


# Safety and Efficacy of Novasil Clay (Calcium Montmorillonite) in Children Exposed to Aflatoxin in Ejura-Sekyedumase District in Ghana

Global Pediatric Health  
Volume 9: 1–9  
© The Author(s) 2022  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/2333794X221121243  
journals.sagepub.com/home/gph  


Justice Kumi, PhD<sup>1,2</sup> , Regina Appiah-Opong, PhD<sup>1</sup>, Dietrich Rein, PhD<sup>3</sup> ,  
Godfred Egbi, PhD<sup>1</sup>, Enoch Aninagyei, PhD<sup>4</sup> , Alex Boye, PhD<sup>5</sup>, and  
Desmond Omane-Acheampong, PhD<sup>2</sup>

## Abstract

**Background.** Aflatoxin levels are very high in animals and humans in places where cereals are poorly stored. In this study, Novasil was evaluated for safety and efficacy in children. **Methods.** Children (200) aged between 2 and 9 years were put into Novasil and placebo group. Participants received either 1.5 g of Novasil or calcium carbonate in their food. Urine samples were analyzed for AFM1 by HPLC, blood samples were assayed for complete blood count and chemistries. **Results.** Aflatoxin M1 levels in the Novasil treated group, significantly reduced to 60% compared to an increase of urine AFM1 in the placebo group. Hematological parameters did not change except for an increase in hemoglobin level in the Novasil group. Biochemical parameters remained unchanged except calcium ions. Glutathione levels in the Novasil increased, compared group to the placebo group. **Conclusion.** Novasil is safe, reduce aflatoxin bioavailability in humans while improving GSH antioxidant capacity as well.

The trial has been registered with Pan African Clinical Trial Registry ([www.pactr.org](http://www.pactr.org)). A WHO registry for clinical trials with a unique identification number PACTR202202797930675.

## Keywords

Children, Novasil, Aflatoxin, Ghana

Received June 23, 2022. Accepted for publication August 7, 2022.

## Introduction

Aflatoxins are toxic secondary metabolites that are produced by molds, mainly *Aspergillus flavus* and *A. parasiticus*. The fungi grow in soil, decaying vegetations and various food crops such as maize, peanuts, wheat and rice. The fungal growth and development of the toxin are facilitated by a combination of inappropriate food storage conditions and hot humid climates. The toxins subsequently contaminate cereals and animal products such as eggs, meat, and milk.<sup>1</sup>

Aflatoxins are mutagenic, cause immunosuppression and have severe detrimental effects on the human liver.<sup>2</sup> The most toxic aflatoxin compound, B<sub>1</sub>, can permeate the skin,<sup>3</sup> while the liver can convert the aflatoxin to a reactive epoxide intermediate or hydroxylated to form the less dangerous aflatoxin M1. Aflatoxin B1-8,

9-epoxide binds to reduced glutathione (GSH), which is essential in protecting tissues from the harmful effects of bioactivated aflatoxin.<sup>4</sup>

<sup>1</sup>University of Ghana, Noguchi Memorial Institute for Medical Research, Legon, Accra, Ghana

<sup>2</sup>University of Cape Coast, Department of Biomedical Science, Cape Coast, Ghana

<sup>3</sup>BASF SE Food Fortification, Human Nutrition, Ludwigshafen, Germany

<sup>4</sup>University of Allied Health Sciences, Department of Biomedical Sciences, Ho, Ghana

<sup>5</sup>University of Cape Coast, Department of Medical Laboratory Science, Cape Coast, Ghana

### Corresponding Author:

Desmond Omane-Acheampong, University of Cape Coast School of Allied Sciences, Department of biomedical Sciences, Cape Coast, Ghana.  
Emails: [dacheampong@ucc.edu.gh](mailto:dacheampong@ucc.edu.gh); [do.acheampong@gmail.com](mailto:do.acheampong@gmail.com)



One potential factor contributing to poor child health is increased exposure to aflatoxin-contaminated foods.<sup>5</sup> Cereal-based complementary foods for infants and young children may be contaminated with high levels of aflatoxin in Ghana.<sup>6,7</sup> Besides the hepatotoxic and immunosuppressive effect of aflatoxin, the toxin has also been found to be an anti-nutrient and has been associated with growth disorders in infants and children.<sup>8</sup> Thus, it is necessary to conduct intervention studies that present therapeutic strategies to prevent aflatoxin bioavailability in children. Due to its high cation exchange, large surface area, mechanical stability, and layered structure, clay has the potential to be a strong absorbent.<sup>9</sup>

The discovery of a healthy enterosorbent-based clay (Novasil), a natural clay containing calcium montmorillonite, presents an intervention strategy to reduce aflatoxin exposure and eradicate aflatoxicosis in animals and humans. Novasil reduces aflatoxin exposure by attracting molecules of positive charges due to the negative charge of the clay compound.<sup>10</sup> Human Novasil intervention trials in Ghanaian adults, previously recorded a reduction of aflatoxin bioavailability.<sup>11,12</sup> In a study by Mitchell et al.,<sup>13</sup> reported that Novasil inclusion in food was safe at a dose of 1.5 g/day and showed a 52% reduction in aflatoxin bioavailability compared to the placebo group in a 14-day safety and efficacy study in Ghanaian children. However, it is appropriate to broaden the scope of the study by Nicole and her group to assess the safety and efficacy of Novasil for a longer period in children. Therefore, we conducted the assessment for 60 days.

