

The temporal specificity of BOLD fMRI is systematically related to anatomical and vascular features of the human brain

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Abstract

The ability to detect fast responses with functional MRI depends on the speed of hemodynamic responses to neural activity, because hemodynamic responses act as a temporal low-pass filter smoothing out rapid changes. However, hemodynamic responses (their shape and timing) are highly variable across the brain and across stimuli. This heterogeneity of responses implies that the temporal specificity of fMRI signals, or the ability of fMRI to preserve fast information, should also vary substantially across the cortex. In this work we investigated how local differences in hemodynamic response timing impact the temporal specificity of fMRI. We conducted our research using ultra-high field (7T) fMRI at high spatiotemporal resolution, using the primary visual cortex (V1) as a model area for investigation. We used visual stimuli oscillating at slow and fast frequencies to probe the temporal specificity of individual voxels. As expected, we identified substantial variability in temporal specificity, with some voxels preserving their responses to fast neural activity more effectively than others. We investigated which voxels had the highest temporal specificity and related those to anatomical and vascular features of V1. We found that low temporal specificity is only weakly explained by the presence of large veins or cerebral cortical depth. Notably, however, temporal specificity depended strongly on a voxel's position along the anterior-posterior anatomical axis of V1, with voxels within the calcarine sulcus being capable of preserving close to 25% of their amplitude as the frequency of stimulation increased from 0.05-Hz to 0.20-Hz, and voxels nearest to the occipital pole preserving less than 18%. These results indicate that detection biases in high-resolution fMRI will depend on the anatomical and vascular features of the area being imaged, and that these biases will differ depending on the timing of the underlying neuronal activity. Importantly, this spatial heterogeneity of temporal specificity suggests that it could be exploited to achieve higher specificity in some locations, and that tailored data analysis strategies may help improve the detection and interpretation of fast fMRI responses.

Keywords: neurovascular coupling, fast fMRI, high resolution, 7T, visual cortex, oscillations

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11 1. Introduction

12 Blood-oxygenation-level-dependent (BOLD) functional MRI (fMRI) is the most commonly used imaging tech-
13 nique for the non-invasive study of cognitive processes in the human brain because of its submillimeter and
14 subsecond resolution. Many ongoing advancements in fMRI techniques have increased the spatial resolution
15 and sensitivity of BOLD fMRI ([Blazejewska et al., 2019](#); [Dumoulin et al., 2018](#); [Mareyam et al., 2020](#); [Polimeni et al., 2015](#); [Viessmann and Polimeni, 2021](#) and other references therein). However, improving temporal resolution
16 is often considered more challenging. The temporal constraints of BOLD functional imaging are ultimately deter-
17 mined by the inherent biological timescales of neurovascular coupling and the resulting hemodynamics ([Buxton
18 et al., 2014](#)) Understanding the temporal precision of the BOLD response is therefore important for extracting
19 precise timing information about the neuronal activity of interest from fMRI data.

20 Although the neuronal activity of interest often happens at millisecond scales, BOLD fMRI only measures
21 the subsequent hemodynamic changes that unfold on the scale of seconds. Common expectations based on
22 hemodynamic response models developed in the early days of fMRI ([Glover, 1999](#)) suggested that neurovascular
23 coupling is slow, yielding BOLD responses expected to be below ~ 0.15 Hz. However, resting-state fMRI studies
24 have demonstrated meaningful spatial structure in high-frequency BOLD signals ([Boubela et al., 2013](#); [Chen
25 and Glover, 2015](#)). Furthermore, task-based fMRI studies have shown that the BOLD responses can track
26 external stimuli with frequencies as high as 0.75 Hz ([Lewis et al., 2016](#)), demonstrating that hemodynamic
27 responses to neuronal activity can be surprisingly faster than previously believed. Imaging fast BOLD responses
28 could open a window to probe rapidly evolving neuronal activity, such as those associated with decision-making,
29 language-processing, attention, and sensory-motor integration ([Polimeni and Lewis, 2021](#)). However, a deeper
30 understanding of the temporal properties of hemodynamic responses is needed to analyze and interpret these
31 fast signals.

32 A challenge for identifying precise timing information in fMRI is that the temporal properties of hemodynamic
33 responses are highly heterogeneous across the brain ([Bailes et al., 2023](#); [Handwerker et al., 2004](#); [Pfeuffer et al., 2003](#)) and across experimental contexts ([Chen et al., 2021](#); [Friston et al., 1998](#); [Handwerker et al., 2012](#)).
34 BOLD responses are influenced by the vascular architecture of the cortex: they are weaker but have an earlier
35 onset in the parenchyma, and stronger in the large draining veins that collect large volumes of deoxygenated
36 blood from multiple capillary beds ([Turner, 2002](#)). Since the vascular architecture is organized such that veins
37 drain upwards towards the pial surface ([Duvernoy et al., 1981](#)), this effect is not only visible when comparing
38 parenchyma and venous responses ([Gati et al., 1997](#); [Kay et al., 2019](#); [Lai et al., 1993](#); [Siero et al., 2009](#);
39 [Uludağ and Blinder, 2018](#); [de Zwart et al., 2005](#)), but by proxy also when comparing responses across cortical
40 depths ([Siero et al., 2014](#); [Siero et al., 2011](#)) given that responses at the pial surface are more likely to include
41 larger veins. Since hemodynamic properties act as a filter on the underlying neuronal activity, they thus determine
42 the timing and amplitude of measurable BOLD responses: areas with slower, sluggish hemodynamic responses
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44

45 should produce correspondingly slow signals, whereas sharper and faster responses should be able to robustly
46 track high-frequency signals. The intrinsic temporal resolution of fMRI should therefore vary across the brain
47 depending on local hemodynamic timing.

48 In addition to modulating the temporal precision of fMRI, this vascular heterogeneity can introduce detection
49 biases (Polimeni et al., 2018) in BOLD responses that becomes increasingly more pronounced as the spatial
50 and temporal resolution of imaging is increased. These biases can have profound effects depending on an fMRI
51 study's experimental design. For studies with slow task designs, small timing differences can sometimes be neg-
52 ligible since they can be accounted for during statistical analysis—using a GLM, for example—by modelling not
53 only the response predicted by a convolution with a standard HRF, but also by modelling its temporal derivatives,
54 or using flexible modelling approaches. However, for rapid task designs (e.g. event-related or naturalistic stim-
55 ulus designs), these biases can lead to several issues, such as undetected responses (false negatives), spatial
56 errors when creating topographic maps; and misinterpretation of the BOLD signal when comparing responses to
57 stimuli with different timings. Any biases that are imparted by vascular anatomy will likely impact distinct cortical
58 regions differently (Handwerker et al., 2004), and indeed differences in hemodynamic response delays on the
59 order of seconds and large response amplitude differences have also been observed (Amemiya et al., 2020).

60 Here, we investigated how the temporal precision of BOLD fMRI signals varies across the human visual
61 cortex, to test whether vascular or other anatomical features may determine, in part, where fast responses can be
62 detected. We used ultra-high field fMRI (7 Tesla) at high spatiotemporal resolution (1.06 mm isotropic resolution
63 with 0.874 s temporal resolution) to measure responses to rapidly varying visual stimuli across cortical depths
64 and close to large veins. Investigating these fast responses at high spatiotemporal resolution requires careful
65 experimental considerations: high-frequency (> 0.20 Hz) responses to rapidly fluctuating stimuli are typically too
66 small in amplitude to measure at the single-voxel level, demanding an impractical amount of trial-averaging for
67 reliable detection above the noise floor, hindering a direct analysis of local response properties at the single-voxel
68 level. For this reason, we used 0.20 Hz as our fastest stimulus frequency and analyzed the frequency response
69 of each voxel to infer how well it can preserve fast information. We defined a metric for "temporal specificity",
70 by calculating the amplitude at faster frequencies relative to the amplitude at a slower reference frequency (in
71 this study, 0.05 Hz). This "temporal specificity" metric represents how well a voxel preserves information about
72 responses to fast stimuli (as compared to slow stimuli). For sharp HRFs, the amplitude at fast frequencies is
73 expected to be higher, since sharp HRFs preserve more high-frequency information. Based on prior work on
74 the relationship between responses and vascular-anatomical features (Siero et al., 2011; de Zwart et al., 2005),
75 we hypothesized that deep layers of cortical gray matter would have increased temporal specificity since they
76 have earlier responses, and for the same reason parenchyma would have increased specificity when compared
77 to large veins. We found that oscillatory stimulation elicited strong single-voxel BOLD responses across all
78 frequencies tested, allowing us to robustly stratify responses according to cortical depth, vascular compartment,
79 and anatomical position along V1. Our results demonstrated that temporal specificity differences were weakly

80 related to cortical depth and vascular compartment yet varied strongly along the anterior-posterior axis of V1.
81 Importantly, our results indicate that fast BOLD responses will be more readily detectable in regions with higher
82 temporal specificity, which is spatially structured and linked to brain anatomy.

83 **2. Methods**

84 *2.1. Subject population*

85 All subjects provided informed written consent, and all procedures were approved by Massachusetts General
86 Hospital's Institutional Review Board. A total of 16 subjects with previous MRI experience were recruited and
87 scanned, and data from 12 subjects (age 26.6 ± 3.2 years, 6F/6M) were analyzed. Subjects were excluded if
88 motion was larger than 2.5 mm throughout the session, or if the behavioral performance on the behavior task
89 (explained below) in any run was found below 75%. Three subjects were excluded for motion and one for not
90 completing the full experiment.

