



Review

# Chromophobe renal cell carcinoma: Novel molecular insights and clinicopathologic updates



Reza Alaghebandan <sup>a,\*</sup>, Christopher G. Przybycin <sup>b</sup>,  
Virginie Verkarre <sup>c</sup>, Rohit Mehra <sup>d,e,f,\*\*</sup>

<sup>a</sup> Department of Pathology, Faculty of Medicine, University of British Columbia, Royal Columbian Hospital, Vancouver, BC, Canada

<sup>b</sup> Robert J Tomsich Pathology and Laboratory Medicine Institute, Cleveland Clinic, Cleveland, OH, USA

<sup>c</sup> Department of Pathology, Georges Pompidou European Hospital, AP-HP, Paris University, INSERM UMR 970, PARCC, Paris, France

<sup>d</sup> Department of Pathology, University of Michigan Medical School, Ann Arbor, MI, USA

<sup>e</sup> Michigan Center for Translational Pathology, Ann Arbor, MI, USA

<sup>f</sup> Rogel Cancer Center, Michigan Medicine, Ann Arbor, MI, USA

Received 24 August 2021; received in revised form 21 October 2021; accepted 21 October 2021

Available online 1 December 2021

## KEYWORDS

Renal cell carcinoma;  
Chromophobe;  
Immunohistochemistry;  
RNA *in situ* hybridization;  
Next-generation sequencing;  
Oncocytic tumors;  
Molecular

**Abstract** Chromophobe renal cell carcinoma (ChRCC) is the third most common renal cell carcinoma (RCC) subtype, which predominantly occurs in sporadic setting. ChRCCs are considered to originate from the intercalated cell of distal tubules with two main morphological variants, classic and eosinophilic. Most ChRCCs carry a favorable clinical outcome. Histology alone is limited in predicting the behavior of ChRCCs that do not have overtly aggressive morphologic findings such as necrosis and sarcomatoid features. Along with positive CD117 expression, classic ChRCCs generally express diffuse and uniform CK7, while eosinophilic variant demonstrates more heterogeneous CK7 expression (rare or patchy). Multiple losses of chromosomes 1, 2, 6, 10, 13, 17, and 21 are considered to be the genetic hallmarks of classic and eosinophilic ChRCCs, while chromosomal gains are known to be associated with sarcomatoid ChRCCs. *TP53* and *PTEN* are the two most frequently mutated genes in ChRCCs. The major challenge in the differential diagnosis of ChRCCs includes considerations around the eosinophilic variant (of ChRCCs), where it may share overlapping features with oncocytoma or other recent emergent oncocytic tumors. Most eosinophilic ChRCCs share expression of the recently described biomarkers, *LINC01187* and *FOXI1*, with classic ChRCCs, however, a subset of eosinophilic-like

\* Corresponding author.

\*\* Corresponding author. Department of Pathology, University of Michigan Medical School, Ann Arbor, MI, USA.

E-mail addresses: [reza.alagh@gmail.com](mailto:reza.alagh@gmail.com) (R. Alaghebandan), [mrohit@med.umich.edu](mailto:mrohit@med.umich.edu) (R. Mehra).

Peer review under responsibility of Tongji University.

ChRCCs with lower biomarker expression have been demonstrated to harbor *MTOR* gene mutations. Overall, the morphologic features of ChRCCs and genetic profile with combinations of chromosomal losses and gains suggest this tumor entity to represent a distinct, yet heterogeneous group of renal neoplasms.

© 2022 Editorial Office of Asian Journal of Urology. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

Chromophobe renal cell carcinoma (ChRCC) is the third most common renal cell carcinoma (RCC) subtype, accounting for 5%–7% of all RCCs. Although ChRCC was first reported and published in 1985 by Thoenes et al. [1], it appears that it was, in fact, first illustrated and described by Dr. Pierre Masson from Montreal, Quebec in 1955 [2]. In contrast to clear cell RCCs (CCRCCs) and papillary RCCs (PRCCs), ChRCCs originate from the intercalated cell of distal tubules [3]. Two main morphological variants of ChRCCs, the classic and eosinophilic, have been described; ChRCCs may also be associated with sarcomatoid dedifferentiation, a kind of an epithelial–mesenchymal transition [4].

Over the last few decades, a large body of literature on ChRCCs, their morphologic variants, clinical behavior, molecular, and histogenetic studies has been published. In this perspective, we present most updated and pertinent information on clinicopathologic and translational aspects of ChRCCs, and their applicability and potential relevance in clinical practice.

## 2. Clinical spectrum of ChRCCs

Most ChRCCs occur sporadically, and predominantly earlier than CCRCCs at an average age of 59 years (range 17–88 years), with a slight male predominance [5–7]. Three rare hereditary diseases that may predispose to the development of ChRCCs are the Birt-Hogg-Dubé syndrome (BHD) with germline mutation of the folliculin gene (*FLCN*), mapped at 17p11.2 [8–11], the tuberous sclerosis complex (TSC) with mutations involving either *TSC1* (9q34) or *TSC2* (16p13) genes [8,12,13], and the Cowden syndrome associated with germline mutation of *PTEN* (10q23) gene [14]. In such rare cancer susceptibility syndromes, genetic testing may be offered to patients and their families, when clinical and/or pathologic features are concerning (for example, multiple oncocytic renal tumors).

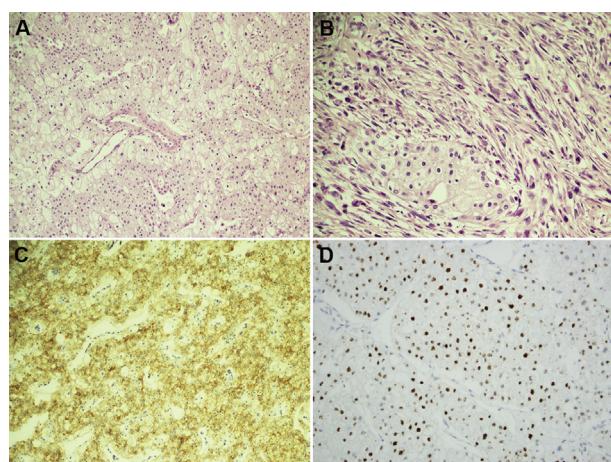
ChRCCs generally carry a favorable clinical outcome with a low tendency to progress or metastasize. Their prognosis is postulated to be better than CCRCCs, with a higher 5- to 10-year cancer-specific survival of 93% and 87%, respectively [7,15,16]. Subsequent disease specific events including recurrence/metastasis/death have been reported in approximately 1.5% and 6.6% of cases, at 5 years and 10 years respectively [7,16,17]. Patients with metastatic ChRCCs have a similar median overall survival compared to patients with CCRCCs, when treated with conventional targeted therapies [17]. Unlike non-sarcomatoid ChRCCs,

sarcomatoid ChRCCs are associated with poor prognosis and worse clinical outcome [18]. Although given their rarity, the prognosis of rare subtypes of ChRCCs is currently unclear, we speculate their clinical behavior to be comparable to those of non-sarcomatoid ChRCCs.

## 3. Morphological correlates of ChRCCs

ChRCCs are characterized by neoplastic cells with prominent cell membranes and pale to eosinophilic cytoplasm, arranged in a solid-alveolar pattern. Characteristic features that facilitate the recognition of most ChRCCs include hyperchromatic and irregular wrinkled (raisinoid) nuclei, perinuclear clearing (halos), and binucleation (Fig. 1A). Of note, electron microscopic analyses have revealed perinuclear halos and raisinoid nuclei to reflect cytoplasmic microvesicles and peripheral condensation of mitochondria, which may not be seen on cytologic imprints [19,20].

