

RESEARCH ARTICLE

Open Access



A study on combination of daptomycin with selected antimicrobial agents: in vitro synergistic effect of MIC value of 1 mg/L against MRSA strains

Yi-Chien Lee^{1,2}, Pao-Yu Chen³, Jann-Tay Wang^{3,4*} and Shan-Chwen Chang³

Abstract

Background: Daptomycin is an important drug used in the treatment of methicillin-resistant *Staphylococcus aureus* (MRSA) infection. A high dose of daptomycin is indicated for an MRSA infection with a minimum inhibitory concentration (MIC) of 1 mg/L for daptomycin. Combination therapies with daptomycin and other antimicrobial agents, including fosfomycin, display in vitro synergism potentially. This study was conducted to investigate the in vitro synergistic effect of daptomycin-based combination therapy against MRSA strains with high daptomycin MIC.

Method: The synergistic effects of daptomycin in combination with fosfomycin, gentamicin, linezolid, oxacillin, or rifampicin against MRSA with an MIC of 1 mg/L for daptomycin were measured using the microbroth checkerboard assay in vitro.

Result: A total of 100 MRSA isolates was tested. The synergistic interactions of the drugs were evaluated using the fractional inhibitory concentration index. The MIC values revealed that all isolates (100%) were found to be susceptible to linezolid, 85% to fosfomycin, 8% to gentamicin, 69% to rifampicin, and no isolate was susceptible to oxacillin. The in vitro synergism rates of daptomycin in combination with fosfomycin, oxacillin, gentamicin, linezolid, and rifampicin were 37, 11, 5, 3, and 1%, respectively.

Conclusion: The combination of daptomycin plus fosfomycin may be an effective therapeutic option for MRSA infection.

Keywords: Combination therapy, Daptomycin, MRSA, Checkerboard assays

Background

For the past few decades, methicillin-resistant *Staphylococcus aureus* (MRSA) continued to be a major human pathogen causing various dangerous infections, such as bacteremia, endocarditis, and abscess, in both community and hospital (nosocomial infection, health-care associated infection or hospital associated infection) settings [1]. The definition of MRSA nosocomial infection was classified “hospital onset”, if culture confirmed to be MRSA isolates was obtained after hospital day 3 with admission being day 1 [2]. In the United States,

MRSA contributes to 25.8% of all *S. aureus* infections and it is associated with approximately 11,000 deaths annually [2]. Its prevalence is particularly higher in Taiwan, as ever more than 80% in intensive care units [3]. Alarming, the emergence of multi-drug resistant *S. aureus* has created an international issue that needs to be solved through new treatment options [4]. Based on the treatment guidelines for MRSA infection, vancomycin remained the mainstay of parenteral therapy. However, daptomycin was considered at least as effective as vancomycin in treating MRSA bacteremia, and high-dose daptomycin in combination with another agent, including gentamicin, rifampicin, linezolid, was suggested to be the option against persistent MRSA bacteremia with vancomycin treatment failures [5]. Hence, combinations of various therapeutic strategies

* Correspondence: wangjt1124@ntu.edu.tw

³Department of Internal Medicine, National Taiwan University Hospital, 7 Chung-Shan South Road, 100 Taipei, Taiwan

⁴Institute of Infectious Diseases and Vaccinology, National Health Research Institutes, Tsu-Nan County, Taiwan

Full list of author information is available at the end of the article



may be effective in improving the clinical outcome of patients with severe staphylococcal diseases [6, 7]. Broadly, the rationale of using combination therapy is as follows: provision of wide-spectrum benefits, acquisition of the synergistic effect, and low risk of emergence of drug-resistant strains [8].

