



Review

# Connections between Immune-Derived Mediators and Sensory Nerves for Itch Sensation

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**Abstract:** Although histamine is a well-known itch mediator, histamine H<sub>1</sub>-receptor blockers often lack efficacy in chronic itch. Recent molecular and cellular based studies have shown that non-histaminergic mediators, such as proteases, neuropeptides and cytokines, along with their cognate receptors, are involved in evocation and modulation of itch sensation. Many of these molecules are produced and secreted by immune cells, which act on sensory nerve fibers distributed in the skin to cause itching and sensitization. This understanding of the connections between immune cell-derived mediators and sensory nerve fibers has led to the development of new treatments for itch. This review summarizes current knowledge of immune cell-derived itch mediators and neuronal response mechanisms, and discusses therapeutic agents that target these systems.

**Keywords:** cytokines; immune cell; itch mediator and modulator; sensory neuron



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## 1. Introduction

Itch (or pruritus) is an unpleasant sensation inducing the desire to scratch [1], as well as being a major and distressing symptom of many skin and systemic diseases. Chronic itch represents a significant clinical problem resulting from renal [2], liver [3], and bowel diseases [4], as well as several serious skin diseases, such as atopic dermatitis (AD). Histamine is one of the best-evaluated itch mediators. If an itch is caused by histamine, antihistamines (histamine H<sub>1</sub>-receptor blockers) can be used to control it. However, recent studies have suggested that histamine-independent pathways are involved in chronic itch, making antihistamines ineffective in the treatment of these patients [5–7]. Thus, the mechanisms of itch development and enhancement other than through histamine remain to be determined. Analyses of the interactions between immune cells and sensory neurons have shown that cytokines produced by immune cells during inflammation enhance itch, and that they act directly on sensory nerve fibers to induce and/or sensitize itch sensation.

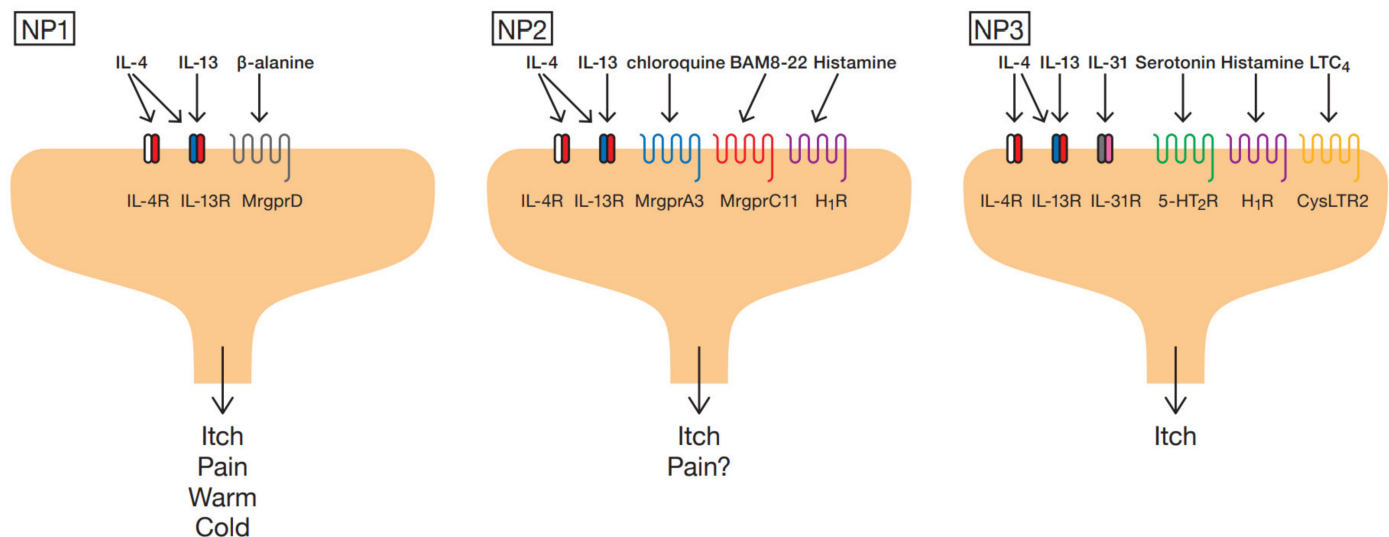
This review focuses on immune cell-derived itch mediators and describes the mechanisms by which they connect to sensory nerves to produce and enhance itch.

## 2. Subtype of Sensory Neurons

Generally, itch sensation is generated by the binding of itch-inducing substances (pruritogens) to their cognate receptors (pruriceptors) on peripheral sensory afferents, especially unmyelinated C-fibers [8]. Single-cell RNA-seq has classified the sensory neuron system into five neurofilament (NF)-containing clusters, two peptidergic (PEP) nociceptor clusters, a tyrosine hydroxylase (TH)-containing cluster and three non-peptidergic (NP) nociceptor clusters [9]. The NF clusters were shown to express neurofilament heavy chain (*Nefh*) and parvalbumin (*Pvalb*), molecules previously associated with myelinated dorsal

root ganglion (DRG) neurons. The PEP clusters were found to express substance P (SP, also known as *Tac1*), TRKA (*Ntrk1*) and calcitonin gene-related peptide (CGRP, also known as *Calca*), molecules previously associated with peptidergic nociceptors. The TH cluster showed distinct expression of tyrosine hydroxylase (*Th*), which is also expressed in a distinct subclass of unmyelinated neurons. Finally, the NP clusters were found to express Mas-related G protein coupled receptor D (*Mrgprd*) and *P2rx3*, molecules previously associated with nonpeptidergic nociceptors. Notably, NP clusters express receptor genes for itch mediators.

NP1 expresses the  $\beta$ -alanine receptor *Mrgprd* [10] and the lysophosphatidic acid receptors *Lpar3* and *Lpar5*. Chloroquine (CQ) receptor (*Mrgpra3*) and bovine adrenal medulla (BAM) 8–22 receptor (*Mrgprx1*:human, *Mrgprc11*:mice) [11] are expressed on NP2; whereas the interleukin (IL)-31 receptor *Il31ra*, the oncostatin M receptor (*OSM*), the leukotriene (LT) C<sub>4</sub> receptor *Cysltr2* [12] and the serotonin receptors *Htr1f* and *Htr2a* are expressed on NP3. Histamine receptor (*H1R*) was detected on NP2 and NP3 [9] (Figure 1). In addition, NP1, NP2, and NP3 were found to be more enriched in neurons that express *Il4ra* and *Il13ra1* than in other types of neurons such as NF and PEP [13].



**Figure 1.** NP clusters of itch-related sensory nerves and itch-related receptors expressed on them. NP1 neurons are positive for IL-4R $\alpha$ , IL-13 R $\alpha$  and MrgprD (left). NP2 neurons are positive for IL-4R $\alpha$ , IL-13 R $\alpha$ , MrgprA3, MrgprC11 and H<sub>1</sub>R (middle). NP3 neurons are positive for IL-4R $\alpha$ , IL-13 R $\alpha$ , IL-31R, 5-HT<sub>2</sub>R, H<sub>1</sub>R and CysLTR2 (right).

### 3. Itch Mediators and Modulators from Immune Cells

Tables 1 and 2 summarize the immune cell-derived itch mediators and modulators, and the therapeutic agents that target them. This section describes the itch mediators and modulators produced by immune cells. As detailed above, the primary sensory nerves associated with itch have been classified into at least three subtypes, each of which has its own response profile. Based on the subtypes of nerve cells, the itch mediators and modulators derived from immune cells are also summarized (Figure 2).

#### 3.1. Amines

##### 3.1.1. Histamine

Histamine, the most well-known pruritogen, is produced by mast cells, basophils and keratinocytes [14–21]. Histamine evokes itch via histamine H<sub>1</sub> and H<sub>4</sub> receptors [19,22]. Histamine H<sub>1</sub> receptor (H<sub>1</sub>R) is a G protein-coupled receptor (GPCR) [20,23–25], a class of receptors globally expressed in various tissues, including sensory nerves [17,21]. Histamine H<sub>4</sub> receptor (H<sub>4</sub>R) is also a GPCR [20,24,25] and is mainly expressed in immunocompetent cells, including mast cells, eosinophils, neutrophils, monocytes, dendritic cells (DCs) and T cells; as well as in intestinal epithelia, spleen, lung, synovial tissue, the central nervous

system (CNS), sensory neurons, and cancer cells [21,24,26]. H<sub>1</sub>R and H<sub>4</sub>R on histaminergic nerves bind histamine and then activate transient receptor potential vanilloid (TRPV) 1 [17,27]. The H<sub>4</sub>R antagonist, ZPL-3893787, improved AD symptoms including itch [28].

A H<sub>3</sub>R inverse agonist was found to induce strong itch in mice. This H<sub>3</sub>R inverse agonist induced pruritus but could be completely blocked by combined treatment with an H<sub>1</sub>R and an H<sub>4</sub>R antagonist, whereas the H<sub>2</sub>R antagonist failed to inhibit the scratch response. The decreased threshold in response to H<sub>3</sub>R antagonism is thought to activate H<sub>1</sub>R and H<sub>4</sub>R on sensory neurons, leading to the excitation of histamine-sensitive afferents and eliciting a sensation of itch [29].

### 3.1.2. Serotonin

Serotonin (5-hydroxytryptamine; 5-HT), which is produced by mast cells, basophils and platelets [15,30–33], evokes scratching in rodents via the 5-HT<sub>2</sub> receptor, which is TRPV4-dependent [34–37]. The 5-HT<sub>2</sub> receptor is expressed in immunocompetent cells, including macrophages, DCs, Langerhans cells, CD3<sup>+</sup> T cells, melanocytes, vascular smooth muscle cells, endothelial cells, central and peripheral neurons including primary sensory neurons (DRG/trigeminal ganglion cells) [31,38–40]. Activation of the 5-HT<sub>2</sub> receptor, which belongs to the GPCR super-family and is coupled to the Gq/11 protein, leads to increases in inositol trisphosphate (IP<sub>3</sub>) and diacylglycerol (DG) levels, generating an antinociceptive effect [38,40]. Sertraline, a selective serotonin reuptake inhibitor, has been found to be effective in treating serotonin-targeted itch [41]. In addition, existing drugs, such as the selective 5-HT<sub>2</sub> receptor antagonist sarpogrelate, may have expanded future clinical application in the treatment of itch.

