comparative	
Notes	Unclear whether the RDT used by the study is eligible for this review because the specific brand name was not mentioned in the main study publication, contacted authors for further information.

Reda 2016

Patient Sampling	This was a cross-sectional study. Patients were recruited between November and December 2014 in two health centres in Adam and Amaya, Oromia region, Ethiopia. Inclusion/ exclusion criteria or sampling method were not reported.
Patient characteristics and setting	Febrile patients with symptoms of malaria who visited the two health facilities were recruited. The study abstract did not include the prevalence of malaria or the characteristics of patients recruited. "A total of 547 febrile patients were diagnosed, of which 127 were microscopy positive for Pf (n=38) and Pv (n=85)".
Index tests	CareStart Malaria Ag Pf/Pv combo test (Access Bio Inc., Somerset, NJ) and SD BIOLINE malaria AG PF/PV test. The RDTs were performed following manufacturer's instructions. In the study abstract, there was no information on batch number or lot testing. The study abstract did not mention blinding.
Target condition and reference standard(s)	The target conditions were *P falciparum* and *P vivax*. The reference standard was microscopy. Microscopic examination was done under 100x magnifications. No information on blinding or who performed the microscopic examination in the abstract.
Flow and timing	Unclear whether the index test and reference standard were performed at the same time and if blood sample was taken at the same time for the tests.
Comparative	
Notes	Unable to deduce the number of true positives, false positives, false negatives and true negatives for *P vivax*. Contacted authors for more details on methodology and results.

DATA

Presented below are all the data for all of the tests entered into the review.

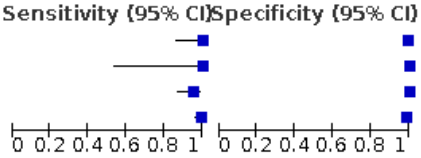
Table Tests. Data tables by test

Test	No. of studies	No. of participants
1 CareStart Malaria Pf/Pv Combo test (Access Bio Inc, New Jersey, USA) (Microscopy)	4	2398
2 Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (Microscopy)	2	710
3 Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil) (Microscopy)	1	181
4 OnSite Pf/Pv test (CTK Biotech Inc, USA) (Microscopy)	1	338
5 SD Bioline Malaria Ag Pf/Pv test (Microscopy)	1	200
6 Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (PCR)	1	338
7 OnSite Pf/Pv test (CTK Biotech Inc, USA) (PCR)	1	338
8 SD Bioline Malaria Ag Pf/Pv test (PCR)	1	200
9 Test Malaria Pf/Pv rapid test (Alltest Biotech, China) (PCR)	1	59
10 SD Bioline Malaria Ag Pf/Pv test (Microscopy corrected by PCR)	1	383

Test 1. CareStart Malaria Pf/Pv Combo test (Access Bio Inc, New Jersey, USA) (Microscopy)

CareStart Malaria Pf/Pv Combo test (Access Bio Inc, New Jersey, USA) (Microscopy)

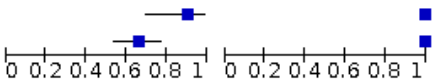
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Chanie 2011	25	6	0	1061	1.00 [0.86, 1.00]	0.99 [0.99, 1.00]		
Hailu 2014	6	0	0	392	1.00 [0.54, 1.00]	1.00 [0.99, 1.00]		
Mekonnen 2010	61	0	3	176	0.95 [0.87, 0.99]	1.00 [0.98, 1.00]		
Sharew 2009	155	9	1	503	0.99 [0.96, 1.00]	0.98 [0.97, 0.99]		



Test 2. Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (Microscopy)

Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (Microscopy)

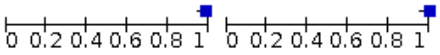
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Alam 2011	19	1	2	316	0.90 [0.70, 0.99]	1.00 [0.98, 1.00]		
Singh 2010	45	3	23	301	0.66 [0.54, 0.77]	0.99 [0.97, 1.00]		



Test 3. Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil) (Microscopy)

Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil) (Microscopy)

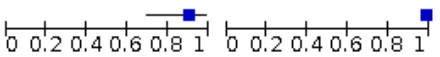
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Costa 2019	94	0	1	86	0.99 [0.94, 1.00]	1.00 [0.96, 1.00]		



Test 4. OnSite Pf/Pv test (CTK Biotech Inc, USA) (Microscopy)

OnSite Pf/Pv test (CTK Biotech Inc, USA) (Microscopy)

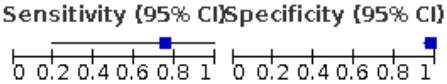
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Alam 2011	19	4	2	313	0.90 [0.70, 0.99]	0.99 [0.97, 1.00]		



Test 5. SD Bioline Malaria Ag Pf/Pv test (Microscopy)

SD Bioline Malaria Ag Pf/Pv test (Microscopy)

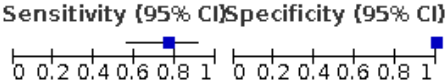
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Saha 2017	3	4	1	192	0.75 [0.19, 0.99]	0.98 [0.95, 0.99]		



Test 6. Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (PCR)

Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals Goa) (PCR)

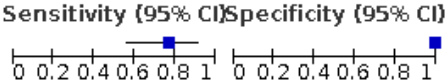
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Alam 2011	20	0	6	312	0.77 [0.56, 0.91]	1.00 [0.99, 1.00]		



Test 7. OnSite Pf/Pv test (CTK Biotech Inc, USA) (PCR)

OnSite Pf/Pv test (CTK Biotech Inc, USA) (PCR)

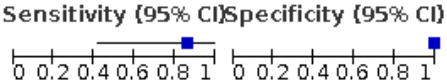
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Alam 2011	20	3	6	309	0.77 [0.56, 0.91]	0.99 [0.97, 1.00]		



Test 8. SD Bioline Malaria Ag Pf/Pv test (PCR)

SD Bioline Malaria Ag Pf/Pv test (PCR)

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Saha 2017	6	1	1	192	0.86 [0.42, 1.00]	0.99 [0.97, 1.00]		



Test 9. Test Malaria Pf/Pv rapid test (Alltest Biotech, China) (PCR)

Test Malaria Pf/Pv rapid test (Alltest Biotech, China) (PCR)

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Mussa 2019	0	4	0	55	Not estimable	0.93 [0.84, 0.98]	0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

Test 10. SD Bioline Malaria Ag Pf/Pv test (Microscopy corrected by PCR)

SD Bioline Malaria Ag Pf/Pv test (Microscopy corrected by PCR)

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Mendoza 2013	73	0	6	304	0.92 [0.84, 0.97]	1.00 [0.99, 1.00]	0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

ADDITIONAL TABLES

Table 1. Summary of key study characteristics

Study	Country	Sample size	Sex	Age	RDT brand	Personnel performing RDT	Storage conditions of RDT	Reference standard	Personnel performing reference standard	Parasite density of positive cases
Alam 2011	Bangladesh	338	49.7% male 50.3% female	Median (range): 14 years (18 months to 82 years)	OnSite Pf/Pv test (CTK Biotech Inc, USA) Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa)	An experienced medical technologist	Unclear, although study stated that the instructions of the manufacturers were followed.	PCR and microscopy (separately)	Slides assessed by two independent microscopists	Of 21 *P vivax* positive slides, parasite count ranged from 32 to 25,120 parasites/µL of blood, with a median of 5,040 (IQR 520 to 17,160) parasites/µL blood.
Chanie 2011	Ethiopia	1092	51.4% male 48.6% female	Mean (SD): 22 (12.8) years	CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ)	Experienced malaria technicians	Kept at the local temperature of the region without any controlling system of the storage temperature during data collection	Microscopy	Experienced technicians examined the slides	Not reported
Costa 2019	Brazil	181	64.1% male 35.9% female	Mean (SD): 41.7 (14.4) years	Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil)	Hospital laboratory staff	According to manufacturer's instructions (2°C to 30°C until the expiration date)	Microscopy	Experienced microscopists then examined the slides	Mean parasitaemia detected by TBS for *P vivax* malaria was 1,206.5 parasites/mm^3 blood
Hailu 2014	Ethiopia	398	44.2% male 55.8% female	Range: 1 to 70 years	CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ)	Not reported	Stored at room temperature according to manufacturer's instructions	Microscopy	Two experienced malaria technologists per-	Not reported