Daptomycin is a cyclic lipopeptide antimicrobial agent produced by *Streptomyces roseosporus*. This agent targets the bacterial cell membrane via a calcium-dependent pathway, disrupting electrical potential, altering cell membrane permeability, opening ion channels, and eventually causing cell death [9]. This drug is FDA-approved for adult patients with *S. aureus* infection, including MRSA infection, bloodstream infections, right-sided infective endocarditis, complicated skin and soft tissue infections [5]. Daptomycin kills MRSA more rapidly than glycopeptides. Several studies have reported that daptomycin exhibited a high efficacy against bacteremia caused by MRSA with a high minimum inhibitory concentration (MIC) for vancomycin [10]. Generally, daptomycin at 6 mg/kg/day is recommended for bacteremia caused by MRSA [9]; however, several studies have shown that up to 8 mg/kg/day daptomycin is required for patients with difficult-to-treat MRSA infection, including that caused by MRSA with an MIC of 1 mg/L for daptomycin [11, 12]. In such a clinical scenario, co-administration of daptomycin and other antimicrobial agents with synergistic interaction may be an effective therapeutic strategy.

Several in vitro studies using checkerboard methods have shown that combination therapies with daptomycin and other antimicrobials exhibit various degrees of synergism [13–15]. Moreover, there is limited information on whether similar synergistic effects are successful against MRSA with a high daptomycin MIC. Hence, in the present study, we aimed to determine the synergistic effects of therapy with daptomycin in combination with fosfomycin, gentamicin, linezolid, oxacillin, or rifampicin by measuring the in vitro antibacterial effects of these combination therapies against MRSA strains with a daptomycin MIC of 1 mg/L using the checkerboard method.

Methods

Bacterial isolates and identification

A total of 1353 MRSA specimens from sterile sites, including blood, cerebrospinal fluid, ascites, and pleural effusion, were isolated consecutively between January 2012 and December 2015 from 14 participating hospitals. These hospitals included 12 medical centers and 2 regional hospitals located in northern (6 hospitals), central (3 hospitals), southern (4 hospitals), and eastern (1 hospital) Taiwan. The present numbers of bacterial isolates enrolled each year was 300. When the total number of collected MRSA isolates reached 300 in each year, the

participating hospital would be informed to stop submitting further samples for that year. Duplicate isolates were excluded. All these strains were identified as *S. aureus* by performing gram staining, catalase-activity test, and coagulase latex agglutination test (automated VITEK-2 system, Biomerieux, France). Methicillin resistance of the cultured *S. aureus* was determined using agar disk diffusion (Kirby-Bauer), according to the guidelines established by the Clinical and Laboratory Standards Institute (CLSI) [16]. These MRSA isolates were sent to the central laboratory of the National Taiwan University Hospital for preservation, until use in the subsequent microbiological studies. The study was approved by the Ethical Committee of the National Taiwan University Hospital (NTUH-IRB No. 201110043RD).

Antimicrobials and measurement of MIC

The tested antimicrobials were daptomycin (Cubist, Pharmaceuticals, Lexington, MA), fosfomycin (Sigma-Aldrich, St. Louis, USA), gentamicin (Sigma-Aldrich), linezolid (Pfizer, New York, USA), oxacillin (Sigma-Aldrich), and rifampicin (USP; Twinbrook Parkway, Rockville, MD, USA), and they were prepared according to the manufacturers' instructions. MIC values were determined by broth microdilution in cation-adjusted Mueller-Hinton broth with an inoculum of 5×10^5 CFU/ml in the wells of the microplates. The results were interpreted according to the CLSI guidelines [16]. MIC values represent the lowest drug concentration at which complete inhibition of visible microbial growth is observed. Calcium was added to the growth media containing daptomycin to a final concentration of 50 µg/mL, whereas 25 µg/mL glucose-6-phosphate was added to the medium containing fosfomycin [16]. One hundred MRSA isolates were randomly selected from those with MIC of 1 mg/L to daptomycin using a random digital table for the subsequent experiments. The MIC₅₀ and MIC₉₀ of and the susceptibilities to each tested drug except daptomycin were calculated.