91 *2.2. MRI Data Acquisition*

92 Participants were scanned on a 7 T Siemens (Magnetom "Classic", Erlangen, Germany) whole-body scan-
93 ner with a custom-built 32-channel head coil array and birdcage head coil for transmit. Each session began
94 with a 0.75-mm isotropic dual-echo MPRAGE ([Van der Kouwe et al., 2008](#)) as an anatomical reference with the
95 following parameter values: TR = 2530 ms, TI = 1100 ms, TE = 1.74 and 3.7 ms, flip angle = 7° , outer-loop ac-
96 celeration = 2 and no partial Fourier, bandwidth=651 Hz/px, and a duration of 7:20 minutes. After the anatomical
97 reference scan, eleven BOLD functional runs were acquired using a single-band single-echo gradient-echo 2D
98 EPI protocol. Coronal slices were positioned over the occipital pole targeting the calcarine sulcus. The protocol
99 had following parameter values: 16 slices with 1.06-mm isotropic resolution (R = 4 in-plane acceleration and
100 no partial Fourier), TR = 874 ms, TE = 24 ms, flip angle = 52° , matrix size = 164×164 , bandwidth = 1452
101 Hz/px, nominal echo spacing 0.81 ms, phase-encoding direction = Left \rightarrow Right. Each functional run collected
102 300 volumes for a total acquisition time of 257 s. Between the 4th and 5th run a larger-FOV EPI protocol with
103 40 slices of same resolution was acquired to aid the registration between the small-FOV BOLD data and the
104 full-brain anatomical scan. Between the 7th and 8th run a 3-minute inversion-recovery EPI ([Renvall et al., 2016](#))
105 was acquired, but not used in this analysis.

106 *2.3. Visual stimulus*

107 Visual stimuli were presented using a DLP projector (Psychology Software Tools), with timing synchronized
108 to the 60-Hz refresh rate of the stimulus delivery computer, onto a screen placed at the back of the scanner bore.
109 Subjects viewed the stimulus through an angled mirror placed above their eyes. Stimulus presentation code
110 was written in Lua using the Löve 2D framework (<https://love2d.org>), and timing accuracy was checked

111 against a 120-Hz iPhone 12 camera by filming the presentation outside of the scanner as it would be displayed
112 in the scanner and comparing its timing to the experimental design. The visual stimuli consisted of counterphase
113 flickering radial “checkerboards” presented continuously, beginning 14 s after the start of the first volume of
114 the scan. The luminance contrast of the checkerboards was sinusoidally modulated at a frequency of interest
115 throughout the runs as previously described (Lewis et al., 2016). For the experiment, eleven functional runs
116 were acquired: 2 with luminance modulated at 0.05 Hz, the first of which was used as a functional localizer, 3
117 modulated at 0.10 Hz, and 6 modulated at 0.20 Hz. The order of the stimuli was set to [0.05, 0.20, 0.10, 0.20,
118 0.20, 0.10, 0.20, 0.05, 0.20, 0.10, 0.20] Hz to avoid order effects. Multiple runs were acquired for averaging.
119 During visual stimulation runs, subjects performed a simple visual fixation task: a red dot at the center of the
120 screen alternated between light and dark red with switch times drawn from a uniform distribution between 0.8
121 and 3 s; subjects were instructed to press a button on an MR-compatible USB button box every time the dot
122 changed color.

123 2.4. Data Preprocessing

124 2.4.1. Anatomical preprocessing

125 As described in (Van der Kouwe et al., 2008), multiple echoes of the MP-RAGE acquisition were combined
126 using root sum of squares before any further processing.

127 1. Bias field correction

128 Anatomical images were then bias field corrected using the joint bias-field and class segmentation esti-
129 mation provided by SPM 12 (<https://www.fil.ion.ucl.ac.uk/spm/software/spm12/>). The bias field
130 correction requires setting two parameters: the smoothness of the bias-field, and the bias regularization.
131 These parameter values have a strong impact on the trade-off between estimating the bias field or the
132 segmented tissue classes. Because we were interested in the portion of the visual cortex covered by
133 the 16-mm FOV where contrast between gray and white matter is subtle, we chose conservative param-
134 eters that penalize large variations in the bias field (`biasreg = 0.15`, `biasfwhm = 20`). This choice opts
135 to underestimate the true bias field but reduces the chance of inadvertently misclassifying the GM/WM
136 intensity contrast as local bias field. These values were chosen after visual inspection of the bias field and
137 reconstructed cortical surfaces when preprocessing data.

138 2. Cerebral cortical surface reconstruction

139 Cortical surfaces were automatically reconstructed after bias-field correction using FreeSurfer 7.1.1 (<https://surfer.nmr.mgh.harvard.edu>). Because V1 intensities at 7T are higher than average brain gray mat-
140 ter values, `recon-all` was run with flags `-seg-wmlo` and `-seg-grayhi` set to 95 and 105, respectively.
141 This reduces the likelihood of surfaces being incorrectly placed in the middle of the GM ribbon. The brain
142 mask generated by `recon-all` was also refined by removing areas which the SPM bias-field and segmen-
143 tation algorithm had already classified as belonging to bone or other tissues. This was done after noticing
144

145 that the automatically generated brain mask was at times too relaxed and did not fully remove the skull nor
146 sagittal sinus.

147 *2.4.2. Functional preprocessing*

148 1. Slice time and motion correction

149 Slice time correction was performed using `filtershift`, interpolating the data to the middle slice of each
150 volume (Parker et al., 2017). Motion correction was performed subsequently using AFNI's `3dVolReg` via
151 FreeSurfer's `mc-afni2` wrapper.

152 2. Within-subject inter-run coregistration and run averaging

153 All functional runs were co-registered to each subject's functional localizer run. Transformations were
154 estimated using ANTs (<https://github.com/ANTsX/ANTs>) in a hierarchical approach whereby first a
155 rigid transform was estimated and used as input to estimate a similarity (rigid + scaling) transform. This
156 was in turn used to estimate an affine transform which was used to co-register runs. (This approach is
157 customary and recommended within the ANTs documentation to improve accuracy.) Co-registered runs
158 were visually inspected, and runs of the same stimulus frequency were averaged using `fs1maths`.

159 3. Registration to anatomical reference

160 A transformation between the functional localizer space and the anatomical space was computed to assign
161 cortical depth estimates and anatomical ROI labels to each voxel in the functional run, using information
162 from the FreeSurfer cortical surface reconstruction and parcellation in anatomical space. A transformation
163 between the functional localizer space and the anatomical space was computed in two steps. The first
164 step was identical to the inter-run coregistration using ANTs, but registers the localizer run to the larger-
165 FOV reference EPI (functional ↔ large FOV). The second step co-registers the larger-FOV reference to
166 the anatomical data using FreeSurfer's boundary-based registration (larger-FOV ↔ anatomical) (Greve
167 and Fischl, 2009). These two transforms were then concatenated to generate a mapping from functional
168 localizer space to anatomical space (functional ↔ anatomical).

169 4. Voxelwise cortical depth estimates

170 Once a transformation between functional and anatomical space had been established, it was possible to
171 obtain the distance from the centroid of each voxel in functional space to the pial and white matter surfaces
172 computed in anatomical space. Depth values are given in millimeters from the centroid of a voxel to the
173 white matter surface (Polimeni et al., 2018, 2010). A value of 0 represents a voxel exactly at the WM/GM
174 interface, and higher values represent distances upwards into the GM and CSF. These values can be
175 normalized by the cortical thickness to obtain normalized cortical depths ranging from 0 (WM/GM interface
176 or "white surface") to 1 (GM/CSF interface or "pial surface").

177 5. Laminar smoothing

178 Finally, because of the small amplitude of oscillatory responses in the parenchyma, especially at the 0.20-
179 Hz condition, a small amount of “laminar” smoothing (Blazejewska et al., 2019) was performed using
180 LAYNII’s LAYERSMOOTH (Huber et al., 2021). For this, the gray matter ribbon was divided into 4 depths and
181 smoothing was applied within voxels of the same depth using a gaussian filter with a full-width at half-max
182 of 1.5mm.

