

Supplementary Information

Sublethal systemic LPS in mice enables gut-luminal pathogens to bloom through oxygen species-mediated microbiota inhibition

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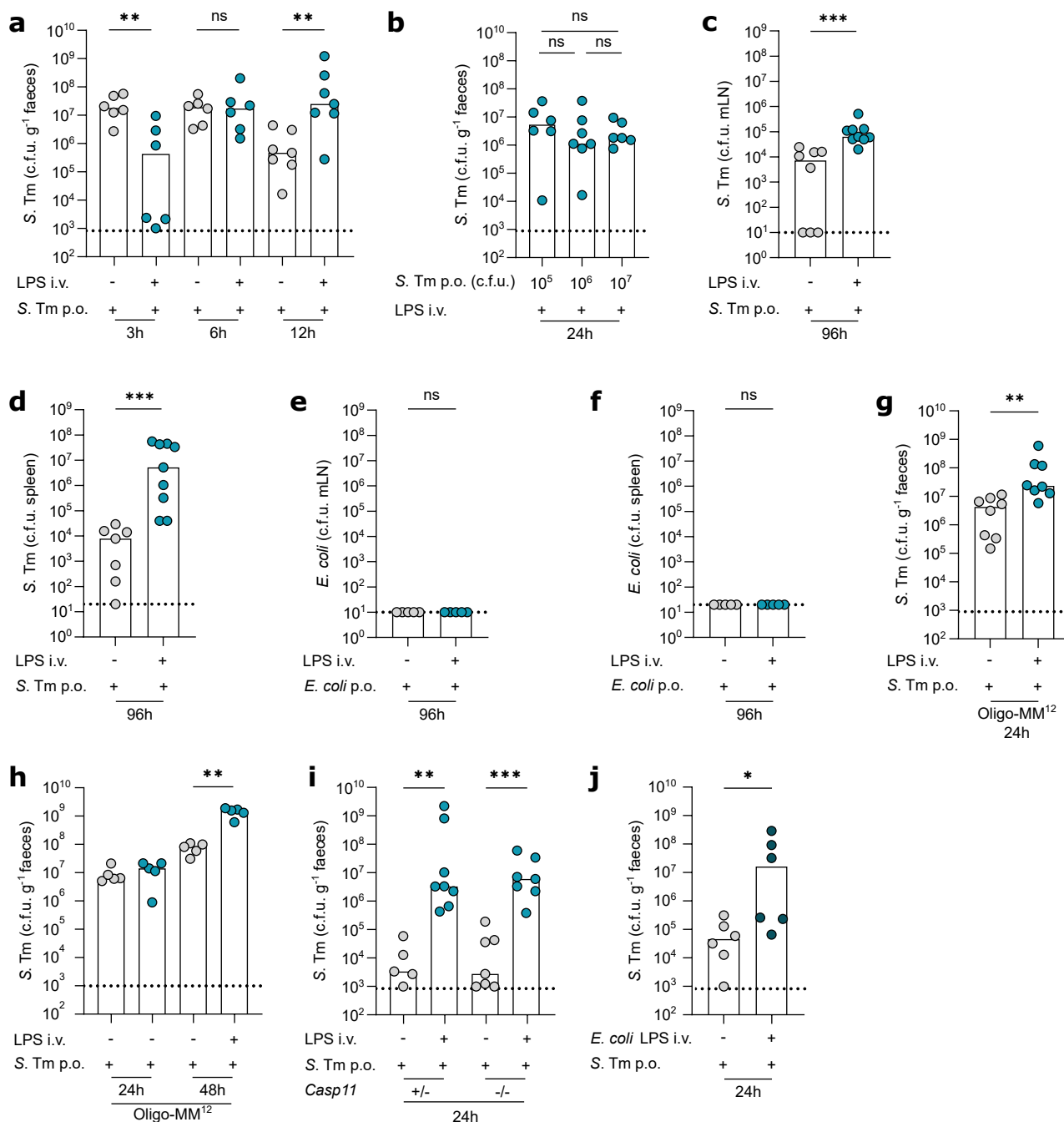
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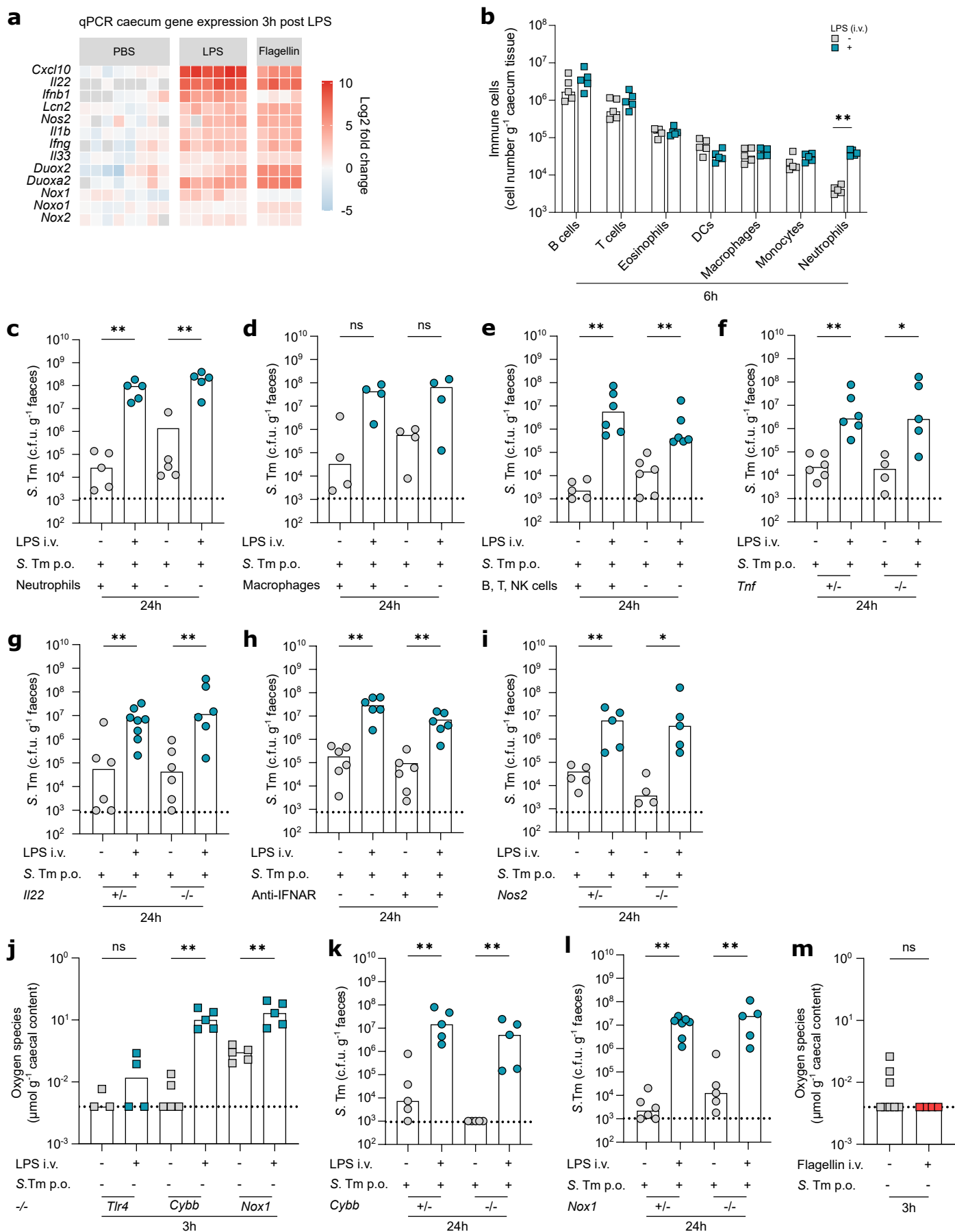
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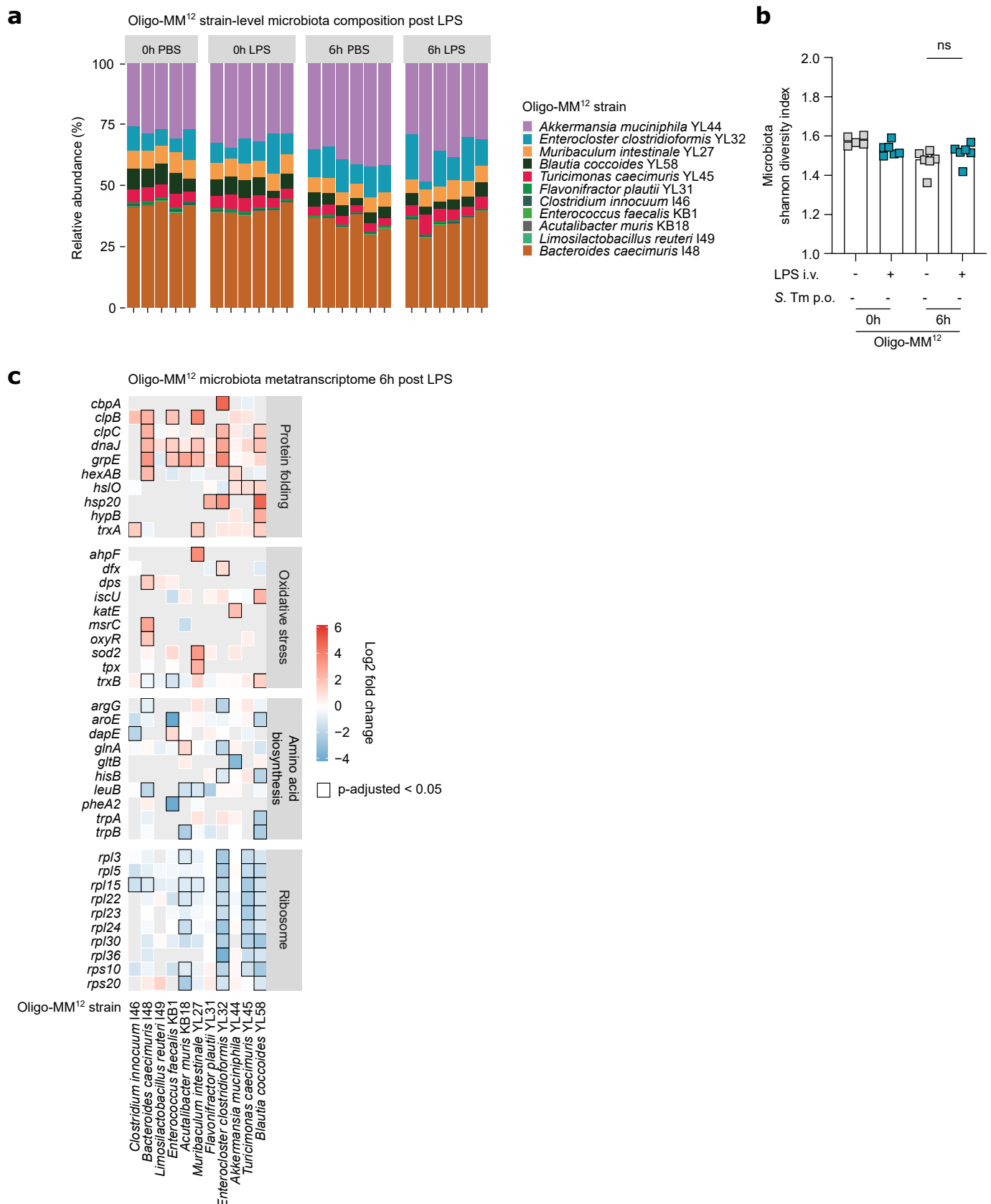
Supplementary Figure 1 Systemic LPS exposure promotes gut-luminal *S. Tm* to bloom. (a) Faecal *S. Tm* loads at 3, 6 or 12 h.p.i. in mice systemically exposed to PBS or LPS (minimum mice n=6, at least two independent replicates). (b) Faecal *S. Tm* loads at 24 h.p.i. in mice systemically exposed to LPS and orally infected with 10⁵, 10⁶ or 10⁷ c.f.u. *S. Tm* (minimum mice n=6, at least two independent replicates). (c) Mesenteric lymph node (mLN) *S. Tm* loads at 24 h.p.i. in mice systemically exposed