Synergy testing using the checkerboard assay

We used the 2-dimensional microbroth checkerboard method to assess the in vitro effects of combinations of various antimicrobial agents against MRSA strains [17]. The tested antimicrobials were daptomycin in combination with fosfomycin, gentamicin, linezolid, oxacillin, or rifampicin. Each antimicrobial agent in the combinations was four dilutions above and four dilutions below the MIC of every drug. The first antimicrobial agent in the combination was serially diluted 2-fold along the ordinate, whereas the second was diluted along the abscissa. Each well of the microtiter plate contained an inoculum of 5×10^5 CFU/mL that had been incubated for 24 h at 35 °C. The MIC of every antimicrobial agent alone and in combination represented the lowest dilution that

completely inhibited the growth of the bacterium. The interaction of the drugs in a combination was expressed quantitatively as a fractional inhibitory concentration (FIC) index (FICI) and calculated for each drug combination using the following equation: $FICI = FIC_A + FIC_B$, where $FIC_A = MIC$ of drug A in a combination/ MIC of drug A alone, and $FIC_B = MIC$ of drug B in a combination/ MIC of drug B alone. The FICI results were interpreted as synergistic (≤ 0.5), additive (> 0.5 to ≤ 1), or indifferent (> 1) [17]. FICI results were expressed as percentage.

Results

MIC values, susceptibility rates, and FICI results of the antimicrobials

There were 144 MRSA isolates with MIC of 1 mg/L for daptomycin among the 1353 MRSA isolates. For the 100 randomly selected clinical isolates of MRSA with MIC of 1 mg/L for daptomycin, 57 were from six hospitals in northern Taiwan, 14 from three hospitals in central Taiwan, 25 from four hospitals in southern Taiwan, and 4 from one hospital in eastern Taiwan.

The MIC ranges, MIC_{50} s, MIC_{90} s, and the susceptible rates of the tested antimicrobials for the MRSA strains are listed in Table 1 and the detailed MICs are shown in Additional file 1. All strains (100%) were susceptible to linezolid, 85% to fosfomycin, 8% to gentamicin, 69% to rifampicin, and no strain was susceptible to oxacillin. The MIC_{50} , MIC_{90} , and MIC_{range} values of the antimicrobials are provided in Table 1.

The FICI values of the combination therapies against the MRSA strains are listed in Table 2 and the in vitro antibacterial effects are demonstrated in Fig. 1. The highest synergistic effect (FICI: ≤ 0.5) was observed for daptomycin in combination with fosfomycin (37%), which was significantly higher than that in the combinations with oxacillin (11%, $p < 0.0001$), that with gentamicin (5%, $p < 0.0001$), that with linezolid (3%, $p < 0.0001$), and that with rifampicin (1%, $p < 0.0001$). In addition, the additive effect (FICI: > 0.5 to ≤ 1) in increasing order was 2% for oxacillin, 38% for gentamicin, 44% for fosfomycin, 51% for rifampicin, and 74% for linezolid. The combination of daptomycin and fosfomycin exhibited the highest synergistic/additive effect (81%) against the

MRSA strains. A majority (87%) of indifference effect (FICI: > 1) was observed for the daptomycin and oxacillin combination.