Two distinct morphological variants of ChRCCs are classic and eosinophilic. Classic ChRCCs exhibit mostly pale cells with raisinoid nuclei in an alveolar, solid to trabecular architectural growth pattern. Eosinophilic variant of ChRCCs was described in 1988 [21], following the initial description of classic ChRCCs. The most recent 2016 World Health Organization (WHO) renal tumor classification formally recognizes the eosinophilic variant of ChRCCs and acknowledges the challenges associated with making a



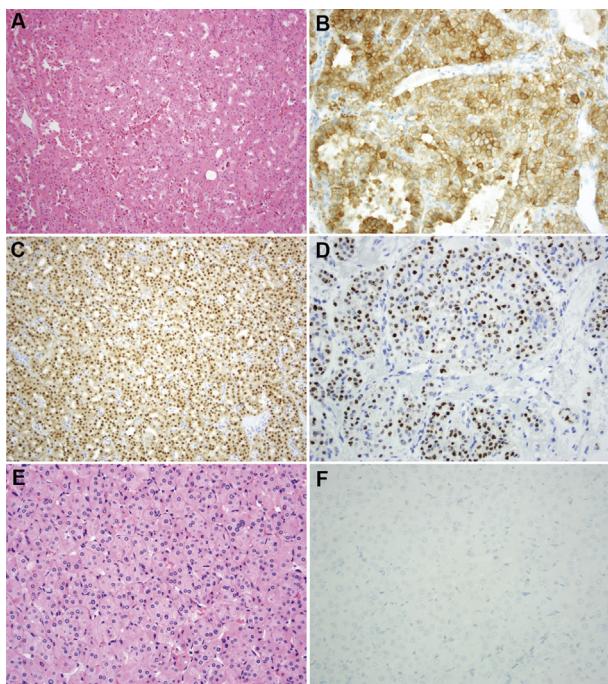
**Figure 1** Classic and sarcomatoid ChRCCs. (A) Classic ChRCC H & E stain; (B) Sarcomatoid ChRCC H & E stain; (C) Membranous staining for CD117; (D) High-level nuclear LINC01187 expression. All images are at 200×. ChRCC, chromophobe renal cell carcinoma; H & E, hematoxylin and eosin.

morphologic distinction from benign oncocytoma but does not provide precise diagnostic criteria [4]. The 2016 WHO classification only stipulates that eosinophilic ChRCCs should be “almost purely eosinophilic”. Eosinophilic ChRCCs typically demonstrate acinar architecture, abundant eosinophilic cytoplasm (almost purely eosinophilic), focal nuclear wrinkling, and subtle perinuclear clearing/halos (Fig. 2A). It is important to recognize that eosinophilic ChRCCs can present diagnostic challenges in daily routine urology and pathology workup, and that not all tumors mimicking eosinophilic ChRCCs belong to the ChRCC family.

Similar to other RCCs, ChRCCs may show sarcomatoid features (Fig. 1B) where the incidence of this epithelial–mesenchymal transition in primary ChRCCs has been estimated at 2%–8% [7, 16, 18, 22–24], which reaches a substantial 26% in metastatic disease [18]. Furthermore, in recent years, multiple studies have shown ChRCCs to display morphologic heterogeneity including those with pigmented/adenomatoid, multi-cystic, neuroendocrine, and papillary features [25–31].

#### 4. Immunohistochemical overview and novel biomarkers

ChRCCs, similar to oncocytomas, originate from the intercalated cells and hence exhibit positive CD117 immunohistochemical expression, known to be constitutionally



**Figure 2** Eosinophilic variant of ChRCC. (A) Eosinophilic variant of ChRCC H&E stain; (B) Predominantly membranous staining for CD117; (C) The nuclear staining for FOXI1 in eosinophilic variant of ChRCC; (D) High-level nuclear LINC01187 expression; (E) Another example of eosinophilic variant of ChRCC H & E stain; (F) With negative CD117 expression and *MTOR* gene mutation (data not shown). All images are at 200×. ChRCC, chromophobe renal cell carcinoma; H & E, hematoxylin and eosin.

expressed within some cells in the distal nephron. Along with positive CD117 expression (Figs. 1C and 2B), classic ChRCCs generally express diffuse and uniform CK7, which may be of diagnostic utility for a practicing pathologist. In most cases, classic ChRCCs can be readily diagnosed on basis of a thorough histologic evaluation. However, in some instances such as limited samples (i.e., core biopsy), this may be challenging; confounding factors include other differential diagnoses and potential mimickers such as CCRCCs and miscellaneous tumors (such as adrenal neoplasms). Key histologic features such as nuclear atypia and prominent cell membranes along with pertinent immunohistochemical stains (i.e., positive expression for PAX8, CD117, and CK7) can assist in such settings. Other helpful markers, whose negative expression may support a ChRCC diagnosis where CCRCC is in the differential diagnosis, include vimentin and carbonic anhydrase-IX (CA-IX).

Distinguishing ChRCCs from other oncocytic neoplasms can be difficult, especially the eosinophilic variant, which can mimic other neoplasms. Eosinophilic variants of ChRCCs demonstrate rare or patchy CK7 expression, similar to oncocytoma [32,33]. A morphologic assessment is often most helpful in distinguishing oncocytoma from classic and eosinophilic variants of ChRCCs, with immunohistochemical markers needed in occasional cases to support morphologic impression. Of note, Hale colloidal iron stain has historically been used in this differential diagnosis, with ChRCCs showing cytoplasmic positivity while oncocytoma is generally negative. The utility of this stain is limited due to potential technical process and interpretation challenges [34].

In terms of cell of origin, data from The Cancer Genome Atlas (TCGA) and others have demonstrated ChRCCs to exhibit a distal nephron-based mRNA expression signature (based on in silico data and immunohistochemistry); CCRCCs, on the other hand, displayed a proximal nephron based signature [35–37]. TCGA also showed ChRCCs to express genes driven by the transcription factor FOXI1, which labels intercalated cells, a finding further validated by follow-up studies (Fig. 2C) [38]. Based on recently published data, ChRCCs are enriched for expression of intercalated cells markers such as FOXI1 protein and long noncoding RNA *LINC01187* (Fig. 2D) in both primary and metastatic sites, where these may serve as useful biomarkers especially in the metastatic setting (Fig. 1D) [37,39]. Sarcomatoid ChRCCs interestingly tend to lose expression of these novel markers in the high-grade spindle cell areas of such tumors, which otherwise tend to lose CK7 and in contrast acquire vimentin expression. Currently, there are no clinically available cancer- or lineage-specific biomarkers for oncocytoma, ChRCCs including the eosinophilic variant, or other related oncocytic tumors. The ongoing in-depth proteogenomic assessment of renal tumors by Clinical Proteomic Tumor Analysis Consortium (CPTAC) is likely to have a major impact on RCC protein biomarker identification [40].

#### 5. Grading considerations

Most ChRCCs do not exhibit aggressive behavior, with only a small subset (approximately 5%) at risk of progression and

metastasis after surgery. Accurate prediction of the clinical behavior and trajectory of ChRCCs by histologic features has proven to be challenging. Historically, risk assessment of ChRCCs has not been successful with a number of studies attempting to address the value of histological grading for these tumors [6, 16, 41–43]. The clinical significance of traditional grading systems applied to other RCC subtypes, such as Fuhrman nuclear grade and the WHO/International Society of Urological Pathology (ISUP) nucleolar system, could not be demonstrated for ChRCCs [44]. In contrast to CCRCs and PRCCs, the WHO/ISUP grading system is not applicable to ChRCCs [44, 45].

Evaluation of a large ChRCC cohort previously demonstrated pT stage, tumor necrosis, and sarcomatoid change to be independently predictive of aggressive behavior on multivariable analysis [23]. The largest single-institution morphology based study showed significant association of clinical outcome with tumor size, small-vessel invasion, sarcomatoid features, and microscopic necrosis [7]. Since the prominent nuclear atypia and relative infrequency of conspicuous nucleoli inherent to ChRCCs pose challenges to applying conventional nuclear grading, a three-tiered grading system [46] has been proposed, correlating geographic nuclear crowding and anaplasia in predicting clinical outcome for patients with ChRCCs. A subsequent study, however, found that this grading system did not provide additional prognostic information once tumor stage and sarcomatoid features were included [43]. Similarly, Ohashi et al. [6] were not able to validate the proposed three-tiered chromophobe grading system or the four-tiered WHO/ISUP grading system for outcome determination in ChRCC. They instead proposed a two-tiered grading system, based on sarcomatoid features and necrosis, which was successful at the multivariate level. In another large study, a modified tumor grading scheme using mitotic index, cytologic eosinophilia, and architecture was not significantly associated with outcome [7].

Overall, histology alone seems to be limited in predicting the behavior of ChRCCs that do not have overtly aggressive morphologic findings (e.g., necrosis and sarcomatoid features). Accordingly, supplementation with data from ancillary studies assessing other biomarkers, as they are discovered, may be necessary to accurately predict behavior in the majority of ChRCCs that do not otherwise have overtly adverse histologic findings.

Hopefully, discovery of novel molecular aberrations and genomic pathway derangements associated with aggressive ChRCCs can act as a surrogate to morphology for determination of clinical outcome of this disease.