## 3.2. Proteases

### 3.2.1. Tryptase

Tryptase, a serine protease with trypsin-like specificity, consists of seven distinct isoforms,  $\alpha$ ,  $\beta$ I,  $\beta$ II,  $\beta$ III,  $\delta$ ,  $\epsilon$  and  $\gamma$ , encoded by a set of protease genes clustered together on chromosome 16p13.3 [42–45]. The tryptase best characterized to date is  $\beta$ -tryptase, and the term “tryptase” is often used as a synonym for this molecule [45]. Tryptase is expressed in mast cells and basophils [45–49]. Intradermal injection of tryptase elicits scratching in mice [50]. Proteases, including tryptase, activate protease-activated receptors (PARs) by cleaving a part of their extracellular domain. PARs are GPCRs, characterized by a unique mechanism of self-activation following specific proteolytic cleavage of their extracellular domains. To date, four PARs have been identified, PAR-1, PAR-2, PAR-3, and PAR-4, which are encoded by the *F2R*, *F2RL1*, *F2RL2*, and *F2RL3* genes, respectively [51,52]. PAR-2 is activated by trypsin-like serine proteases and is widely distributed throughout the mammalian body. In the skin, PAR-2 is expressed by almost all cell types, especially by keratinocytes. In addition, endothelial cells, fibroblasts, sensory neurons, and inflammatory cells such as mast cells, T lymphocytes, eosinophils, neutrophils, monocytes, macrophages, and DCs have been reported to express functional PAR-2 [52]. Tethered ligands, such as the PAR-2 agonist SLIGRL-NH<sub>2</sub>, have been shown to elicit scratching in mice, but not rats [53]. Activated PAR-2 coactivates TRPV1 channels stimulating the release of the neuropeptides SP and CGRP from nerve terminals [54,55]. In addition, SLIGRL-NH<sub>2</sub> enhances CQ- and BAM8-22-induced itch and acts as a modulator [56].

### 3.2.2. Chymase

Chymase is a chymotrypsin-like serine endopeptidase stored in mast cell secretory granules [18]. Human chymase, encoded by the *CMA1* gene located on chromosome 14q11.2, co-localizes with clusters formed by cathepsin G, granzyme B and granzyme C/H [46,57,58]. In rats, the chymase-encoding gene is located on chromosome 15p12/13, and in mice on chromosome 14C1/2 [58–62]. Chymase also activates PAR-2 [63,64]. The chymase specific inhibitor Y-40613 was found to suppress scratching behavior in

a mouse model of pruritus [65]. In the eyes, chymase also induced scratching behavior, which was suppressed by the selective chymase inhibitor ONO-WH-236 [64].

### 3.2.3. Cathepsin S

Cathepsin S is a cysteine protease produced by DCs, macrophages, basophils and keratinocytes [19,66,67]. Cathepsin S activates PAR-2, PAR-4 and MrgprC11 to produce itch [68–70]. Intradermal injection of the selective PAR-4 agonist AYPGKF-NH<sub>2</sub> (AYP) elicited scratching behavior in mice [56,71], which was prevented by the selective PAR-4 antagonist (pepducin P4pal-10) [71]. AYP-induced itch was reduced by gastrin-releasing peptide (GRP), NK-1, TRPV1 and a TRPA1 antagonist. These results indicated that PAR-4-activated itch is induced via TRPV1/TRPA1 in mice [71]. Moreover, touch-evoked scratching (alloknesis) was observed following intradermal injection of AYP, but not PAR-2 [56]. Cathepsin S also evoked a calcium response in mouse DRG neurons, which is reduced by PAR-2 antagonists and in TRPV1-/-or TRPA1-/-mouse-derived DRGs. In addition, intradermal injection of cathepsin S induced scratching behavior, which was inhibited by the cathepsin S inhibitor LHVS [70].

## 3.3. Peptides

### 3.3.1. Substance P

Substance P (SP) is a short neuropeptide of the tachykinin family, consisting of 11 amino acids (Arg-Pro-Lys-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>), and is one of most potent pruritogens identified to date [72,73]. SP is expressed by many cell types, including sensory neurons, astrocytes, microglia, epithelial cells, endothelial cells and immune cells, including T cells, macrophages, DCs and eosinophils [11,20,74]. SP binds to neurokinin 1 receptor (NK-1R) and another class of receptors involved in itch signaling, consisting of mouse MrgprA1, mouse MrgprB2 and human MrgprX2. NK-1R is a tachykinin receptor belonging to the GPCR family and expressed in the CNS, keratinocytes, fibroblasts and mast cells [72,73]. In humans, SP promotes degranulation by binding to mast cell NK-1R, releasing histamine and LTB<sub>4</sub> and causing itch [22,73]. In mice, SP induces itch through direct action on primary sensory neurons, as well as by release of nitric oxide (NO) and LTB<sub>4</sub> from keratinocytes, rather than by mast cell degranulation [22,75].

### 3.3.2. Endothelin-1

Endothelin (ET)-1 is a 21 amino-acid peptide member of the endothelin family and a potent pruritogen that can elicit scratching at low concentration (10–400 pmol/site) [76–78]. ET-1 is produced by mast cells, endothelial cells and keratinocytes in the skin [54,76–78]. ETs have two active receptors, ET<sub>A</sub> and ET<sub>B</sub>, which belong to the GPCR superfamily [78–80]. Endothelin receptors are widely expressed in all tissues [81], and ET-1-evoked scratching is mediated by ET<sub>A</sub> [76]. In addition, the endothelin receptor antagonist bosentan inhibited symptoms including itch in AD model mice [82].

## 3.4. Cytokines

### 3.4.1. IL-2

Interleukin (IL)-2 is a 15.5 kDa cytokine secreted by antigen-activated CD4<sup>+</sup> T cells and mast cells [83–86]. It was first described as a T cell growth factor and later also found to have the ability to act on natural killer (NK) cells and NKT cells, to activate B cells, and to induce the proliferation of regulatory T cells (Tregs), innate lymphoid cells (ILCs) and effector T cells. IL-2 has three receptors, each of which is composed of three subunits: IL-2 receptor  $\alpha$  (IL-2R $\alpha$ , CD25), IL-2R $\beta$  (CD122), and IL-2R $\gamma$  (CD132). IL-2R $\alpha$  is expressed by several types of immune cells, including Tregs, ILC2, activated CD4<sup>+</sup> and CD8<sup>+</sup> T cells, B cells, CD56<sup>hi</sup> NK cells, mature DCs, and endothelial cells. IL-2R $\beta$  is mainly expressed by multiple lymphoid populations, such as Tregs, memory CD8<sup>+</sup> T cells, NK cells, and NKT cells, and to some extent, by monocytes and neutrophils. IL-2R $\gamma$  is expressed mostly by hematopoietic cells [83,86,87]. The binding of IL-2 to its receptors induces trans-phosphorylation of Janus

kinase (JAK) 1 and JAK3. This, in turn, activates the JAK/signal transducer and activator of transcription (STAT), phosphoinositide (PI) 3-kinase and MAPK signaling pathways [86,87]. Intravenous IL-2 treatment has been approved for the treatment of patients with metastatic melanoma and renal cell carcinoma, with beneficial results in a subset of patients, although severe pruritus is a known side effect [83,86–89]. Moreover, intradermal injection of IL-2 in either healthy subjects or patients with AD induced pruritus and erythema [89,90]. The calcineurin inhibitor cyclosporine A has been shown to downregulate IL-2 synthesis, reducing pruritus in patients with treatment resistant Sezary syndrome, as well as in patients with AD [89].

#### 3.4.2. IL-4

IL-4 is a type 2 cytokine produced by T helper (Th) 2 cells, lymph node T follicular helper (Tfh) cells, mast cells, basophils, eosinophils and ILC2 [91–94]. IL-4 has two receptors, IL-4R $\alpha$  (CD124) and the more common IL-4R $\gamma$ , with IL-4 having higher affinity to IL-4R $\alpha$  [95]. IL-4 signals through the IL-4R $\alpha$ / $\gamma$ C complex in hematopoietic cells, such as lymphocytes and DCs. IL-4 binds IL-4R $\alpha$ / $\gamma$ C and activates the downstream signaling molecules JAK1/JAK3 and then STAT6. Non-hematopoietic cells including keratinocytes also express IL-4R $\alpha$ /IL-13R $\alpha$ 1 complex, which binds both IL-4 and IL-13, resulting in the downstream activation of JAK1/TYK2/JAK2 and then STAT6/STAT3 [93]. IL-4-evoked mouse DRG neurons respond to calcium, and deletion of IL-4Ra on sensory neurons was found to disrupt scratching behavior in a mouse model of AD. Moreover, IL-4 has been suggested as a modulator of itch because it enhances itch by increasing the neural responses induced by histamine, chloroquine, TSLP, and IL-31 [13,91]. Intradermal administration of IL-4 has also been reported to induce itching and allodynia [96,97]. Dupilumab, a monoclonal antibody that binds specifically to the shared alpha chain subunit of the IL-4 and IL-13 receptors, was associated with improvements in clinical end points, including reduced pruritus in AD [98].

#### 3.4.3. IL-13

IL-13 is another type 2 cytokine produced by Th2, ILC2, mast cells, basophils, and eosinophils [91–94]. It has two receptors, IL-13R $\alpha$ 1 (CD213 $\alpha$ 1) and IL-13R $\alpha$ 2 (CD213 $\alpha$ 2). IL-13R $\alpha$ 1 alone binds IL-13 with low affinity, but when paired with IL-4R $\alpha$  it binds IL-13 with high affinity and forms a functional IL-13 receptor that signals and results in activation of STAT3/6 [93,99]. Similar to IL-4, intradermal administration of IL-13 has been reported to induce itching and allodynia [96,97].

To date, the role of IL-13R $\alpha$ 2 in itch has been unclear. However, a recent study showed that the expression of IL-13R $\alpha$ 2 is upregulated in the skin of patients with AD, but not in the skin of patients with psoriasis, in a disease activity-dependent manner. In keratinocytes, IL-13 regulated IL-13R $\alpha$ 2 expression level and promoted IL-13R $\alpha$ 2 signaling. In addition, TLR2 activation was found to increase IL-13 mediated itch by potentiating IL-13R $\alpha$ 2, suggesting that IL-13R $\alpha$ 2 signaling promotes AD symptoms including itch [100]. Monoclonal antibodies that target and neutralize IL-13, Tralokinumab and Lebrikizumab, both improved AD symptoms including itch [28].