183 2.5. Data Analysis

184 2.5.1. Defining a localizer ROI to estimate oscillatory responses

185 A localizer ROI was defined for each subject based on both anatomical and functional considerations. The
186 mask was created based on the following criteria: (1.) only voxels located within anatomically defined V1; and (2.)
187 above-threshold positive activation in the functional localizer run. For this, first a V1 anatomical mask was cre-
188 ated by registering the V1 prediction (i.e., the surface based `V1_exvivo` label) from FreeSurfer to each subject’s
189 functional space. The mask resampling was performed with `antsApplyTransforms` using ITK’s GenericLa-
190 bel interpolator with linear interpolation (Schaerer et al., 2014). Second, an activation mask was generated by
191 thresholding Z (Gaussianized F) statistics at $Z > 3.5$ from activation maps from the averaged 0.05-Hz runs. Ac-
192 tivation maps were obtained using FSL FEAT (<https://fsl.fmrib.ox.ac.uk/fsl/fslwiki/FEAT>) (Woolrich
193 et al., 2001), and using as regressors a sine and cosine waveform with the frequency of the localizer run, 0.05
194 Hz, and performing an F-test on the response to both sinusoid regressors. The analysis was run after discarding
195 the initial 60 volumes of the acquisition, to minimize not only T1 transient magnetization effects but also transient
196 hemodynamic effects related to the onset of the continuous stimulation. A high-pass filter with a cutoff of 1/25 s
197 was included in the GLM for localizer definition, but high-pass filtered data were not used in any further analysis to
198 avoid biasing results towards any frequency (Simon and Buxton, 2015) (results did not qualitatively change when
199 high-pass filtering was used). No additional regressors were added as nuisance components. The mask was
200 constrained to only include positive BOLD responses: the delay of all voxels from the 0.05-Hz localizer run was
201 estimated (estimation described below) and a mask was defined from all voxels whose onset delays were within
202 1–6 seconds after the stimulus. This interval was chosen based on our estimates of delays from conventional
203 HRFs which suggest that delays should be well within that range. Negative BOLD responses were expected to
204 be out of phase and thus have delays outside of that window. The localizer ROI mask was thus defined as the
205 intersection of the anatomically defined V1 mask and the functionally-defined positive BOLD mask.

206 Finally, since more voxels were expected to be active at the pial surface of GM than in deeper GM due to
207 the larger amplitude of pial-vein BOLD signals (Polimeni et al., 2010), the mask was further refined by only
208 keeping voxels that were active at the pial surface that had a corresponding voxel active deep within the gray
209 matter ribbon. To do that we used the cortical surface mesh vertex coordinates that represent cortical geometry:
210 because FreeSurfer establishes a one-to-one vertex correspondence between the white and pial surfaces, it is
211 straightforward to identify voxels intersecting the white surface that correspond to any given intersecting the pial

212 surface; this allowed voxels to be considered in pairs at both the inner and outer boundaries of the gray matter
213 ribbon. All active voxels not intersecting the cortical ribbon (occasional false positives), or where only a single
214 voxel of the pair was active, were removed to avoid an analysis biased towards surface voxels. Thus, the final
215 functional ROI was created from the localizer ROI using this criterion.

216 *2.5.2. Estimating delay, amplitude and temporal specificity of oscillatory responses*

217 To estimate delay and amplitude of the BOLD responses to visual stimulation, the time-series of each voxel
218 in each averaged functional run (one per frequency per subject) was interpolated to a 100-ms grid using cubic
219 interpolation. Subsequently percent signal changes were calculated within each voxel by subtracting the mean
220 and dividing the result by the mean and multiplying that value by 100. All oscillation cycles within each frequency
221 were then averaged to yield a cycle-average response for each voxel at each frequency. Finally, the amplitude
222 of the oscillation was defined as the peak-to-through magnitude of this cycle-averaged response. The delay
223 was defined as the time of the trough of the response. The temporal specificity was defined as the ratio of the
224 amplitude of responses at different stimulus frequencies. For example, the amplitude ratio “0.20 Hz/0.05 Hz”
225 means the measured oscillation amplitude of a voxel to the 0.20-Hz stimulus divided by the estimated amplitude
226 to the 0.05-Hz stimulus condition. This metric thus captured the ability of individual voxels to preserve their
227 response magnitude to fast stimuli.

228 *2.5.3. Excluding false-positive voxels with a control analysis*

229 As a control analysis to remove “false-positive” voxels that appeared to be responding to the stimulus but were
230 doing so by chance, all data were re-analyzed as if the underlying stimulus had a frequency of 0.19 Hz. This
231 resulted in a null distribution for amplitudes for each stimulus condition (0.05Hz, 0.10Hz and 0.20Hz). Voxels that
232 had a true response amplitude at any frequency below the 50% of the null-distribution for that frequency were
233 excluded from the analysis (we chose 50% inspired by the mixture modelling approaches used by FSL Melodic
234 (Beckmann and Smith, 2004) to separate noise from signal, which uses 0.5 as a threshold to equally balance
235 false-positives and false-negatives). This control excluded $3.0 \pm 2.6\%$ of voxels within the localizer mask for each
236 subject, such that the final localizer masks contained 2907 ± 1129 voxels for each subject.

237 *2.5.4. Classifying voxels into veins and parenchyma*

238 Voxels were classified as veins if the amplitude of measured oscillations was within the top 10% of all mea-
239 sured responses, subject to two constraints: (1.) the responses were in voxels in the pial surface, and (2.) the
240 inverse tSNR of the voxel was below a threshold of 0.05 (Zhang et al., 2009), to avoid the accidental inclusion
241 of bright voxels unlikely to be veins but that also displayed strong oscillations. This thresholding approach was
242 visually inspected and compared against a high-resolution susceptibility-weighted image (SWI) acquired during
243 pilot sessions in separate participants (data not shown).

244 *2.5.5. Estimating the spatial spread of delays across V1*

245 For each subject, the x , y and z coordinates of the V1 mask, in functional space, were mean centered and
246 converted to millimeters. This yielded a position estimate from the center of V1, in millimeters, for each voxel
247 within the mask. Since our imaging was performed such that slices were positioned parallel to the anterior-
248 posterior axis of V1, we used the y -coordinate estimates as a proxy for the location along the anterior-posterior
249 axis of V1.

250 *2.5.6. Statistical analysis*

251 Statistical analysis was conducted using the R programming language (<https://www.r-project.org>) and
252 the `brms` package (<https://cloud.r-project.org/web/packages/brms>). In particular, the `brm` function was
253 used to fit Bayesian linear mixed models to the data to: (1.) estimate the effect of anatomical correlates on the
254 observed amplitude, delay and temporal specificity of responses, and (2.) to estimate whether the amplitude
255 and delay of responses at slow stimuli are related to observed amplitudes at fast stimuli. Weakly informative
256 Gaussian priors were used by default and four Markov chains were run, each with 2000 iterations (discarding the
257 first 1000 as warm-up), resulting in 4000 samples to approximate the posterior distribution of model parameters.
258 The convergence of the Markov chains was assessed by computing the \hat{R} statistic (Gelman-Rubin diagnostic)
259 for each parameter. All chains for all models run in the current manuscript fully converged, as indicated by \hat{R}
260 statistics for all parameters almost identical to 1.

261 *2.6. Simulating voxels with fast and slow frequency response*

262 In order to simulate single-voxel frequency responses and compare them to experimental data, we simulated
263 a range of HRFs modelled by a sum of two Gamma functions by sweeping across a range of peak delays (ranging
264 from 3 to 6.5 s) and full width at half maximum (FWHM) (ranging from 2.25 – 3.75s). For each combination
265 of peak delay and FWHM, an HRF was sampled at 10-ms intervals, the result of which was then convolved
266 with stimuli of different frequencies to generate a frequency response amplitude curve. We also simulated the
267 frequency response of the canonical HRF (as described by (Glover, 1999)) and two HRFs that approximate the
268 impulse responses for the pial and parenchymal visual cortex (as shown by (Siero et al., 2011)).

269 **3. Results**

270 *3.1. Oscillatory stimuli enable measuring the temporal specificity of voxels in V1*

271 To measure how well voxels in V1 can preserve high-frequency signals, we measured BOLD responses
272 in the visual cortex induced by sinusoidally modulated visual stimuli at either slow or fast frequencies. This
273 design allowed us to directly measure the ability of the vasculature to respond to increasingly faster stimuli,
274 estimating the frequency response of each individual voxel. Oscillatory stimuli also have other advantages, such
275 as minimizing neuronal nonlinearities at stimulus onset (Grill-Spector et al., 2006) and improving the separation

276 between stimulus-evoked responses from background fluctuations (Kalatsky and Stryker, 2003). They are also
277 simpler to analyze (Regan, 1966), since delays and amplitude can be unambiguously defined, whereas the
278 onset and peak of transient responses, such as in blocked or event-related designs, are usually defined ad
279 hoc and harder to compare across conditions. From the amplitude of oscillatory responses to different stimulus
280 frequencies, we defined a “temporal specificity” metric, as a proxy of how much high frequency information is
281 preserved in response to fast stimuli (Fig. 1).