to PBS or LPS (minimum mice n=8, at least two independent replicates). (d) Spleen *S. Tm* loads at 24 h.p.i. in mice systemically exposed to PBS or LPS (minimum mice n=7, at least two independent replicates). (e) mLN *E. coli* loads at 24 h.p.i. in mice systemically exposed to PBS or LPS (mice n=5, at least two independent replicates). (f) Spleen *E. coli* loads at 24 h.p.i. in mice systemically exposed to PBS or LPS (mice n=5, at least two independent replicates). (g) Faecal *S. Tm* loads at 24 h.p.i. in Oligo-MM12 mice systemically exposed to PBS or LPS (mice n=8, at least two independent replicates). (h) Faecal *S. Tm* loads at 24 h and 48 h.p.i. in Oligo-MM12 mice orally infected with *S. Tm* and systemically exposed to PBS or LPS at 24 h.p.i. (mice n=5, at least two independent replicates). (i) Faecal *S. Tm* loads at 24 h.p.i. in *Casp11*^{+/-} and *Casp11*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (j) Faecal *S. Tm* loads at 24 h.p.i. in mice systemically exposed to PBS or 5 µg *E. coli* LPS (mice n=6, at least two independent replicates). Bars indicate median values. Dotted lines indicate

conservative average limit of detection. P values were calculated using the two-sided Mann-Whitney U test (a,c-j) or two-sided Kruskal-Wallis test with Dunn's multiple comparisons adjustment (b). ns, not significant; * $P<0.05$ and ** $P<0.01$. Source data are provided in the Source Data file.



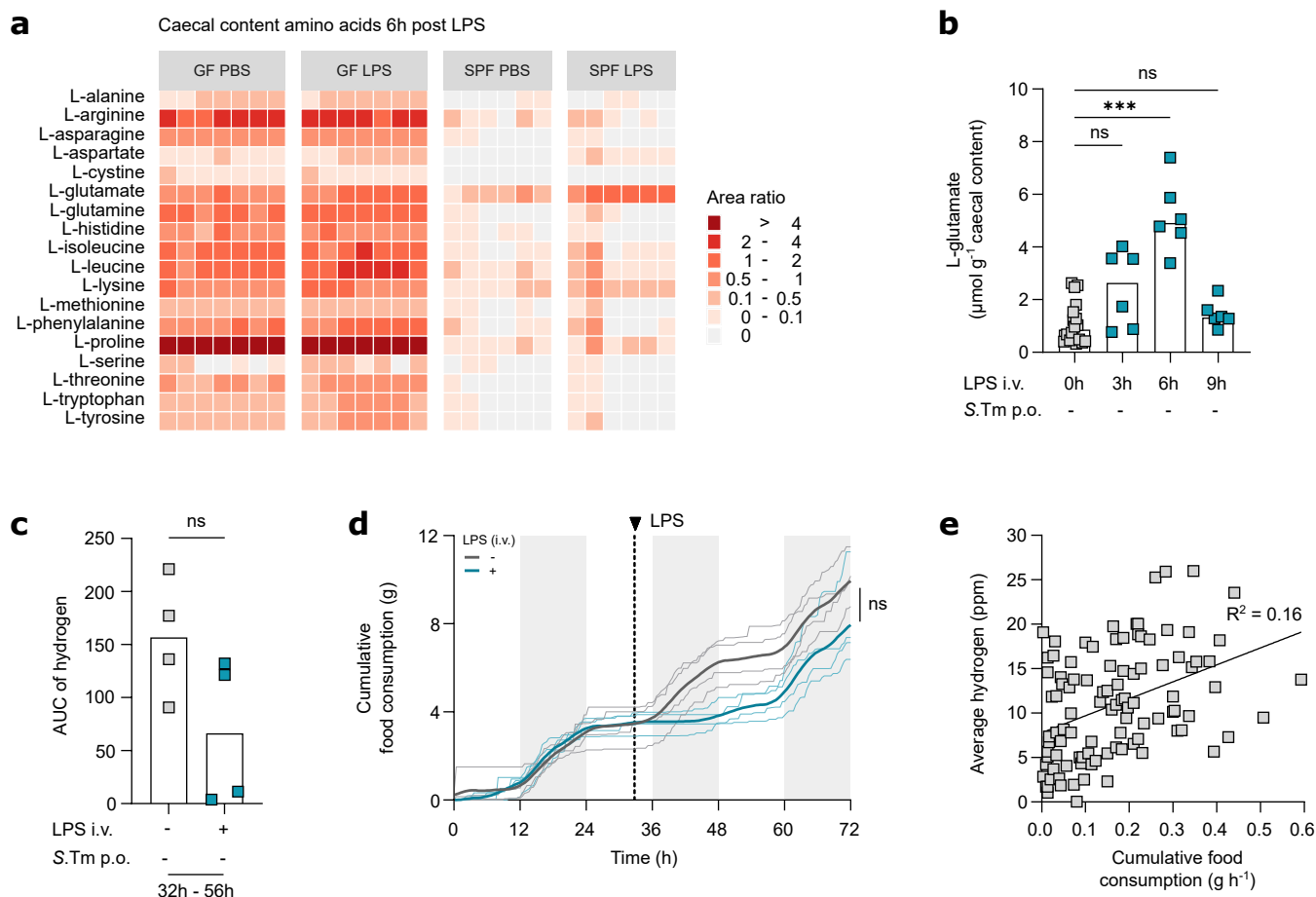
Supplementary Figure 2 Contribution of immune mediators to LPS-induced gut-luminal *S. Tm* bloom. (a) qPCR log₂ fold change as determined of classical inflammatory response genes in mice systemically exposed to PBS or LPS at 3 h.p.i (minimum mice n=4, at least two independent replicates). (b) Caecum tissue immune cell composition at 6 h.p.i. in mice systemically exposed to PBS or LPS (mice