#### 3.4.4. IL-17

IL-17A, also called IL-17, is produced by various cell types of T cells, including the Th17 subset of CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells,  $\gamma\delta$  T cells, and NKT cells, as well as by immune cells such as lymphoid tissue inducer (LTi)-like cells and neutrophils, and nonimmune cells such as Paneth cells. IL-17 has two receptors, IL-17RA and IL-17RC, which form a heterodimer. Binding of IL-17A or an IL-17F heterodimer to IL-17R induces Act1 activation, which, in turn, activates multiple signaling cascades that operate through different TNF receptor-associated factor (TRAF) proteins. Subsequently, the complex associates with TRAF6, leading to the activation of NF- $\kappa$ B, MAPK-AP-1, and C/EBP. ERK1/2 mediates the phosphorylation of C/EBP $\beta$  at Thr188, with the CBAD of IL-17R also required for



IL-17-mediated inducible phosphorylation of C/EBP $\beta$  at Thr179 through GSK3 $\beta$ . IL-17 can also induce different feedback regulatory responses by inducing and/or recruiting deubiquitinase enzymes (A20 and USP25) or kinases (TBK1) [101,102]. Three randomized, controlled, phase 3 trials reported that brodalumab, an IL-17 receptor A antagonist, is safe and effective in treating moderate-to-severe psoriasis. In addition, brodalumab demonstrated improved itch responses in psoriasis [103]. These results suggest that IL-17 may act as an itch mediator and/or modulator. Other studies, however, have reported that IL-17 is neither a mediator nor a modulator of itching, [104] leading to the need for additional research.

#### 3.4.5. IL-23

IL-23 belongs to the IL-12 family of proinflammatory cytokines. IL-23 is heterodimeric, being composed of IL-12p40 and p19 molecules. It is produced by activated DCs and macrophages in response to microbial pathogens, with its production enhanced by interactions between the costimulatory molecule CD40 and its ligand. IL-23 signals via IL-12R $\beta$ 1 and IL-23R and mediates the phosphorylation of STAT3 and STAT4 by JAK2 and Tyk2 [105,106]. Intradermal injection of IL-23 did not induce scratching behavior, but calcium imaging showed that about 5% of DRG neurons in mice responded to IL-23. IL-23 was also found to attenuate histamine-induced itch, suggesting that this cytokine may function as a desensitizer [104]. In addition, IL-23 might play a role in regulating histaminergic itch by modulating TRPV1 activity [104].

#### 3.4.6. IL-31

IL-31, which belongs to the IL-6 family of cytokines, is produced by Th2 cells, mast cells, eosinophils, basophils, macrophages and DCs [19,89,107–111]. IL-31 binds to its receptor, a complex composed of IL-31 receptor A (IL-31RA) and oncostatin M (OSM) receptor, which is expressed on keratinocytes, epithelial cells, mast cells, basophils, eosinophils, macrophages, sensory neurons, DRG and the dorsal horn of the spinal cord [13,89,91,111]. IL31RA/OSMR is activated with similar affinities by OSM and IL-31. Binding of IL-31 leads to activation of diverse kinase pathways, including the JAK1/2/STAT3, ERK1/2, PI3K/Akt, p38 MAPK and JNK cascades [111–114]. *IL-31Tg* mice showed a marked and significant increase in cutaneous nerve fiber density in lesional skin compared with uninvolved or healthy skin [115]. In addition, cutaneous and intrathecal injections of IL-31 evoked intense itch, which was TRPV1 and TRPA1-dependent [113]. Moreover, a more recent study showed that transmembrane protein 184B (TMEM184B) is necessary for IL-31-induced itch [116]. Thus, the details of the mechanism of IL-31-induced itch are becoming clearer, and target molecules that can lead to treatment are being identified one after another.

### 3.5. Lipid Mediators

#### 3.5.1. PAF

Platelet-activating factor (PAF) is produced and released by mast cells, basophils, neutrophils, eosinophils, monocytes, macrophages, fibroblasts, platelets, endothelial cells, and cardiac muscle cells, all of which play important roles in inflammatory and thrombotic diseases. PAF is an inflammatory factor and has important functions in acute and chronic inflammation [117,118]. PAF receptor (PAFR) has been found in a host of cell membranes, including those of platelets, neutrophils, macrophages, mononuclear leukocytes, and eosinophils, as well as on hippocampal nerves, microglia, astrocytes, and oligodendrocyte progenitor cells [118]. Intradermal PAF injection evoked scratching behavior [35,119] and induced histamine release through degranulation of mast cells, contributing to itch accompanied by flare and wheal reactions [120].

#### 3.5.2. LTB<sub>4</sub>

Leukotrienes (LTs) are eicosanoid lipid mediators generated upon activation of both immune and structural cells such as epithelial cells and endothelial cells. LTB<sub>4</sub>, a 5-lipoxygenase

metabolite, is increased in the skin of AD model mice [121]. This molecule is produced and released by various types of immune cells, including mast cells, basophils, eosinophils, and macrophages [122–125]. LTB<sub>4</sub> has two receptors, BLT1 and BLT2, which are GPCR and present on cell surfaces, with BLT1 having higher affinity and activity than BLT2. BLT1 is mainly expressed by leukocytes and DRG neurons, whereas BLT2 is expressed on many tissues [126–128]. LTB<sub>4</sub>-induced DRG neurons respond to calcium, an effect inhibited by the LTB<sub>4</sub> antagonist ONO-4057 [128]. Intradermal LTB<sub>4</sub> injection induces scratching via TRPA1 and TRPV1 [129]. Moreover, the LTB<sub>4</sub> receptor antagonist CMHVA attenuated IL-31-induced scratching [130].

### 3.5.3. LTC<sub>4</sub>

LTC<sub>4</sub> is a cysteinyl LT produced and released by mast cells, basophils, and eosinophils [131–133]. Its receptors, CysLTR1 and CysLTR2, are widely expressed by hematopoietic and structural cells [12]. Basophils have been shown to release LTC<sub>4</sub> upon stimulation with antigen-specific IgE, which binds to CysLTR2 expressed on sensory nerve fibers (mainly NP3 nerves), evoking acute severe itch (itch flares) of AD [132]. Moreover, the LTC<sub>4</sub>/CysLTR2 pathway was shown to contribute not only to acute but also to chronic itch [12].

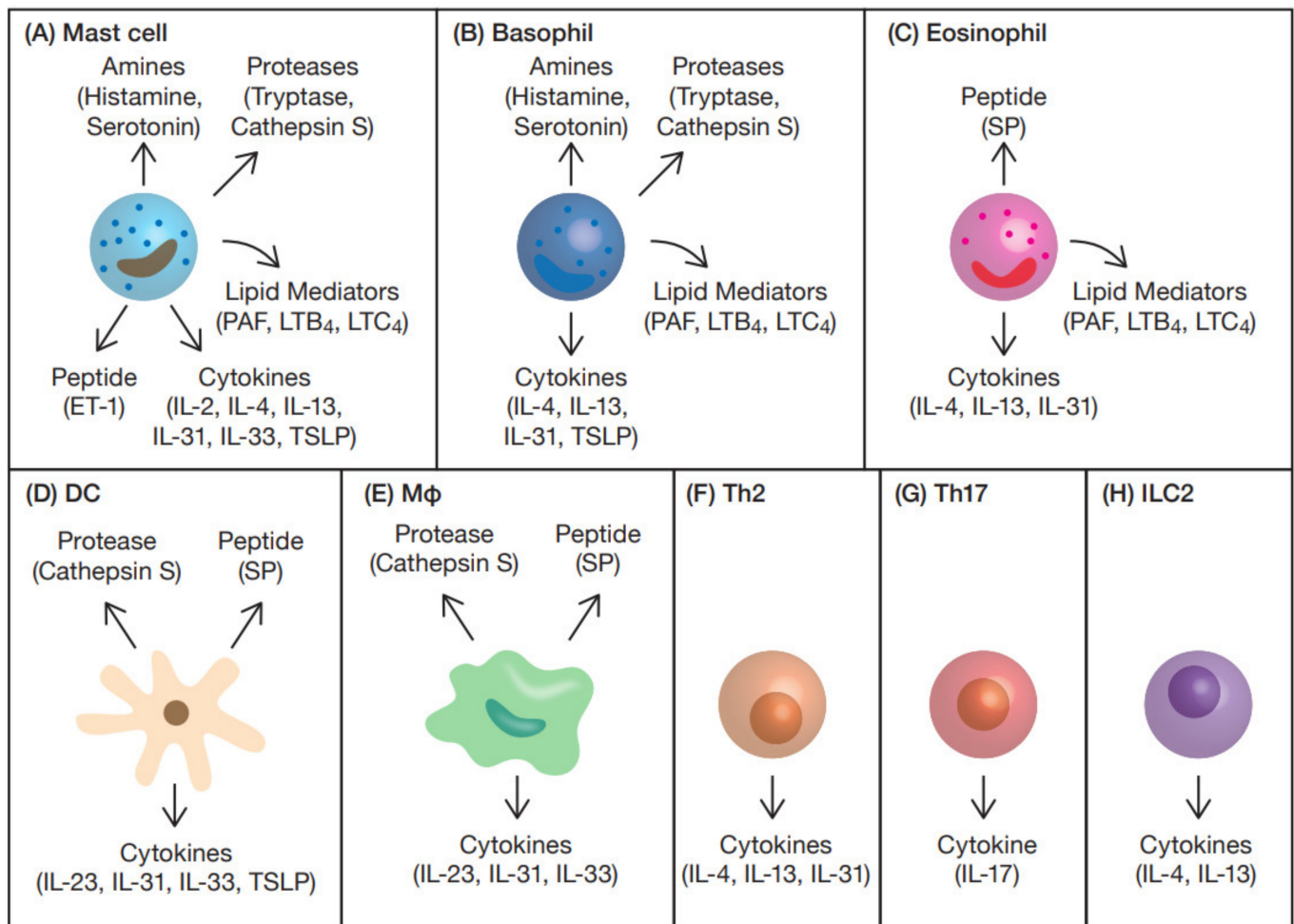
## 3.6. Others

### 3.6.1. IL-33

IL-33, a member of the IL-1 cytokine family, is considered important for host defenses and allergy by inducing Th2 cytokine production via the IL-33 receptor. This receptor is a heterodimer composed of IL-1 receptor-like 1 (IL-1RL1; also called ST2) and IL-1 receptor accessory protein (IL-1RAcP) molecules. IL-33 was first identified as a nuclear protein expressed in endothelial cell nuclei and was shown to be constitutively expressed in the nuclei of various cell types, such as endothelial and epithelial cells [134,135]. IL-33 was also recently shown to be constitutively expressed in other cells, including DCs, macrophages, mast cells, fibroblasts, smooth muscle cells, platelets and megakaryocytes [135,136]. ST2 expressing cells include basophils, mast cells, eosinophils, macrophages, DCs, NK cells, NKT cells, Th2 cells, cytotoxic T cells, Tregs, B cells, ILCs, microglia, astrocytes, neurons, epithelial cells, endothelial cells, and fibroblasts [135,137,138]. Treatment of AD model mice with anti-IL-33 antibody improved AD-like symptoms, including scratching behavior [139]. Moreover, IL-33/ST2 signaling was found to mediate chronic itch in a mouse model of contact hypersensitivity through the astrocytic JAK2/STAT3 cascade [140]. IL-33 was also shown to evoke calcium responses in neurons, with enhanced CQ evoking calcium responses [138]. Taken together, these findings suggested that IL-33 also functions as a modulator to enhance itch.