Discussion

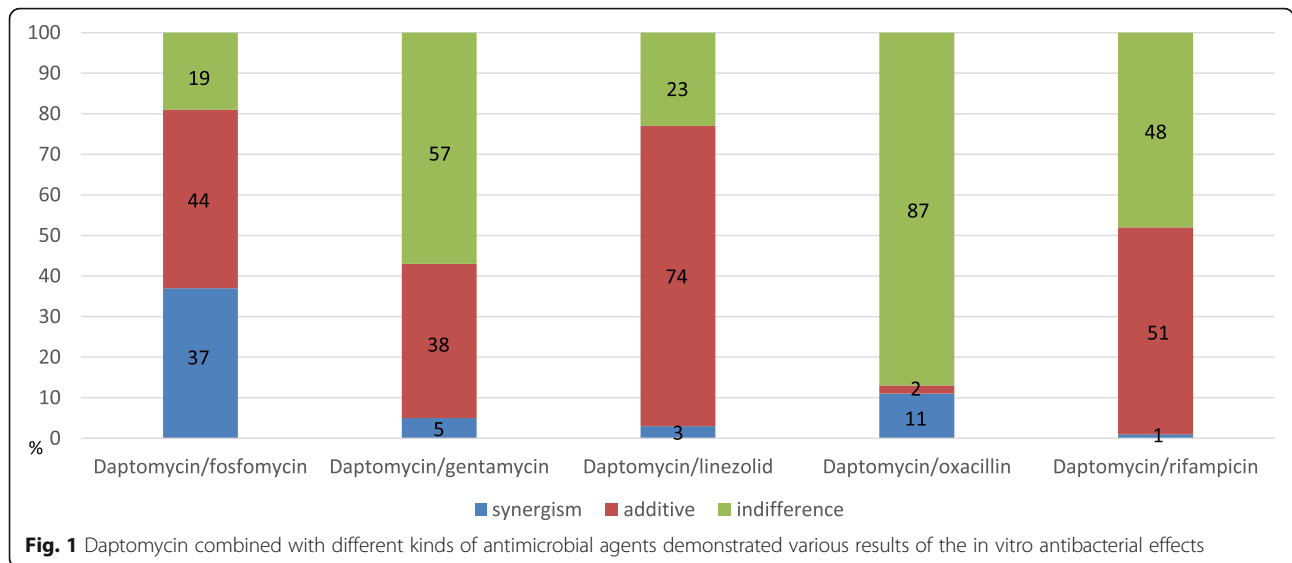
The development of optimal therapeutic strategies for serious MRSA infections remains of great interest because glycopeptide monotherapy has multiple disadvantages, including varied tissue penetration, slow bacterial killing, and emergence of drug-resistant strains [6]. Daptomycin, which has a rapid bactericidal effect against MRSA, may be an ideal treatment option [11]; however, it needs to be administered at high doses [12] and has poor prognosis [18] for patients with high MIC for daptomycin. Thus, combination therapy with daptomycin might overcome the aforementioned disadvantages. To our knowledge, this is the first in vitro study that evaluated the potential effects of a combination therapy with daptomycin and other antimicrobial agents against MRSA strains with MIC of 1 mg/L for daptomycin. In the present study, combination therapy with daptomycin and fosfomycin had the highest synergistic/additive effects against the 100 MRSA strains.

Fosfomycin, a phosphonic acid derivative first identified in 1969 [19], has a good antibacterial effect against various drug-resistant gram-positive cocci, including MRSA. In clinical settings, it is usually combined with other antimicrobials because an in vitro study has reported the rapid development of resistance to this drug [20]. Several studies have shown the synergistic effects of combination therapy with daptomycin and fosfomycin in vitro and in vivo [21, 22]. Despite limited clinical experience, several patients with MRSA bacteremia have shown successful outcomes via such a therapy [23]. In addition, our findings were supported by those of a similar study, which reported that the in vitro synergistic effect of daptomycin combined with fosfomycin is 100% [15]. The difference in synergistic effects between the previous and present studies may be due to the differences in susceptibility to fosfomycin and MIC for daptomycin. The exact mechanism underlying the synergism is still unclear. However, fosfomycin has a unique inhibitory effect on the early stage of peptidoglycan synthesis by inhibiting the formation of N-acetylmuramic acid, which may increase daptomycin binding because of alteration in the electrical charge of the outer bacterial membrane [24]. Additionally, daptomycin combined with fosfomycin could preserve daptomycin susceptibility against MRSA strains with high MIC, which made maintenance of prior therapeutic dose of daptomycin for MRSA possible clinically. However, further studies need to be conducted to confirm the above-mentioned hypothesis.

Linezolid, the first marketed synthetic oxazolidinone drug, has an inhibitory effect on protein synthesis by

Table 1 MIC values of antimicrobial agents for 100 MRSA strains (daptomycin MIC = 1) and susceptibility rates

	MIC values ($\mu\text{g/mL}$)			Susceptibility (%)
	MIC_{50}	MIC_{90}	MIC_{range}	
Fosfomycin	4	> 128	$< 0.5 - > 128$	85
Gentamicin	> 256	> 256	$0.125 - > 256$	8
Linezolid	2	2	0.5–4	100
Oxacillin	> 256	> 256	$8 - > 256$	0
Rifampicin	0.008	32	$< 0.004 - > 64$	69



affecting the 50S subunit of the bacterial ribosome [25]. It is an FDA-approved drug for skin and soft tissue infection and nosocomial pneumonia caused by MRSA, although the bacteriostatic entity may preclude its clinical use [5]. A study involving the addition of carbapenem to the parent linezolid showed a prominent synergistic effect against MRSA infection [26]. Moreover, the effects of therapy with daptomycin in combination with linezolid have been investigated. Parra-Ruiz et al. [27] reported that a daptomycin/linezolid combination is more effective than a single drug in an in vitro biofilm model. Similar results were observed using a model of simulated endocardial vegetations [28]. However, indifferent and antagonistic interactions between drugs in a combination were shown by Kelesidis et al. [29], whose results were comparable to our findings: a small proportion (3%) of synergism and large proportion (74%) of additive effects. Hence, further studies need to be performed to elucidate the efficacy of daptomycin/linezolid combination against MRSA.