## Methodology

### *Study Design, Study Participants, and Study Period*

This study was done on children with detectable urinary aflatoxin in the Ejura-Sekyedumase District of the Ashanti Region of Ghana.

### *Participants' Selection*

The cluster sampling method was used to select qualified participants for the study. Ejura-Sekyedumase District was divided into 4 clusters (communities). A cluster was chosen using a simple random sampling technique, and all eligible subjects within households whose parents/guardians consented were included. Fifty-five (55) participants from each cluster were selected to obtain a total of 220 participants. Two hundred healthy children of both sexes (age: 2-9 years old) who fulfilled the inclusion criteria were selected for the

intervention study and assign to 2 groups of 100 each for placebo and Novasil treatment groups.

### *Experimental Design Framework and Participants Inclusion Criteria*

Figure 1 details the experimental framework of the study. Any child participant with urinary aflatoxin level of < 0.5 µg/kg was excluded from the study. Participants with detectable urinary aflatoxin (>0.5 µg/kg) were randomly placed on Novasil and placebo (calcium carbonate). Participants placed on either treatment arm were screened for full blood count, kidney and liver function tests, serum electrolyte assay (Na, K, Ca), urinary aflatoxin and GSH at baseline and at day 60.

### *Preparing the Novasil Clay for the Experiment*

Novasil used in this study was obtained from Engelhard/BASF Chemical Corporation (Jackson, MS, USA). The product was donated by Texas A&M University in the USA. Novasil clay was tested analogously for its effect on various environmental contaminants such as dioxins and heavy metals. The metals and dioxin analyses of Novasil were below the tolerable daily intake in accordance with the limit set up by the World Health Organization (WHO) and the Joint Food and Agriculture Organization/WHO Expert Committee on Food Additives. Novasil was sterilized by electron beam irradiation to prevent any possible bacterial or viral contamination before trial initiation.

### *Recruitment and Training of Field Assistants*

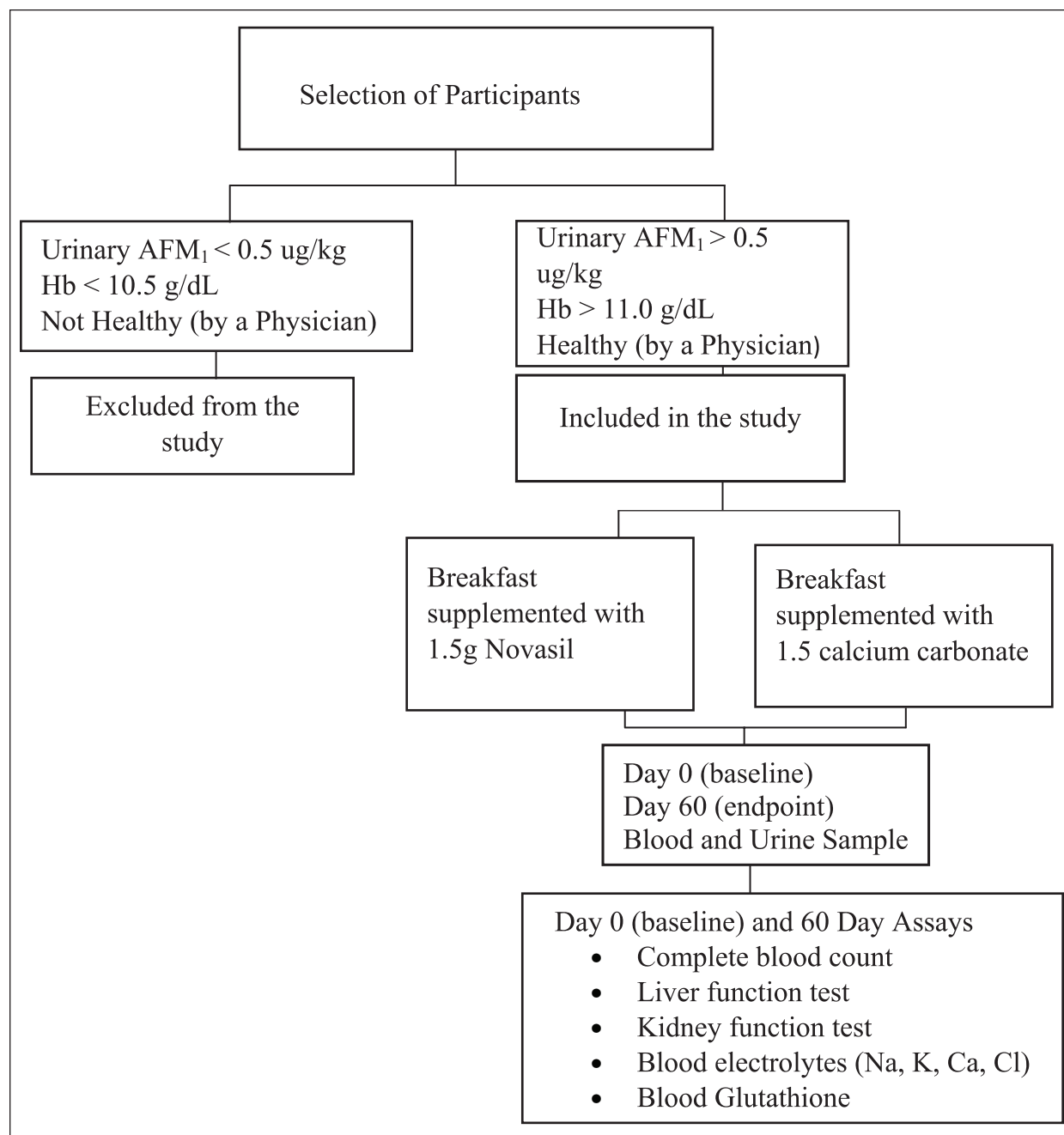
The study recruited 5 field assistants. The field assistants included a Physician, 2 Nurses and 2 Laboratory Scientists. They were trained on sampling and data collection by the Principal Investigator.