### 3.6.2. TSLP

Thymic stromal lymphopoietin (TSLP) is a IL-7 like cytokine belonging to the IL-2 cytokine family [110,141]. It is primarily produced by epithelial cells, including keratinocytes, fibroblasts and stromal cells, as well as by DCs, mast cells, and basophils [110,142]. Its receptor, TSLPR, is expressed on monocytes/macrophages, T cells, B cells, mast cells, eosinophils, NK cells, DCs, keratinocytes and sensory neuronal endings [143–148]. TSLPR is activated upon binding of TSLP, which activates JAK1/2 and STAT1/3/4/5/6 [149,150]. Intradermal injection of TSLP evoked scratching behavior. This is initiated by the binding of TSLP to TSLPR expressed on sensory nerve fibers. The TSLP-induced itch also required TRPA1, with the expression and release of keratinocyte-derived TSLP depending on the ORAI1/NFAT calcium signaling pathway [148]. Epithelial cell-derived cytokines, including TSLP and IL-33, strongly activate ILC2 and recruit Th2 cells into the skin. ILC2 and Th2 cells are rich sources of type 2 cytokines, which can initiate and perpetuate allergic skin inflammation, including itch, by recruiting basophils and eosinophils [91].



**Figure 2.** Immune cells and itch mediators and modulators. (A) Mast cells produce amines (histamine and serotonin), proteases (tryptase and cathepsin S), peptide (ET-1), cytokines (IL-2, IL-4, IL-13, IL-31, IL-33 and TSLP) and lipid mediators (PAF, LTB<sub>4</sub>, and LTC<sub>4</sub>). (B) Basophils produce amines (histamine and serotonin), proteases (tryptase and cathepsin S), cytokines (IL-4, IL-13, IL-31 and TSLP) and lipid mediators (PAF, LTB<sub>4</sub> and LTC<sub>4</sub>). (C) Eosinophils produce peptide (SP), cytokines (IL-4, IL-13 and IL-31) and lipid mediators (PAF, LTB<sub>4</sub>, LTC<sub>4</sub>). (D) DCs produce protease (cathepsin S), peptide (SP) and cytokines (IL-23, IL-31, IL-33 and TSLP). (E) Macrophages produce protease (cathepsin S), peptide (SP) and cytokines (IL-23, IL-31 and IL-33). (F) Th2 cells produce cytokines (IL-4, IL-13 and IL-31). (G) Th17 cells produce the cytokine IL-17. (H) ILC2 cells produce cytokines (IL-4 and IL-13).

#### 4. Immune System-Targeted Antipruritic Drugs

##### 4.1. Therapeutic Drugs for Amines

As described above, conventional treatments such as anti-histamines are often ineffective in patients with chronic pruritus. Therapeutic drugs other than antihistamines that target histamine consist of topical or systemic anti-inflammatory and immunomodulatory agents (e.g., cyclosporine A, pimecrolimus, tacrolimus and corticosteroids) [6]. Serotonin-targeted itch treatments include sertraline [41], but the clinical application of existing drugs such as sarpgrelate may also expand in the future.

##### 4.2. Therapeutic Drugs for Proteases

Protease-targeted therapies for itch are thought to be similar to histamine [6]. Furthermore, the selective chymase inhibitor ONO-WH-236 and the cathepsin S inhibitor LHSV were found to suppress scratching behavior [64,70]. In the future, protease inhibitors may become a more established method of treating itch.



#### 4.3. Therapeutic Drugs for Peptides

Gabapentin, pregabalin and capsaicin are effective for the treatment of neuropathic itch [6]. A phase II randomized clinical trial showed that a NK-1R (a receptor for SP) inhibitor was effective for treating itch in patients with psoriasis [151].

#### 4.4. Therapeutic Drugs for Cytokines

More recently, a variety of monoclonal antibodies have been shown to be effective in the treatment of itch. For example, dupilumab was found to improve AD symptoms and itch [152]. Most cytokines are activated via JAK/STAT signaling. Recently, a JAK inhibitor, delgocitinib, was reported to improve symptoms and itching of AD and was approved in Japan [153]. Moreover, Baricitinib, which inhibits JAK1 and JAK2, and Abrocitinib, which inhibits JAK1, improved AD symptoms including itch [28]. JAK inhibitors will be used for the treatment of itch in AD in the future.

#### 4.5. Therapeutic Drugs for Lipid Mediators

CMHVA, a LTB<sub>4</sub> receptor antagonist, was found to improve itch [130], suggesting it may be targeted as a lipid mediator to treat itch in the future.

**Table 1.** Immune cell-derived itch mediators and therapeutic methods.

Category	Puritogens	Receptors	Therapeutic Methods	Reference
Amines	Histamine	H <sub>1</sub> R/H <sub>4</sub> R	Anti-histamine/Anti-inflammatory, immuno-modulatory topical and systemic therapy (Cyclosporine A, Pimecrolimus, Tacrolimus and Corticosteroids)	[6,28]
	Serotonin	5-HT <sub>2</sub> receptor	Sertraline	[41]
Proteases	Tryptase	PAR-2	Anti-histamine/Cyclosporine A/Pimecrolimus/Tacrolimus/Corticosteroids	[6]
	Chymase	PAR-2	ONO-WH-236/Anti-histamine/Cyclosporine A/Pimecrolimus/Tacrolimus/Corticosteroids	[6,63]
	Cathepsin S	PAR-2/PAR-4	LHVS/Anti-histamine/Cyclosporine A/Pimecrolimus/Tacrolimus/Corticosteroids	[6,70]
Peptides	Substance P	NK-1R	Serlopitant/Gabapentin/Pregabalin/Capsaicin	[6,151]
	Endothelin-1	ET <sub>A</sub>	Bosentan	[82]
Cytokines	IL-2	IL-2R	Cyclosporine A/Delgocitinib/Baricitinib/Abrocitinib	[28,86,87,89,153]
	IL-4	IL-4Rα/γC IL-4Rα/IL-13Rα1	Dupilumab/Delgocitinib/Baricitinib/Abrocitinib	[28,93,99,152,153]
	IL-13	IL-4Rα/IL-13Rα1	Dupilumab/Tralokinumab/Lebrikizumab	[28,93,99]
	IL-17	IL-17RA/IL-17RC	Brodalumab	[103]
	IL-23	IL-12Rβ1/IL-23R	Delgocitinib/Baricitinib	[28,105,106,153]
	IL-31	IL-31RA/OSMR	Nemolizumab/Delgocitinib/Baricitinib/Abrocitinib	[28,111–114,153,154]
	IL-33	ST2/IL-1RAcP	Etokimab/Delgocitinib/Baricitinib	[28,140,153]
Lipid mediators	TSLP	TSLPR	Tezepelumab/Delgocitinib/Baricitinib/Abrocitinib	[28,149,150,153,155]
	PAF	PAFR	PAF antagonist	[118,156]
	LTB <sub>4</sub>	BLT1/BLT2	CMHVA	[128,130]
	LTC <sub>4</sub>	CysLTR1/CysLTR2	CysLTR2 antagonist	[157]

**Table 2.** Itch modulators from immune cells.

Ligands	Receptors	Source	Modulation
SLIGRL-NH <sub>2</sub>	PAR-2	mast cells, basophils	Enhances CQ and BAM8-22 induced itch
IL-4	IL-4Rα/γC IL-4Rα/IL-13Rα1	Th2, Tfh, ILC2, mast cells, basophils, eosinophils	Enhanced neuronal responsiveness to histamine, CQ, TSLP and IL-31
IL-13	IL-13Rα1/IL-13Rα2	Th2, ILC2, mast cells, basophils, eosinophils	May enhance neuronal responsiveness to histamine, CQ, TSLP and IL-31, as well as IL-4
IL-23	IL-12Rβ1/IL-23R	DCs, macrophages	Reduced histamine-induced itch
IL-33	ST2/IL-1RAcP	DCs, macrophages, mast cells	Enhanced CQ evoked calcium responses

## 5. Conclusions

This review presents recent knowledge regarding immune cell-derived mediators and modulators of itch. Many of these mediators cause nerve firing via their respective receptors expressed on sensory nerves, affecting the induction and modulation of itch. The variety of immune-derived itch mediators alone suggests that the mechanisms of itch are diverse. Although it is practical to focus on a common molecule such as JAK as a therapeutic target for itch, in fact, the development of therapeutic agents that target individual itch mediators and their receptors is ongoing. Thus, in clinical practice, however, in the future, due to the diverse molecules involved, a combination of therapies may be required to treat itch. It would be ideal to develop a system to test for itch mediators in each individual patient to determine the best treatment or appropriate combination therapy for each individual patient.

