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# Case Report

# Response to immunotherapy in a patient with adult onset Leigh syndrome and T9176C mtDNA mutation



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## ABSTRACT

Leigh syndrome is a mitochondrial disease caused by mutations in different genes, including ATP6A for which no known therapy is available. We report a case of adult-onset Leigh syndrome with response to immunotherapy. A twenty year-old woman with baseline learning difficulties was admitted with progressive behavioral changes, diplopia, headaches, bladder incontinence, and incoordination. Brain MRI and PET scan showed T2 hyperintensity and increased uptake in bilateral basal ganglia, respectively. Autoimmune encephalitis was suspected and she received plasmapheresis with clinical improvement. She was readmitted 4 weeks later with dysphagia and aspiration pneumonia. Plasmapheresis was repeated with resolution of her symptoms. Given the multisystem involvement and suggestive MRI changes, genetic testing was done, revealing a homoplasmic T9176C ATPase 6 gene mtDNA mutation. Monthly IVIG provided clinical improvement with worsening when infusions were delayed. Leigh syndrome secondary to mtDNA T9176C mutations could have an autoimmune mechanism that responds to immunotherapy.

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# 1. Introduction

Subacute necrotizing encephalomyelopathy was first described by Leigh in 1951 and has since been referred to as Leigh disease or Leigh syndrome. Leigh syndrome is a devastating, neurodegenerative disorder with almost identical radiological and pathological changes but marked clinical and genetic heterogeneity. Patients usually present with progressive decline of central nervous system function due to focal, necrotizing lesions of the basal ganglia, diencephalon, cerebellum or brainstem. Clinical features include regression or psychomotor delay, weakness, hypotonia, truncal ataxia, intention tremor, lactic acidosis in blood, cerebrospinal fluid or urine [1]. Leigh syndrome is usually a disorder of infancy and early childhood although rare adolescent and adult cases have been reported. The prognosis is usually poor and most patients usually die before age 5 [1]. There is no known treatment. We report a patient with juvenile-adult onset of Leigh syndrome and apparent response to immunotherapy.

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## 2. Material and methods

Case report.

# 3. Results

A 20 year old woman with learning disability and problems during school, non-athletic and described by family members as "clumsy" suffered a car accident in February 2015. This was followed by development of hypersomnia, frequent falls, increased headaches with migraine features, intermittent diplopia, bladder incontinence, behavioral changes with apathy, poor hygiene, irritability and disinhibition. She could not perform her activities of daily living (ADL) independently.

Her past medical history was significant for asthma, migraines and attention deficit hyperactivity disorder (ADHD) for which she used dextroamphetamine when she was 11 years old (discontinued due to development of paranoid behavior). She had normal motor milestones and was toilet trained at 18 months. She had speech problems since early age, with stuttering that needed speech therapy. Her school performance was below her peer levels and she had an individualized education program (IEP) until high school. Her family history was significant for migraines in her mother. The patient had 2 halfbrothers on her father side, one had learning disability, delayed speech milestones and exercise-induced asthma. The other half brother had

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diagnosis of glycogen-storage disease type 3A (Cori disease). Home medications included amitryptiline 10 mg at bedtime, sumatriptan 25 mg PRN and medroxyprogesterone acetate 400 mg IM q3 months.

On admission on 03/24/15 she was oriented ×3, had Medical Research Council (MRC) 3/5 strength in proximal upper and lower extremities and 4/5 in distal lower extremities and she could not do tandem gait. Cranial nerves II–XII, sensation and deep tendon reflexes were normal. Brain MRI showed bilateral T2/FLAIR hyperintensity in the basal ganglia (Fig. 1A, B). Cerebrospinal fluid (CSF) showed 21 mg/dl protein, 64 mg/dl glucose, 0 white blood cells (WBCs) and 3 red blood cells (RBCs). Oligoclonal bands were absent. Other tests such as VDRL, antinuclear (ANA) panel, Mayo Clinic paraneoplastic panel, serum protein electrophoresis, CSF Herpes simplex virus polymerase chain reaction (PCR) were normal or negative. Nerve conduction study in March 2015 showed small distal peroneal and tibial compound muscle action potentials (CMAP) amplitudes bilaterally. Electromyography was not tolerated by the patient. Computed tomography of chest, abdomen and pelvis was normal.