### *Sample Size Estimation*

The parameters for the sample size calculation were derived from a previous study which found that 52% of subjects treated with Novasil (drug A) was efficacious.<sup>13</sup> and a clinically significant difference of 19% when compared to a placebo is acceptable. Level of significance = 5%, Power = %, Type of test = two-sided.

The following formula was used for calculating the sample size:

$$n = \left[ (Z_{\alpha/2} + Z_{\beta})^2 \times \left\{ \frac{p_1(1-p_1) + p_2(1-p_2)}{(p_1 - p_2)^2} \right\} \right]$$



**Figure 1.** Experimental and sampling framework for the study.

Where:

$n$  = sample size required in each group,

$p_1$  = proportion of subject cured by Drug A = 0.52,

$p_2$  = proportion of subject perceived to be cured by Placebo = 0.33,

$p_1 - p_2$  = clinically significant difference = 0.19

The sample size required per group, according to the above formula, was 100. As a result, the total sample

size required was 200, using a 2-tailed z-test of proportions between 2 groups with 80% power and a 5% level of significance.

#### **Recruitment and Assigning Study Participants to Treatment Group**

Study participants were recruited from the Ejura-Sekyedumase district of the Ashanti Region of Ghana using the cluster sampling techniques. The community is a

rural area where residents are predominantly subsistence farmers. All recruited participants were between 2 and 9 years of age. Consent was obtained from the parents or legal guardians resident in the community. Consent documents were explained to each participant, and then signed by participant's guardian before initiation of the study. Participants were randomly assigned to the 2 arms of study, namely, treatment with Novasil or placebo group (treatment with calcium carbonate). The treatment arm consisted of 1.5 g of Novasil and the placebo group took 1.5 g of calcium carbonate. Both arms of the study received 1.5 g of Novasil or 1.5 g of placebo once daily. Doses were weighed into identical packages at the Noguchi Memorial Institute for Medical Research (NMIMR) to ensure that monitors and participants were blinded to their treatment. Trained study monitors mixed each participant's treatment into their normal maize porridge breakfast meal. Monitors visit participants each day to record any adverse effects and report to the study physician for treatment.

### **Sample Collection, Processing, and Analysis**

Three milliliters of baseline blood and urine samples and another at day 60 of treatment were collected from each child. Blood samples were collected by trained phlebotomists at the Ejura District Hospital while early morning urine samples were at day 0 and day 60 by trained monitors. Whole blood samples were used for hematological analysis whereas serum samples stored at  $-20^{\circ}\text{C}$ , were used for biochemical analysis. Urine samples were used for detecting aflatoxins.

### **Laboratory Analysis**

*Detecting of aflatoxin in the urine (AFM<sub>1</sub>).* Aflatoxin Vicam AflaTest kits were used according to the Association of Official Analytical Chemists' method (AOAC) for aflatoxin analysis. AflaTest columns were purchased from VICAM (Watertown, MA, USA). Analysis of urinary AFM<sub>1</sub> levels followed previously published method (Wang et al<sup>11</sup>). After the immunoaffinity column (AflatestWB; VICAM) clean up, high-pressure liquid chromatography (HPLC) system with fluorescence detection was used to analyze urine aflatoxin M<sub>1</sub> concentrations (Shimadzu Corp., Kyoto, Japan). Urinary aflatoxin concentrations were expressed as picograms per milligram creatinine to correct for variations in urine dilution among samples.