A synergistic effect between beta-lactams and daptomycin against MRSA strains was observed in a previous in vitro study [6]. Several in vitro [13, 30] and animal studies [31], have shown that daptomycin/oxacillin

combination has a synergistic effect on most tested strains, and the addition of oxacillin to daptomycin in a case series led to the rapid eradication of MRSA bacteremia [32]. The following are proposed mechanisms that may explain these synergistic effects: a decline in the surface charge of MRSA by oxacillin increases daptomycin membrane binding, death of bacteria [32], and activity of oxacillin-mediated innate host defense peptides against MRSA [33]. However, currently, beta-lactam combination therapy is not suggested in the Infectious Diseases Society of America guidelines [5]. In our study, only 11% of the tested MRSA strains were consistent with the synergistic findings, and other strains with indifferent results may have to be further examined in an in vitro study to elucidate this difference.

Co-administration of daptomycin and gentamicin had potential synergistic activities against *S. aureus* in time-killing analyses, as verified by Debbia et al. [21]. Nevertheless, several in vitro and in vivo studies have reported varied results: some reported synergy [13, 34], whereas others did not [35, 36]. Synergistic interaction of up to 68% was reported by Aktas et al. [15], which contradicts the 5% synergism observed in our study. This distinction may be due to the differences in the bacterial strains used. A previous random control clinical trial for evaluating the efficacy of daptomycin with and without gentamicin against *S. aureus* infective endocarditis was terminated prematurely after only 24 patients were recruited [6], making the effects of this combination even more inconclusive.

The use of rifampicin as a single agent against MRSA infection is associated with the rapid development of drug resistance [37], and the effectiveness of the rifampicin and daptomycin combination for increasing the clearance of MRSA biofilm has been reported previously

Table 2 Interpreted FICI results of antimicrobial agents for 100 MRSA strains (daptomycin MIC = 1)

Combination	Interpreted FICI results, (%)		
	synergism	additive	indifference
Daptomycin/fosfomycin	37	44	19
Daptomycin/gentamycin	5	38	57
Daptomycin/linezolid	3	74	23
Daptomycin/oxacillin	11	2	87
Daptomycin/rifampicin	1	51	48

[38]. Studies using animal models and retrospective reviews [39, 40] and case reports [41] have also confirmed the synergy and clinical success of daptomycin in combination with rifampicin. Rose et al. [14] showed via checkerboard analysis that the in vitro synergy of this combination was 75%, with 100% predictive value for treatment outcome. However, a similar study [19] showed poor synergism (12%), which is comparable to that observed in our study (1%). Therefore, owing to incomplete clinical data, as no clinical outcome associated with daptomycin combined with rifampicin is demonstrated in our study and probably different epidemiological background of bacterial isolates between our present study and previous ones, the benefits of this combination therapy are unclear.

Our study had certain limitations. First, although the present study showed the synergistic effects of daptomycin in combination with some drugs, the insufficient clinical data limited our interpretation of the correlation between the in vitro studies and treatment outcome. Second, this study was conducted using the population of a single country, Taiwan; thus worldwide generalization should be made carefully. Finally, some antimicrobial agents effectively against MRSA, like ceftaroline (not available in Taiwan when this manuscript was written), were not included for assessment of the in vitro antibacterial effects.

Conclusions

In conclusion, our study showed that daptomycin in combination with fosfomycin had a high synergistic/additive effect against MRSA with an MIC of 1 mg/L for daptomycin. Hence, combination therapy with daptomycin and fosfomycin may be an effective therapeutic strategy for MRSA infection with high MIC for daptomycin as compared to monotherapy. However, further clinical research should be conducted to verify the synergistic mechanism of these antimicrobial agents.