Due to acute/subacute onset and progressive symptoms in a young female patient, an autoimmune etiology was suspected and she underwent 5 sessions of plasmapheresis with improvement of her behavior, bladder incontinence, and muscle strength. She was able to walk with a walker and perform her ADLs on her own. She was discharged to a rehabilitation center on 04/03/15 and readmitted on 04/21/15 for dysphagia leading to aspiration and respiratory distress needing intubation. Examination showed primary gaze horizontal nystagmus, mild bifacial weakness, 3/5 muscle strength in deltoid, iliopsoas, quadriceps, tibialis anterior (with other muscles being 4/5), deep tendon reflexes were 1 + in biceps and triceps and absent in lower extremities. She was unable to walk or stand and failed her bedside swallow evaluation.

Brain MRI showed progression of T2/FLAIR hyperintensity in the bilateral basal ganglia, substantia nigra, midbrain, periaqueductal grey

matter, and 2 foci of restricted diffusion within the left lentiform nucleus (Fig. 1C–E). CSF showed 11 mg/dl protein, 54 mg/dl glucose, with 0 RBC and WBCs. CSF lactic acid was elevated at 2.9 mmol/l. CSF myelin basic protein was elevated at 5.79 ng/ml. Serum lactate and pyruvate were normal. CSF NMDA-R, VGKC, GAD65, GABA-B, AMPA-R, ANNA-1 were negative. CSF and serum samples tested negative for mGluR1, mGluR5, LGI, Caspr2, MOG and AQP4 antibodies and there was no reactivity to suggest antibodies against unknown cell surface antigens. Urine organic acids, acylcarnitine profile, serum aminoacid profile were normal. Urine aminoacid testing showed generalized aminoaciduria. Fluorodexyglucose (FDG) PET scan in April 2015 showed increased uptake within the caudate and basal ganglia bilaterally (Fig. 1F). She received antibiotics for aspiration pneumonia, and 5 sessions of plasmapheresis with improvement of swallowing, and muscle strength to 4/5.

Patient was discharged to rehabilitation center and started receiving IVIG every 4 weeks with continued improvement of her behavior and strength, including going from walking with assistance of a walker to walking independently. She had episodes of paranoia that also improved. At 3 months follow up after discharge patient was oriented  $\times$  3, followed commands, cranial nerves were normal, strength was 4+/5 in hip flexion and otherwise 5/5, reflexes were 2 + all over and she could walk albeit a little unsteady but without support.

Given her baseline learning difficulties, coordination problems and brain MRI pattern suggestive of mitochondrial disease further genetic evaluation was performed. Karyotype was 46, XX. Whole exome array CGH + SNP analysis was normal. Mitochondrial genome testing showed a homoplasmic T9176C mutation in the MT-ATP6A gene which has been described in 1–5% of patients with Leigh Syndrome. Nuclear genome testing showed a new c.3483-7\_3509del34 heterozygous mutation in polymerase gamma (POLG) that destroys the canonical splice acceptor site in exon 22. She also had mutations of unknown clinical significance in DARS2 (c.228-20dupT) and LRPPRC genes (c.1529 C > G).



Fig. 1. (A and B) Brain MRI FLAIR sequence on 1st admission shows bilateral basal ganglia and periaqueductal area hyperintensity. (C) Brain MRI FLAIR sequence on 2nd admission shows worsening hyperintensity in bilateral basal ganglia and periaqueductal area. (D and E) Brain MRI DWI sequence shows diffusion restriction in left lenticular nucleus and periaqueductal area. (F) FDG PET shows increased uptake in bilateral basal ganglia.