### **Determination of Blood Glutathione Levels**

The blood samples were homogenized and diluted 200X with GSH buffer before use. The reaction mixture

comprised 50  $\mu\text{L}$  of GSH buffer (0.1 M  $\text{NaH}_2\text{PO}_4$ , 5 mM EDTA, pH 8.0 with 1 N NaOH) with 50  $\mu\text{L}$  of the blood sample and 10  $\mu\text{L}$  of 0.75 mM O-Phthaldehyde (OPT). The mixture was incubated for 15 minutes at room temperature in the dark, and fluorescence was measured at 340 nm for excitation and 460 nm for emission.<sup>14</sup> The experiment was carried out in triplicate. The reduced glutathione levels were calculated using the calibration curve and expressed as mg/mL glutathione equivalent.

### **Complete Blood Count**

Complete blood count parameters analyzed included hemoglobin level (Hb), total white blood cell count (WBC) and total red blood cell count (RBC). White blood cell count analyzed were lymphocytes, neutrophils, monocytes, and eosinophils (Sysmex KX 21 hematology analyzer from Europe GmbH, Germany).

### **Kidney and Liver Biomarkers**

The levels of kidney and liver biomarkers in serum were analyzed using an automated colorimetric method with Flexor E, endpoint automation (Vital Scientific, Netherlands). Liver markers analyzed included alanine aminotransferase (ALT), aspartate aminotransferase (AST), albumin, gamma-glutamyl transferase (GGT), total protein and total bilirubin. Kidney markers analyzed include creatinine, urea, sodium, chloride, and potassium. Serum calcium was also measured.

### **Statistical Analysis**

Statistical analysis was conducted using Sigma Stat 3 (Jadel Scientific Software, 2003, USA). A descriptive analysis was employed to analyze for mean, standard deviation and the range of variables. Paired *T*-test was used to analyze AFM<sub>1</sub> data, full blood count, biochemical parameters for the placebo and treatment group. The Signed Rank test was used to analyze significance between data where it was not normalized among the intervention group and the placebo group. Statistical significance was defined as a 2-tailed *P*-value  $< .05$ .

### **Ethical Consideration/Informed Consent**

Ethical clearance for this study was obtained from the Institutional Review Board of the Noguchi Memorial Institute for Medical Research (NMIMR), University of Ghana (reference: IRB0001276) and the Institutional Review Board (IRB) of the University of Cape Coast (UCC) (reference: UCC/RB/CHAS/2019/109). Informed written consent were obtained from all participants in

**Table 1.** Characteristics of Study Participants.

Participants	Treatment group	Placebo group	P value	Range
Age (years)	5.6 ± 1.8	5.9 ± 1.9	.261	2 to 9
Gender				
Male (number)	50	48		
Female (number)	50	52		
Height (m)	1.1 ± 0.1	1.1 ± 0.2	.204	0.8 to 1.31
Weight-for-age z-score	-0.4 ± 0.9	-0.3 ± 1.0	.245	-2.35 to 1.9
Height-for-age z-score	-0.4 ± 1.4	-0.4 ± 1.3	.909	-3.6 to 2.5
Body weight (kg)	18.6 ± 5.0	19.4 ± 4.6	.250	10.3 to 40
BMI-for-age	-4.10 ± 1.46	-0.0001 ± 1.48	.05	-5.68 to -3.19

Data are represented as mean ± SD. All the data are baseline values. Significance:  $P < .05$ .

**Table 2.** Analysis of Hematology Parameters in the Treatment Groups.

Parameters	Novasil treatment group		Placebo group	
	Before	After	Before	After
Hemoglobin (g/dL)	11.6 ± 0.8	12.0 ± 0.8*	11.4 ± 0.7	11.3 ± 0.6
WBC (1000/mm <sup>3</sup> )	7.6 ± 2.1	7.5 ± 1.8	7.5 ± 1.7	7.3 ± 1.1
Platelets (1000/mm <sup>3</sup> )	336 ± 89.3	332.0 ± 100	310 ± 85.8	312.1 ± 75.2
Lymphocytes (%)	49.1 ± 10.0	49.0 ± 10.6	47.8 ± 12.2	48.9 ± 9.0
Monocytes (%)	9.4 ± 3.3	9.6 ± 3.2	8.6 ± 2.7	8.2 ± 2.3
Neutrophils (%)	37.1 ± 10.2	37.0 ± 10.9	35.9 ± 10.6	34.4 ± 8.7

Data is represented as mean ± SD.

\*Indicates an increased level of hemoglobin. N=200 before intervention, N= 198 at post intervention.

accordance with human experimental guidelines of the NMIMR (IRB) and UCC (IRB). Confidentiality was observed throughout the research process and the data made accessible only to the Principal Investigator and Study Supervisors.

## Results

Table 1 indicates a total of 200 children (98 males and 102 females) between the ages of 2 and 9 years who were enrolled into the study. Only 2 participants (one from treatment and one from placebo) could not give blood at the post-intervention stage. Also, 2 participants within the placebo group were diagnosed and treated for malaria. All the participants completed the study. Table 1 also shows the average z score analysis of participants. The average z score for weight-for-age for the treatment group was -0.4 SD with a range of -2.35 to 1.9 SD, while the placebo had an average z score for weight-for-age as -0.3SD with a range of -2.35 to 1.9 SD. The average height-for-age z score was -0.4SD in a range of -3.6 to 2.5 SD in the treatment group. The placebo group also showed an average z-score value of -0.4 for height-for-age in a range of -3.6 to 2.5 SD.