Additional file:

Additional file 1: The MIC of the five antimicrobial agents to those 100 MRSA strains. (PDF 283 kb)

Abbreviations

CFU: Colony-forming unit; CLSI: Clinical and Laboratory Standards Institute; FDA: Food and Drug Administration; FICI: Fractional inhibitory concentration index; MIC: Minimum inhibitory concentration; MRSA: Methicillin resistant *Staphylococcus aureus*

Acknowledgements

Not applicable.

Funding

There were no external or internal sources of specific funding for this paper, and the data were generated as part of the department's routine work.

Availability of data and materials

The datasets used during the current study are available from the corresponding author on reasonable request.

Authors' contributions

YL and JW wrote the article and revised it critically for important intellectual content. PC collected the data and did the analysis and interpretation of data. SC was responsible for the conception and design of the study. JW had given the final approval of the version to be published. All authors had read and approved the final manuscript.

Ethics approval and consent to participate

The study was carried out in accordance with the principles stated in the Declaration of Helsinki, and approved by the Ethical Committee of National Taiwan University Hospital (NTUH-IRB No. 201110043RD). The Review Board approved to waive informed consent due to the retrospective study design and the research posing no more than minimal risk.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interest.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Department of Internal Medicine, Fu Jen Catholic University Hospital, Fu Jen Catholic University, New Taipei City, Taiwan. ²School of Medicine, College of Medicine, Fu Jen Catholic University, New Taipei City, Taiwan. ³Department of Internal Medicine, National Taiwan University Hospital, 7 Chung-Shan South Road, 100 Taipei, Taiwan. ⁴Institute of Infectious Diseases and Vaccinology, National Health Research Institutes, Tsu-Nan County, Taiwan.

Received: 26 December 2018 Accepted: 23 April 2019

Published online: 06 May 2019

References

- Lowy FD. Staphylococcus aureus infections. *N Engl J Med.* 1998;339:520–32.
- Dantes R, Mu Y, Belflower R, Aragon D, Dumyati G, Harrison LH, et al. National burden of invasive methicillin-resistant *Staphylococcus aureus* infections, United States, 2011. *JAMA Intern Med.* 2013;173:1970–8.
- Wang JT, Lauderdale TL, Lee WS, Huang JH, Wang TH, Chang SC. Impact of active surveillance and contact isolation on transmission of methicillin-resistant *Staphylococcus aureus* in intensive care units in an area with high prevalence. *J Formos Med Assoc.* 2010;109:258–68.
- Stefani S, Gogliio A. Methicillin-resistant *Staphylococcus aureus*: related infections and antibiotic resistance. *Int J Infect Dis.* 2010;14(Suppl 4):S19–22.
- Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, et al. Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis.* 2011;52:e18–55.
- Davis JS, Van Hal S, Tong SY. Combination antibiotic treatment of serious methicillin-resistant *Staphylococcus aureus* infections. *Semin Respir Crit Care Med.* 2015;36:3–16.
- Chen YS. Guidelines for the treatment of methicillin-resistant *Staphylococcus aureus* infections in Taiwan. *J Microbiol Immunol Infect.* 2013;46:147–50.
- García AB, Candel FJ, Lopez L, Chiarella F, Viñuela-Prieto JM. In vitro ceftaroline combinations against methicillin-resistant *Staphylococcus aureus*. *J Med Microbiol.* 2016;65:1119–22.
- Bayer AS, Schneider T, Sahl HG. Mechanisms of daptomycin resistance in *Staphylococcus aureus*: role of the cell membrane and cell wall. *Ann N Y Acad Sci.* 2013;1277:139–58.
- Moore CL, Oski-Kiyan P, Haque NZ, Perri MB, Donabedian S, Zervos MJ. Daptomycin versus vancomycin for bloodstream infections due to methicillin-resistant *Staphylococcus aureus* with a high vancomycin minimum inhibitory concentration: a case-control study. *Clin Infect Dis.* 2012;54:51–8.

