



Genome Sequences of Serotype A6 *Mannheimia haemolytica* Isolates D174 and D38 Recovered from Bovine Pneumonia

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Here, we report two genomes, one complete and one draft, from virulent bovine strains of *Mannheimia haemolytica* serotype A6 recovered prior to the field usage of modern antimicrobial drugs.

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annheimia haemolytica is a facultative respiratory pathogen of ruminants. Among cattle, serotypes A1, A2, and A6 colonize and are commonly recovered from the nasopharynx (1). In pneumonic disease, it is predominantly serotypes A1 and A6 that are recovered from diseased lung tissues (2). Resistance to nasopharyngeal colonization can be elicited with vaccine products, and such resistance has been shown to be serotype specific (3). Although the serotype of *M. haemolytica* is based on capsule type (2), it is currently unknown whether acquired resistance to nasopharyngeal colonization is based upon anticapsular or other bacterial components. Antimicrobial resistance among bacterial bovine respiratory disease pathogens is of growing concern (4, 5), and multidrug-resistant isolates of Pasteurella multocida and M. haemolytica were recently sequenced (6, 7). Isolates D174 and D38 were recovered from a pneumonic calf lung in January 1984 and December 1982, respectively. The genome sequencing of these strains was undertaken to further our understanding of the basis of acquired resistance to nasopharyngeal colonization and provide insight into the acquisition of antimicrobial resistance.

The genome sequencing of M. haemolytica strain D174 was achieved using 3 platforms: the Roche (454) GS FLX titanium, resulting in 25-fold coverage; Illumina GA IIx, resulting in 1,600fold coverage; and PacBio RS, resulting in 21-fold coverage. The Illumina reads were used to error correct the PacBio reads using CLC Genomics Workbench version 6.0.2. A hybrid assembly using the CLC software was performed, and the resultant contigs were aligned to an optical map (MapSolver software; OpGen, Gaithersburg, MD) to confirm the assembly and generate a single scaffold. Reiterative alignments of the 454 and corrected PacBio reads >1 kb against the scaffold, using the CLC software, closed all gaps and resulted in a single circular chromosome. The completed D174 genome consists of 2.70 Mb, with a G+C content of 41.1%. The draft genome of *M. haemolytica* D38 was determined using the Roche platform alone, which yielded 24-fold coverage. Assembly against the closed D174 reference genome using the CLC software yielded 97 contigs with a total of 2.61 Mb, a G+C content of 41.0%, an N_{50} of 48,311 bp, and 100% contigs >500 bp.

The annotation of both genomes was accomplished with the

NCBI Prokaryotic Genome Annotation Pipeline revision 2.1. Strain D174 contains a total of 2,814 genes, including 2,687 predicted protein-coding genes, 40 frameshifted pseudogenes, 19 rRNA genes, and 66 tRNA genes. Strain D38 contains a total of 2,710 genes, including 2,606 predicted protein-coding genes, 48 frameshifted pseudogenes, 6 rRNA genes, and 50 tRNA genes. One clustered regularly interspaced short palindromic repeat (CRISPR) array was detected in each isolate. In contrast to the multiresistant *M. haemolytica* isolate 42548 (6), genes *aphA1*, *strA*, *strB*, *sul2*, *tetR*, and *tetH* are absent in both strains D174 and D38. The determination of genes potentially involved serotype-specific resistance to nasopharyngeal colonization will require additional analysis.

Nucleotide sequence accession numbers. The genome sequence of *M. haemolytica* strain D174 has been deposited in Gen-Bank under the accession no. CP006574. The genome sequence of *M. haemolytica* D38 has been deposited in GenBank under the accession no. AUNL00000000. The version described in this paper is version AUNL01000000.

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REFERENCES

- 1. Singh K, Ritchey JW, Confer AW. 2011. *Mannheimia haemolytica*: bacterial-host interactions in bovine pneumonia. Vet Pathol **48**:338–348. http://dx.doi.org/10.1177/0300985810377182.
- Al-Ghamdi GM, Ames TR, Baker JC, Walker R, Chase CC, Frank GH, Maheswaran SK. 2000. Serotyping of *Mannheimia (Pasteurella) haemolytica* isolates from the upper Midwest United States. J Vet Diagn Invest 12:576–578. http://dx.doi.org/10.1177/104063870001200617.
- Frank GH, Briggs RE, Loan RW, Purdy CW, Zehr ES. 1994. Serotypespecific inhibition of colonization of the tonsils and nasopharynx of calves after *Pasteurella haemolytica* serotype A1 after vaccination with the organism. Am J Vet Res 55:1107–1110.

- Watts JL, Sweeney MT. 2010. Antimicrobial resistance in bovine respiratory disease pathogens: measures, trends, and impact on efficacy. Vet Clin North Am Food Anim Pract 26:79–88. http://dx.doi.org/10.1016/ j.cvfa.2009.10.009.
- Katsuda K, Kohmoto M, Mikami O. 2013. Relationship between serotype and the antimicrobial susceptibility of *Mannheimia haemolytica* isolates collected between 1991 and 2010. Res Vet Sci 94:205–208. http:// dx.doi.org/10.1016/j.rvsc.2012.09.015.
- 6. Eidam C, Poehlein A, Brenner Michael G, Kadlec K, Liesegang H,

Brzuszkiewicz E, Daniel R, Sweeney MT, Murray RW, Watts JL, Schwarz S. 2013. Complete genome sequence of *Mannheimia haemolytica* strain 42548 from a case of bovine respiratory disease. Genome Announc 1(3): e00318-13. http://dx.doi.org/10.1128/genomeA.00318-13.

 Michael GB, Kadlec K, Sweeney MT, Brzuszkiewicz E, Liesegang H, Daniel R, Murray RW, Watts JL, Schwarz S. 2012. ICEPmu1, an integrative conjugative element (ICE) of *Pasteurella multocida*: analysis of the regions that comprise 12 antimicrobial resistance genes. J Antimicrob Chemother 67:84–90. http://dx.doi.org/10.1093/jac/dkr406.