## Adverse/Side Effects of Novasil

There were no significant side effects recorded with respect to Novasil administration during the 60-day intervention period. No major variations in the number of side effects were found between the treatment and control groups ( $P = .746$ ) or differences in severity of symptoms reported ( $P = .700$ ).

## Analysis of Hematological Parameters

Table 2 shows an analysis of the hematology results obtained before and after each treatment group. With the exception of hemoglobin and platelets levels that were elevated after Novasil treatment, no other hematological parameter was significantly altered after both Novasil and placebo treatment.

## Effect of Novasil and Calcium Carbonate on Liver and Kidney Parameters

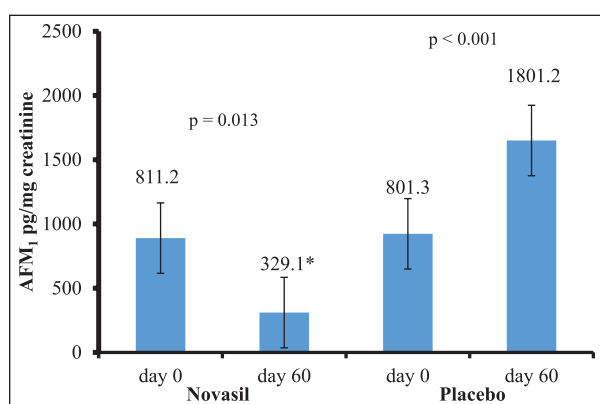
Decrease levels in blood GGT levels were observed after Novasil treatment. In the placebo group, ALT was significantly elevated. Notwithstanding these observations,

**Table 3.** Analysis of serum liver and kidney injury biomarkers.

Biochemical parameters	Novasil treatment group		Placebo group	
	Before	After	Before	After
Albumin(g/L)	45.3 ± 5.9	45.5 ± 2.7	45.3 ± 5.2	44.6 ± 4.0
ALT(U/L)	20.1 ± 7.2	21.4 ± 6.0	20.1 ± 7.8	22.5 ± 7.6 <sup>b</sup>
AST(U/L)	28.3 ± 10.9	29.0 ± 9.4	28.2 ± 15.3	30.0 ± 13.6
GGT(U/L)	19.8 ± 10.1	19.0 ± 9.4 <sup>a</sup>	21.0 ± 8.1	20.8 ± 7.2
T. Bil (µmol/L)	7.4 ± 3.4	7.6 ± 4.3	8.0 ± 4.7	8.3 ± 4.3
T. Protein (g/L)	75.8 ± 9.7	76.1 ± 6.1	76.3 ± 5.4	75.4 ± 4.4
Urea (mmol/L)	3.1 ± 0.9	3.0 ± 0.8	3.2 ± 0.5	3.1 ± 0.5
Creat. (µmol/L)	59.3 ± 7.0	60.5 ± 8.1	61.7 ± 6.5	60.6 ± 6.9
Sodium (mmol/L)	135.4 ± 6.1	135.9 ± 5.3	132 ± 8.3	133 ± 10.0
Potassium (mmol/L)	3.6 ± 0.4	3.7 ± 0.5	3.4 ± 0.4	3.4 ± 0.4
Chloride (mmol/L)	102.0 ± 7.2	101.8 ± 4.5	103 ± 7.9	104.6 ± 7.4
Calcium (mmol/L)	2.2 ± 0.1	2.3 <sup>a</sup> ± 0.2	2.2 ± 0.1	2.2 ± 0.1

Data is showed as mean ±SD.

<sup>a</sup>Indicates decrease levels within the treatment group, while <sup>b</sup> indicates increase in the placebo group. N=200 before intervention, N=198 at post intervention.

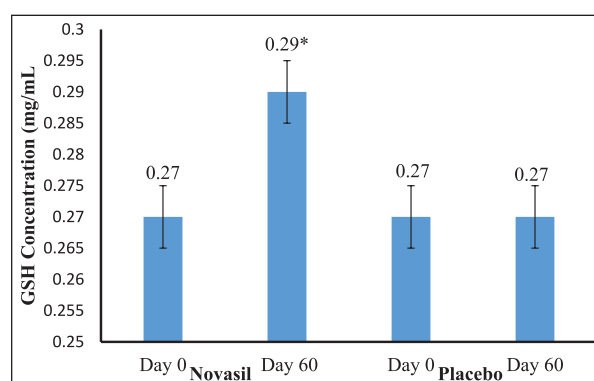


**Figure 2.** Mean AFM<sub>1</sub> levels in pre- and post-treatment groups. \*significant reduction

all other Day-60 parameters did not significantly differ from baseline levels (Table 3).