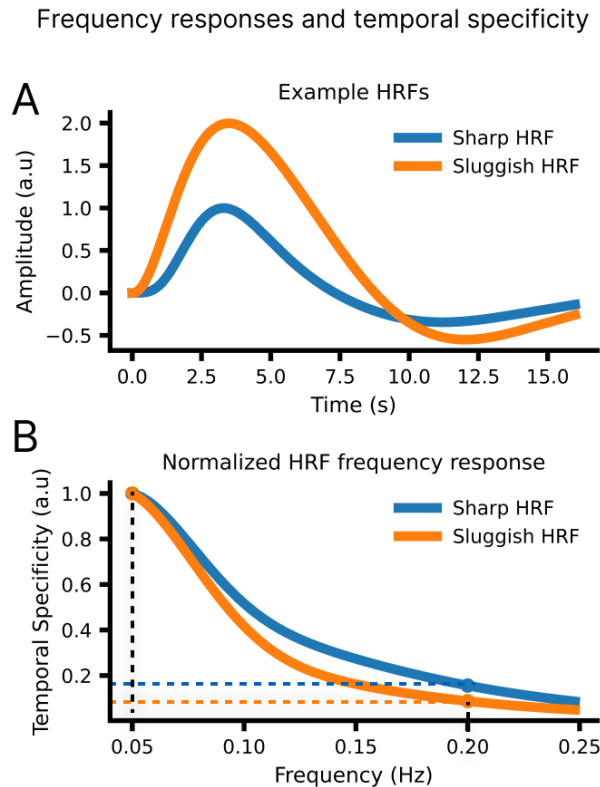


Figure 1: **Temporal specificity is estimated by calculating the ratio of response amplitudes at different stimulus frequencies.** A. A simulation of two example HRFs, one that is slow and has low amplitude (Sluggish HRF) and one that is fast and has a high amplitude (Sharp HRF). B. The frequency response is normalized to the reference response to 0.05-Hz stimulation. The resulting frequency response can be used to compare which HRF produces relatively larger amplitudes at fast frequencies. This simulation illustrates that sharper HRFs should yield higher values of “temporal specificity”, i.e. with relatively high-amplitude responses at high frequencies.

282 3.2. High spatio-temporal resolution fMRI detects BOLD responses at the single-voxel level up to at least 0.20Hz

283 Since we aimed to investigate whether BOLD responses are linked to single-voxel vascular and anatomical
284 properties, we first tested whether we could reliably identify BOLD fMRI responses to our oscillatory stimuli (Fig,
285 2, panel A) at the single-voxel level. We examined BOLD timeseries from all active voxels (Fig. 2, panel B), and
286 the oscillatory patterns seen in individual voxels clearly demonstrated that responses were visible at the single-
287 voxel level up to at least 0.20 Hz. From each voxel time-series, we calculated the mean cycle-locked response

288 and extracted the amplitude and delay of the mean response for each voxel (Fig. 2, panel C). We observed
 289 a large heterogeneity in response properties across voxels, even at the single-subject level (Fig. 2, panel D),
 290 demonstrating the diversity of hemodynamic responses uncovered with high resolution imaging.

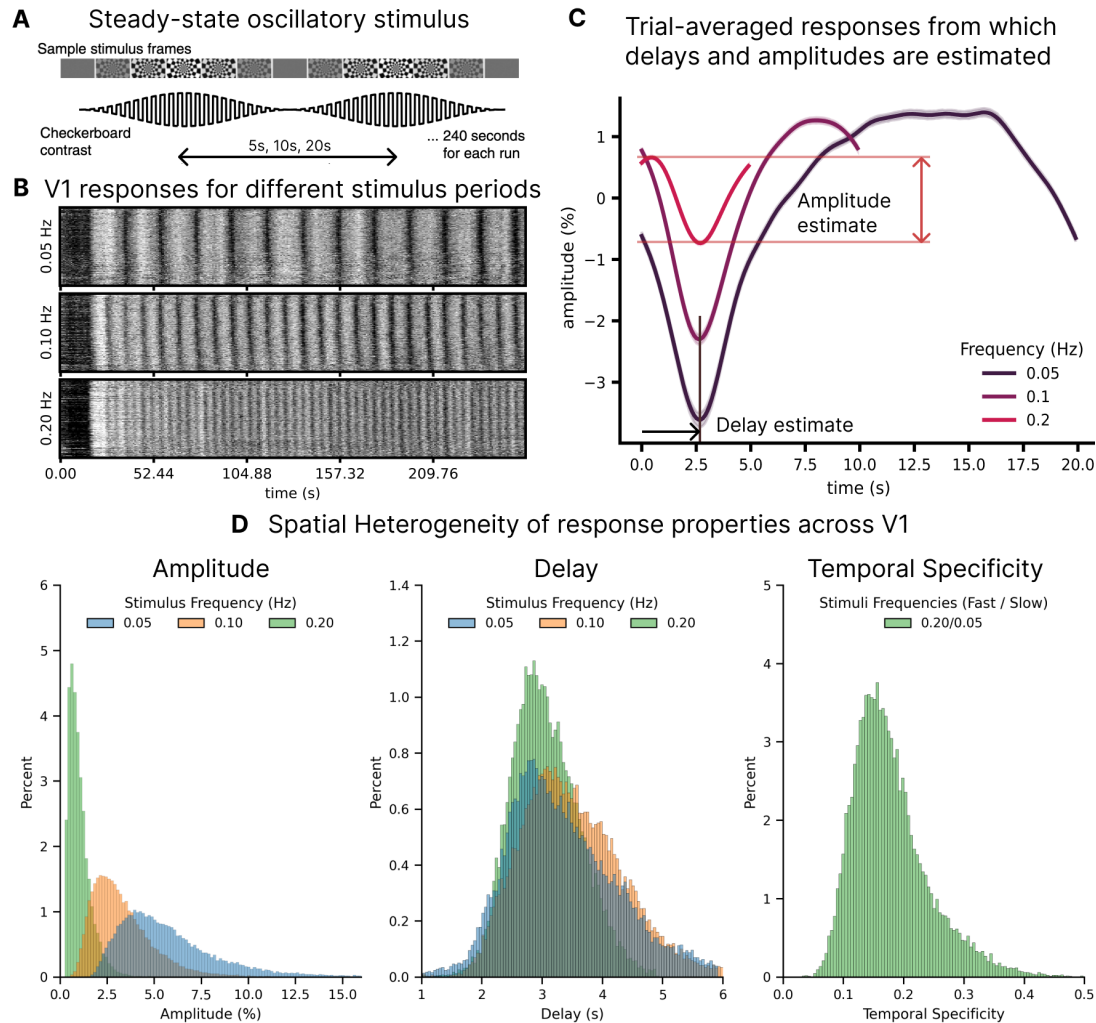


Figure 2: **Single-voxel oscillatory responses can be reliably measured and show large heterogeneity.** A. Continuous stimulus used to study the relationship between delays and hemodynamic variability: a flickering checkerboard with luminance contrast sinusoidally modulated with different frequencies. B. Carpet plots illustrating responses in the V1 ROI in an example subject, for three different frequencies. Each row represents a voxel within the localizer mask. C. Trial-averaged responses for the three stimuli in an example subject. Arrows indicate how delays and amplitudes are estimated. D. Histograms of the amplitude, delay and temporal specificity of responses show large heterogeneity even at the single-subject level.

291 **3.3. Anatomical features influence the spatial patterns of delays and amplitudes and temporal specificity**

292 Having extracted the oscillatory response in individual voxels, we next aimed to test how these responses
 293 were related to anatomical features within V1. We examined whether the estimated delay, amplitude and tem-

294 poral specificity of single-voxel responses was linked to vascular-anatomical properties: cortical depth, vascular
295 compartment (vein vs. parenchyma) and position along V1. We first inspected maps to compare the spatial
296 patterns of anatomy and vasculature against maps of delay and amplitude estimated for the response to different
297 stimulus frequencies (Fig. 3, panel A). Measured patterns aligned with previously reported BOLD properties
298 (Koopmans et al., 2010; Siero et al., 2011, 2009): response amplitude and delay increased towards the pial
299 surface, and responses were substantially larger in veins when compared to the parenchyma. We also observed
300 a gradient of response properties across V1, with delay maps closely following the position along the anterior-
301 posterior axis of V1. To quantify these spatial patterns, we conducted a Bayesian linear mixed effects analysis,
302 testing whether anatomical predictors (cortical depth, position along the anterior-posterior axis of V1, and vascu-
303 lar compartment) significantly predicted fMRI response properties (delay, amplitude and temporal specificity in
304 each voxel). We assumed that the absolute influence of cortical depth and position would vary for each subject
305 (because of e.g. different cortical thickness or orientation of V1 with respect to B0 for each individual), thus
306 modelling those as random effects. The model was fit against data from each frequency separately since the
307 frequency effect is not expected to be linear.