## 6. Molecular underpinnings of ChRCCs

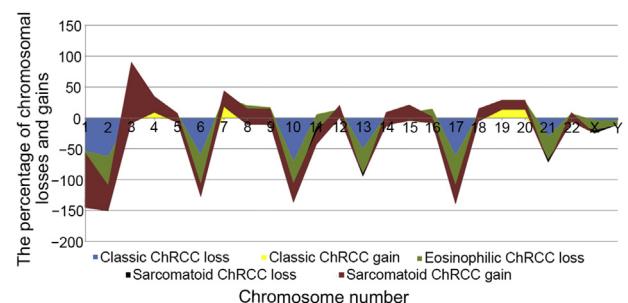
Recent comprehensive molecular characterizations, with identification of genetic aberrations and chromosomal abnormalities, have considerably improved our understanding of ChRCC, however, to date, predictive biomarkers remain to be identified [5, 17].

Multiple losses of chromosomes 1, 2, 6, 10, 13, 17, and 21 have historically been considered the genetic hallmark of ChRCC (both classic and eosinophilic variants). Earlier strategies using classic cytogenetic techniques established classic

ChRCC to be associated with multiple losses of chromosomes 1, 2, 3, 6, 7, 9, 10, 12, 13, 17, 18, and 21 [47–53]. The TCGA analysis of ChRCC further confirmed the characteristic pattern of chromosomal losses of 1, 2, 6, 10, 13, and 17 in 86% of tumors (mainly classic type), with documented additional losses of chromosomes 3, 5, 8, 9, 11 and 18, and 21q in 12%–58% of tumors [54]. In the last two decades, multiple studies have also demonstrated multiple chromosomal gains in ChRCCs [25, 53, 55–61]. Although this has generally been deemed an uncommon phenomenon in ChRCCs, studies with larger cohorts showed a more variable genetic profile of ChRCCs with multiple chromosomal losses as well as multiple gains [25, 53, 55–61]. Most frequently detected chromosomal gains are 4, 7, 15, 19, and 20 (mainly in classic and sarcomatoid types) [25, 53, 55–61]. It is noteworthy that, unlike classic ChRCCs, diploid chromosomal pattern is more commonly found in the eosinophilic variant [62].

It should be noted that although both oncocytomas and ChRCCs arise from the distal nephron, they do not represent a continuum of progression from benign to malignant disease. Even eosinophilic ChRCCs, which share some morphologic features with oncocytoma, have different chromosomal numerical aberrations (CNAs) and gene expression landscape than oncocytomas [36]. On the other hand, sarcomatoid ChRCCs carry multiple chromosomal gains including 1, 2, 6, 10 and 17, which are often different than CNAs seen in the epithelial component; these tumors appear to have lower frequency of chromosomal losses [53, 55–58, 63]. Studies of other rare variants, such as ChRCCs with pigmented microcystic adenomatoid/multi-cystic growth [26–28, 64], ChRCCs with neuroendocrine features [25, 29, 30, 65, 66], and ChRCCs with papillary architecture [31], also show variable CNAs including multiple losses (more common) and gains (less common). A survey of CNAs in metastatic ChRCC suggests distant metastases to demonstrate the same genetic patterns, usually chromosomal losses, as found in the primary tumors [55]. Of note, no correlation between CNAs and the proposed three-tiered grading system for ChRCCs has so far been established [46, 58]. Distribution of chromosomal losses and gains in classic, eosinophilic, and sarcomatoid ChRCCs are presented in Fig. 3 [53].

A diverse molecular phenotype of ChRCCs has been highlighted by recent genetic studies encompassing eosinophilic variant, sarcomatoid, and metastatic ChRCCs. These studies provide valuable insight into characterization and sub-classification of ChRCCs, or evolutionary patterns associated with these malignancies. Classic ChRCCs display



**Figure 3** Distribution of chromosomal losses and gains in classic, eosinophilic and sarcomatoid ChRCC. ChRCC, chromophobe renal cell carcinoma.

fewer hypermethylation events and harbor a lower number of somatic variants (low mutational burden) compared to both CCRCCs and PRCCs [36,67,68]. *TP53* and *PTEN* are the two genes most frequently found to be mutated in respectively 32%–64% and 9%–45% of classic ChRCCs, independent of the presence of sarcomatoid features [18,36,68]. *TERT* promoter mutations/rearrangements have also been detected in 6%–12% of cases and associated with *TERT* upregulation, without any distinct impact on clinical outcome [36,69]. The TCGA analyses of ChRCCs labeled *TP53* and *PTEN* as the most frequently mutated genes in ChRCCs [54,70]. Loss of *CDKN2A* gene or its expression, by either deletion of 9p21.3 or hypermethylation, was the second most common alteration [54,70]. Ricketts et al. [68] examined metabolic analysis of RCC histologic subtypes and found that metabolically divergent ChRCCs to be associated with high stage and lack the chromosomal copy number losses typically associated with ChRCCs; hence were associated with much poorer survival.

Mutations identified in other genes have been identified in less than 5% of ChRCCs (mostly eosinophilic) including *MTOR*, *TSC1*, *TSC2*, *NRAS*, and others [67]. In contrast to CCRCCs and PRCCs, no significant mutations have been identified in the three chromatin remodeling genes *PBRM1*, *BAP1* and *SETD2* [68]. Mutations in mitochondrial DNA within ChRCCs have enabled establishment of a link with alterations of respiration and oxidative phosphorylation pathways [36]. Additionally, pathway analysis demonstrated rare alterations in PI3K-AKT-mTOR pathway genes, including *PTEN*, *TSC1*, *TSC2* and *MTOR*, in ChRCC, which would potentially result in appropriate targets for mTOR inhibitors [54,70,71]. Durinck et al. [67] identified *TP53*, *PTEN*, *FAAH2*, *PDHB*, *PDXDC1* and *ZNF765* to be significantly mutated in ChRCC, with frequent *TP53* mutations being exclusively associated with classic ChRCCs. Most recently, Roldan-Romero et al. [72] found within the eosinophilic variant of ChRCCs an overrepresentation of mTOR pathway (*MTOR*, *TSC1*, and *TSC2*) mutations as well as aberrations in the mitochondrial genes encoding the complex I of the electron respiratory chain.

Interestingly, recent studies have allowed a greater insight into the genomic features of metastatic ChRCCs supporting *TP53* mutations, *PTEN* mutations, *CDKN2A*

alterations and imbalanced chromosome duplication (defined as duplication of ≥three chromosomes) as high-risk features associated with poor survival [68,73]. DNA hypermethylation has also been associated with high-stage disease, *TP53* mutation and poor survival in ChRCCs [68]. Identification of such molecular genotypes provides hope for an integration of such tools into existing models of clinical care.

Overall, ChRCCs show variable morphology and genetic profile with combinations of chromosomal losses and gains, suggesting that they represent a heterogeneous group of neoplasms, both from a morphologic and a molecular perspective (Table 1).

## 7. ChRCCs and contemporary/emerging renal oncocytic neoplasms

The major challenge in the differential diagnosis of ChRCCs would be associated with its eosinophilic variant, especially in distinction from oncocyomas, and particularly in limited samples (i.e., core biopsy), where such a distinction may carry clinical implications. Oncocyomas typically show a relatively nested growth pattern where the neoplastic cells have uniform round nuclei, lack perinuclear halos, and a shared immunohistochemical phenotype with eosinophilic ChRCCs (*CD117+*, *CK7-* or only scattered positive cells). Liu et al. [62] recently examined a scoring system where histological patterns (organoid/nested, tubulocystic and solid/confluent), the quality of stroma (fibromyxoid and edematous), nuclear wrinkling, perinuclear halos, well-defined cell borders with clear cytoplasm, and *CK7* immunohistochemical expression, were features found to be significantly discriminatory between oncocyoma and eosinophilic ChRCCs.

Other “eosinophilic” renal neoplasms that need to be considered in the differential diagnosis include WHO recognized renal entities such as succinate dehydrogenase (SDH)-deficient RCC, as well as provisional/emerging ones including eosinophilic solid and cystic RCC (ESC-RCC), low-grade oncocytic renal tumor (LOT), anaplastic lymphoma kinase rearrangement-associated RCC (ALK-RCC), and eosinophilic vacuolated tumor (EVT) (Table 2).