Table 1. Summary of key study characteristics *(Continued)*

									formed the microscopy	
Mekonnen 2010	Ethiopia	240	57.5% male 42.5% female	Mean (range): 25 years (1 to 60 years)	CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ)	Experienced malaria technicians	According to manufacturer's instructions	Microscopy	Three experienced technicians examined the slides	Not reported
Mendoza 2013	Colombia	383	52.5% male 47.5% female	Range: 6 to 92 years	SD Bioline Malaria Ag Pf/Pv test (Standard Diagnostics Inc)	Conducted by a trained person	According to manufacturer's recommendations (1°C to 40°C)	Microscopy corrected with PCR	Blood films were examined by two experienced readers	Parasitemia for *P vivax* ranged from 40 to 40,000 parasites/µL
Mussa 2019	Sudan	59	45.8% male 54.2% female	Not reported	Test Malaria Pf/Pv rapid test (Alltest Biotech, China)	Not reported	Unclear, although study stated that instructions of the manufacturer were followed	PCR	Not reported	Not reported
Saha 2017	India	200	56.0% male 44.0% female	Mean: 34.6 years 11 to 20 years: 20.5% 21 to 60 years: 68.5% <10 years: 2.5% > 61 years: 8.5%	SD Bioline Malaria Ag Pf/Pv test (Standard Diagnostics Inc)	Microscopy, RDT and PCR done by different technicians	Unclear, although study stated that instructions of the manufacturer were followed	PCR and Microscopy (separately)	Blood films were examined by two microscopists having >15 years of experience	Not reported
Sharew 2009	Ethiopia	668	54.0% males 46.0% females	Range: 6 months to 75 years	CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ)	Experienced malaria technicians	Stored according to manufacturer's instructions	Microscopy	Thick and thin smears determined by two experienced malaria technicians	Not reported

Table 1. Summary of key study characteristics *(Continued)*

Singh 2010	India	372	Not reported	Mean (SD): 15 (14.1) years	Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa)	Two research assistants	Detailed storage information provided	Microscopy	Blood films examined by an experienced microscopist	Not reported

PCR = polymerase chain reaction; RDT = rapid diagnostic test; SD = standard deviation; TBS =thick blood smear

Table 2. Comparison of microscopy and PCR reference standards for P vivax

RDT brand	Microscopy				PCR				Microscopy corrected with PCR			
	Number of studies	Number of participants (*P vivax* malaria cases)	Sensitivity (95% CI) (%)	Specificity (95% CI) (%)	Number of studies	Number of participants (*P vivax* malaria cases)	Sensitivity (95% CI) (%)	Specificity (95% CI) (%)	Number of studies	Number of participants (*P vivax* malaria cases)	Sensitivity (95% CI) (%)	Specificity (95% CI) (%)
CareStart Malaria Pf/Pv Combo test (Access Bio Inc, Somerset, NJ)	4	2398 (251)	99% (94% to 100%)	99% (99% to 100%)	0	-	-	-	0	-	-	-
Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa)	2	710 (89)	77% (53% to 91%)	99% (98% to 100%)	1	338 (26)	77% (56% to 91%)	100% (99% to 100%)	0	-	-	-
Immuno-Rapid Malaria Pf/Pv test (Wama Diagnostica, Sao Paulo, Brazil)	1	181 (95)	99% (94% to 100%)	100% (96% to 100%)	0	-	-	-	0	-	-	-
SD Bioline Malaria Ag Pf/Pv test (Standard Diagnostics Inc)	1	200 (4)	75% (19% to 99%)	98% (95% to 99%)	1	200 (7)	86% (42% to 100%)	99% (97% to 100%)	1	383 (79)	92% (84% to 97%)	100% (99% to 100%)
OnSite Pf/Pv test (CTK Biotech Inc, USA)	1	338 (21)	90% (70% to 99%)	99% (97% to 100%)	1	338 (26)	77% (56% to 91%)	99% (97% to 100%)	0	-	-	-