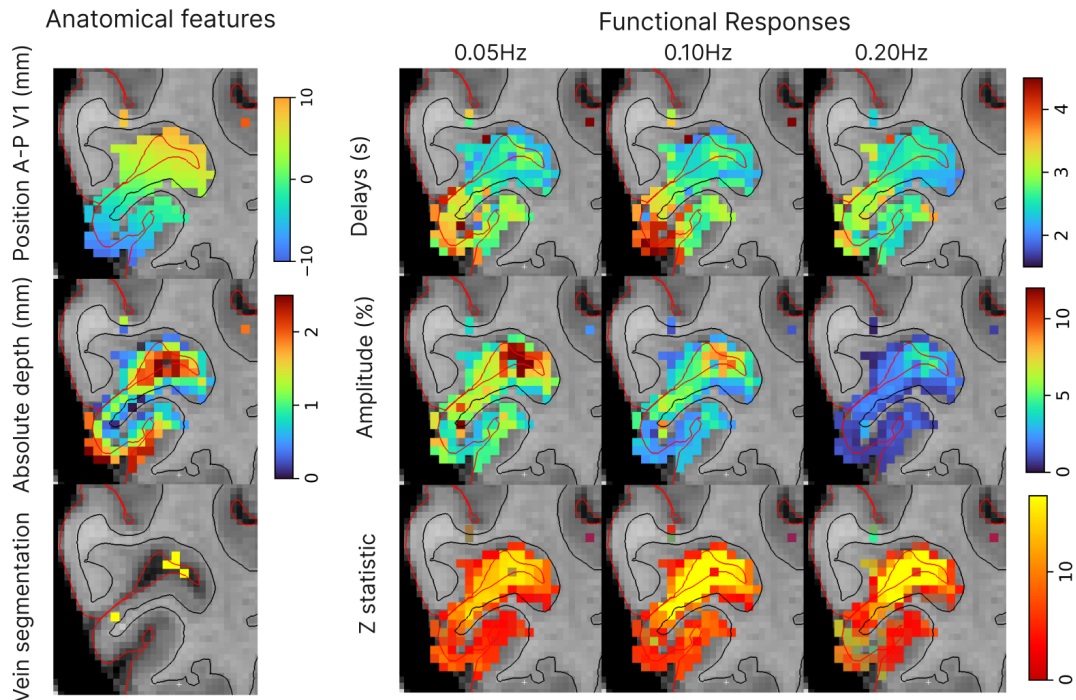
308 *3.3.1. Effect of anatomical covariates on hemodynamic delays*

309 Using these model estimates, we first examined how each anatomical feature related to the delay of the fMRI
310 responses. The effect of cortical depth on delay was similar for all frequencies, with delay increasing from the
311 white to the pial surface by 170 ms/mm on average. The presence of a vein was associated with an additional
312 215 ms delay, beyond the effect of cortical depth. The magnitude of the effect of vascular compartment on
313 delay was also consistent across different stimulus frequencies (Table 1). The anatomical position was also a
314 significant predictor, with delays increasing towards posterior V1 (from the calcarine sulcus towards the occipital
315 pole), however, this relationship showed a dependence on stimulus frequency, with weaker effects at the faster
316 frequencies (from 42 ms/mm at 0.05 and 0.10 Hz to 22 ms/mm at 0.20 Hz, Table 1). These results demonstrated
317 that the depth, vascular anatomy, and position of voxels each contributed to its response delay.

318 *3.3.2. Effect of anatomical covariates on response amplitude*

319 We next investigated the spatial pattern of response amplitudes, investigating which voxels had the largest
320 responses. Amplitudes dropped substantially when using faster stimuli, as expected from prior work (Lewis et
321 al., 2016): the mean amplitude at the white matter at the center of V1 was: 4.38% (95% CI: 3.76 to 5.00%) for the
322 0.05-Hz stimulus, 2.42% (95% CI: 2.00 to 2.83%) for the 0.10-Hz stimulus, and 0.74% (95% CI: 0.62 to 0.86%)
323 for the 0.20-Hz stimulus. The response amplitudes were highly variable, and some voxels showed extremely
324 large responses; notably, individual single-voxel responses reached as high as 60.84% for one participant, with
325 the largest voxelwise amplitudes averaging $33.56\% \pm 16.25\%$ across all 12 participants. Response amplitudes
326 increased substantially as a function of cortical depth and were heavily influenced by the presence of a vein with

A Example maps of anatomical and vascular features, and functional responses



Delays, amplitude and specificity as a function of anatomical correlates

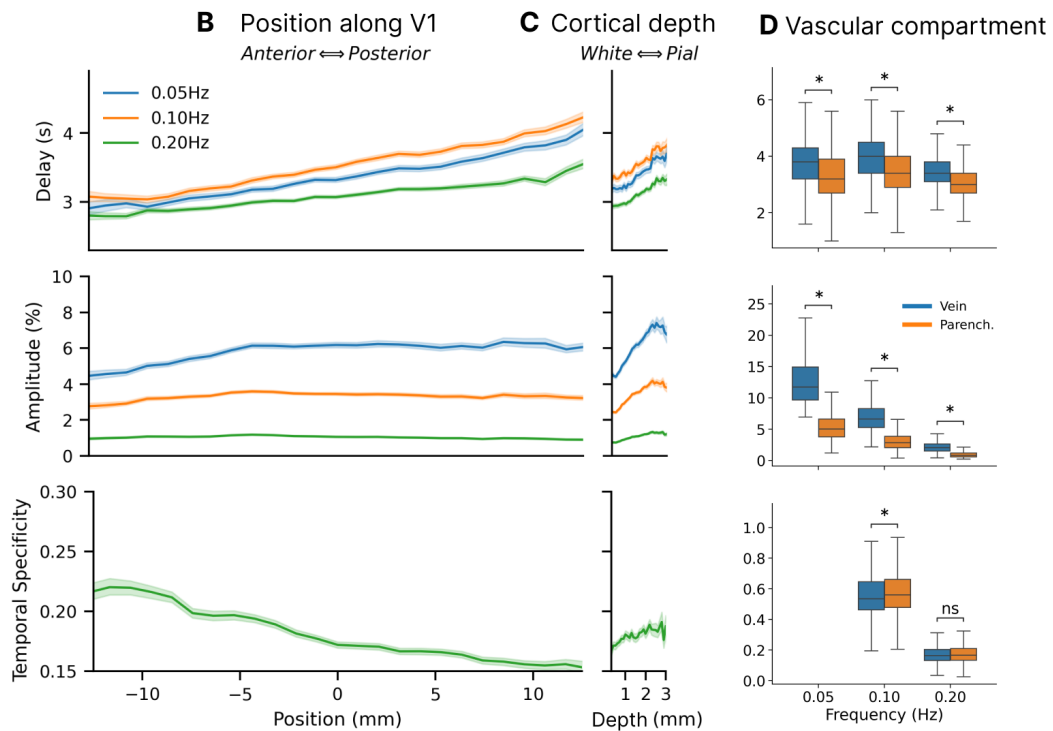


Figure 3: The amplitude, delay and temporal specificity of voxelwise responses are spatially structured. A. (left) Example maps of the three anatomical features used in this study: position along the A–P axis of V1, cortical depth and vein segmentation; (right) maps of derived delay, amplitude and F/Z-statistic maps obtained across the three different stimulus frequencies. Maps are clearly spatially structured. B. Plots of delay, amplitude and temporal specificity (0.05Hz/0.20Hz) as a function of position along V1 (left) and cortical depth (right). Note that x-axis are on the same scale. C. Box plots of delay, amplitude and temporal specificity in veins and parenchyma for the three frequencies imaged. Stars indicate that differences were statistically significant (mixed-effects model, see Table 1).

Table 1: **Mixed-effects model identifies strong effects of anatomical features on single-voxel fMRI responses.** Weight estimates for delay, amplitude and specificity, for different stimulus frequencies as a function of cortical depth, position in V1, vascular compartment. Values in brackets represent 95% CI. (All temporal specificity values and gradients are multiplied by 100 for clarity.)

Effect of anatomical correlates on response onset delay with respect to stimulus presentation

Frequency	Delay at WM (ms)	WM → CSF (ms/mm)	Anterior → Posterior (ms/mm)	Impact of vein (ms)
0.05 Hz	3110 [2740, 3470]	179 [123, 236]	42 [28, 57]	238 [206, 269]
0.10 Hz	3300 [2670, 3010]	178 [127, 230]	42 [33, 52]	208 [181, 236]
0.20 Hz	2840 [2670, 3010]	156 [103, 209]	22 [15, 30]	204 [182, 227]

Effect of anatomical correlates on signal amplitude

Frequency	Amplitude at WM (%)	WM → CSF (%/mm)	Anterior → Posterior (%/mm)	Impact of vein (%)
0.05 Hz	4.38 [3.76, 5.00]	0.77 [0.37, 1.18]	0.04 [0.02, 0.07]	6.99 [6.88, 7.10]
0.10 Hz	2.42 [2.00, 2.83]	0.45 [0.27, 0.61]	0.00 [-0.01, 0.01]	3.74 [3.68, 3.81]
0.20 Hz	0.74 [0.62, 0.86]	0.14 [0.07, 0.22]	-0.007 [-0.012, -0.001]	1.19 [1.16, 1.21]

Effect of anatomical correlates on temporal specificity (multiplied by 100)

Frequency	100 × specificity at WM	WM → CSF (mm ⁻¹)	Anterior → Posterior (mm ⁻¹)	Impact of vein
0.10 Hz	55.07 [50.96, 59.33]	1.04 [0.05, 2.05]	-0.44 [-0.27, -0.62]	-1.06 [-1.61, -0.50]
0.20 Hz	16.88 [15.29, 18.52]	0.39 [-0.06, 0.86]	-0.25 [-0.34, -0.17]	-0.12 [-0.34, 0.11]

327 substantial amplitude increases (Table 1). This is consistent with prior studies observing the largest responses
328 at the cortical surface and veins (Siero et al., 2011, 2009). Importantly, amplitudes were strongest in veins for all
329 stimulus conditions (Figure 3, panel D).