### Analysis of Blood Glutathione and Urinary Aflatoxin M<sub>1</sub>

Baseline mean urinary aflatoxin M<sub>1</sub> levels in the treatment group (AFM<sub>1</sub>=811.2pg/mg creatinine) and the placebo group (AFM<sub>1</sub>=801.3pg/mg creatinine) were statistically not different ( $P>.05$ ). However, after a 60-day Novasil treatment, mean urinary level of aflatoxin significantly reduced to 329.1pg/mg creatinine ( $P=.013$ ) representing 60% reduction. On the other hand, Day-60 mean urinary level of aflatoxin increased to 1801.2pg/mg creatinine increased within the placebo group representing 125% increase (Figure 2).



**Figure 3.** Mean GSH levels in pre- and post-treatment groups. \*increased

Whereas urinary levels of aflatoxin reduced in the treatment group, mean serum levels of GSH increased from 0.27mg/mL (baseline) to 0.29mg/mL (Day-60) representing 7.4% increase in blood GSH. However, the mean baseline and mean Day-60 levels of GSH in the placebo group remained unchanged (Figure 3).

### Discussion

Aflatoxin-related health effects are widespread especially in developing countries that rely on food commodities such as cereals as their primary source of food. Aflatoxin exposure during infancy, through breast milk and complementary foods, may result in growth impairment, immune suppression and micronutrient deficiencies.<sup>15,16</sup> Over the past years, high levels of aflatoxin exposure have been detected in people



from the Ejura Sekyedumase District in the Ashanti Region of Ghana.<sup>11,12,17,18</sup>

Weanimix, made from groundnuts, beans and maize, is a popular semi-solid food used to wean children between the ages of 2 and 9 years in Ghana. The use of weanimix is intended to correct malnutrition in children. However, weanimix has been found to be heavily contaminated with aflatoxin in 2 studies done in the Ejura Sekyedumase District.<sup>7,18</sup> It was not surprising to realize that close to 38% of children between the ages of 6 and 59 months were found to be stunted in the Atwima Kwanwoma District in the Ashanti region of Ghana<sup>19</sup> a district in the same region as this study district. It is noteworthy that farmers in the Ejura Sekyedumase District are leading producers of maize, beans and peanuts in Ghana.<sup>20</sup> When cereals are contaminated with aflatoxins, some of the suggested harmful effects on humans are vomiting, diarrhea, hepatitis, cirrhosis, immunosuppression, miscarriages, mutagenic and teratogenic effects, and different cancers.<sup>21</sup> One of the most fundamental approaches to lowering the risk of aflatoxicosis in humans is to avoid eating foods contaminated with aflatoxin. However, many communities in developing countries simply are poor and ignorant about aflatoxin poisoning. As a result, strategies to reduce aflatoxin poisoning especially in children are required particularly in aflatoxin endemic areas. This will help to reduce the occurrence of aflatoxin related disease in children. Therefore, this study aimed at evaluating the safety and efficacy of Novasil in children who were at risk of developing aflatoxin related diseases in Ghana.

The findings of this study show that giving healthy children (ages 2-9) dietary Novasil powder at a concentration of 1.5 g/day for 60 days was safe and also reduced the bioavailability of aflatoxin metabolite in urine. These findings are consistent with other studies in Ghana and elsewhere.<sup>11-13,22,23</sup>

Novasil exhibited binding capacity to aflatoxin by reducing urinary aflatoxin levels by 60% while increasing the glutathione levels in the children. Novasil clay has been found to have high affinity for aflatoxin due to the opposite charges they possess. Due to the electronegativity of the Novasil clay, it has high binding capacity in the gastrointestinal tract for aflatoxin.<sup>10</sup> Novasil has been found to absorb aflatoxin in both animal models and human studies, reducing the bioavailability of the blood toxin.<sup>17</sup> Additionally, the use of Novasil on humans did not affect the utilization of vitamins and trace elements in the body.<sup>24,25</sup>

Glutathione, a potent antioxidant is critical in protecting tissues from the damaging effects of toxicants such

as bioactivated aflatoxin.<sup>4</sup> In an animal study, aflatoxin induced reduction in the level of blood GSH.<sup>26</sup> The present study observed an increase in blood GSH levels in the treatment group at the end of the Novasil intervention. This study also confirms the potential cytoprotective nature of Novasil. The systemic safety of Novasil has been demonstrated in this study. Novasil did not significantly alter the blood levels of WBC and its subcells, lymphocytes, monocytes and neutrophils. Platelets levels were also not affected however, at day 60 of Novasil use, hemoglobin levels were increased. With the exception of calcium levels that were elevated, all liver and kidney parameters did not change after the 60-day treatment period. The efficacy of Novasil has been previously determined in human and animal models.<sup>13,27</sup> Elevation of total serum Ca<sup>2+</sup> levels post-Novasil treatment supports the studies by Afriyie-Gyawu et al<sup>17</sup>, and Marroquín-Cardona et al.<sup>28</sup> The elevation could be due to the breakdown of calcite and exchangeable Ca<sup>2+</sup> from montmorillonite clay as previously explained<sup>17,28</sup> Since Novasil clay commonly acts as a calcium supplement, it was likely that the increase in total serum calcium levels seen in the treatment group in this study was due to the addition of Novasil to the food. Calcium levels in the placebo group did not see any change in levels although the placebo group also received calcium carbonate. Possibly due to the lack of supplementation in calcium levels in the placebo group. However this development needs to be further investigated.