Table 2. Comparison of microscopy and PCR reference standards for P vivax *(Continued)*

Test Malaria Pf/Pv rapid test (Alltest Biotech, China)	0	-	-	-	1	59 (0)	Not estimable	93% (84% to 98%)	0	-	-	-

PCR = polymerase chain reaction; RDT = rapid diagnostic test.

Table 3. Direct comparisons between OnSite Pf/Pv test and Falcivax Device Rapid test

Study	Reference standard	Sensitivity (true positives/malaria cases) (%)		Difference (95% CI) (percentage points)	P value	Specificity (true negatives/non-cases) (%)		Difference (95% CI) (percentage points)	P value
		OnSite Pf/Pv test (CTK Biotech Inc, USA)	Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa)			OnSite Pf/Pv test (CTK Biotech Inc, USA)	Falcivax Device Rapid test for malaria Pv/Pf (Zephyer Biomedicals, Goa)		
Alam 2011	Microscopy	90 (19/21)	90 (19/21)	0 (-17.8 to 17.8)	P = 1.00	99 (313/317)	100 (316/317)	0.9 (-0.4 to 2.3)	P = 0.18
Alam 2011	PCR	77 (20/26)	77 (20/26)	0 (-22.9 to 22.9)	P = 1.00	99 (309/312)	100 (312/312)	1.0 (-0.1 to 2.0)	P = 0.08

PCR = polymerase chain reaction.

APPENDICES

Appendix 1. Detailed search strategy

Search set	MEDLINE (PubMed)
1	Malaria, vivax [MeSH]
2	Plasmodium vivax [MeSH]
3	“Plasmodium vivax” or “P vivax” or “vivax malaria” or “non-falciparum Malaria” Field: Title/Abstract
4	1 or 2 or 3
5	Exp Reagent kits, diagnostics [MeSH]
6	"Diagnostic Tests, Routine"[Mesh]
7	rapid diagnostic test* Field: Title/Abstract
8	RDT* Field: Title/Abstract
9	Dipstick* Field: Title/Abstract
10	“Rapid diagnostic device*” Field: Title/Abstract
11	MRDD Field: Title/Abstract
12	OptiMal Field: Title/Abstract
13	“Binax NOW” or “NOW-ICT-Malaria” or “NOW-Malaria-ICT” Field: Title/Abstract
14	ParaSight or Parascreen or ParaHIT Field: Title/Abstract
15	“SD Bioline” or Carestart or Falcivax or Malascan Field: Title/Abstract
16	Immunochromatograph* or Immuno-chromatograph* Field: Title/Abstract
17	“Antigen detection” Field: Title/Abstract
18	“Rapid malaria antigen test*” Field: Title/Abstract
19	“Combo card test*” Field: Title/Abstract
20	Immunoassay [MeSH]
21	Chromatography [MeSH]
22	Enzyme-linked immunosorbent assay [MeSH]
23	“Rapid test*” Field: Title/Abstract
24	“Card test*” Field: Title/Abstract

(Continued)

25	Rapid AND (detection* or device* or test* or kit*) Field: Title/Abstract
26	5 or 6 or 7 or 8 or 9 or 10 or 11 or 12 or 13 or 14 or 15 or 16 or 17 or 18 or 19 or 20 or 21 or 22 or 23 or 24 or 25
27	4 and 26