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## References

- Ikoma, A. Updated neurophysiology of itch. *Biol. Pharm. Bull.* **2013**, *36*, 1235–1240. [[CrossRef](#)] [[PubMed](#)]
- Mettang, T.; Kremer, A.E. Uremic pruritus. *Kidney Int.* **2015**, *87*, 685–691. [[CrossRef](#)] [[PubMed](#)]
- Dull, M.M.; Kremer, A.E. Treatment of Pruritus Secondary to Liver Disease. *Curr. Gastroenterol. Rep.* **2019**, *21*, 48. [[CrossRef](#)]
- Iwamoto, S.; Tominaga, M.; Kamata, Y.; Kawakami, T.; Osada, T.; Takamori, K. Association Between Inflammatory Bowel Disease and Pruritus. *Crohns Colitis* **2020**, *2*, otaa012. [[CrossRef](#)]
- Greaves, M.W. Itch in systemic disease: Therapeutic options. *Dermatol. Ther.* **2005**, *18*, 323–327. [[CrossRef](#)] [[PubMed](#)]
- Ikoma, A.; Steinhoff, M.; Stander, S.; Yosipovitch, G.; Schmelz, M. The neurobiology of itch. *Nat. Rev. Neurosci.* **2006**, *7*, 535–547. [[CrossRef](#)]
- Paus, R.; Schmelz, M.; Biro, T.; Steinhoff, M. Frontiers in pruritus research: Scratching the brain for more effective itch therapy. *J. Clin. Investig.* **2006**, *116*, 1174–1186. [[CrossRef](#)]
- Basbaum, A.L.; Bautista, D.M.; Scherrer, G.; Julius, D. Cellular and molecular mechanisms of pain. *Cell* **2009**, *139*, 267–284. [[CrossRef](#)]
- Usoskin, D.; Furlan, A.; Islam, S.; Abdo, H.; Lonnerberg, P.; Lou, D.; Hjerling-Leffler, J.; Haeggstrom, J.; Kharchenko, O.; Kharchenko, P.V.; et al. Unbiased classification of sensory neuron types by large-scale single-cell RNA sequencing. *Nat. Neurosci.* **2015**, *18*, 145–153. [[CrossRef](#)]
- Liu, Q.; Sikand, P.; Ma, C.; Tang, Z.; Han, L.; Li, Z.; Sun, S.; LaMotte, R.H.; Dong, X. Mechanisms of itch evoked by beta-alanine. *J. Neurosci.* **2012**, *32*, 14532–14537. [[CrossRef](#)]
- Choi, J.E.; Di Nardo, A. Skin neurogenic inflammation. *Semin. Immunopathol.* **2018**, *40*, 249–259. [[CrossRef](#)] [[PubMed](#)]
- Voisin, T.; Perner, C.; Messou, M.A.; Shiers, S.; Ualiyeva, S.; Kanaoka, Y.; Price, T.J.; Sokol, C.L.; Bankova, L.G.; Austen, K.F.; et al. The CysLT2R receptor mediates leukotriene C4-driven acute and chronic itch. *Proc. Natl. Acad. Sci. USA* **2021**, *118*. [[CrossRef](#)]
- Oetjen, L.K.; Mack, M.R.; Feng, J.; Whelan, T.M.; Niu, H.; Guo, C.J.; Chen, S.; Trier, A.M.; Xu, A.Z.; Tripathi, S.V.; et al. Sensory Neurons Co-opt Classical Immune Signaling Pathways to Mediate Chronic Itch. *Cell* **2017**, *171*, 217–228.e13. [[CrossRef](#)] [[PubMed](#)]
- MacGlashan, D. Histamine. *J. Allergy Clin. Immunol.* **2003**, *112*, S53–S59. [[CrossRef](#)]
- Ringvall, M.; Ronnberg, E.; Wernersson, S.; Duelli, A.; Henningsson, F.; Abrink, M.; Garcia-Faroldi, G.; Fajardo, I.; Pejler, G. Serotonin and histamine storage in mast cell secretory granules is dependent on serglycin proteoglycan. *J. Allergy Clin. Immunol.* **2008**, *121*, 1020–1026. [[CrossRef](#)]
- Shimizu, K.; Andoh, T.; Yoshihisa, Y.; Shimizu, T. Histamine released from epidermal keratinocytes plays a role in alpha-melanocyte-stimulating hormone-induced itching in mice. *Am. J. Pathol.* **2015**, *185*, 3003–3010. [[CrossRef](#)] [[PubMed](#)]
- Yosipovitch, G.; Rosen, J.D.; Hashimoto, T. Itch: From mechanism to (novel) therapeutic approaches. *J. Allergy Clin. Immunol.* **2018**, *142*, 1375–1390. [[CrossRef](#)]
- Kabashima, K.; Nakashima, C.; Nonomura, Y.; Otsuka, A.; Cardamone, C.; Parente, R.; De Feo, G.; Triggiani, M. Biomarkers for evaluation of mast cell and basophil activation. *Immunol. Rev.* **2018**, *282*, 114–120. [[CrossRef](#)]
- Hashimoto, T.; Rosen, J.D.; Sanders, K.M.; Yosipovitch, G. Possible roles of basophils in chronic itch. *Exp. Dermatol.* **2019**, *28*, 1373–1379. [[CrossRef](#)]
- Nakashima, C.; Ishida, Y.; Kitoh, A.; Otsuka, A.; Kabashima, K. Interaction of peripheral nerves and mast cells, eosinophils, and basophils in the development of pruritus. *Exp. Dermatol.* **2019**, *28*, 1405–1411. [[CrossRef](#)]

21. Moriguchi, T.; Takai, J. Histamine and histidine decarboxylase: Immunomodulatory functions and regulatory mechanisms. *Genes Cells* **2020**, *25*, 443–449. [[CrossRef](#)]
22. Akiyama, T.; Carstens, E. Neural processing of itch. *Neuroscience* **2013**, *250*, 697–714. [[CrossRef](#)] [[PubMed](#)]
23. Yamashita, M.; Fukui, H.; Sugama, K.; Horio, Y.; Ito, S.; Mizuguchi, H.; Wada, H. Expression cloning of a cDNA encoding the bovine histamine H1 receptor. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 11515–11519. [[CrossRef](#)]
24. Oda, T.; Morikawa, N.; Saito, Y.; Masuho, Y.; Matsumoto, S. Molecular cloning and characterization of a novel type of histamine receptor preferentially expressed in leukocytes. *J. Biol. Chem.* **2000**, *275*, 36781–36786. [[CrossRef](#)] [[PubMed](#)]
25. Hough, L.B. Genomics meets histamine receptors: New subtypes, new receptors. *Mol. Pharmacol.* **2001**, *59*, 415–419. [[CrossRef](#)]
26. Ohsawa, Y.; Hirasawa, N. The role of histamine H1 and H4 receptors in atopic dermatitis: From basic research to clinical study. *Allergol. Int.* **2014**, *63*, 533–542. [[CrossRef](#)] [[PubMed](#)]
27. Shim, W.S.; Tak, M.H.; Lee, M.H.; Kim, M.; Kim, M.; Koo, J.Y.; Lee, C.H.; Kim, M.; Oh, U. TRPV1 mediates histamine-induced itching via the activation of phospholipase A2 and 12-lipoxygenase. *J. Neurosci.* **2007**, *27*, 2331–2337. [[CrossRef](#)]
28. Iannone, M.; Tonini, G.; Janowska, A.; Dini, V.; Romanelli, M. Definition of treatment goals in terms of clinician-reported disease severity and patient-reported outcomes in moderate-to-severe adult atopic dermatitis: A systematic review. *Curr. Med. Res. Opin.* **2021**, *37*, 1295–1301. [[CrossRef](#)]
29. Rossbach, K.; Nassenstein, C.; Gschwandtner, M.; Schnell, D.; Sander, K.; Seifert, R.; Stark, H.; Kietzmann, M.; Baumer, W. Histamine H1, H3 and H4 receptors are involved in pruritus. *Neuroscience* **2011**, *190*, 89–102. [[CrossRef](#)]
30. Sommer, C. Serotonin in pain and analgesia: Actions in the periphery. *Mol. Neurobiol.* **2004**, *30*, 117–125. [[CrossRef](#)]
31. Conti, P.; Shaik-Dasthagirisaheb, Y.B. Mast Cell Serotonin Immunoregulatory Effects Impacting on Neuronal Function: Implications for Neurodegenerative and Psychiatric Disorders. *Neurotox. Res.* **2015**, *28*, 147–153. [[CrossRef](#)]
32. Domocos, D.; Selescu, T.; Ceafalan, L.C.; Iodi Carstens, M.; Carstens, E.; Babes, A. Role of 5-HT1A and 5-HT3 receptors in serotonergic activation of sensory neurons in relation to itch and pain behavior in the rat. *J. Neurosci. Res.* **2020**, *98*, 1999–2017. [[CrossRef](#)]
33. Akiyama, T.; Ivanov, M.; Nagamine, M.; Davoodi, A.; Carstens, M.I.; Ikoma, A.; Cevikbas, F.; Kempkes, C.; Buddenkotte, J.; Steinhoff, M.; et al. Involvement of TRPV4 in Serotonin-Evoked Scratching. *J. Investig. Dermatol.* **2016**, *136*, 154–160. [[CrossRef](#)]
34. Yamaguchi, T.; Nagasawa, T.; Satoh, M.; Kuraishi, Y. Itch-associated response induced by intradermal serotonin through 5-HT2 receptors in mice. *Neurosci. Res.* **1999**, *35*, 77–83. [[CrossRef](#)]
35. Thomsen, J.S.; Petersen, M.B.; Benfeldt, E.; Jensen, S.B.; Serup, J. Scratch induction in the rat by intradermal serotonin: A model for pruritus. *Acta Derm. Venereol.* **2001**, *81*, 250–254. [[CrossRef](#)]
36. Jinks, S.L.; Carstens, E. Responses of superficial dorsal horn neurons to intradermal serotonin and other irritants: Comparison with scratching behavior. *J. Neurophysiol.* **2002**, *87*, 1280–1289. [[CrossRef](#)] [[PubMed](#)]
37. Nojima, H.; Carstens, E. 5-Hydroxytryptamine (5-HT)<sub>2</sub> receptor involvement in acute 5-HT-evoked scratching but not in allergic pruritus induced by dinitrofluorobenzene in rats. *J. Pharmacol. Exp. Ther.* **2003**, *306*, 245–252. [[CrossRef](#)] [[PubMed](#)]
38. Hu, W.P.; Guan, B.C.; Ru, L.Q.; Chen, J.G.; Li, Z.W. Potentiation of 5-HT<sub>3</sub> receptor function by the activation of coexistent 5-HT<sub>2</sub> receptors in trigeminal ganglion neurons of rats. *Neuropharmacology* **2004**, *47*, 833–840. [[CrossRef](#)]
39. Machida, T.; Iizuka, K.; Hirafuji, M. Recent Advances in 5-Hydroxytryptamine (5-HT) Receptor Research: How Many Pathophysiological Roles Does 5-HT Play via Its Multiple Receptor Subtypes? *Biol. Pharm. Bull.* **2013**, *36*, 1416–1419. [[CrossRef](#)] [[PubMed](#)]
40. Cortes-Altamirano, J.L.; Olmos-Hernandez, A.; Jaime, H.B.; Carrillo-Mora, P.; Bandala, C.; Reyes-Long, S.; Alfaro-Rodriguez, A. Review: 5-HT<sub>1</sub>, 5-HT<sub>2</sub>, 5-HT<sub>3</sub> and 5-HT<sub>7</sub> Receptors and their Role in the Modulation of Pain Response in the Central Nervous System. *Curr. Neuropharmacol.* **2018**, *16*, 210–221. [[CrossRef](#)]
41. Bolier, A.R.; Peri, S.; Oude Elferink, R.P.; Beuers, U. The challenge of cholestatic pruritus. *Acta Gastroenterol. Belg.* **2012**, *75*, 399–404.
42. Caughey, G.H.; Raymond, W.W.; Blount, J.L.; Hau, L.W.; Pallaoro, M.; Wolters, P.J.; Verghese, G.M. Characterization of human gamma-tryptases, novel members of the chromosome 16p mast cell tryptase and prostasin gene families. *J. Immunol.* **2000**, *164*, 6566–6575. [[CrossRef](#)]
43. Wong, G.W.; Yasuda, S.; Madhusudhan, M.S.; Li, L.; Yang, Y.; Krilis, S.A.; Sali, A.; Stevens, R.L. Human tryptase epsilon (PRSS22), a new member of the chromosome 16p13.3 family of human serine proteases expressed in airway epithelial cells. *J. Biol. Chem.* **2001**, *276*, 49169–49182. [[CrossRef](#)] [[PubMed](#)]
44. Caughey, G.H. Tryptase genetics and anaphylaxis. *J. Allergy Clin. Immunol.* **2006**, *117*, 1411–1414. [[CrossRef](#)] [[PubMed](#)]
45. Hernandez-Hernandez, L.; Sanz, C.; Garcia-Solaesa, V.; Padron, J.; Garcia-Sanchez, A.; Davila, I.; Isidoro-Garcia, M.; Lorente, F. Tryptase: Genetic and functional considerations. *Allergol. Immunopathol.* **2012**, *40*, 385–389. [[CrossRef](#)]
46. Caughey, G.H. The structure and airway biology of mast cell proteinases. *Am. J. Respir. Cell. Mol. Biol.* **1991**, *4*, 387–394. [[CrossRef](#)] [[PubMed](#)]
47. Nadel, J.A. Biologic effects of mast cell enzymes. *Am. Rev. Respir. Dis.* **1992**, *145*, S37–S41. [[CrossRef](#)]
48. Xia, H.Z.; Kepley, C.L.; Sakai, K.; Chelliah, J.; Irani, A.M.; Schwartz, L.B. Quantitation of Tryptase, Chymase, Fc $\gamma$ R1, and Fc $\epsilon$ R1 mRNAs in Human Mast Cells and Basophils by Competitive Reverse Transcription-Polymerase Chain Reaction. *J. Immunol.* **1995**, *154*, 5472–5480. [[PubMed](#)]
49. Jogie-Brahim, S.; Min, H.K.; Fukuoka, Y.; Xia, H.Z.; Schwartz, L.B. Expression of alpha-tryptase and beta-tryptase by human basophils. *J. Allergy Clin. Immunol.* **2004**, *113*, 1086–1092. [[CrossRef](#)]