**Table 1** Molecular aberrations and mutation summary for ChRCC including a comparison with CCRCC and PRCC.

Genetic alteration	ChRCC	CCRCC	PRCC
Main chromosomal numerical variation	- Multiple chromosomal losses [1,2,6,10,13,17,21] and gains [1,2,6,10,17] more frequent in sarcomatoid ChRCC	- Loss of 3p (91%), gains of 5q (67%), loss of 14q (45%), del 9p21 ( <i>CDKN2A</i> locus), and del 10q23 ( <i>PTEN</i> locus)	- Trisomy 7 (50%–100%), Trisomy 17 (66%–100%), and loss of Y (77%–100%)
Main genetic aberration	- <i>TP53</i> (32%–64%), <i>PTEN</i> (9%–45%), and <i>TERT</i> promoter mutations/rearrangements (6%–12%)	- <i>VHL</i> (75%), <i>PBRM1</i> (33%–40%), <i>BAP1</i> (10%), and <i>SETD2</i>	- <i>MET</i> (17%–21% in type 1), and some with mutations in <i>BAP1</i> , <i>SETD2</i> , <i>ARID2</i> , <i>KEAP1</i> , <i>TERT</i> promoter, <i>CDKN2A/B</i> , and <i>NF2</i>
Methylation status	- Hypomethylated, less frequently hypermethylated (mostly in advanced stage)	- Hypermethylated (subset)	- Hypermethylated (subset)

ChRCC, chromophobe renal cell carcinoma; CCRCC, clear cell renal cell carcinoma; PRCC, papillary renal cell carcinoma.

**Table 2** Clinical, histologic, immunophenotypic, and molecular summary of ChRCC, eosinophilic variant, and overlapping entities.

Renal tumor subtype	Clinical feature	Morphologic feature	Ancillary feature	Molecular aberration
ChRCC	- Majority indolent, 3%–10% metastasis; mostly sporadic; rare hereditary setting (TSC, BHD, and Cowden)	- Large pale cells with prominent cell membranes, +/– admixed with granular eosinophilic cytoplasm and wrinkled nuclei with perinuclear haloes, solid/large alveolar architecture with incomplete vascular septa	- CD117+; usually diffuse CK7+ (sometimes focal or negative); positive expression of <i>LINC01187</i> and <i>FOXI1</i> ; vimentin–	- Multiple chromosomal losses and gains; <i>TP53</i> and <i>PTEN</i> gene mutations; other less common mutations
Eosinophilic variant of ChRCC	- Similar to ChRCC	- Granular eosinophilic cytoplasm, perinuclear haloes, and nested architecture	- CD117+; more focal CK7+; expression of <i>LINC01187</i> and <i>FOXI1</i> ; vimentin–; SDHB retained	- Diploid or multiple chromosomal losses
Oncocytoma	- Benign and mostly sporadic	- Granular eosinophilic cytoplasm, central “archipelagos”, uniform round nuclei with minimal atypia and no mitotic activity	- CD117+; focal to negative CK7; positive for <i>LINC01187</i> and <i>FOXI1</i>	- Diploid or hypodiploid (−1, X, Y, 14, 21); 11q13 ( <i>CCND1</i> locus) rearrangement; mutation in mitochondrial gene
Hybrid oncocytic tumor	- Exceptional metastases; usually associated with BHD	- Features of oncocytoma and ChRCC in the same tumor	- Variable; generally CD117+; CK7 focal to patchy	- Germline mutations in <i>FLCN</i> encoding folliculin
Low-grade oncocytic RCC, unclassified or oncocytic renal neoplasm of low malignant potential	- Presumably indolent	- Resembles oncocytoma with greater nuclear atypia and absence of diagnostic features of another oncocytic neoplasm	- Generally CD117+; CK7 variable	- Unknown
ESC-RCC <sup>a</sup>	- Established, metastatic potential; sporadic or germline	- Solid and cystic architecture, abundant cytoplasm with coarse basophilic granules	- Usually CK20+; CK7– or focal; CD117–; vimentin+	- <i>TSC1/TSC2</i> somatic or germline mutations
LOT <sup>a</sup>	- Indolent <sup>b</sup> and mostly sporadic	- Solid architecture with edematous areas, monomorphic round to oval nuclei with delicate perinuclear haloes	- CK7+ diffusely; CD117–; vimentin–; SDHB retained	- Disomic or del 19p13, 19q13, 1p36; <i>MTOR</i> gene mutations in borderline tumor or closely similar tumor
EVT <sup>a</sup>	- Indolent <sup>b</sup> ; sporadic or germline	- Solid/nested growth, oncocytic cytoplasm with large vacuoles, and prominent nucleoli	- CD117+; CK7– or focal+; vimentin–; cathepsin-K+; CK20–; SDHB retained	- <i>TSC2/MTOR</i> gene mutations; loss of chromosomes 1 and 19, LOH of 16p11.2–11.1; 7q31.31
SDH-deficient RCC	- Established metastatic potential; germline	- Eosinophilic cytoplasm with vacuoles containing eosinophilic to pale flocculent material	- Loss of SDHB; CD117–; CK7–	- Germline mutations in one of the SDH genes
ALK-RCC	- Established metastatic potential	- Eosinophilic cytoplasm, variable and admixed architecture, mucinous/myxoid background	- ALK+; SDHB and FH retained	- Rearrangement of ALK with various partners

ALK, anaplastic lymphoma kinase; ALK-RCC, anaplastic lymphoma kinase rearrangement-associated renal cell carcinoma; BHD, Birt-Hogg-Dubé syndrome; ChRCC, chromophobe renal cell carcinoma; ESC-RCC, eosinophilic solid and cystic renal cell carcinoma; EVT, eosinophilic vacuolated tumor; FH, fumarate hydratase; LOH, loss of heterozygosity; LOT, low-grade oncocytic renal tumor; *MTOR*, mammalian target of rapamycin; SDH, succinate dehydrogenase; SDHB, succinate dehydrogenase complex subunit B; SDH-deficient RCC, succinate dehydrogenase deficient renal cell carcinoma; TSC, tuberous sclerosis complex.

<sup>a</sup> Emerging entity under investigation.

<sup>b</sup> Limited follow-up data available.

## 7.1. SDH-deficient RCCs

These are rare renal neoplasms which occur in setting of germline mutation of one of the subunits of *SDH* gene (mostly *SDHB* and *SDHA*). Histologically, they exhibit solid, nested or tubular growth pattern, mimicking neuroendocrine tumors. Characteristic findings of this entity include neoplastic cells with flocculent cytoplasmic vacuoles with a pale eosinophilic or bubbly appearance and low-grade nuclei [74]. In contrast to ChRCCs, CD117 and CK7 are usually negative [75]. Loss of *SDHB* immunohistochemical staining is diagnostic [74] and should lead to genetic consultation, if germline setting is not previously established or known.

## 7.2. ESC-RCCs

Most ESC-RCCs are small, solitary with low stage that are predominantly found in females, and generally exhibit indolent behavior [76]. Approximately 10% of these neoplasms occur in TSC patients. ESC-RCCs show solid and cystic growth pattern, with often scattered histiocytes and lymphocytes. The neoplastic cells are voluminous and eosinophilic with cytoplasmic coarse granularity (stippling) [76]. Unlike conventional renal tumors, most ESC-RCCs (about 85%) are positive for CK20 (at least focal), while negative for CD117 and may rarely/focally express CK7 [76].

## 7.3. LOTs

LOTs are typically single, small, low-stage tumors demonstrating an overlapping morphology with oncocytoma and eosinophilic ChRCCs [77,78]. LOTs show solid growth pattern with sharply delineated edematous areas containing single cells and irregular cell cords. The eosinophilic cells show “low-grade”, round to oval nuclei, often with delicate perinuclear clearing. Immunohistochemically, these tumors consistently show CD117-/CK7 diffuse+ profile [78]. Importantly, these tumors tend to share morphologic, immunophenotypic and molecular overlap with the eosinophilic-like variant of ChRCCs harboring *MTOR* mutations [37,79]. Rare cases have been recently described in TSC setting [80,81].

## 7.4. ALK-RCCs

ALK-RCCs are solitary tumors with slight male predilection, and a significant potential for metastasis (up to 30%) [82,83]. By definition, ALK-RCCs are associated with an ALK rearrangement at the genomic level. ALK-RCCs may show variable and admixed growth patterns (e.g., solid, tubular or tubulo-cystic, papillary, cribriform, trabecular, and signet-ring individual cell growth), often in a mucinous background [77]. The neoplastic cells have eosinophilic cytoplasm and may show variable morphologies, including rhabdoid, vacuolated, pleomorphic, and giant cell [77]. Diffuse cytoplasmic and membranous ALK protein expression by immunohistochemistry is diagnostic and can aid in screening suspicious cases, particularly in tumors difficult

to classify (where immunohistochemistry for conventional markers is nonspecific or negative).