### Table 1

Ref #	Sex	Clinical features	Mutation load	MRI /NCS/EMG	Treatment	Prognosis
	Age of onset					
5	Girl 3 yo	Girl: Ataxia, nystagmus, slurred	100% homoplasmic	CT: bilateral basal ganglia	None described	Girl: Recovered over 12 months. No
		speech after febrile illness		hypodensities		more episodes up to 8 yo
	Mother 29 yo	Mother: sudden ataxia, headache,	Mother 96%	Mother: normal MRI, serum		Mother: improved over 9 months
		blurry vision, nystagmus		lactate		
	Boy 5 yo from	Boy: ataxia, slurred speech, lethargy		MRI:hyperintensities cauda,		Boy: improved but has residual
	different family	after flu like illness, hypotonia,		globus pallidus, pons		coordination and lethargy problems,
		hyperreflexia				"absence-like" events at 10 yo
6	3 females	Maternally-inherited, late-onset	100% homoplasmic	MRI: Normal	None	All alive, men more disabled than
	(30-48 yo)	hereditary spastic paraparesis		EMG: normal or axonal		women
		Spastic paraparesis, lower extremity		neuropathy		
	2 males	hyperreflexia and distal weakness,		Normal serum lactate and		
	(30–50 yo)	pain, reduced vibration sense		pyruvate		
7	Girl: 21 m	Cerebellar ataxia, speech delay,	Siblings: 90–95%	CT: Bilateral hypodensities	Boy and girl that	1 girl died
		dystonia, respiratory distress,	Mother: 25%	cerebellum, pons, midbrain	survived received vitamin	
	Boy: 24 m	exacerbated by viral infections with			B complex, vitamin E,	Boy: 8 yo cerebellar ataxia,
		attacks of paraparesis, dystonia,			carnitine	dystonia, pyramidal syndrome, only
		sighing, dyspnea				speaks a few words, mental
						retardation
	Girl: 24 m					Girl 6yo: mild speech delay, ataxia
8	Воу 3 уо	Ataxia, lethargy, apnea after febrile	Boy 100% homoplasmic	MRI hyperintensities basal	None described	Boy died after 1 month
		illness		ganglia, thalamus, periaqueductal,		
				periventricular, medulla		
	Mother 22 yo	Mother: developmental delay,	Mother 93%	Mother MRI: mild cerebellar		Mother: ataxia, nystagmus, mental
		progressive mental retardation,	Asymptomatic maternal	atrophy		retardation
		ataxia, nystagmus, pyramidal signs	uncle: 88%			
			Asymptomatic maternal			
			grandmother: 76%			
9	2 brothers;	Poor sucking, hypotonia, brisk	>95%	Brothers: leukodystrophy, diffuse	None	Died at 7 and 10 months of age
	4 and 5 m	reflexes, hearing loss, no visual	Asymptomatic mother:	cerebral and cerebellar white		
		contact	80%	matter hyperintensity, posterior		
				limb internal capsule		
10	Woman 22 yo	Acute diplopia, blurry vision, eyelid	>99% homoplasmic	MRI bilateral hyperintensity pons,	None described	Woman: not described
		ptosis, tachycardia, respiratory		midbrain, diencephalon		
		distress, seizures, ataxia		PET: reduction glucose		
				metabolism in cerebral and		
				cerebellar cortex		
	Brother 4 yo	Brother: Leigh syndrome				Brother died 7 yo
	Mother	Mother: mild mental retardation	Mother: 30.5%			

(continued on next page)