Web of Science

Search set	Web of Science
# 6	#5 AND #1 Indexes=SCI-EXPANDED, CPCI-S Timespan=All years
# 5	#4 OR #3 OR #2 Indexes=SCI-EXPANDED, CPCI-S Timespan=All years
# 4	**TOPIC:** (("alere trueline" or "Rapigen biocredit" or "SD bioline" or "standard Q" or VISITECT* or PA-LUTOP*)) *OR* **TOPIC:** (((necviparum or "one step" or meriscreen or "onsite malaria" or paraHIt* or Quickprofile))) Indexes=SCI-EXPANDED, CPCI-S Timespan=All years
# 3	**TOPIC:** (("ADVANCED QUALITY ONE STEP" or Tri-line or BIOCREDIT or Biosynex or BioTracer or Carestart or Aspenmal)) *OR* **TOPIC:** (("combo RDT" or careUS or Coretests* or EGENS or EzDx or Falcivax or "first response" or Humasis or Karwa or KHB* or "malaria Pf (HRPII)/ PV")) Indexes=SCI-EXPANDED, CPCI-S Timespan=All years
# 2	**TOPIC:** ((("rapid diagnostic test*" or RDT* or dipstick or MRDD) OR (“Binax NOW” or “NOW-ICT-Malaria” or “NOW-Malaria-ICT”))) *OR* **TOPIC:** (((ParaSight or Parascreen or ParaHIT or “SD Bioline” or Carestart or Falcivax or Malascan))) *OR* **TOPIC:** (: ((Immunochromatograph* or Immuno-chromatograph* or "card test" or chromatography))) Indexes=SCI-EXPANDED, CPCI-S Timespan=All years
# 1	**TOPIC:** (("plasmodium vivax" or "vivax malaria")) Indexes=SCI-EXPANDED, CPCI-S Timespan=All years

Database: Embase (OVID)

Search Strategy:

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1 malaria vivax.mp. or Plasmodium vivax malaria/

2 plasmodium vivax.mp. or Plasmodium vivax/

3 ("P vivax" or "non-falciparum Malaria").ab. or ("P vivax" or "non-falciparum Malaria").ti.

4 1 or 2 or 3

5 diagnostic procedure/

6 "rapid diagnos$ test$ ".ab. or "rapid diagnos$ test$".ti.

7 RDT$.ab. or RDT$.ti.

8 Dipstick$.ab. or Dipstick$.ti.

9 "Rapid diagnos$ device$ ".ab. or "Rapid diagnos$ device$ ".ti.

10 MRDD.ab. or MRDD.ti.

11 ("Binax NOW" or "NOW-ICT-Malaria" or "NOW-Malaria-ICT").ab. or ("Binax NOW" or "NOW-ICT-Malaria" or "NOW-Malaria-ICT").ti.

12 (ParaSight or Parascreen or ParaHIT).ab. or (ParaSight or Parascreen or ParaHIT).ti.

13 ("SD Bioline" or Carestart or Falcivax or Malascan).ab. or ("SD Bioline" or Carestart or Falcivax or Malascan).ti.

14 ("ADVANCED QUALITY ONE STEP" or Tri-line or BIOCREDIT or Biosynex or BioTracer or Carestart or Aspenmal).mp.

15 ("combo RDT" or careUS or Coretests* or EGENS or EzDx or Falcivax or "first response" or Humasis or Karwa or KHB* or "malaria Pf (HRPII)/ PV").mp.

16 (necviparum or "one step" or meriscreen or "onsite malaria" or paraHIt* or Quickprofile).mp.

17 ("alere trueline" or "Rapigen biocredit" or "SD bioline" or "standard Q" or VISITECT* or PALUTOP*).mp.