## 7.5. EVTs

EVTs present a unique morphology, relatively consistent immunophenotypic profile and distinct molecular/genetic features [75,77,84,85]. These tumors demonstrate a solid to nested growth pattern. The neoplastic cells exhibit eosinophilic cytoplasm, with large intracytoplasmic vacuoles, prominent cell membranes, and round to oval nuclei, with enlarged nucleoli. Normal renal tubules and thick-walled vessels are frequently found at the periphery. The neoplastic cells are typically immunoreactive for cathepsin-K, CD117, and CD10. CK7 is variably positive, and negative stains include vimentin, CK20, HMB45, and Melan-A. EVTs have been shown to harbor *TSC2* or *MTOR* somatic mutations [85,86].

## 7.6. Other “difficult to classify” oncocytic tumors

It is worth noting that the surgical pathology community, at large, still considers oncocytic neoplasms and their clinicopathologic spectrum as one of the most challenging areas in routine clinical practice. The diagnosis of tumors with overlapping morphologic features spanning a spectrum of entities like oncocytoma, ChRCC, and others, mentioned above, not surprisingly, presents a frequent clinical management dilemma. The Genitourinary Pathology Society (GUPS) has recently proposed the term “oncocytic renal neoplasm of low malignant potential, not further classified” for such “borderline” cases, which should be reserved for solitary, sporadic tumors with overlapping features [75]. We believe this entity encompasses the currently so called “low-grade oncocytic renal cell carcinoma, type unclassified” tumor subtype. This approach recognizes the fact that this “clinical management” category likely includes a heterogeneous group of renal neoplasms that most likely have a significantly low risk for developing metastasis. Of importance, the term “hybrid oncocytic tumor” should be reserved for hereditary cases (*i.e.*, BHD syndrome), as per the GUPS recommendations [75].

## 8. A practical diagnostic approach to ChRCCs

Most ChRCCs can be accurately diagnosed on the basis of a thorough histologic evaluation. However, the diagnosis of ChRCCs at times remains a challenge, particularly in limited samples, and when confounded by the presence of morphologic features overlapping with oncocytoma. The role of immunohistochemistry in ChRCCs (with its limitations) is evolving and may aid in challenging cases. It should be noted that a subset of *TSC/MTOR*-aberration associated RCCs have been demonstrated to harbor features morphologically similar to eosinophilic ChRCC, where, interestingly, these tumors may be negative for CD117 expression (Figs. 2E and 2F) [13,79]. Most eosinophilic ChRCCs share expression of the recently described biomarkers, *LINC01187* and *FOXI1*, with classic ChRCCs, however, a subset of eosinophilic-like ChRCCs with lower biomarker expression

were demonstrated to harbor *MTOR* gene mutations (Figs. 2E and 2F) [37].

Presence of a renal tumor in general warrants dedicated cross sectional imaging for further characterization to facilitate treatment planning. It should be noted that oncocytoma cannot reliably be distinguished from RCC, but techniques such as diffusion-weighted (DW) MRI show promising results [87]. Renal mass biopsy demonstrates a reliable ability to determine the presence of malignancy and characterize histology in small renal masses [88]. The current clinical guideline panels suggest offering renal mass biopsy as an adjunctive option in the evaluation of patients with localized RCCs [88]. Unfortunately, an accurate diagnosis of oncocytic renal tumors may be challenging upon renal mass biopsy; a limited sampling of tumor at the time of renal biopsy may or may not be representative of the entire lesion [89]. This is most pronounced in distinguishing eosinophilic ChRCCs from oncocytoma, given the fact that this differential often carries significant clinical and management implications. As such, the field of urologic pathology is still relatively unclear as to whether it is preferable to issue an outright diagnosis of oncocytoma (when features are typical in the biopsy sample) or to use more general terminology, such as "low-grade oncocytic neoplasm" with a comment that the features are compatible with oncocytoma [90]. At times, this distinction is impossible upon renal biopsy and a general terminology of "low-grade oncocytic neoplasm" is advisable, with comment that the differential diagnosis includes entities like renal oncocytoma and eosinophilic ChRCCs.

In perplexing cases, genomic profiling may be helpful in guiding towards the correct renal tumor categorization. In a comprehensive genomic study of non-clear cell RCCs, Durinck et al. [67] identified *ADAP1*, *SDCBP2*, *HOOK2*, *BAIAP3*, and *SPINT1* as the top five genes with high level of expression in ChRCCs, while *ITGB3*, *MINOS1-NBL1*, and *ASB1* were found to be upregulated in oncocytomas. Furthermore, a set of five genes including *ASB1*, *GLYAT*, *PDK1IP1*, *PLCG2*, and *SDCBP2* was reported to be sufficient to separate ChRCC, oncocytoma, and PRCC [67]. These findings shed light on developing of companion diagnostic panels for renal neoplasms with oncocytic features. It is worth noting that such findings would require further validation studies before they are considered for application in clinical practice.

Concerning the utility of CNAs, it should be noted that classic ChRCCs show frequent loss of chromosomes 1, 2, 6, 8, 10, 13, 17 and 21, while the eosinophilic variant of ChRCCs appears to be almost completely diploid [53]. Oncocytomas, on the other hand, showed very few copy number alterations, with chromosome 1 deletion being the most frequent [53,67]. We believe that in distinguishing ChRCC from its mimickers such as oncocytoma, careful histologic examination along with immunohistochemistry (as needed) may serve as a baseline approach.

Finally, other ancillary methods such as novel and emerging markers (*i.e.*, *FOXI1* and *LINC01187*) and molecular testing may be helpful, while a concise discussion of pertinent clinicopathologic features with clinicians is paramount in ambiguous and unresolved cases.

## 9. Management implications for ChRCCs and related oncocytic neoplasms

The clinical management of ChRCCs is relatively well established, while in some areas it continues to evolve. Similar to CCRCs, the general approach for the treatment of localized and resectable ChRCCs is surgery, with no role for adjuvant therapy after definitive surgery for clinical Stages I–III [91]. For advanced or metastatic ChRCCs, systemic therapies with vascular endothelial growth factor (VEGF) inhibitors (*i.e.*, sunitinib) or mTOR inhibitors (*i.e.*, everolimus) are currently preferred first-line options [91]. Enrollment to clinical trials, immunotherapy, and cytoreductive nephrectomy could also be considered, whenever feasible, as well as molecular characterization of metastatic ChRCCs, to improve systemic therapy strategy.

There are currently limited clinical data available on the recently described and emerging oncocytic renal neoplasms which fall within the differential diagnosis of ChRCC, such as EVTs and LOTs, as well as borderline oncocytic tumors. From a practical standpoint, we believe that it would be appropriate, for the time being, to clinically manage such tumors similar to those with low malignant potential for recurrence/metastasis, such as eosinophilic ChRCCs.

## 10. Conclusion

ChRCCs represent a heterogeneous group of neoplasms demonstrating unique morphologic and genetic profiles. Most ChRCCs carry a favorable clinical outcome, with the exception of those with sarcomatoid features. Conventional WHO/ISUP grading system does not apply to ChRCCs as histology alone is limited in predicting the behavior of these tumors in the absence of known adverse factors such as necrosis and sarcomatoid features. To distinguish ChRCCs from its mimickers, careful histologic examination along with a small set of immunohistochemical stains including CD117/CK7/CA-IX is likely to serve as an optimum baseline approach. In perplexing cases, genomic profiling may be helpful.

## Author contributions

*Study concept and design:* Reza Alaghehbandan, Rohit Mehra.

*Data acquisition:* Reza Alaghehbandan, Christopher G. Przybycin.

*Data analysis:* Reza Alaghehbandan, Christopher G. Przybycin, Virginie Verkarre.

*Drafting of manuscript:* Reza Alaghehbandan, Rohit Mehra.

*Critical revision of the manuscript:* Reza Alaghehbandan, Rohit Mehra.

## Conflicts of interest

The authors declare no conflict of interest.