#### Table 1 (continued)

11	Boy 12 m	Hyperpnea, microcephaly, external	100%	Familial bilateral striatal necrosis	None described	6 yo: needs to hold onto furniture,
		ophthalmoplegia, dystonia,		MRI hyperintensity		severe language delay, static
		developmental delay		putamen, periaqueductal region		
	Воу 8 уо	Headache, altered mental status after	98%			11 yo: learning disabled, poor
		viral illness, language regression,	Unaffected mother: 7%			coordination, motor tics, static
		dystonia, chorea, myoclonus	Unaffected brother 55%			
			Unaffected sister 76%			
12	Boy 9 m	Seizures, hypotonia, coma during	>95%	CT hypodensity basal ganglia,	None described	Died within hours
		febrile illness	Unaffected mother 50%	brainstem		
	Boy 9 m	Coma, periodic breathing, seizures,	Unaffected maternal			Died within 3 weeks
		pyramidal signs, following febrile	uncle: 70%			
		illness				
13	Boy 6 m	Hypotonia, developmental delay,	>95%	MRI Hyperintensity basal ganglia,	None described	Bedridden at age 10
		seizure, retinitis pigmentosa		midbrain, pons		
	Воу 8 уо	Ataxia, chorea, neuropathy,	>97%			Mental retardation age 9
		abnormal eye movements				
	Воу 3 уо	Ataxia, pyramidal signs, dysarthria,	97%			Slowly progressive
		oculomotor palsy, mental retardation	Unaffected mother 52%			
This	20 уо	Long history clumsiness, language	100% homoplasmic	MRI Hyperintensity bilateral basal	IVIG, plasmapheresis	Improvement after plasmapheresis
patient		and learning problems, worse after		ganglia, periaqueductal area	Carnitine 1500 mg BID	and IVIG infusions
		car accident			Coenzyme Q10 200 mg	Deterioration when IVIG is delayed
					TID	
					Vitamin B50 complex	
					Lipoic acid 400 mg TID	
					Vitamin E 200 IU qday	
					Vitamin C 100 mg TID	
					Selenium 25 µg qday	

Ref: reference number; m: months; yo: years-old; BID: twice a day; IU: international units.

## 4. Discussion

Leigh syndrome is associated with defects in the pyruvate dehydrogenase complex, electron transport chain complexes I, III, IB, ATP synthase and mitochondrial electron transport assembly proteins. These abnormalities are due to mutations in mitochondrial and nuclear genes. The most commonly mutated mitochondrial gene is ATPase 6, with the most frequent mutation being 8993T > G. The nuclear gene most frequently mutated in Leigh syndrome is SURF1 [1].

Our patient met criteria for clinical diagnosis of Leigh syndrome due to her progressive neurologic disease with cognitive, behavioral and motor symptoms, signs of brainstem and basal ganglia disease, elevated CSF lactate, and compatible brain MRI pattern [2].

Leigh syndrome has characteristic neuroimaging findings with symmetrical T2 hyperintensities in deep grey matter including thalamus, lentiform and caudate nuclei, periaqueductal grey matter and midbrain tegmentum [1].

Our patient history and presentation is peculiar because although she had prolonged history of learning problems, clumsiness and poor athletic performance, she developed new symptoms of diplopia, behavioral changes, incontinence, generalized weakness and headache rather subacutely after a motor vehicle accident, all of which responded to immunomodulatory therapies such as plasmapheresis and IVIG. Although the subacute clinical presentation, increased uptake on PET scan and improvement with plasmapheresis or IVIG suggested a possible autoimmune mechanism; the MRI pattern was more in link with metabolic or mitochondrial disease, which led us to do genetic testing that showed the known T9176C mutation in ATP6A associated with Leigh syndrome.

Our patient is also heterozygous for a POLG mutation affecting the splice acceptor site at exon 22. POLG mutations can cause conditions such as Alpers syndrome, progressive external ophthalmoplegia (PEO) or sensory ataxic neuropathy with dysarthria and ophthalmoparesis (SANDO). Our patient did not have clinical evidence of POLG-related disease, including liver failure, myoclonic seizures or PEO; although she had small peroneal and tibial CMAP amplitudes which can be secondary to myopathy or motor neuropathy (she refused repeated nerve studies and muscle biopsy). Although our patient's POLG deletion has not been described before, and it affects a splice site, it is not necessarily pathogenic on its own. Skipping exon 22 would either cause nonsense mediated decay of the transcript or, if translated, make a dysfunctional DNA polymerase [3]. An Alpers syndrome patient compound heterozygous for A467T and 3482 + 2T->C splicing POLG mutations has been reported. His asymptomatic father had the 3482 + 2T-> C mutation, which affects the splicing of intron 21 and is very close to our patient's mutation [4]. Based on this report, we think that our patient's POLG mutation is likely recessive and non-clinically manifest on its own, as she had no mutation in the other allele.