18 5 or 6 or 7 or 8 or 9 or 10 or 11 or 12 or 13 or 14 or 15 or 16 or 17

19 4 and 18

Search Name: Cochrane Central Register of Controlled Trials

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ID Search Hits

#1 vivax malaria

#2 MeSH descriptor: [Malaria, Vivax] explode all trees

#3 MeSH descriptor: [Plasmodium vivax] explode all trees

#4 #1 or #2 or #3

#5 rapid diagnostic test*

#6 RDT*

#7 “ADVANCED QUALITY™ ONE STEP” or Tri-line or “Aspen® Mal” or BIOCREDIT or Biosynex or BioTracer or Carestart or “combo RDT” or careUS or Coretests* or EGENS or EzDx™ or Falcivax or “first response”

#8 Humasis or Karwa or KHB* or necviparum or “one step” or meriscreen or “onsite malaria” or “paraHIt*” or Quickprofile or “alere trueline” or “Rapigen biocredit” or “SD bioline” or “standard Q” or VISITECT* or PALUTOP*

#9 “Binax NOW” or “NOW-ICT-Malaria” or “NOW-Malaria-ICT”

#10 ParaSight or Parascreen or ParaHIT

#11 “SD Bioline” or Carestart or Falcivax or Malascan

#12 Immunochromatography or Immuno-chromatography

#13 antigen detection

#14 combo card

#15 immunoassay or chromatography

#16 Enzyme-linked immunosorbent assay

#17 #5 or #6 or #7 or #8 or #9 or #10 or #11 or #12 or #13 or #14 or #15 or #16

#18 #17 and #4

Database :	LILACS
Search on :	vivax malaria [Words] and "rapid test$" or PCR or diagnosis [Words]

Appendix 2. QUADAS-2 tool tailored to the context of the review

Domain	Patient selection	Index test	Reference standard	Flow and timing
Description	**Methods of patient selection**	**How index test was conducted and reported**	**How reference standard was conducted and reported**	**Describe patients that did not receive and time interval between index test or reference standard**
Signalling questions (yes, no, or unclear)	Consecutive or random sample of patients? • 'Yes' if the study reported consecutive enrolment or random sampling of patients presenting with uncomplicated malaria symptoms. • 'No' if patients were purposefully selected, for example based on previous test results (such as using Rapid diagnostic tests (RDTs) only on those who tested positive for *P vivax* by microscopy/PCR). • 'Unclear' if the study did not explicitly state consecutive enrolment or random sampling, and it was unclear how patients were sampled.	Index test results interpreted without knowledge of the results of reference standard? • 'Yes' if RDT was performed fully blinded to reference standard result. • 'No' if reference standard result was known prior to interpretation of RDT result. • 'Unclear' if blinding was no explicitly stated.	**PCR** PCR likely to correctly classify the target condition? We will answer this question as 'yes' for all studies because PCR is an objective test with binary outcomes. Thus, there is no room for subjective interpretation of test results or poor performance of the test leading to false negatives or false positives. 'Yes' if reference standard was PCR. **Microscopy** Microscopy likely to correctly classify the target condition? 'Yes' if microscopy was performed for one sample by two independent trained microscopist examining 100 high-power fields. 'No' if microscopy was performed: • by insufficiently trained individuals; • by one individual only; • with inadequate equipment;	Was there an appropriate interval between index test and reference standard? • 'Yes' if samples for RDT and microscopy or PCR were taken at the same time. We felt this was important given the transient parasitaemia associated with malaria. • 'No' if the samples for RDT and microscopy or PCR were taken at different times. • 'Unclear' if insufficient or no information on the time interval.

(Continued)

- by viewing less than 100 microscopic fields before declaring negative.

'Unclear' if insufficient information was provided.