50. Ui, H.; Andoh, T.; Lee, J.B.; Nojima, H.; Kuraishi, Y. Potent pruritogenic action of tryptase mediated by PAR-2 receptor and its involvement in anti-pruritic effect of nafamostat mesilate in mice. *Eur. J. Pharmacol.* **2006**, *530*, 172–178. [[CrossRef](#)]
51. Lee, S.E.; Jeong, S.K.; Lee, S.H. Protease and protease-activated receptor-2 signaling in the pathogenesis of atopic dermatitis. *Yonsei Med. J.* **2010**, *51*, 808–822. [[CrossRef](#)]
52. Heuberger, D.M.; Schuepbach, R.A. Protease-activated receptors (PARs): Mechanisms of action and potential therapeutic modulators in PAR-driven inflammatory diseases. *Thromb. J.* **2019**, *17*, 4. [[CrossRef](#)] [[PubMed](#)]
53. Klein, A.; Carstens, M.I.; Carstens, E. Facial injections of pruritogens or algogens elicit distinct behavior responses in rats and excite overlapping populations of primary sensory and trigeminal subnucleus caudalis neurons. *J. Neurophysiol.* **2011**, *106*, 1078–1088. [[CrossRef](#)]
54. Gupta, K.; Harvima, I.T. Mast cell-neural interactions contribute to pain and itch. *Immunol. Rev.* **2018**, *282*, 168–187. [[CrossRef](#)]
55. Thapaliya, M.; Chompunud Na Ayudhya, C.; Amponnawarat, A.; Roy, S.; Ali, H. Mast Cell-Specific MRGPRX2: A Key Modulator of Neuro-Immune Interaction in Allergic Diseases. *Curr. Allergy Asthma. Rep.* **2021**, *21*, 3. [[CrossRef](#)] [[PubMed](#)]
56. Akiyama, T.; Carstens, M.I.; Ikoma, A.; Cevikbas, F.; Steinhoff, M.; Carstens, E. Mouse model of touch-evoked itch (alloknesis). *J. Invest. Dermatol.* **2012**, *132*, 1886–1891. [[CrossRef](#)]
57. Gallwitz, M.; Enoksson, M.; Hellman, L. Expression profile of novel members of the rat mast cell protease (rMCP)-2 and (rMCP)-8 families, and functional analyses of mouse mast cell protease (mMCP)-8. *Immunogenetics* **2007**, *59*, 391–405. [[CrossRef](#)]
58. Atiakshin, D.; Buchwalow, I.; Tiemann, M. Mast cell chymase: Morphofunctional characteristics. *Histochem. Cell Biol.* **2019**, *152*, 253–269. [[CrossRef](#)]
59. Caughey, G.H. Mast cell tryptases and chymases in inflammation and host defense. *Immunol. Rev.* **2007**, *217*, 141–154. [[CrossRef](#)]
60. Caughey, G.H. Mast cell proteases as protective and inflammatory mediators. *Adv. Exp. Med. Biol.* **2011**, *716*, 212–234. [[CrossRef](#)] [[PubMed](#)]
61. Wasse, H.; Naqvi, N.; Husain, A. Impact of Mast Cell Chymase on Renal Disease Progression. *Curr. Hypertens. Rev.* **2012**, *8*, 15–23. [[CrossRef](#)]
62. De Souza Junior, D.A.; Santana, A.C.; da Silva, E.Z.; Oliver, C.; Jamur, M.C. The Role of Mast Cell Specific Chymases and Tryptases in Tumor Angiogenesis. *Biomed. Res. Int.* **2015**, *2015*, 142359. [[CrossRef](#)] [[PubMed](#)]
63. Sharma, R.; Prasad, V.; McCarthy, E.T.; Savin, V.J.; Dileepan, K.N.; Stechschulte, D.J.; Lianos, E.; Wiegmann, T.; Sharma, M. Chymase increases glomerular albumin permeability via protease-activated receptor-2. *Mol. Cell. Biochem.* **2007**, *297*, 161–169. [[CrossRef](#)]
64. Nabe, T.; Kijitani, Y.; Kitagawa, Y.; Sakano, E.; Ueno, T.; Fujii, M.; Nakao, S.; Sakai, M.; Takai, S. Involvement of chymase in allergic conjunctivitis of guinea pigs. *Exp. Eye Res.* **2013**, *113*, 74–79. [[CrossRef](#)] [[PubMed](#)]
65. Imada, T.; Komorita, N.; Kobayashi, F.; Naito, K.; Yoshikawa, T.; Miyazaki, M.; Nakamura, N.; Kondo, T. Therapeutic potential of a specific chymase inhibitor in atopic dermatitis. *Jpn. J. Pharmacol.* **2002**, *90*, 214–217. [[CrossRef](#)]
66. Schwarz, G.; Boehncke, W.H.; Braun, M.; Schroter, C.J.; Burster, T.; Flad, T.; Dressel, D.; Weber, E.; Schmid, H.; Kalbacher, H. Cathepsin S activity is detectable in human keratinocytes and is selectively upregulated upon stimulation with interferon-gamma. *J. Invest. Dermatol.* **2002**, *119*, 44–49. [[CrossRef](#)]
67. Viode, C.; Lejeune, O.; Turlier, V.; Rouquier, A.; Casas, C.; Mengeaud, V.; Redoules, D.; Schmitt, A.M. Cathepsin S, a new pruritus biomarker in clinical dandruff/seborrheic dermatitis evaluation. *Exp. Dermatol.* **2014**, *23*, 274–275. [[CrossRef](#)]
68. Reddy, V.B.; Shimada, S.G.; Sikand, P.; Lamotte, R.H.; Lerner, E.A. Cathepsin S elicits itch and signals via protease-activated receptors. *J. Invest. Dermatol.* **2010**, *130*, 1468–1470. [[CrossRef](#)]
69. Reddy, V.B.; Sun, S.; Azimi, E.; Elmariah, S.B.; Dong, X.; Lerner, E.A. Redefining the concept of protease-activated receptors: Cathepsin S evokes itch via activation of Mrgprs. *Nat. Commun.* **2015**, *6*, 7864. [[CrossRef](#)] [[PubMed](#)]
70. Chung, K.; Pitcher, T.; Grant, A.D.; Hewitt, E.; Lindstrom, E.; Malcangio, M. Cathepsin S acts via protease-activated receptor 2 to activate sensory neurons and induce itch-like behaviour. *Neurobiol. Pain* **2019**, *6*, 100032. [[CrossRef](#)]
71. Patricio, E.S.; Costa, R.; Figueiredo, C.P.; Gers-Barlag, K.; Bicca, M.A.; Manjavachi, M.N.; Segat, G.C.; Gentry, C.; Luiz, A.P.; Fernandes, E.S.; et al. Mechanisms Underlying the Scratching Behavior Induced by the Activation of Proteinase-Activated Receptor-4 in Mice. *J. Invest. Dermatol.* **2015**, *135*, 2484–2491. [[CrossRef](#)]
72. Lotts, T.; Stander, S. Research in practice: Substance P antagonism in chronic pruritus. *J. Dtsch. Dermatol. Ges.* **2014**, *12*, 557–559. [[CrossRef](#)] [[PubMed](#)]
73. Stander, S.; Yosipovitch, G. Substance P and neurokinin 1 receptor are new targets for the treatment of chronic pruritus. *Br. J. Dermatol.* **2019**, *181*, 932–938. [[CrossRef](#)] [[PubMed](#)]
74. Mashaghi, A.; Marmalidou, A.; Tehrani, M.; Grace, P.M.; Pothoulakis, C.; Dana, R. Neuropeptide substance P and the immune response. *Cell Mol. Life Sci.* **2016**, *73*, 4249–4264. [[CrossRef](#)]
75. Andoh, T.; Nagasawa, T.; Satoh, M.; Kuraishi, Y. Substance P induction of itch-associated response mediated by cutaneous NK1 tachykinin receptors in mice. *J. Pharmacol. Exp. Ther.* **1998**, *286*, 1140–1145. [[PubMed](#)]
76. McQueen, D.S.; Noble, M.A.; Bond, S.M. Endothelin-1 activates ETA receptors to cause reflex scratching in BALB/c mice. *Br. J. Pharmacol.* **2007**, *151*, 278–284. [[CrossRef](#)]
77. Gomes, L.O.; Hara, D.B.; Rae, G.A. Endothelin-1 induces itch and pain in the mouse cheek model. *Life Sci.* **2012**, *91*, 628–633. [[CrossRef](#)]
78. Davenport, A.P.; Hyndman, K.A.; Dhaun, N.; Southan, C.; Kohan, D.E.; Pollock, J.S.; Pollock, D.M.; Webb, D.J.; Maguire, J.J. Endothelin. *Pharmacol. Rev.* **2016**, *68*, 357–418. [[CrossRef](#)] [[PubMed](#)]