## Acknowledgements

The authors of this manuscript were previously members of the Genitourinary Pathology Society (GUPS) working group for editing the chromophobe RCC section in the consensus GUPS update on renal neoplasia articles [75,92]. We would like to acknowledge and thank the GUPS executives, particularly, Dr. Kiril Trpkov (GUPS past-president) and Dr. Ondrej Hes, for leading the joint GUPS initiatives and forming various working groups including ours on chromophobe RCC. The statements and views expressed in this review article reflect the opinions of all co-authors only.

## References

- [1] Thoenes W, Storkel S, Rumpelt HJ. Human chromophobe cell renal carcinoma. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1985;48:207–17.
- [2] MacLennan GT, Cheng L. Five decades of urologic pathology: The accelerating expansion of knowledge in renal cell neoplasia. *Hum Pathol* 2020;95:24–45.
- [3] Storkel S, Steart PV, Drenckhahn D, Thoenes W. The human chromophobe cell renal carcinoma: Its probable relation to intercalated cells of the collecting duct. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1989;56:237–45.
- [4] Moch H, Humphrey PA, Ulbright TM, Reuter VE. WHO classification of tumours of the urinary system and male genital organs. Lyon, France: International Agency for Research on Cancer; 2016. p27–8.
- [5] Colomba E, Le Teuff G, Eisen T, Stewart GD, Fife K, Larkin J, et al. Metastatic chromophobe renal cell carcinoma treated with targeted therapies: A Renal Cross Channel Group study. *Eur J Cancer* 2017;80:55–62.
- [6] Ohashi R, Martignoni G, Hartmann A, Calio A, Segala D, Stohr C, et al. Multi-institutional re-evaluation of prognostic factors in chromophobe renal cell carcinoma: Proposal of a novel two-tiered grading scheme. *Virchows Arch* 2020;476:409–18.
- [7] Przybycin CG, Cronin AM, Darvishian F, Gopalan A, Al-Ahmadie HA, Fine SW, et al. Chromophobe renal cell carcinoma: A clinicopathologic study of 203 tumors in 200 patients with primary resection at a single institution. *Am J Surg Pathol* 2011;35:962–70.
- [8] Linehan WM, Ricketts CJ. The metabolic basis of kidney cancer. *Semin Cancer Biol* 2013;23:46–55.
- [9] Benusiglio PR, Giraud S, Deveaux S, Mejean A, Correas JM, Joly D, et al. Renal cell tumour characteristics in patients with the Birt-Hogg-Dubé cancer susceptibility syndrome: A retrospective, multicentre study. *Orphanet J Rare Dis* 2014;9:163. <https://doi.org/10.1186/s13023-014-0163-z>.
- [10] Pavlovich CP, Walther MM, Eyler RA, Hewitt SM, Zbar B, Linehan WM, et al. Renal tumors in the Birt-Hogg-Dubé syndrome. *Am J Surg Pathol* 2002;26:1542–52.
- [11] Iribar Y, Yao M, Tanaka R, Kuroda N, Nagashima Y, Nakatani Y, et al. Genome-wide uniparental disomy and copy number variations in renal cell carcinomas associated with Birt-Hogg-Dubé syndrome. *Am J Pathol* 2016;186:337–46.
- [12] Yang P, Cornejo KM, Sadow PM, Cheng L, Wang M, Xiao Y, et al. Renal cell carcinoma in tuberous sclerosis complex. *Am J Surg Pathol* 2014;38:895–909.
- [13] Guo J, Tretiakova MS, Troxell ML, Osunkoya AO, Fadare O, Sangui AR, et al. Tuberous sclerosis-associated renal cell carcinoma: A clinicopathologic study of 57 separate carcinomas in 18 patients. *Am J Surg Pathol* 2014;38:1457–67.
- [14] Shuch B, Ricketts CJ, Vocke CD, Komiya T, Middleton LA, Kauffman EC, et al. Germline PTEN mutation Cowden syndrome: An underappreciated form of hereditary kidney cancer. *J Urol* 2013;190:1990–8.
- [15] Cheville JC, Lohse CM, Zincke H, Weaver AL, Blute ML. Comparisons of outcome and prognostic features among histologic subtypes of renal cell carcinoma. *Am J Surg Pathol* 2003;27:612–24.
- [16] Volpe A, Novara G, Antonelli A, Bertini R, Billia M, Carmignani G, et al. Chromophobe renal cell carcinoma (RCC): Oncological outcomes and prognostic factors in a large multicentre series. *BJU Int* 2012;110:76–83.
- [17] Yip SM, Ruiz Morales JM, Donskov F, Fraccon A, Basso U, Rini BI, et al. Outcomes of metastatic chromophobe renal cell carcinoma (chrRCC) in the targeted therapy era: Results from the International Metastatic Renal Cell Cancer Database Consortium (IMDC). *Kidney Cancer* 2017;1:41–7.
- [18] Ged Y, Chen YB, Knezevic A, Casuscelli J, Redzematovic A, DiNatale RG, et al. Metastatic chromophobe renal cell carcinoma: Presence or absence of sarcomatoid differentiation determines clinical course and treatment outcomes. *Clin Genitourin Cancer* 2019;17:e678–88. <https://doi.org/10.1016/j.clgc.2019.03.018>.
- [19] Lee W. Imprint cytology of the chromophobe renal cell carcinoma: Correlation with the histological and ultrastructural features. *J Cytol* 2011;28:77–80.
- [20] Tickoo SK, Lee MW, Eble JN, Amin M, Christopherson T, Zarbo RJ, et al. Ultrastructural observations on mitochondria and microvesicles in renal oncocytoma, chromophobe renal cell carcinoma, and eosinophilic variant of conventional (clear cell) renal cell carcinoma. *Am J Surg Pathol* 2000;24:1247–56.
- [21] Thoenes W, Storkel S, Rumpelt HJ, Moll R, Baum HP, Werner S. Chromophobe cell renal carcinoma and its variants—a report on 32 cases. *J Pathol* 1988;155:277–87.
- [22] Cheville JC, Lohse CM, Zincke H, Weaver AL, Leibovich BC, Frank I, et al. Sarcomatoid renal cell carcinoma: An examination of underlying histologic subtype and an analysis of associations with patient outcome. *Am J Surg Pathol* 2004;28:435–41.
- [23] Amin MB, Paner GP, Alvarado-Cabrero I, Young AN, Stricker HJ, Lyles RH, et al. Chromophobe renal cell carcinoma: Histomorphologic characteristics and evaluation of conventional pathologic prognostic parameters in 145 cases. *Am J Surg Pathol* 2008;32:1822–34.
- [24] de Peralta-Venturina M, Moch H, Amin M, Tamboli P, Hailemariam S, Mihatsch M, et al. Sarcomatoid differentiation in renal cell carcinoma: A study of 101 cases. *Am J Surg Pathol* 2001;25:275–84.
- [25] Peckova K, Martinek P, Ohe C, Kuroda N, Bulimbasic S, Condom Mundo E, et al. Chromophobe renal cell carcinoma with neuroendocrine and neuroendocrine-like features. Morphologic, immunohistochemical, ultrastructural, and array comparative genomic hybridization analysis of 18 cases and review of the literature. *Ann Diagn Pathol* 2015;19:261–8.
- [26] Michal M, Hes O, Svec A, Ludvikova M. Pigmented microcystic chromophobe cell carcinoma: A unique variant of renal cell carcinoma. *Ann Diagn Pathol* 1998;2:149–53.
- [27] Hes O, Vanecek T, Perez-Montiel DM, Alvarado Cabrero I, Hora M, Suster S, et al. Chromophobe renal cell carcinoma with microcystic and adenomatous arrangement and pigmentation—a diagnostic pitfall. Morphological, immunohistochemical, ultrastructural and molecular genetic report of 20 cases. *Virchows Arch* 2005;446:383–93.
- [28] Durdik P, Pesl M, Povysil C, Tvrđík D, Pavlik I, Soukup V, et al. Pigmented microcystic chromophobe renal cell carcinoma. *Pathol Res Pract* 2007;203:593–7.
- [29] Kuroda N, Tamura M, Hes O, Michal M, Gatalica Z. Chromophobe renal cell carcinoma with neuroendocrine differentiation and sarcomatoid change. *Pathol Int* 2011;61:552–4.