11. Gonzalez-Ruiz A, Seaton RA, Hamed K. Daptomycin: an evidence-based review of its role in the treatment of gram-positive infections. *Infect Drug Resist.* 2016;15(9):47–58.
12. Soon RL, Turner SJ, Forrest A, Tsuji BT, Brown J. Pharmacokinetic/ pharmacodynamic evaluation of the efficacy and safety of daptomycin against *Staphylococcus aureus*. *Int J Antimicrob Agents.* 2013;42:53–8.
13. Snyderman DR, McDermott LA, Jacobus NV. Evaluation of in vitro interaction of daptomycin with gentamicin or beta-lactam antibiotics against *Staphylococcus aureus* and enterococci by FIC index and timed-kill curves. *J Chemother.* 2005;17:614–21.
14. Rose WE, Berti AD, Hatch JB, Maki DG. Relationship of in vitro synergy and treatment outcome with daptomycin plus rifampin in patients with invasive methicillin-resistant *Staphylococcus aureus* infections. *Antimicrob Agents Chemother.* 2013;57:3450–2.
15. Aktas G, Derbentli S. In vitro activity of daptomycin combinations with rifampin, gentamicin, fosfomicin and fusidic acid against MRSA strains. *J Glob Antimicrob Resist.* 2017;10:223–7.
16. Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing: Twenty-eighth informational supplement M100-S20. Wayne: CLSI; 2018.
17. Pillai SK, Moellering RC Jr, Eliopoulos GM. Antimicrobial combinations. In: Lorian V, editor. *Antibiotics in laboratory medicine*. Philadelphia: Lippincott Williams and Wilkins; 2005. p. 365–73.
18. Ruiz J, Ramirez P, Concha P, Salavert-Lletí M, Villarreal E, Gordon M, et al. Vancomycin and daptomycin minimum inhibitory concentrations as a predictor of outcome of methicillin-resistant *Staphylococcus aureus* bacteraemia. *J Glob Antimicrob Resist.* 2018;14:141–4.
19. Hendlin D, Stapley EO, Jackson M, Wallick H, Miller AK, Wolf FJ, et al. Phosphonomycin, a new antibiotic produced by strains of streptomycetes. *Science.* 1969;166:122–3.
20. Falagas ME, Roussos N, Kgekkes ID, Rafailidis PI, Karageorgopoulos DE. Fosfomicin for the treatment of infections caused by gram-positive cocci with advanced antimicrobial drug resistance: a review of microbiological, animal and clinical studies. *Expert Opin Investig Drugs.* 2009;18:921–44.
21. Debbia E, Pesce A, Schito GC. In vitro activity of LY146032 alone and in combination with other antibiotics against gram-positive bacteria. *Antimicrob Agents Chemother.* 1988;32:279–81.
22. Lingscheid T, Poeppl W, Bernitzky D, Veletzky L, Kussmann M, Plasenzotti R, et al. Daptomycin plus fosfomicin, a synergistic combination in experimental implant-associated osteomyelitis due to methicillin-resistant *Staphylococcus aureus* in rats. *Antimicrob Agents Chemother.* 2015;59:859–63.
23. Miró JM, Entenza JM, Del Río A, Velasco M, Castañeda X, García de la Mària C, et al. High-dose daptomycin plus fosfomicin is safe and effective in treating methicillin-susceptible and methicillin-resistant *Staphylococcus aureus* endocarditis. *Antimicrob Agents Chemother.* 2012;56:4511–5.
24. Popovic M, Steinort D, Pillai S, Joukhadar C. Fosfomicin: an old, new friend? *Eur J Clin Microbiol Infect Dis.* 2010;29:127–42.
25. MacGowan AP. Pharmacokinetic and pharmacodynamic profile of linezolid in healthy volunteers and patients with gram-positive infections. *J Antimicrob Chemother.* 2003;51(Suppl 2):ii17–25.
26. Jang HC, Kim SH, Kim KH, Kim CJ, Lee S, Song KH, et al. Salvage treatment for persistent methicillin-resistant *Staphylococcus aureus* bacteremia: efficacy of linezolid with or without carbapenem. *Clin Infect Dis.* 2009;49:395–401.
27. Parra-Ruiz J, Bravo-Molina A, Peña-Monje A, Hernández-Quero J. Activity of linezolid and high-dose daptomycin, alone or in combination, in an in vitro model of *Staphylococcus aureus* biofilm. *J Antimicrob Chemother.* 2012;67:2682–5.