107. Kunsleben, N.; Rudrich, U.; Gehring, M.; Novak, N.; Kapp, A.; Raap, U. IL-31 Induces Chemotaxis, Calcium Mobilization, Release of Reactive Oxygen Species, and CCL26 in Eosinophils, Which Are Capable to Release IL-31. *J. Investig. Dermatol.* **2015**, *135*, 1908–1911. [[CrossRef](#)]
108. Hashimoto, T.; Kursewicz, C.D.; Fayne, R.A.; Nanda, S.; Shah, S.M.; Nattkemper, L.; Yokozeki, H.; Yosipovitch, G. Mechanisms of Itch in Stasis Dermatitis: Significant Role of IL-31 from Macrophages. *J. Investig. Dermatol.* **2020**, *140*, 850–859.e3. [[CrossRef](#)] [[PubMed](#)]
109. Xu, J.; Zanvit, P.; Hu, L.; Tseng, P.Y.; Liu, N.; Wang, F.; Liu, O.; Zhang, D.; Jin, W.; Guo, N.; et al. The Cytokine TGF-beta Induces Interleukin-31 Expression from Dermal Dendritic Cells to Activate Sensory Neurons and Stimulate Wound Itching. *Immunity* **2020**, *53*, 371–383. [[CrossRef](#)] [[PubMed](#)]
110. Ruppenstein, A.; Limberg, M.M.; Loser, K.; Kremer, A.E.; Homey, B.; Raap, U. Involvement of Neuro-Immune Interactions in Pruritus With Special Focus on Receptor Expressions. *Front. Med.* **2021**, *8*, 627985. [[CrossRef](#)]
111. Datsi, A.; Steinhoff, M.; Ahmad, F.; Alam, M.; Buddenkotte, J. Interleukin-31: The “itchy” cytokine in inflammation and therapy. *Allergy* **2021**, *76*, 2982–2997. [[CrossRef](#)]
112. Zhang, Q.; Putheti, P.; Zhou, Q.; Liu, Q.; Gao, W. Structures and biological functions of IL-31 and IL-31 receptors. *Cytokine Growth Factor Rev.* **2008**, *19*, 347–356. [[CrossRef](#)]
113. Cevikbas, F.; Wang, X.; Akiyama, T.; Kempkes, C.; Savinko, T.; Antal, A.; Kukova, G.; Buhl, T.; Ikoma, A.; Buddenkotte, J.; et al. A sensory neuron-expressed IL-31 receptor mediates T helper cell-dependent itch: Involvement of TRPV1 and TRPA1. *J. Allergy Clin. Immunol.* **2014**, *133*, 448–460. [[CrossRef](#)]
114. Furue, M.; Yamamura, K.; Kido-Nakahara, M.; Nakahara, T.; Fukui, Y. Emerging role of interleukin-31 and interleukin-31 receptor in pruritus in atopic dermatitis. *Allergy* **2018**, *73*, 29–36. [[CrossRef](#)]
115. Feld, M.; Garcia, R.; Buddenkotte, J.; Katayama, S.; Lewis, K.; Muirhead, G.; Hevezi, P.; Plessner, K.; Schruppf, H.; Krjutskov, K.; et al. The pruritus- and TH2-associated cytokine IL-31 promotes growth of sensory nerves. *J. Allergy Clin. Immunol.* **2016**, *138*, 500–508.e524. [[CrossRef](#)] [[PubMed](#)]
116. Larsen, E.G.; Cho, T.S.; McBride, M.L.; Feng, J.; Manivannan, B.; Madura, C.; Klein, N.E.; Wright, E.B.; Wickstead, E.S.; Garcia-Verdugo, H.D.; et al. Transmembrane protein TMEM184B is necessary for interleukin-31-induced itch. *Pain* **2021**. publish ahead of print. [[CrossRef](#)]
117. Palgan, K.; Bartuzi, Z. Platelet activating factor in allergies. *Int. J. Immunopathol. Pharmacol.* **2015**, *28*, 584–589. [[CrossRef](#)] [[PubMed](#)]
118. Liu, Y.; Shields, L.B.E.; Gao, Z.; Wang, Y.; Zhang, Y.P.; Chu, T.; Zhu, Q.; Shields, C.B.; Cai, J. Current Understanding of Platelet-Activating Factor Signaling in Central Nervous System Diseases. *Mol. Neurobiol.* **2017**, *54*, 5563–5572. [[CrossRef](#)]
119. Thomsen, J.S.; Sonne, M.; Benfeldt, E.; Jensen, S.B.; Serup, J.; Menne, T. Experimental itch in sodium lauryl sulphate-inflamed and normal skin in humans: A randomized, double-blind, placebo-controlled study of histamine and other inducers of itch. *Br. J. Dermatol.* **2002**, *146*, 792–800. [[CrossRef](#)] [[PubMed](#)]
120. Petersen, L.J.; Church, M.K.; Skov, P.S. Platelet-activating factor Induces histamine release from human skin mast cells in vivo, which is reduced by local nerve blockade. *J. Allergy Clin. Immunol.* **1997**, *99*, 640–647. [[CrossRef](#)]
121. Andoh, T.; Haza, S.; Saito, A.; Kuraishi, Y. Involvement of leukotriene B4 in spontaneous itch-related behaviour in NC mice with atopic dermatitis-like skin lesions. *Exp. Dermatol.* **2011**, *20*, 894–898. [[CrossRef](#)]
122. Miyahara, N.; Ohnishi, H.; Miyahara, S.; Takeda, K.; Matsubara, S.; Matsuda, H.; Okamoto, M.; Loader, J.E.; Joetham, A.; Tanimoto, M.; et al. Leukotriene B4 release from mast cells in IgE-mediated airway hyperresponsiveness and inflammation. *Am. J. Respir. Cell Mol. Biol.* **2009**, *40*, 672–682. [[CrossRef](#)]
123. Bando, T.; Fujita, S.; Nagano, N.; Yoshikawa, S.; Yamanishi, Y.; Minami, M.; Karasuyama, H. Differential usage of COX-1 and COX-2 in prostaglandin production by mast cells and basophils. *Biochem. Biophys. Res. Commun.* **2017**, *487*, 82–87. [[CrossRef](#)]
124. Pal, K.; Feng, X.; Steinke, J.W.; Burdick, M.D.; Shim, Y.M.; Sung, S.S.; Teague, W.G.; Borish, L. Leukotriene A4 Hydrolase Activation and Leukotriene B4 Production by Eosinophils in Severe Asthma. *Am. J. Respir. Cell Mol. Biol.* **2019**, *60*, 413–419. [[CrossRef](#)]
125. Finney-Hayward, T.K.; Bahra, P.; Li, S.; Poll, C.T.; Nicholson, A.G.; Russell, R.E.; Ford, P.A.; Westwick, J.; Fenwick, P.S.; Barnes, P.J.; et al. Leukotriene B4 release by human lung macrophages via receptor- not voltage-operated Ca<sup>2+</sup> channels. *Eur. Respir. J.* **2009**, *33*, 1105–1112. [[CrossRef](#)] [[PubMed](#)]
126. Yokomizo, T.; Izumi, T.; Chang, K.; Takawa, Y.; Shimizu, T. A G-protein-coupled receptor for leukotriene B4 that mediates chemotaxis. *Nature* **1997**, *387*, 620–624. [[CrossRef](#)]
127. Yokomizo, T.; Kato, K.; Terawaki, K.; Izumi, T.; Shimizu, T. A second leukotriene B(4) receptor, BLT2. A new therapeutic target in inflammation and immunological disorders. *J. Exp. Med.* **2000**, *192*, 421–432. [[CrossRef](#)]
128. Andoh, T.; Kuraishi, Y. Expression of BLT1 leukotriene B4 receptor on the dorsal root ganglion neurons in mice. *Mol. Brain Res.* **2005**, *137*, 263–266. [[CrossRef](#)]
129. Fernandes, E.S.; Vong, C.T.; Quek, S.; Cheong, J.; Awal, S.; Gentry, C.; Aubdool, A.A.; Liang, L.; Bodkin, J.V.; Bevan, S.; et al. Superoxide generation and leukocyte accumulation: Key elements in the mediation of leukotriene B(4)-induced itch by transient receptor potential ankyrin 1 and transient receptor potential vanilloid 1. *FASEB J.* **2013**, *27*, 1664–1673. [[CrossRef](#)] [[PubMed](#)]
130. Andoh, T.; Harada, A.; Kuraishi, Y. Involvement of Leukotriene B4 Released from Keratinocytes in Itch-associated Response to Intradermal Interleukin-31 in Mice. *Acta Derm. Venereol.* **2017**, *97*, 922–927. [[CrossRef](#)] [[PubMed](#)]