- [30] Parada DD, Pena KB. Chromophobe renal cell carcinoma with neuroendocrine differentiation. *APMIS* 2008;116:859–65.
- [31] Michalova K, Tretiakova M, Pivovarcikova K, Alaghehbandan R, Perez Montiel D, Ulamec M, et al. Expanding the morphologic spectrum of chromophobe renal cell carcinoma: A study of 8 cases with papillary architecture. *Ann Diagn Pathol* 2019;44:151448. <https://doi.org/10.1016/j.anndiagpath.2019.151448>.
- [32] Taylor AS, Spratt DE, Dhanasekaran SM, Mehra R. Contemporary renal tumor categorization with biomarker and translational updates: A practical review. *Arch Pathol Lab Med* 2019;143:1477–91.
- [33] Williamson SR, Gill AJ, Argani P, Chen YB, Egevad L, Kristiansen G, et al. Report from the International Society of Urological Pathology (ISUP) consultation conference on molecular pathology of urogenital cancers: III: Molecular pathology of kidney cancer. *Am J Surg Pathol* 2020;44:e47–65. <https://doi.org/10.1097/PAS.0000000000001476>.
- [34] Skinnider BF, Jones EC. Renal oncocyтома and chromophobe renal cell carcinoma. A comparison of colloidal iron staining and electron microscopy. *Am J Clin Pathol* 1999;111:796–803.
- [35] Chen F, Zhang Y, Senbabaoglu Y, Ciriello G, Yang L, Reznik E, et al. Multilevel genomics-based taxonomy of renal cell carcinoma. *Cell Rep* 2016;14:2476–89.
- [36] Davis CF, Ricketts CJ, Wang M, Yang L, Cherniack AD, Shen H, et al. The somatic genomic landscape of chromophobe renal cell carcinoma. *Cancer Cell* 2014;26:319–30.
- [37] Skala SL, Wang X, Zhang Y, Mannan R, Wang L, Narayanan SP, et al. Next-generation RNA sequencing-based biomarker characterization of chromophobe renal cell carcinoma and related oncocytic neoplasms. *Eur Urol* 2020;78:63–74.
- [38] Lindgren D, Eriksson P, Krawczyk K, Nilsson H, Hansson J, Veerla S, et al. Cell-type-specific gene programs of the normal human nephron define kidney cancer subtypes. *Cell Rep* 2017;20:1476–89.
- [39] Drevik J, Abbosh P. Novel biomarkers in chromophobe renal cell carcinoma: Distinguishing it from its mimics. *Eur Urol* 2020;78:75–6.
- [40] Clark DJ, Dhanasekaran SM, Petralia F, Pan J, Song X, Hu Y, et al. Integrated proteogenomic characterization of clear cell renal cell carcinoma. *Cell* 2020;180:207. <https://doi.org/10.1016/j.cell.2019.12.026>.
- [41] Lopez-Beltran A, Montironi R, Cimadamore A, Cheng L. Grading of chromophobe renal cell carcinoma: Do we need it? *Eur Urol* 2021;79:232–3.
- [42] Avulova S, Cheville JC, Lohse CM, Gupta S, Potretzke TA, Tsivian M, et al. Grading chromophobe renal cell carcinoma: Evidence for a four-tiered classification incorporating coagulative tumor necrosis. *Eur Urol* 2021;79:225–31.
- [43] Cheville JC, Lohse CM, Sukov WR, Thompson RH, Leibovich BC. Chromophobe renal cell carcinoma: The impact of tumor grade on outcome. *Am J Surg Pathol* 2012;36:851–6.
- [44] Delahunt B, Cheville JC, Martignoni G, Humphrey PA, Magi-Galluzzi C, McKenney J, et al. The International Society of Urological Pathology (ISUP) grading system for renal cell carcinoma and other prognostic parameters. *Am J Surg Pathol* 2013;37:1490–504.
- [45] Delahunt B, McKenney JK, Lohse CM, Leibovich BC, Thompson RH, Boorjian SA, et al. A novel grading system for clear cell renal cell carcinoma incorporating tumor necrosis. *Am J Surg Pathol* 2013;37:311–22.
- [46] Paner GP, Amin MB, Alvarado-Cabrerо I, Young AN, Stricker HJ, Moch H, et al. A novel tumor grading scheme for chromophobe renal cell carcinoma: Prognostic utility and comparison with Fuhrman nuclear grade. *Am J Surg Pathol* 2010;34:1233–40.
- [47] Akhtar M, Chantziantoniou N. Quantitative image cell analysis of cytologic smears for DNA ploidy in renal parenchymal neoplasms. *Diagn Cytopathol* 1999;21:223–9.
- [48] Crotty TB, Lawrence KM, Moertel CA, Bartelt DH, Batts KP, Dewald GW, et al. Cytogenetic analysis of six renal oncocytomas and a chromophobe cell renal carcinoma. Evidence that –Y, –1 may be a characteristic anomaly in renal oncocytomas. *Cancer Genet Cytogenet* 1992;61:61–6.
- [49] Kovacs A, Kovacs G. Low chromosome number in chromophobe renal cell carcinomas. *Genes Chromosomes Cancer* 1992;4:267–8.
- [50] Kovacs A, Storkel S, Thoenes W, Kovacs G. Mitochondrial and chromosomal DNA alterations in human chromophobe renal cell carcinomas. *J Pathol* 1992;167:273–7.
- [51] Kovacs G, Soudah B, Hoene E. Binucleated cells in a human renal cell carcinoma with 34 chromosomes. *Cancer Genet Cytogenet* 1988;31:211–5.
- [52] Speicher MR, Schoell B, du Manoir S, Schrock E, Ried T, Cremer T, et al. Specific loss of chromosomes 1, 2, 6, 10, 13, 17, and 21 in chromophobe renal cell carcinomas revealed by comparative genomic hybridization. *Am J Pathol* 1994;145:356–64.
- [53] Alaghehbandan R, Trpkov K, Tretiakova M, Luis AS, Rogala JD, Hes O. Comprehensive review of numerical chromosomal aberrations in chromophobe renal cell carcinoma including its variant morphologies. *Adv Anat Pathol* 2021;28:8–20.
- [54] Linehan WM, Ricketts CJ. The cancer genome atlas of renal cell carcinoma: Findings and clinical implications. *Nat Rev Urol* 2019;16:539–52.
- [55] Brunelli M, Gobbo S, Cossu-Rocca P, Cheng L, Hes O, Delahunt B, et al. Chromosomal gains in the sarcomatoid transformation of chromophobe renal cell carcinoma. *Mod Pathol* 2007;20:303–9.
- [56] Kang XL, Zou H, Pang LJ, Hu WH, Zhao J, Qi Y, et al. Chromosomal imbalances revealed in primary renal cell carcinomas by comparative genomic hybridization. *Int J Clin Exp Pathol* 2015;8:3636–47.
- [57] Ren Y, Liu K, Kang X, Pang L, Qi Y, Hu Z, et al. Chromophobe renal cell carcinoma with and without sarcomatoid change: A clinicopathological, comparative genomic hybridization, and whole-exome sequencing study. *Am J Transl Res* 2015;7:2482–99.
- [58] Sperga M, Martinek P, Vanecek T, Grossmann P, Bauleth K, Perez-Montiel D, et al. Chromophobe renal cell carcinomas—chromosomal aberration variability and its relation to Paner grading system: An array CGH and FISH analysis of 37 cases. *Virchows Arch* 2013;463:563–73.
- [59] Tan MH, Wong CF, Tan HL, Yang XJ, Ditlev J, Matsuda D, et al. Genomic expression and single-nucleotide polymorphism profiling discriminates chromophobe renal cell carcinoma and oncocytoma. *BMC Cancer* 2010;10:196. <https://doi.org/10.1186/1471-2407-10-196>.
- [60] Verdorfer I, Hobisch A, Hittmair A, Duba HC, Bartsch G, Utermann G, et al. Cytogenetic characterization of 22 human renal cell tumors in relation to a histopathological classification. *Cancer Genet Cytogenet* 1999;111:61–70.
- [61] Vieira J, Henrique R, Ribeiro FR, Barros-Silva JD, Peixoto A, Santos C, et al. Feasibility of differential diagnosis of kidney tumors by comparative genomic hybridization of fine needle aspiration biopsies. *Genes Chromosomes Cancer* 2010;49:935–47.
- [62] Liu YJ, Ussaklı C, Antic T, Liu Y, Wu Y, True L, et al. Sporadic oncocytic tumors with features intermediate between oncocytoma and chromophobe renal cell carcinoma: Comprehensive clinicopathological and genomic profiling. *Hum Pathol* 2020;104:18–29.
- [63] Gunawan B, Bergmann F, Braun S, Hemmerlein B, Ringert RH, Jakse G, et al. Polyploidization and losses of chromosomes 1, 2, 6, 10, 13, and 17 in three cases of chromophobe renal cell carcinomas. *Cancer Genet Cytogenet* 1999;110:57–61.
- [64] Foix MP, Dunatov A, Martinek P, Mundo EC, Suster S, Sperga M, et al. Morphological, immunohistochemical, and chromosomal