The ATPase 6 gene encodes part of the mitochondrial F0-F1 ATP synthese (complex V of oxidative phosphorylation). Complex V is

composed of at least 16 subunits, of which 2 (ATP6 and 8) are mtDNAencoded. Complex V synthesize ATP utilizing the proton gradient created by the respiratory chain complexes I–IV [5,6]. The T9176C mutation in ATPase 6 results in the replacement of a highly conserved leucine residue (aminoacid 270) by proline. In the literature there are reported cases of patients with the T9176C mutation and most of them presented with Leigh syndrome; although other presentations include bilateral striatal necrosis, hereditary spastic paraparesis, ataxia and mental retardation (Table 1) [5–13]. The prognosis is usually poor in cases of Leigh syndrome, although the 3 cases described by Wilson only had one episode of neurological deterioration, triggered by a viral or febrile illness, followed by spontaneous improvement over 9–12 months with a very good prognosis [5].

The reason for our patient's clinical improvement after plasmapheresis and IVIG treatment is unclear since Leigh syndrome is a mitochondrial disease. Although her CSF was normal (including negative oligoclonal bands) and serum and CSF testing for cell surface antibodies was also negative; her brain PET scan showed hypermetabolism in bilateral basal ganglia corresponding to her MRI hyperintensities. Mitochondrial disorders such as POLG mutations, mitochondrial neurogastrointestinal encephalopathy (MNGIE), mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes (MELAS) and Leigh syndrome have been reported to have hypometabolism in PET scan [14-18]; although acute MELAS lesions can also have hypermetabolism [19]. It is known that PET hypermetabolism can also be seen in autoimmune encephalitis [20] and we believe that our patient's PET findings are related to an underlying autoimmune process triggered by her mitochondrial disease which can explain her improvement after immunotherapy. There are published cases of mitochondrial diseases such as Leber's hereditary optic neuropathy (LHON), PEO and POLG mutations mimicking multiple sclerosis or acute demyelinating encephalomyelitis (ADEM) including the presence of oligoclonal bands [21,22]. There is also a case of inflammatory mitochondrial myopathy secondary to a tRNA Leu mtDNA mutation that responded to IVIG infusions [23]. Although we do not know the actual mechanism for our patient's response immunotherapy, we can postulate a hypothesis. The T9176C is a known pathogenic mtDNA mutation and affects ATP synthesis [6,24]. Reduced ATP production can increase the mitochondrial transmembrane potential (with resultant hyperpolarization) and increase reactive oxygen species (ROS) production [25]. ROS are known damage-associated molecular patterns (DAMPs) that can activate the necroptosis pathway [26]. Impaired ATP synthesis can also induce necroptosis [27]. Cell necrosis can lead to release of immunogenic material and activate immune and inflammation pathways (inflammasome) [26,28]. In ischemic stroke models, it has been shown that IVIG can suppress inflammasome-mediated neuronal death [29].

#### 5. Conclusions

Leigh syndrome secondary to T9176C mutations may have an underlying autoimmune mechanism amenable to immunotherapy. Further research on the role of the inflammation/immune pathway activation in patients with Leigh syndrome and T9176C mtDNA mutations, other ATP synthase mutations and other mitochondrial diseases is needed in order to explore the role of immunotherapy in these conditions.

#### References

- J. Finsterer, Leigh and Leigh-like syndrome in children and adults, Pediatr. Neurol. 39 (2008) 223–235.
- [2] S. Rahman, R.B. Blok, H.H. Dahl, D.M. Danks, D.M. Kirby, C.W. Chow, J. Christodoulou, D.R. Thorburn, Leigh syndrome: clinical features and biochemical and DNA abnormalities, Ann. Neurol. 39 (1996) 343–351.
- [3] S.S. Chan, W.C. Copeland, DNA polymerase gamma and mitochondrial disease: understanding the consequence of POLG mutations, Biochim. Biophys. Acta 1787 (2009) 312–319.
- [4] G. Ferrari, E. Lamantea, A. Donati, M. Filosto, E. Briem, F. Carrara, R. Parini, A. Simonati, R. Santer, M. Zeviani, Infantile hepatocerebral syndromes associated with mutations in the mitochondrial DNA polymerase-gammaA, Brain 128 (2005) 723–731.