131. Murakami, M.; Matsumoto, R.; Urade, Y.; Austen, K.F.; Arm, J.P. c-kit ligand mediates increased expression of cytosolic phospholipase A2, prostaglandin endoperoxide synthase-1, and hematopoietic prostaglandin D2 synthase and increased IgE-dependent prostaglandin D2 generation in immature mouse mast cells. *J. Biol. Chem.* **1995**, *270*, 3239–3246. [[CrossRef](#)] [[PubMed](#)]
132. Wang, F.; Trier, A.M.; Li, F.; Kim, S.; Chen, Z.; Chai, J.N.; Mack, M.R.; Morrison, S.A.; Hamilton, J.D.; Baek, J.; et al. A basophil-neuronal axis promotes itch. *Cell* **2021**, *184*, 422–440.e417. [[CrossRef](#)]
133. Takafuji, S.; Bischoff, S.C.; De Weck, A.L.; Dahinden, C.A. IL-3 and IL-5 prime normal human eosinophils to produce leukotriene C4 in response to soluble agonists. *J. Immunol.* **1991**, *147*, 3855–3861.
134. Ohno, T.; Morita, H.; Arae, K.; Matsumoto, K.; Nakae, S. Interleukin-33 in allergy. *Allergy* **2012**, *67*, 1203–1214. [[CrossRef](#)]
135. Nakae, S.; Morita, H.; Ohno, T.; Arae, K.; Matsumoto, K.; Saito, H. Role of interleukin-33 in innate-type immune cells in allergy. *Allergol. Int.* **2013**, *62*, 13–20. [[CrossRef](#)]
136. Takeda, T.; Unno, H.; Morita, H.; Futamura, K.; Emi-Sugie, M.; Arae, K.; Shoda, T.; Okada, N.; Igarashi, A.; Inoue, E.; et al. Platelets constitutively express IL-33 protein and modulate eosinophilic airway inflammation. *J. Allergy Clin. Immunol.* **2016**, *138*, 1395–1403. [[CrossRef](#)] [[PubMed](#)]
137. Toyama, S.; Moniaga, C.S.; Nakae, S.; Kurosawa, M.; Ogawa, H.; Tominaga, M.; Takamori, K. Regulatory T Cells Exhibit Interleukin-33-Dependent Migratory Behavior during Skin Barrier Disruption. *Int. J. Mol. Sci.* **2021**, *22*, 7443. [[CrossRef](#)] [[PubMed](#)]
138. Trier, A.M.; Mack, M.R.; Fredman, A.; Tamari, M.; Ver Heul, A.M.; Zhao, Y.; Guo, C.J.; Avraham, O.; Ford, Z.K.; Oetjen, L.K.; et al. IL-33 Signaling in Sensory Neurons Promotes Dry Skin Itch. *J. Allergy Clin. Immunol.* **2021**, in press. [[CrossRef](#)]
139. Peng, G.; Mu, Z.; Cui, L.; Liu, P.; Wang, Y.; Wu, W.; Han, X. Anti-IL-33 Antibody Has a Therapeutic Effect in an Atopic Dermatitis Murine Model Induced by 2, 4-Dinitrochlorobenzene. *Inflammation* **2018**, *41*, 154–163. [[CrossRef](#)]
140. Du, L.; Hu, X.; Yang, W.; Yasheng, H.; Liu, S.; Zhang, W.; Zhou, Y.; Cui, W.; Zhu, J.; Qiao, Z.; et al. Spinal IL-33/ST2 signaling mediates chronic itch in mice through the astrocytic JAK2-STAT3 cascade. *Glia* **2019**, *67*, 1680–1693. [[CrossRef](#)]
141. Kahremany, S.; Hofmann, L.; Gruzman, A.; Cohen, G. Advances in Understanding the Initial Steps of Pruritoceptive Itch: How the Itch Hits the Switch. *Int. J. Mol. Sci.* **2020**, *21*, 4883. [[CrossRef](#)] [[PubMed](#)]
142. Dewas, C.; Chen, X.; Honda, T.; Junntila, I.; Linton, J.; Udey, M.C.; Porcella, S.F.; Sturdevant, D.E.; Feigenbaum, L.; Koo, L.; et al. TSLP expression: Analysis with a ZsGreen TSLP reporter mouse. *J. Immunol.* **2015**, *194*, 1372–1380. [[CrossRef](#)]
143. Reche, P.A.; Soumelis, V.; Gorman, D.M.; Clifford, T.; Liu, M.; Travis, M.; Zurawski, S.M.; Johnston, J.; Liu, Y.J.; Spits, H.; et al. Human thymic stromal lymphopoietin preferentially stimulates myeloid cells. *J. Immunol.* **2001**, *167*, 336–343. [[CrossRef](#)]
144. Allakhverdi, Z.; Comeau, M.R.; Jessup, H.K.; Yoon, B.R.; Brewer, A.; Chartier, S.; Paquette, N.; Ziegler, S.F.; Sarfati, M.; Delespesse, G. Thymic stromal lymphopoietin is released by human epithelial cells in response to microbes, trauma, or inflammation and potently activates mast cells. *J. Exp. Med.* **2007**, *204*, 253–258. [[CrossRef](#)]
145. Hirano, R.; Hasegawa, S.; Hashimoto, K.; Haneda, Y.; Ohsaki, A.; Ichiyama, T. Human thymic stromal lymphopoietin enhances expression of CD80 in human CD14+ monocytes/macrophages. *Inflamm. Res.* **2011**, *60*, 605–610. [[CrossRef](#)]
146. Cook, E.B.; Stahl, J.L.; Schwantes, E.A.; Fox, K.E.; Mathur, S.K. IL-3 and TNFalpha increase Thymic Stromal Lymphopoietin Receptor (TSLPR) expression on eosinophils and enhance TSLP-stimulated degranulation. *Clin. Mol. Allergy* **2012**, *10*, 8. [[CrossRef](#)]
147. Kubo, T.; Kamekura, R.; Kumagai, A.; Kawata, K.; Yamashita, K.; Mitsushashi, Y.; Kojima, T.; Sugimoto, K.; Yoneta, A.; Sumikawa, Y.; et al. DeltaNp63 controls a TLR3-mediated mechanism that abundantly provides thymic stromal lymphopoietin in atopic dermatitis. *PLoS ONE* **2014**, *9*, e105498. [[CrossRef](#)] [[PubMed](#)]
148. Wilson, S.R.; The, L.; Batia, L.M.; Beattie, K.; Katibah, G.E.; McClain, S.P.; Pellegrino, M.; Estandian, D.M.; Bautista, D.M. The epithelial cell-derived atopic dermatitis cytokine TSLP activates neurons to induce itch. *Cell* **2013**, *155*, 285–295. [[CrossRef](#)] [[PubMed](#)]
149. Rochman, Y.; Kashyap, M.; Robinson, G.W.; Sakamoto, K.; Gomez-Rodriguez, J.; Wagner, K.U.; Leonard, W.J. Thymic stromal lymphopoietin-mediated STAT5 phosphorylation via kinases JAK1 and JAK2 reveals a key difference from IL-7-induced signaling. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 19455–19460. [[CrossRef](#)]
150. Arima, K.; Watanabe, N.; Hanabuchi, S.; Chang, M.; Sun, S.C.; Liu, Y.J. Distinct signal codes generate dendritic cell functional plasticity. *Sci. Signal.* **2010**, *3*, ra4. [[CrossRef](#)]
151. Pariser, D.M.; Bagel, J.; Lebwohl, M.; Yosipovitch, G.; Chien, E.; Spellman, M.C. Serlopitant for psoriatic pruritus: A phase 2 randomized, double-blind, placebo-controlled clinical trial. *J. Am. Acad. Dermatol.* **2020**, *82*, 1314–1320. [[CrossRef](#)]
152. Silverberg, J.I.; Yosipovitch, G.; Simpson, E.L.; Kim, B.S.; Wu, J.J.; Eckert, L.; Guillemin, I.; Chen, Z.; Ardeleanu, M.; Bansal, A.; et al. Dupilumab treatment results in early and sustained improvements in itch in adolescents and adults with moderate to severe atopic dermatitis: Analysis of the randomized phase 3 studies SOLO 1 and SOLO 2, AD ADOL, and CHRONOS. *J. Am. Acad. Dermatol.* **2020**, *82*, 1328–1336. [[CrossRef](#)]
153. Nakagawa, H.; Nemoto, O.; Igarashi, A.; Saeki, H.; Kaino, H.; Nagata, T. Delgocitinib ointment, a topical Janus kinase inhibitor, in adult patients with moderate to severe atopic dermatitis: A phase 3, randomized, double-blind, vehicle-controlled study and an open-label, long-term extension study. *J. Am. Acad. Dermatol.* **2020**, *82*, 823–831. [[CrossRef](#)] [[PubMed](#)]
154. Kabashima, K.; Matsumura, T.; Komazaki, H.; Kawashima, M.; Nemolizumab, J.P.S.G. Trial of Nemolizumab and Topical Agents for Atopic Dermatitis with Pruritus. *N. Engl. J. Med.* **2020**, *383*, 141–150. [[CrossRef](#)] [[PubMed](#)]
155. Ratchataswan, T.; Banzon, T.M.; Thyssen, J.P.; Weidinger, S.; Guttman-Yassky, E.; Phipatanakul, W. Biologics for Treatment of Atopic Dermatitis: Current Status and Future Prospect. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 1053–1065. [[CrossRef](#)]

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156. Abeck, D.; Andersson, T.; Grosshans, E.; Jablonska, S.; Kragballe, K.; Vahlquist, A.; Schmidt, T.; Dupuy, P.; Ring, J. Topical application of a platelet-activating factor (PAF) antagonist in atopic dermatitis. *Acta Derm. Venereol.* **1997**, *77*, 449–451. [[CrossRef](#)] [[PubMed](#)]
  157. Itadani, S.; Takahashi, S.; Ima, M.; Sekiguchi, T.; Fujita, M.; Nakayama, Y.; Takeuchi, J. Discovery of Highly Potent Dual CysLT1 and CysLT2 Antagonist. *ACS Med. Chem. Lett.* **2014**, *5*, 1230–1234. [[CrossRef](#)] [[PubMed](#)]