## Conclusion

This study has shown that the addition of Novasil to cereal-based weaning foods is beneficial in reducing aflatoxin bioavailability in children. This has the potential to reduce the negative effects of aflatoxin exposure. Therefore, we recommend that a longer safety and efficacy trial be established to ascertain whether Novasil could reduce urinary aflatoxin level to less than 0.5 µg/kg. Furthermore, it is essential to evaluate the impact of Novasil on nutrients, growth and anthropometric indicators for improvement in children's health in a longer study. Furthermore, a longer safety trial is necessary to establish whether Novasil could affect micronutrient absorption, while controlling for critical dietary nutrient intake. The study showed that levels of aflatoxin exposure in children at Ejura-Sekyedumase continues to be a major problem. Thus, parents of the study participants were educated on the dangers of aflatoxin exposure to the development of their children. Parents were encouraged to feed them on uncontaminated cereals.

## Acknowledgment

We appreciate the unmeasurable support from BASF SE food fortification, Human Nutrition Germany, for providing financial support for the field work. We are grateful to Ghana Education Trust Fund (GETFUND) for providing financial sponsorship for the academic work. To parents who voluntarily agreed for their children to be part of the study we say thank you. We sincerely appreciate the children who took their time to be part of the research, and also the kindness of the Nurses and Laboratory Scientist who helped in data and sample collection. To Dr. Seidu Zakari of Ejura-Sekyedumase Hospital, we say thank you.

## Author Contributions

Justice Kumi, contributed to the conception and design, drafted the manuscript.

Regina Appiah-Opong, contributed to the design and critically reviewed manuscript.

Dietrich Rein, contributed to interpretation and critically reviewed manuscript.

Godfred Egbi, Contributed to conception and manuscript review. Enoch Aninagyei, contributed to analysis and critically reviewed manuscript.

Alex Boye, contributed to interpretation and manuscript review.

Desmond Omane-Acheampong, contributed to conception and design, critically reviewed manuscript.

## Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: The author(s) received financial support from BASF SE Food Fortification, Human Nutrition, Ludwigshafen, Germany for the research.

## Ethical Approval

Ethical Approval for the study was obtained from the Institutional Review Board of the Noguchi Memorial Institute for Medical Research (NMIMR), University of Ghana (reference: IRB0001276) and the Institutional Review Board (IRB) of the University of Cape Coast (UCC) (reference: UCC/RB/CHAS/2019/109).

## Consent

Informed written consent were obtained from all participants through their parents or guardian in accordance with human experimental guidelines of the NMIMR (IRB) and UCC (IRB). Confidentiality was observed throughout the research process

and the data made accessible only to the Principal Investigator and Study Supervisors.

## ORCID iDs

Justice Kumi  <https://orcid.org/0000-0002-5929-2322>

Dietrich Rein  <https://orcid.org/0000-0003-0076-1865>

Enoch Aninagyei  <https://orcid.org/0000-0002-6489-167X>

## References

1. Fratamico PM, Bhunia AK, Smith JL. *Foodborne Pathogens: Microbiology and Molecular Biology*. Horizon Scientific Press; 2008.
2. Benkerroum N. Chronic and acute toxicities of aflatoxins: mechanisms of action. *Int J Environ Res Public Health*. 2020;17(2):423. doi:10.3390/ijerph17020423
3. Boonen J, Malysheva SV, Taevernier L, Diana Di, Mavungu J, De Saeger S, De Spiegeleer B. Human skin penetration of selected model mycotoxins. *Toxicology*. 2012;301(1-3):21-32. doi:10.1016/j.tox.2012.06.012
4. Iyer RS, Coles BF, Raney KD, Thier R, Guengerich FP, Harris TM. DNA adduction by the potent carcinogen aflatoxin B1: mechanistic studies. *J Am Chem Soc*. 1994;116(5):1603-1609. doi:10.1021/ja00084a001
5. Gong YY, Cardwell K, Hounsa A, et al. Dietary aflatoxin exposure and impaired growth in young children from Benin and Togo: cross sectional study. *BMJ*. 2002;325(7354):20-21. doi:10.1136/bmj.325.7354.20
6. Blankson GK, Mill-Robertson FC. Aflatoxin contamination and exposure in processed cereal-based complementary foods for infants and young children in greater Accra, Ghana. *Food Control*. 2016;64:212-217. doi:10.1016/j.foodcont.2015.12.032
7. Kumi J, Mitchell NJ, Asare GA, et al. Aflatoxins and fumonisins contamination of home-made food (Weanimix) from cereal-legume blends for children. *Ghana Med J*. 2014;48(3):121-126. doi:10.4314/gmj.v48i3.1
8. Gong Y, Hounsa A, Egal S, et al. Post-weaning exposure to aflatoxin results in impaired child growth: a longitudinal study in Benin, West Africa. *Environ Health Perspect*. 2004;112(13):1334-1338. doi:10.1289/ehp.6954
9. Sheng G, Dong H, Li Y. Characterization of diatomite and its application for the retention of radiocobalt: role of environmental parameters. *J Environ Radioact*. 2012;113:108-115. doi:10.1016/j.jenvrad.2012.05.011
10. Phillips TD, Wang M, Elmore SE, Hearon S, Wang JS. NovaSil clay for the protection of humans and animals from aflatoxins and other contaminants. *Clays Clay Miner*. 2019;67(1):99-110. doi:10.1007/s42860-019-0008-x
11. Wang P, Afriyie-gyawu E, Tang Y, et al. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis: II. Reduction in biomarkers of aflatoxin exposure in blood and urine. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*. 2008;25:622-634. doi:10.1080/02652030701598694