330 While response amplitudes were consistently related to cortical depth and compartment, the effect of position
331 differed qualitatively across stimulus frequencies. For the 0.05-Hz stimulus, the amplitude significantly increased
332 towards the posterior part of V1 by 0.04%/mm (95% CI: 0.02 to 0.07%) being largest closer to the occipital pole.
333 For the 0.10-Hz frequency, no significant trend was observed in the amplitude as a function of position (0.00%,
334 95% CI: -0.01 to 0.01%). For the 0.20 Hz frequency, the amplitude significantly decreased towards the posterior
335 part of V1 by 0.007% (95% CI: 0.001 to 0.012%) being largest deep into the calcarine sulcus (Figure 3, panels
336 B and C). This reversal effect under different stimulus frequencies is unlikely to be explained by neuronal effects,
337 since the amplitude of neuronal activity is not expected to invert across stimulus frequencies (Lewis et al., 2016).
338 Instead, this reversal of the spatial pattern is likely caused by varying hemodynamic and vascular properties
339 within V1, suggesting that regions with fast responses may be spatially structured in such a way as to yield
340 distinct spatial maps under different stimulus frequencies.

341 3.3.3. Effect of anatomical covariates on temporal specificity

342 Finally, to test our hypothesis that the temporal precision of functional MRI is also spatially structured, we in-
343 vestigated the spatial structure of temporal specificity. A temporal specificity value of 0 indicates a zero amplitude
344 at the fast frequency, and a value of 1 signifies that the response amplitude remains constant across slow and
345 fast frequencies. Higher values imply voxels with relatively stronger responses to the faster stimulus, indicating
346 that they more effectively retain temporal information and are thus more “temporally specific”.

347 We found no significant effect of cortical depth on temporal specificity, with a change of $+0.0039 \text{ mm}^{-1}$ (95%
348 CI: -0.0006 to 0.0086), a small number that suggests voxels sampling the pial surface preserves only about 0.4%
349 more amplitude at 0.20-Hz compared to 0.05-Hz than those that sample the white matter. We point out, however,
350 that the magnitude of the effect of cortical depth should be interpreted with care. We found no statistically
351 significant linear effect of cortical depth, but cortical depth could have a more nuanced effect on specificity. The
352 trend in these data suggested that temporal specificity may increase slightly towards the pial surface until depths
353 of about 1 mm (our imaging resolution), an effect likely related to partial volume effects, but then flatten out
354 (Fig. 3, panel B). This suggests that the effect of cortical depth on specificity is not strictly linear, and also that
355 comparing temporal specificity differences between “deep” and “superficial” bins of the cortical ribbon may lead
356 to statistically significant results based on where the threshold is placed that defines whether a voxel is in the
357 deep or superficial bin.

358 We next examined the effects of veins, and found that voxels with veins had moderately lower temporal
359 specificity in the 0.10-Hz stimulus (reduction of -0.0106 (95%CI: -0.0161 to -0.0050)), and a similar trend was
360 seen in the response to the 0.20-Hz stimulus (reduction of -0.0012 (95%CI: -0.0034 to 0.0011)). These results

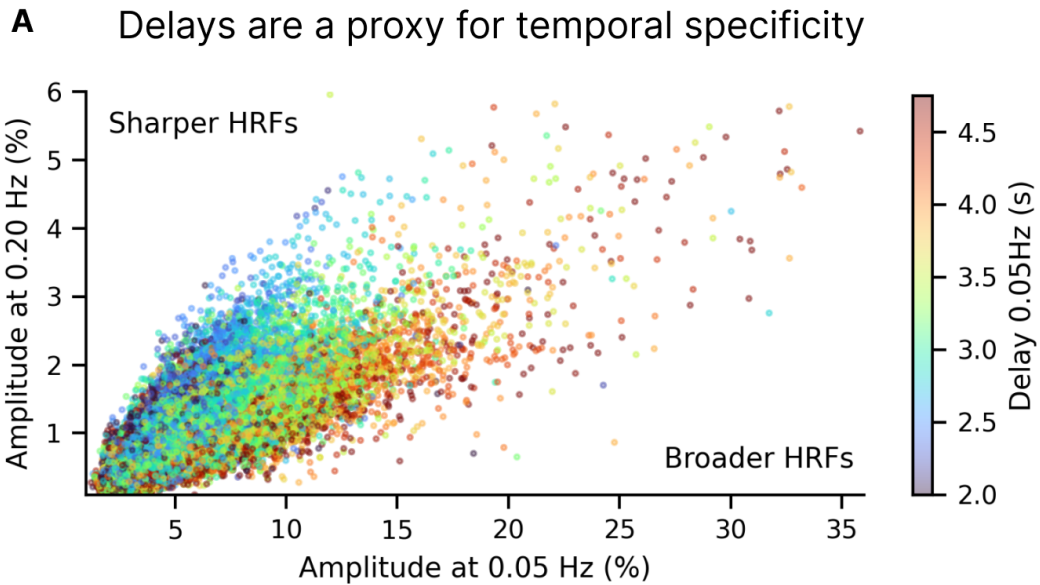
361 demonstrated a subtle effect of veins on temporal specificity, with slightly decreased temporal specificity in the
362 veins.

363 In contrast to the subtle effects of depth and vascular anatomy, we observed a strong relationship between
364 anatomical location and temporal precision, with a steep reduction in specificity from anterior to posterior V1
365 by 0.0025 mm^{-1} (95%CI: 0.0017 to 0.0034). To exemplify the size of this effect we can consider two arbitrary
366 voxels, a voxel A in anterior V1 about 2cm away along the A–P axis of V1 from a voxel B in posterior V1. If
367 they show the same response amplitude of 10% at 0.05-Hz stimulation, our analysis suggests that at 0.20 Hz,
368 voxel A would have an amplitude of about 2%, and voxel B an amplitude of 1.5%, a large difference due to
369 hemodynamic effects alone. Our analysis also revealed significant between-subject variability (0.0283, 95%CI:
370 0.0192 to 0.0425, values larger than the estimated effect of depth or compartment) in mean temporal specificity,
371 which suggests that fast fMRI responses may be more prominent in some but not other individuals, perhaps due
372 to variability in properties of blood and vasculature or perhaps other aspects of baseline physiology.

373 *3.4. Functional responses to slow stimuli are linked to temporal specificity*

374 These results demonstrated that there is a strong relationship between vascular-anatomical properties and
375 BOLD amplitude and delay estimates, and a more complex relationship to temporal specificity, which is mostly
376 influenced by anatomical location along the A–P axis of V1. Nonetheless, in practice not all experiments can
377 be conducted at a sufficiently high resolution to allow for cortical depth analysis or vascular segmentation, nor
378 does every experimental session provide sufficient time to acquire multiple runs to map out temporal specificity.
379 Additionally, it would be helpful to find ways to generalize results to other brain areas outside of V1. Furthermore,
380 the fact that there is strong subject variability suggests that using anatomical covariates alone is not sufficient to
381 fully explain responses at the single-subject level. Therefore, we also investigated whether the responses to a
382 fast stimulus were related to the responses to the slower stimulus, since slow task designs are ubiquitous in fMRI
383 experiments, and would enable subject-specific analyses to account for local temporal specificity. To do so, we
384 fit a mixed-effects linear model to test whether the amplitude at the 0.20-Hz stimulus was linked to the amplitude
385 and delay from the 0.05-Hz stimulus (which are simple to obtain in a short experiment).

386 This analysis confirmed a significant positive relationship between the amplitude at the slow stimulus (0.05-
387 Hz) and the fast stimulus (0.20-Hz): the population-level estimate for the effect of amplitude was 0.24 (95% CI:
388 0.22 to 0.27), meaning that response amplitudes at 0.20-Hz stimulus will range between approximately 22–27%
389 of the response amplitude at 0.05 Hz. The effect of delay alone was not significant (0.01; 95% CI: -0.03 to 0.06).
390 Importantly, however, there was a strong negative interaction between the delay and amplitude at each voxel in
391 the 0.05 Hz condition, of -0.020 (95% CI: -0.022 to -0.019). This suggests that the relationship between the
392 amplitude at the slow and the fast condition is modulated by the value of delay, with short delays correlating with
393 higher temporal specificity, and thus better preservation of fast responses, and long delays with lower specificity.
394 This means that, for two voxels with identical response amplitudes at 0.05 Hz, the shorter their delay at 0.05Hz,



B Delay and temporal specificity maps present strong anti-correlation

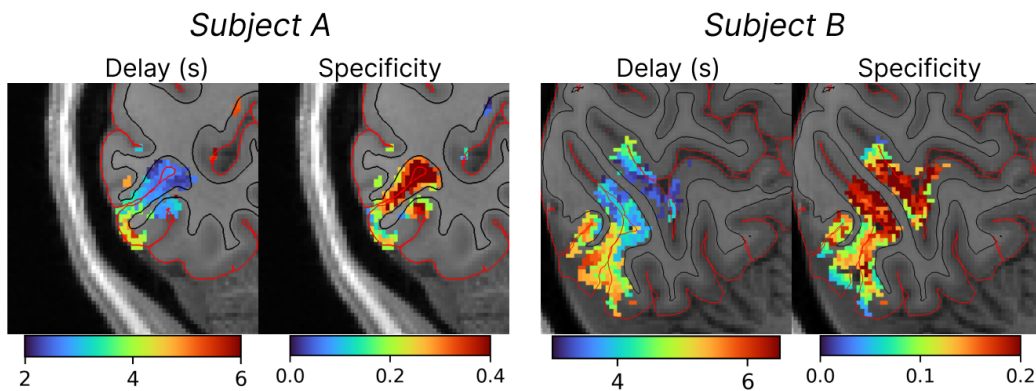


Figure 4: **Relationship between delays and temporal specificity.** A. The amplitude values at two different stimuli color-coded by the delay (a measure of hemodynamic latency). The delay and amplitude of the response to a slow stimulus are thus linked to the amplitude of the response to a fast stimulus (n=12 subjects). B. Maps of the hemodynamic delays measured with the 0.10-Hz stimulus and temporal specificity estimated as the ratio of amplitudes of the responses to fast and slow stimuli (0.20 Hz / 0.05 Hz), shown for two example subjects with distinct cortical folding. A clear anti-correlation can be seen where voxels with the lowest delays tend to also have the highest specificities.