- [5] C.J. Wilson, N.W. Wood, J.V. Leonard, R. Surtees, S. Rahman, Mitochondrial DNA point mutation T9176C in Leigh syndrome, J. Child Neurol. 15 (2000) 830–833.
- [6] C. Verny, N. Guegen, V. Desquiret, A. Chevrollier, A. Prundean, F. Dubas, J. Cassereau, M. Ferre, P. Amati-Bonneau, D. Bonneau, P. Reynier, V. Procaccio, Hereditary spastic paraplegia-like disorder due to a mitochondrial ATP6 gene point mutation, Mitochondrion 11 (2011) 70–75.
- [7] L.J. Jacobs, I.F. de Coo, J.G. Nijland, R.J. Galjaard, F.J. Los, K. Schoonderwoerd, M.F. Niermeijer, J.P. Geraedts, H.R. Scholte, H.J. Smeets, Transmission and prenatal diagnosis of the T9176C mitochondrial DNA mutation, Mol. Hum. Reprod. 11 (2005) 223–228.
- [8] Y. Campos, M.A. Martín, J.C. Rubio, L.G. Solana, C. García-Benayas, J.L. Terradas, J. Arenas, Leigh syndrome associated with the T9176C mutation in the ATPase 6 gene of mitochondrial DNA, Neurology 49 (1997) 595–597.
- [9] P.C. Hung, H.S. Wang, A previously undescribed leukodystrophy in Leigh syndrome associated with T9176C mutation of the mitochondrial ATPase 6 gene, Dev. Med. Child Neurol. 49 (2007) 65–67.
- [10] D. Ronchi, A. Bordoni, A. Cosi, M. Rizzuti, E. Fassone, A. Di Fonzo, M. Servida, M. Sciacco, M. Collotta, M. Ronzoni, V. Lucchini, M. Mattioli, M. Moggio, N. Bresolin, S. Corti, G.P. Comi, Unusual adult-onset Leigh syndrome presentation due to the mitochondrial m.9176T > C mutation, Biochem. Biophys. Res. Commun. 412 (2011) 245–248.
- [11] D. Thyagarajan, S. Shanske, M. Vazquez-Memije, D. De Vivo, S. DiMauro, A novel mitochondrial ATPase 6 point mutation in familial bilateral striatal necrosis, Ann. Neurol. 38 (1995) 468–472.
- [12] C. Dionisi-Vici, S. Seneca, M. Zeviani, G. Fariello, M. Rimoldi, E. Bertini, L. De Meirleir, Fulminant Leigh syndrome and sudden unexpected death in a family with the T9176C mutation of the mitochondrial ATPase 6 gene, J. Inherit. Metab. Dis. 21 (1998) 2–8.
- [13] M. Makino, S. Horai, Y. Goto, I. Nonaka, Confirmation that a T-to-C mutation at 9176 in mitochondrial DNA is an additional candidate mutation for Leigh's syndrome, Neuromuscul. Disord. 8 (1998) 149–151.
- [14] C. Tzoulis, G.T. Tran, T. Schwarzlmüller, K. Specht, K. Haugarvoll, N. Balafkan, P.K. Lilleng, H. Miletic, M. Biermann, L.A. Bindoff, Severe nigrostriatal degeneration without clinical parkinsonism in patients with polymerase gamma mutations, Brain 136 (2013) 2393–2404.
- [15] P. McKelvie, B. Infeld, R. Marotta, J. Chin, D. Thorburn, S. Collins, Late-adult onset Leigh syndrome, J. Clin. Neurosci. 19 (2012) 195–202.
- [16] M.J. Molnár, A. Valikovics, S. Molnár, L. Trón, P. Diószeghy, F. Mechler, B. Gulyás, Cerebral blood flow and glucose metabolism in mitochondrial disorders, Neurology 55 (2000) 544–548.
- [17] M.J. Shelly, P. Kelly, M.J. O'Connell, FDG-PET imaging in the investigation of homonymous hemianopia in a patient with MELAS syndrome, Clin. Nucl. Med. 32 (2007) 479–480.