28. Steed ME, Vidaillic C, Rybak MJ. Novel daptomycin combinations against daptomycin-nonsusceptible methicillin-resistant *Staphylococcus aureus* in an in vitro model of simulated endocardial vegetations. *Antimicrob Agents Chemother.* 2010;54:5187–92.
29. Kelesidis T, Humphries R, Ward K, Lewinski MA, Yang OO. Combination therapy with daptomycin, linezolid, and rifampin as treatment option for MRSA meningitis and bacteremia. *Diagn Microbiol Infect Dis.* 2011;71:286–90.
30. Leonard SN, Rolek KM. Evaluation of the combination of daptomycin and nafcillin against vancomycin-intermediate *Staphylococcus aureus*. *J Antimicrob Chemother.* 2013;68:644–7.
31. Garrigós C, Murillo O, Lora-Tamayo J, Verdaguier R, Tubau F, Cabellos C, et al. Efficacy of daptomycin-cloxacillin combination in experimental foreign-body infection due to methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother.* 2012;56:3806–11.
32. Dhand A, Bayer AS, Pogliano J, Yang SJ, Bolaris M, Nizet V, et al. Use of antistaphylococcal beta-lactams to increase daptomycin activity in eradicating persistent bacteremia due to methicillin-resistant *Staphylococcus aureus*: role of enhanced daptomycin binding. *Clin Infect Dis.* 2011;53:158–63.
33. Sakoulas G, Okumura CY, Thienphrapa W, Olson J, Nonejuie P, Dam Q, et al. Nafcillin enhances innate immune-mediated killing of methicillin-resistant *Staphylococcus aureus*. *J Mol Med (Berl).* 2014;92:139–49.
34. Credito K, Lin G, Appelbaum PC. Activity of daptomycin alone and in combination with rifampin and gentamicin against *Staphylococcus aureus* assessed by time-kill methodology. *Antimicrob Agents Chemother.* 2007;51:1504–7.
35. LaPlante KL, Woodmansee S. Activities of daptomycin and vancomycin alone and in combination with rifampin and gentamicin against biofilm-forming methicillin-resistant *Staphylococcus aureus* isolates in an experimental model of endocarditis. *Antimicrob Agents Chemother.* 2009;53:3880–6.
36. Rose WE, Leonard SN, Rybak MJ. Evaluation of daptomycin pharmacodynamics and resistance at various dosage regimens against *Staphylococcus aureus* isolates with reduced susceptibilities to daptomycin in an in vitro pharmacodynamic model with simulated endocardial vegetations. *Antimicrob Agents Chemother.* 2008;52:3061–7.
37. Perlroth J, Kuo M, Tan J, Bayer AS, Miller LG. Adjunctive use of rifampin for the treatment of *Staphylococcus aureus* infections: a systematic review of the literature. *Arch Intern Med.* 2008;168:805–19.
38. Raad I, Hanna H, Jiang Y, Dvorak T, Reitzel R, Chaiban G, et al. Comparative activities of daptomycin, linezolid, and tigecycline against catheter-related methicillin-resistant staphylococcus bacteremic isolates embedded in biofilm. *Antimicrob Agents Chemother.* 2007;51:1656–60.
39. Saleh-Mghir A, Muller-Serieys C, Dinh A, Massias L, Crémieux AC. Adjunctive rifampin is crucial to optimizing daptomycin efficacy against rabbit prosthetic joint infection due to methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother.* 2011;55:4589–93.
40. Lora-Tamayo J, Parra-Ruiz J, Rodríguez-Pardo D, Barberán J, Ribera A, Tornero E, et al. High doses of daptomycin (10 mg/kg/d) plus rifampin for the treatment of staphylococcal prosthetic joint infection managed with implant retention: a comparative study. *Diagn Microbiol Infect Dis.* 2014;80:66–71.
41. Hagiya H, Terasaka T, Kimura K, Satou A, Asano K, Waseda K, et al. Successful treatment of persistent MRSA bacteremia using high-dose daptomycin combined with rifampicin. *Intern Med.* 2014;53:2159–63.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