- analysis of multicystic chromophobe renal cell carcinoma, an architecturally unusual challenging variant. *Virchows Arch* 2016;469:669–78.
- [65] Ohe C, Kuroda N, Matsuura K, Kai T, Moriyama M, Sugiguchi S, et al. Chromophobe renal cell carcinoma with neuroendocrine differentiation/morphology: A clinicopathological and genetic study of three cases. *Hum Pathol: Case Reports* 2014; 1:31–9.
- [66] Mokhtar GA, Al-Zahrani R. Chromophobe renal cell carcinoma of the kidney with neuroendocrine differentiation: A case report with review of literature. *Urol Ann* 2015;7:383–6.
- [67] Durinck S, Stawiski EW, Pavia-Jimenez A, Modrusan Z, Kapur P, Jaiswal BS, et al. Spectrum of diverse genomic alterations define non-clear cell renal carcinoma subtypes. *Nat Genet* 2015;47:13–21.
- [68] Ricketts CJ, De Cubas AA, Fan H, Smith CC, Lang M, Reznik E, et al. The cancer genome atlas comprehensive molecular characterization of renal cell carcinoma. *Cell Rep* 2018;23: 313–326 e5. <https://doi.org/10.1016/j.celrep.2018.03.075>.
- [69] Casuscelli J, Becerra MF, Manley BJ, Zabor EC, Reznik E, Redzematovic A, et al. Characterization and impact of TERT promoter region mutations on clinical outcome in renal cell carcinoma. *Eur Urol Focus* 2019;5:642–9.
- [70] Ricketts CJ, De Cubas AA, Fan H, Smith CC, Lang M, Reznik E, et al. The cancer genome atlas comprehensive molecular characterization of renal cell carcinoma. *Cell Rep* 2018;23: 3698. <https://doi.org/10.1016/j.celrep.2018.06.032>.
- [71] Ohashi R, Schraml P, Angori S, Batavia AA, Rupp NJ, Ohe C, et al. Classic chromophobe renal cell carcinoma incur a larger number of chromosomal losses than seen in the eosinophilic subtype. *Cancers* 2019;11:1492. <https://doi.org/10.3390/cancers11101492>.
- [72] Roldan-Romero JM, Santos M, Lanillos J, Caleiras E, Anguera G, Maroto P, et al. Molecular characterization of chromophobe renal cell carcinoma reveals mTOR pathway alterations in patients with poor outcome. *Mod Pathol* 2020;33:2580–90.
- [73] Casuscelli J, Weinhold N, Gundem G, Wang L, Zabor EC, Drill E, et al. Genomic landscape and evolution of metastatic chromophobe renal cell carcinoma. *JCI Insight* 2017;2:e92688. <https://doi.org/10.1172/jci.insight.92688>.
- [74] Gill AJ, Hes O, Papathomas T, Sedivcova M, Tan PH, Agaimy A, et al. Succinate dehydrogenase (SDH)-deficient renal carcinoma: A morphologically distinct entity: A clinicopathologic series of 36 tumors from 27 patients. *Am J Surg Pathol* 2014; 38:1588–602.
- [75] Trpkov K, Williamson SR, Gill AJ, Adeniran AJ, Agaimy A, Alaghehbandan R, et al. Novel, emerging and provisional renal entities: The Genitourinary Pathology Society (GUPS) update on renal neoplasia. *Mod Pathol* 2021;34:1167–84.
- [76] Trpkov K, Hes O, Bonert M, Lopez JI, Bonsib SM, Nesi G, et al. Eosinophilic, solid, and cystic renal cell carcinoma: Clinicopathologic study of 16 unique, sporadic neoplasms occurring in women. *Am J Surg Pathol* 2016;40:60–71.
- [77] Siadat F, Trpkov K. ESC, ALK, HOT and LOT: Three letter acronyms of emerging renal entities knocking on the door of the WHO classification. *Cancers* 2020;12:168. <https://doi.org/10.3390/cancers12010168>.
- [78] Trpkov K, Williamson SR, Gao Y, Martinek P, Cheng L, Sangi AR, et al. Low-grade oncocytic tumour of kidney (CD117-negative, cytokeratin 7-positive): A distinct entity? *Histopathology* 2019;75:174–84.
- [79] Tjota M, Chen H, Parilla M, Wanjari P, Segal J, Antic T. Eosinophilic renal cell tumors with a TSC and MTOR gene mutations are morphologically and immunohistochemically heterogenous: Clinicopathologic and molecular study. *Am J Surg Pathol* 2020;44:943–54.
- [80] Lerma LA, Schade GR, Tretiakova MS. Co-existence of ESC-RCC, EVT, and LOT as synchronous and metachronous tumors in six patients with multifocal neoplasia, but without clinical features of tuberous sclerosis complex. *Hum Pathol* 2021;116: 1–11.
- [81] Kravtsov O, Gupta S, Cheville JC, Sukov WR, Rowsey R, Herrera-Hernandez LP, et al. Low-grade oncocytic tumor of kidney (CK7-positive, CD117-negative): Incidence in a single institutional experience with clinicopathological and molecular characteristics. *Hum Pathol* 2021;114:9–18.
- [82] Sukov WR, Hodge JC, Lohse CM, Akre MK, Leibovich BC, Thompson RH, et al. ALK alterations in adult renal cell carcinoma: Frequency, clinicopathologic features and outcome in a large series of consecutively treated patients. *Mod Pathol* 2012;25:1516–25.
- [83] Kuroda N, Trpkov K, Gao Y, Tretiakova M, Liu YJ, Ulamec M, et al. ALK rearranged renal cell carcinoma (ALK-RCC): A multi-institutional study of twelve cases with identification of novel partner genes CLIP1, KIF5B and KIAA1217. *Mod Pathol* 2020;33: 2564–79.
- [84] He H, Trpkov K, Martinek P, Isikci OT, Maggi-Galuzzi C, Alaghehbandan R, et al. "High-grade oncocytic renal tumor": Morphologic, immunohistochemical, and molecular genetic study of 14 cases. *Virchows Arch* 2018;473:725–38.
- [85] Chen YB, Mirsadraei L, Jayakumaran G, Al-Ahmadi HA, Fine SW, Gopalan A, et al. Somatic mutations of TSC2 or MTOR characterize a morphologically distinct subset of sporadic renal cell carcinoma with eosinophilic and vacuolated cytoplasm. *Am J Surg Pathol* 2019;43:121–31.
- [86] Kapur P, Gao M, Zhong H, Rakheja D, Cai Q, Pedrosa I, et al. Eosinophilic vacuolated tumor of the kidney: A review of evolving concepts in this novel subtype with additional insights from a case with MTOR mutation and concomitant chromosome 1 loss. *Adv Anat Pathol* 2021;28:251–7.
- [87] van Oostenbrugge TJ, Futterer JJ, Mulders PFA. Diagnostic imaging for solid renal tumors: A pictorial review. *Kidney Cancer* 2018;2:79–93.
- [88] Sanchez A, Feldman AS, Hakimi AA. Current management of small renal masses, including patient selection, renal tumor biopsy, active surveillance, and thermal ablation. *J Clin Oncol* 2018;36:3591–600.
- [89] Wobker SE, Williamson SR. Modern pathologic diagnosis of renal oncocytoma. *J Kidney Cancer VHL* 2017;4:1–12.
- [90] Williamson SR, Gadde R, Trpkov K, Hirsch MS, Srigley JR, Reuter VE, et al. Diagnostic criteria for oncocytic renal neoplasms: A survey of urologic pathologists. *Hum Pathol* 2017;63:149–56.
- [91] Garje R, Elhag D, Yasin HA, Acharya L, Vaena D, Dahmoush L. Comprehensive review of chromophobe renal cell carcinoma. *Crit Rev Oncol Hematol* 2021;160:103287. <https://doi.org/10.1016/j.critrevonc.2021.103287>.
- [92] Trpkov K, Hes O, Williamson SR, Adeniran AJ, Agaimy A, Alaghehbandan R, et al. New developments in existing WHO entities and evolving molecular concepts: The Genitourinary Pathology Society (GUPS) update on renal neoplasia. *Mod Pathol* 2021;34:1392–424.