- [18] F.G. Lehnhardt, R. Horvath, R. Ullrich, L. Kracht, J. Sobesky, W. Möller-Hartmann, A.H. Jacobs, W.F. Haupt, Altered cerebral glucose metabolism in a family with clinical features resembling mitochondrial neurogastrointestinal encephalomyopathy syndrome in association with multiple mitochondrial DNA deletions, Arch. Neurol. 65 (2008) 407–411.
- [19] M. Ikawa, H. Okazawa, K. Arakawa, T. Kudo, H. Kimura, Y. Fujibayashi, M. Kuriyama, M. Yoneda, PET imaging of redox and energy states in stroke-like episodes of MELAS, Mitochondrion 9 (2009) 144–148.
- [20] M. Sekigawa, A. Okumura, S. Niijima, M. Hayashi, K. Tanaka, T. Shimizu, Autoimmune focal encephalitis shows marked hypermetabolism on positron emission tomography, J. Pediatr. 156 (2010) 158–160.
- [21] A. Echaniz-Laguna, M. Chassagne, J. de Sèze, M. Mohr, P. Clerc-Renaud, C. Tranchant, B. Mousson de Camaret, POLG1 variations presenting as multiple sclerosis, Arch. Neurol. 67 (2010) 1140–1143.
- [22] M. Slee, J. Finkemeyer, M. Krupa, R. Raghupathi, J. Gardner, P. Blumbergs, M. Agzarian, D. Thyagarajan, A novel mitochondrial DNA deletion producing progressive external ophthalmoplegia associated with multiple sclerosis, J. Clin. Neurosci. 18 (2011) 1318–1324.
- [23] M. Mancuso, D. Orsucci, E.C. Ienco, G. Ricci, G. Ali, A. Servadio, G. Fontanini, M. Filosto, V. Vielmi, A. Rocchi, L. Petrozzi, A. Logerfo, G. Siciliano, An "inflammatory" mitochondrial myopathy. A case report, Neuromuscul. Disord. 23 (2013) 907–910.
- [24] R. Kucharczyk, N. Ezkurdia, E. Couplan, V. Procaccio, S.H. Ackerman, M. Blondel, J.P. di Rago, Consequences of the pathogenic T9176C mutation of human mitochondrial DNA on yeast mitochondrial ATP synthase, Biochim. Biophys. Acta 1797 (2010) 1105–1112.
- [25] J. Houstek, A. Pícková, A. Vojtísková, T. Mrácek, P. Pecina, P. Jesina, Mitochondrial diseases and genetic defects of ATP synthase, Biochim. Biophys. Acta 1757 (2006) 1400–1405.
- [26] D.V. Krysko, P. Agostinis, O. Krysko, A.D. Garg, C. Bachert, B.N. Lambrecht, P. Vandenabeele, Emerging role of damage-associated molecular patterns derived from mitochondria in inflammation, Trends Immunol. 32 (2011) 157–164.
- [27] M.J. Koo, K.T. Rooney, M.E. Choi, S.W. Ryter, A.M. Choi, J.S. Moon, Impaired oxidative phosphorylation regulates necroptosis in human lung epithelial cells, Biochem. Biophys. Res. Commun. 464 (2015) 875–880.
- [28] M. Pasparakis, P. Vandenabeele, Necroptosis and its role in inflammation, Nature 517 (2015) 311–320.
- [29] D.Y. Fann, S.Y. Lee, S. Manzanero, S.C. Tang, M. Gelderblom, P. Chunduri, C. Bernreuther, M. Glatzel, Y.L. Cheng, J. Thundyil, A. Widiapradja, K.Z. Lok, S.L. Foo, Y.C. Wang, Y.I. Li, G.R. Drummond, M. Basta, T. Magnus, D.G. Jo, M.P. Mattson, C.G. Sobey, T.V. Arumugam, Intravenous immunoglobulin suppresses NLRP1 and NLRP3 inflammasome-mediated neuronal death in ischemic stroke, Cell Death Dis. 4 (2013), e790.