Was a case-control design avoided? This will always be 'yes' because case control studies will be excluded from this review.	Pre-specified threshold used? As the threshold is prespecified by the manufacturer in all RDTs, we will answer this question 'yes' for all studies.	Reference standard results interpreted without knowledge of the results of index test? We will answer this question 'yes' for all studies using only PCR as the reference standard because PCR is an objective test with binary outcomes. Thus, there is no room for subjective interpretation of test results. • 'Yes' if results of microscopy were interpreted without knowledge of RDT results • 'No' if results of microscopy were interpreted with knowledge of RDT results • 'Unclear' if there is insufficient information on whether or not microscopy results were interpreted with knowledge of RDT results	Did all patients receive a reference standard? • 'Yes' if all participants received a microscopy or PCR. • 'No' if one or more participants did not receive microscopy or PCR. Or if the reference standard was applied depending on index test results • 'Unclear' if there is insufficient information to determine whether or not all patients received microscopy/PCR.
Did the study avoid inappropriate exclusions? • 'Yes' if no patients were excluded after inclusion in the study or if exclusions are adequately described. • 'No' if specific populations were excluded (for example, pregnant patients, children or immunocompromised patients), • 'Unclear' if unreported or insufficient information given to make a decision.			Did all patients receive the same reference standard? • We will answer this question 'yes' if all participants in the study or a subset of participants in the study received the acceptable reference standard (microscopy, PCR, or both), which we specified as a criterion for inclusion in the review. • 'No' if participants did not receive the same reference standard. • 'Unclear' if there is insufficient information to determine whether or not all patients received the same reference standard.
			Were all patients included in the analysis? • 'Yes' if the number of participants in the two-by-two table matches the number of participants recruited into the study or if sufficient explanation was provided for any discrepancy. • 'No' if some participants recruited into the study were unaccounted for.

(Continued)

				• 'Unclear' if unreported or insufficient information given to make a decision.
Risk of bias (high, low, or unclear)	Could the selection of patients have introduced bias?	Could the conduct or interpretation of the index test have introduced bias?	Could the reference standard, its conduct, or its interpretation has introduced bias?	Could the patient flow have introduced bias?
Applicability concerns (high, low, or unclear)	Not applicable	Are there concerns that the index test, its conduct, or interpretation differs from the review question? • 'High' if the study describes inappropriate storage conditions for the index test, or if the index test has not been lot tested • 'Low' if the study describes suitable storage conditions for the index test that meet manufacturer's requirements and if the study has reported the index test has been lot tested • 'Unclear' if insufficient information to make a decision	Are there concerns that the target condition as defined by the reference standard does not match the review question? We will answer this question 'low' for all studies because *P vivax* diagnosed by light microscopy or PCR does match the review question	Not applicable

HISTORY

Protocol first published: Issue 2, 2019
Review first published: Issue 11, 2020

CONTRIBUTIONS OF AUTHORS

RA, LC, and SJ screened the searches and assessed studies for inclusion, extracted data and performed methodological quality assessment. RA conducted the analyses and drafted the review under supervision by YT. LC and YT provided content expertise. YT critically revised the draft. All review authors read and approved the final draft of the review.

DECLARATIONS OF INTEREST

RA has no known conflicts of interest.
LC has no known conflicts of interest.
SJ has no known conflicts of interest.
YT has no known conflicts of interest.

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DIFFERENCES BETWEEN PROTOCOL AND REVIEW

In the protocol, the stated secondary objectives were to assess (1) the effect of transmission setting (perennial, seasonal, or epidemic) and type of malaria present in the region on the accuracy of RDTs for detecting *P vivax* malaria parasitaemia; (2) the effect of different generations of an RDT on test accuracy; and (3) the impact of level of training for studies that used microscopy as the reference standard. However, we were unable to conduct comparative meta-analyses, investigations of heterogeneity, and sensitivity analyses due to the limited number of included studies.

We only assessed the certainty of the evidence using GRADE methods where there were sufficient studies for meta-analyses.

INDEX TERMS

Medical Subject Headings (MeSH)

Ambulatory Care [statistics & numerical data]; Antigens, Protozoan [blood]; Bias; *Endemic Diseases; False Negative Reactions; False Positive Reactions; Malaria, Vivax [blood] [*diagnosis] [epidemiology]; Microscopy [standards]; Plasmodium vivax [immunology]; Point-of-Care Testing [standards]; Polymerase Chain Reaction [standards]; *Reagent Kits, Diagnostic [statistics & numerical data]; Reference Standards; Sensitivity and Specificity; Species Specificity

MeSH check words

Humans