12. Mitchell NJ, Kumi J, Johnson NM, et al. Reduction in the urinary aflatoxin M1 biomarker as an early indicator of the efficacy of dietary interventions to reduce exposure to aflatoxins as an early indicator of the efficacy of dietary interventions to reduce exposure to aflatoxins. *Biomarkers*. 2013;18:391-398. doi:10.3109/1354750X.2013.798031
13. Mitchell NJ, Ankrah NA, Zychowski KE, et al. Short-term safety and efficacy of calcium montmorillonite clay (UPSN) in children. *Am J Trop Med*. 2014;91(4):777-785. doi:10.4269/ajtmh.14-0093
14. Hissin PJ, Hilf R. A fluorometric method for determination of oxidized and reduced glutathione in tissues. *Anal Biochem*. 1976;74(1):214-226.
15. Wild CP. Aflatoxin exposure in developing countries: the critical interface of agriculture and health. *Food Nutr Bull*. 2017;28(2 Suppl):S372-S380. doi:10.1177/15648265070282S217
16. Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. Human aflatoxicosis in developing countries: a review of toxicology, exposure, potential health consequences, and interventions. *Am J Clin Nutr*. 2004;80(5):1106-1122.
17. Afriyie-Gyawu E, Mackie J, Dash B, et al. Chronic toxicological evaluation of dietary NovaSil clay in Sprague-Dawley rats. *Food Addit Contam*. 2005;22(3):259-269. doi:10.1080/02652030500110758
18. Kumi J, Dotse E, Asare G, et al. Urinary aflatoxin M1 exposure in Ghanaian children weaned on locally prepared nutritional food. *Afr J Sci Res*. 2015;4(6):28-32.
19. Delle F. *Malnutrition among children 6-59 months in the Atwima Kwanwoma District, Ashanti Region-Ghana*. Unpublished master's thesis, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana, 2016.
20. Cudjoe GP, Antwi-Agyei P, Gyampoh BA. The effect of climate variability on maize production in the Ejura-Sekyedumase Municipality, Ghana. *Climate*. 2021;9:145. doi:10.3390/cli9100145
21. Sarma UP, Bhetaria PJ, Devi P, Varma A. Aflatoxins: Implications on Health. *Indian J Clin Biochem*. 2017;32:124-133. doi:10.1007/s12291-017-0649-2
22. Awuor AO, Yard E, Daniel JH, et al. Evaluation of the efficacy, acceptability and palatability of calcium montmorillonite clay used to reduce aflatoxin B1 dietary exposure in a crossover study in Kenya. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*. 2017;34(1):93-102.
23. Afriyie-Gyawu E, Ankrah NA, Huebner HJ, et al. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis. I. Study design and clinical outcomes. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*. 2008;25(1):76-87. doi:10.1080/02652030701458105
24. Mayura K, Abdel-Wahhab MA, McKenzie KS, et al. Prevention of maternal and developmental toxicity in rats via dietary inclusion of common aflatoxin sorbents: potential for hidden risks. *Toxicol Sci*. 1998;41(2):175-182. doi:10.1006/toxs.1997.2405
25. Phillips TD, Afriyie-Gyawu E, Williams J, et al. Reducing human exposure to aflatoxin through the use of clay: A review. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*. 2008;25(2):134-145. doi:10.1080/02652030701567467
26. Ankrah NA, Sittie A, Addo PG, Ekuban FA. Enhanced depletion of glutathione and increased liver oxidative damage in aflatoxin-fed mice infected with *Plasmodium berghei*. *Trans R Soc Trop Med Hyg*. 1995;89(1):59-61. doi:10.1177/030006059402200306
27. Phillips TD, Kubena LF, Harvey RB, Taylor DR, Heidelbaugh ND. Hydrated sodium calcium aluminosilicate: A high affinity sorbent for aflatoxin. *Poult Sci*. 1988;67(2):243-247. doi:10.3382/ps.0670243
28. Marroquin-Cardona A, Deng Y, Garcia-Mazcorro JF, et al. Characterization and safety of uniform particle size Novasil clay as a potential aflatoxin enterosorbent. *Appl Clay Sci*. 2011;54(3-4):248-257. doi:10.1016/j.clay.2011.09.009