395 the larger the response amplitude at 0.20 Hz is expected to be. This is illustrated in the scatter plot of figure
396 4 (panel A), which shows, for each voxel, amplitudes at 0.05 Hz and 0.20 Hz color-coded by their delay at
397 0.05 Hz. In this plot, temporal specificity, or the ratio of the responses to the 0.20-Hz and 0.05-Hz stimuli is
398 related to the slope, and datapoints with different delay values appear to lie along straight lines with different
399 slopes, with the shorter delay values exhibiting a steeper slope (higher temporal specificity) and longer delay
400 values exhibiting a shallower slope (lower temporal specificity), indicating an anti-correlation between delay and
401 temporal specificity. The effect is also shown qualitatively in two individual subject maps in figure 4 (panel B), with
402 delays maps showing similar spatial patterns to those of temporal specificity. This holds even though these two
403 example subjects have clearly different anatomy. These results demonstrate that in addition to our observation
404 that anatomical covariates can explain substantial variance in temporal specificity, the properties of the functional
405 response to slow stimuli are also strongly related to temporal specificity, which in future studies could provide a
406 complementary approach to identify fast voxels in addition to anatomical criteria.

407 *3.5. Temporal precision may be enhanced by hemodynamic non-linearity*

408 Overall, our results demonstrated relatively robust responses to the fastest 0.20 Hz stimulus, indicating that
409 voxels that respond fast are widespread in V1. The robust, fast responses could reflect HRFs that are generally
410 sharper and more precise than expected, or could reflect voxels with nonlinear hemodynamics, such that faster
411 stimuli elicit more temporally precise HRFs. We therefore investigated whether a single HRF (i.e., a linear model)
412 could be used to predict the amplitude behavior observed in our experimental data. HRFs become faster and
413 sharper as the stimuli become shorter (Lewis et al., 2018; Yeşilyurt et al., 2008), so we hypothesized that re-
414 sponses to rapidly oscillating stimuli would in part reflect a nonlinear change in voxelwise HRFs. To answer this
415 question we compared the temporal specificity at 0.20 Hz and at 0.10 Hz against a family of physiologically plau-
416 sible HRFs parameterized by their full-width at half-maximum and their peak delay, the canonical HRF (Glover,
417 1999) and two compartment-specific HRFs, parenchymal and venous (Siero et al., 2011). As hypothesized, we
418 found that no single HRF was capable of satisfactorily predicting the amplitude across frequencies observed in
419 our data, as the measured responses to the high-frequency stimulus were larger than models predicted. Instead,
420 we found an apparent "acceleration" of HRF as frequency increases (Figure 5): slower HRFs (Fig. 5, in blue)
421 better predicted the responses to the 0.10-Hz stimulus, and faster HRFs (in red) better predicted the responses
422 to the 0.20-Hz stimulus. In particular, the canonical HRF clearly failed to predict responses to either stimulus,
423 and the Siero HRFs best predicted responses to 0.20-Hz. These results suggest that a nonlinearity manifesting
424 as an acceleration of the HRF with faster stimulus frequencies may have contributed in part to the fast responses
425 observed in our study.

Normalized amplitudes for a family of Gamma HRFs

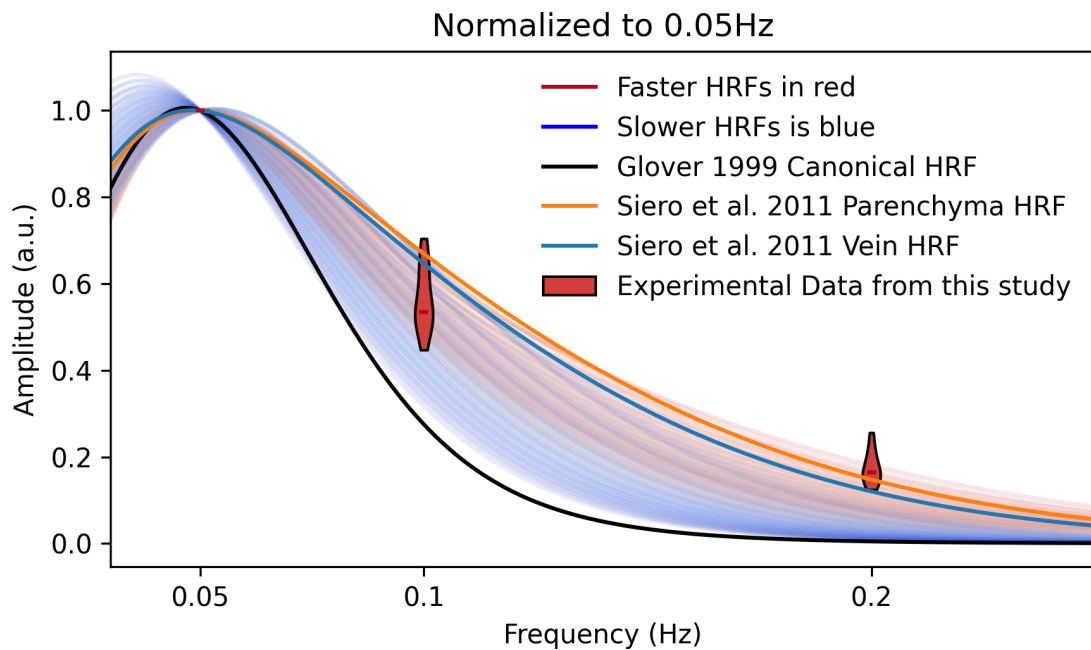


Figure 5: **Different HRFs predict the fMRI responses to 0.10-Hz and 0.20-Hz stimuli.** Curves of normalized amplitudes for a family of HRFs generated by varying the FWHM (2.25–3.75 s) and onset delay (2.3–5.4 s) of a double Gamma function. No combination of FWHM or onset delay generates a frequency response that matches the observed amplitude ratios. Faster HRFs are shown in red, slower in blue, with the gradient of colors changing from darkest blue to darkest red as the HRFs become faster. The red barplots show the temporal specificity estimates for each frequency. The dark line represents the canonical HRF, which clearly underestimates response amplitudes for both the 0.10- and 0.20-Hz stimuli. In orange and blue are the Siero HRFs which best predict the 0.20-Hz responses but overestimate the amplitude of 0.10-Hz responses.

426 4. Discussion

427 The interpretation of fMRI studies is critically influenced by hemodynamic response properties: if voxels with
428 different response properties are spatially organized in distinct regions of the cortex, this can be reflected in
429 statistical activation maps. Here we showed how the properties of V1 responses are influenced by anatomical
430 features. Cortical depth, veins, and anatomical position all influenced single-voxel response properties. Cortical
431 depth and the presence of veins strongly influenced delays and amplitudes, but only weakly influenced temporal
432 specificity. Surprisingly, we found that the strongest predictor of temporal specificity in V1 was anatomical position
433 across the A–P axis. This spatial gradient strongly dominated temporal specificity and produced an apparent
434 reversal of the spatial pattern of response amplitudes when using fast, as opposed to slow, stimuli.

435 4.1. Anatomical features associated with single-voxel responses

436 In accordance with previous literature, we found that hemodynamic delays measured with BOLD within the
437 cortical gray matter increase from the white matter to the pial surface ([Havlicek and Uludağ, 2020](#); [Markuerkiaga
438 et al., 2016](#); [de Zwart et al., 2005](#)), and increase in the presence of veins. Response amplitudes also increased
439 from the white matter to the pial surface and increased substantially in the presence of veins ([Koopmans et al.,
440 2010](#); [Siero et al., 2011, 2009](#)). It is worth noting, however, that the amplitude of parenchymal voxels—both those
441 intersecting the white matter and those intersecting the pial surface—can be strikingly similar when compared to
442 amplitude values found in veins. This suggests that, despite a correlation between cortical depth and vascular
443 compartment, not all voxels at the pial surface should be expected to have a large BOLD amplitude—only those
444 in the proximity of large pial veins. The effects of pial veins could be relatively sparse when imaging at high
445 spatial resolution, because not all voxels at the pial surface will contain large draining veins due to the sparsity of
446 these large veins on the surface of the brain.

447 Our analysis also measured the temporal specificity of fMRI responses, testing how robustly each voxel could
448 retain temporal information at faster frequencies. Our results demonstrated that spatial position within V1 has a
449 large effect on temporal specificity, and far outweighs the effects of veins or cortical depth, with voxels in anterior
450 V1 being relatively more sensitive to fast stimuli. Even though positional effects had been observed in the context
451 of the entire human cortex ([Handwerker et al., 2004](#)), we find local hemodynamic differences to be substantial
452 even within distances as small as 2–3 cm within the primary visual cortex.

453 The local hemodynamic variability suggests that biases could emerge when the intrinsic temporal properties
454 of voxels align or misalign with the stimulus frequency or experimental design. While designs that use high
455 frequencies or approaches like event-related and naturalistic designs may be more biased towards fast voxels,
456 so are designs that use low frequencies or long duration blocked stimuli likely to be biased towards slower
457 voxels. Therefore, careful consideration must be given to positional effects, as they could lead to discrepancies
458 in amplitude estimates across regions that are unrelated to the neuronal effects of interest. Our results focus on

459 V1 and identify a spatial gradient of temporal specificity within this region. A key next step will be to investigate
460 whether analogous gradients are present in other cortical regions. One potential approach our results suggest is
461 to use delay maps as a proxy for temporal specificity.

462 *4.2. On the striking and widespread heterogeneity of responses within V1*

463 The spatial heterogeneity of temporal properties of BOLD responses across V1 was striking. Indeed, earlier
464 studies had already shown a structured spatial patterns of BOLD responses across the visual cortex, suggesting
465 these effects may be broadly present in other datasets. For example, analysis of resting-state and task-driven
466 fMRI datasets have identified multiple visual networks in the human brain (Smith et al., 2009; Yeo et al., 2011),
467 namely a medial, an occipital and a lateral network. Since functional connectivity estimates are delay-sensitive,
468 it could be that the exact boundaries at which networks segregate are influenced by differences in their hemo-
469 dynamic response properties even if the underlying neuronal activity were synchronous. Although prior studies
470 have not specifically tested this effect, their results often display a spatial pattern that resembles our observed
471 delays (Amemiya et al., 2020; Bailes et al., 2023; Lewis et al., 2016, fig. 7). These experimental observations all
472 converge on the notion that local hemodynamic differences are widespread, particularly when imaging at high
473 resolution. A distinctive aspect of our work that allowed us to study these effects was the use of high temporal
474 and spatial resolution imaging, and an analysis pipeline focused on the single subject, extracting data in each
475 subject's native space to minimize loss of specificity due to smoothing and interpolation (Wang et al., 2022).

476 *4.3. Potential causes for the heterogeneity in specificity and its relationship to response delays*

477 What mechanisms could underlie this diversity in temporal specificity, and why is it that differences in delays
478 across cortical depths and vascular compartments are pronounced but these anatomical features only weakly
479 relate to specificity? Our data are not able to address this, but some possibilities can be considered. One
480 explanation in-line with anatomical properties of the angioarchitecture of the human brain, is that measurable
481 BOLD response variability reflects two distinct mechanisms. First, a macrovascular delay mechanism could
482 contribute: the time that it takes for arterial blood to reach a voxel or for venous blood to drain into a vein,
483 which varies due to the varying structure of regional blood supply and drainage across the cortex. If the voxel
484 is just parenchyma or capillaries, the delay of the arterial supply would affect timing, but the large veins may
485 not play a role. If the voxel, however, contains a large vein, then the arterial supply is still just as relevant, but
486 the time it takes for the blood from the upstream capillary bed to drain into the vein is what is relevant. This
487 mechanism is likely to influence delays across and even within regions, but probably not across cortical depths.
488 This is because diving arterioles and ascending venules run perpendicular to the layers, making arterial arrival
489 and venous draining times similar at all depths within a single cortical column. Second, a microvascular delay
490 mechanism could contribute: the time it takes for blood to pass through the capillary bed and into a draining
491 venule. This mechanism is likely to influence delays across cortical depths, as the deoxygenated blood travels

492 upwards from deep layers onto the larger draining veins where it pools, but this mechanism is not likely to vary
493 much within regions of V1, since we expect its angioarchitecture to be similar through the entire ROI (Weber
494 et al., 2008). Our observations suggest that macrovascular dynamics could play a more pronounced role in
495 temporal specificity variations along the A–P axis of V1 compared to microvascular dynamics. Nonetheless,
496 although arterial arrival times, which do follow the A–P axis as blood is delivered to the visual cortex via the
497 posterior cerebral artery, may explain our findings, functional hyperemia is known to propagate upstream from
498 the sites of activation, thus BOLD delays should not unambiguously track blood delivery times. Thus, it is unclear
499 exactly how arterial blood delivery times would affect the BOLD response dynamics, which mainly reflects local
500 vascular reactivity. Delays could also reflect venous effects towards the inferior and superior sagittal sinuses,
501 which also run in the A–P direction, although again the exact reason why these drainage effects influence BOLD
502 response dynamics is unclear.

503 4.4. Implications for laminar fMRI

504 The results we presented regarding the impact of cortical depth and vascular compartment on responses
505 agree with many previous reports from the literature (Gati et al., 1997; Lai et al., 1993; Siero et al., 2011, 2009).
506 They corroborate the importance of accounting for venous effects and depth-dependent biases on the BOLD
507 signal. Nonetheless, despite potentially biasing analysis, since the amplitude of veins can be almost an order
508 of magnitude larger than those of parenchymal voxels, the venous signal could still be exploited as a sentinel
509 for neuronal activity at high frequencies, even if their spatial location is non-specific. Veins are slower, but their
510 larger amplitude may balance out their lower temporal specificity, and as such, at least up until frequencies of
511 0.20Hz, and potentially even higher, the largest amplitudes should still be expected in venous voxels.

512 4.5. Implications for fast fMRI

513 While the heterogeneity of single-voxel responses can pose analytic challenges, it also reflects an opportu-
514 nity for fast fMRI: fast hemodynamics are widespread and common in the cortex. Identifying the location of fast
515 voxels could therefore be highly advantageous for conducting successful fast fMRI experiments. Due to the small
516 amplitude of fast responses, an ideal imaging approach would focus on areas where responses are fast enough
517 to be visible or where the baseline BOLD amplitude is overwhelmingly large, such as in individual venous vox-
518 els. Our results demonstrate how anatomy and delayed functional responses are linked to temporal specificity
519 and could guide localizer and ROI selection to enhance detectability of fast responses. Taking advantage of
520 the location of fast voxels may require revising strategies for data preprocessing and analysis, possibly requiring
521 more detailed anatomical imaging to create finer masks and regions-of-interest (ROIs) and data registration and
522 averaging techniques informed by anatomical and vascular segmentation, which are currently being developed
523 in the context of high spatial resolution imaging as well (Blazejewska et al., 2019; Wang et al., 2022). More-
524 over, when modelling fast fMRI using linear models adopting a single HRF, our data supports employing faster

525 hemodynamic response functions HRFs for capturing the dynamics, such as those measured by Siero et al.
526 2011. Those could be used as updated “canonical-fast” response functions for high-resolution fast fMRI, since
527 the conventional canonical HRFs clearly underestimate the amplitude of responses to faster frequencies.

528 *4.6. Identifying mechanisms of fast responses and avenues for further study*

529 In future studies, it would be crucial to explore mechanisms contributing to the fast and slow responses
530 observed here in the visual cortex. One possible avenue of investigation includes the use of fast functional
531 Magnetic Resonance Angiography (fMRA) to image blood velocity, flow and potentially vessel dilation (Bizeau et
532 al., 2017; Cho et al., 2012, 2008) to understand the underlying physiology. Furthermore, examining perfusion
533 or blood volume changes could provide insights into how the various hemodynamic components interact to
534 produce the fast BOLD responses. Expanding the application of fast fMRI to other brain regions, as proposed by
535 Hodono et al. 2022, may help elucidate the generalizability of these findings. Additionally, further investigations
536 on the impact of baseline blood flow, as in combining gas challenges with task fMRI, could shed light on the
537 interaction between baseline brain activity and the speed of the hemodynamic responses. For instance, it would
538 be worthwhile to test whether using fast stimuli with hypocapnia (which has been shown to accelerate BOLD
539 responses (Cohen et al., 2002)) could lead to even stronger responses to fast stimuli. Lastly, it is worth noting
540 that recent advances in MR instrumentation (Bates et al., 2023; Feinberg et al., 2023; Vizioli et al., 2021) could
541 soon allow the direct imaging of even faster frequencies. These advances in imaging, when combined with
542 advances in modelling of fast responses (Polimeni and Lewis, 2021), will further extend the capabilities of fMRI
543 towards its biological limits.

544 **5. Data and Code Availability Statement**

545 Datasets used for this study will be made available upon publication in anonymized and deidentified form.
546 Analysis code will be made available under <http://github.com/dangom/gomez2023temporal>. In case of
547 issues or data related queries please contact the corresponding author.

548 **6. Author Contributions**

549 **Conceptualization** DG, JP, LL

550 **Methodology** DG, JP, LL

551 **Software** DG, JP

552 **Validation** DG, LL

553 **Formal Analysis** DG, JP, LL

554 **Investigation** DG, JP, LL

555 **Resources** JP, LL

556 **Data Curation** DG

557 **Writing - Original Draft** DG, LL

558 **Writing - Review and Editing** DG, JP, LL

559 **Visualization** DG

560 **Supervision** JP, LL

561 **Project Administration** DG, LL

562 **Funding Acquisition** JP, LL

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567 **8. Declaration of competing interests**

568 The authors have no competing interests to declare.

569 **9. Acknowledgements**

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572 **10. Bibliography**

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