n=5, at least two independent replicates). (c) Faecal S. Tm loads at 24 h.p.i. in mice treated with PBS or anti-Ly6G and systemically exposed to PBS or LPS (mice n=5, at least two independent replicates). (d) Faecal S. Tm loads at 24 h.p.i. in mice treated with PBS or anti-CSF1R and systemically exposed to PBS or LPS (mice n=4, at least two independent replicates). (e) Faecal S. Tm loads at 24 h.p.i. in *Rag2*^{+/-} *Il2rg*^{+/-} and *Rag2*^{-/-} *Il2rg*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (f) Faecal S. Tm loads at 24 h.p.i. in *Tnf*^{+/-} and *Tnf*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=4, at least two independent replicates). (g) Faecal S. Tm loads at 24 h.p.i. in *Il22*^{+/-} and *Il22*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=6, at least two independent replicates). (h) Faecal ^S. Tm loads at 24 h.p.i. in mice treated with PBS or anti-IFNAR and systemically exposed to PBS or LPS (minimum mice n=6, at least two independent replicates). (i) Faecal ^S. Tm loads at 24 h.p.i. in *Nos2*^{+/-} and *Nos2*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=4, at least two independent replicates). (j) Caecal oxygen species levels at 3 h.p.inj. in *Tlr4*^{-/-}, *Cybb*^{-/-}, and *Nox1*^{-/-} mice systemically exposed to PBS or LPS (minimum mice n=3, at least two independent replicates). (k) Faecal S. Tm loads at 24 h.p.i. in *Cybb*^{+/-} and *Cybb*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (l) Faecal S. Tm loads at 24 h.p.i. in *Nox1*^{+/-} and *Nox1*^{-/-} littermates systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (m) Caecal oxygen species levels at 3 h.p.inj. in mice systemically exposed to PBS or flagellin (minimum mice n=4, at least two independent replicates). Bars indicate median values. Dotted lines indicate conservative average limit of detection. *P* values were calculated using the two-sided Mann-Whitney U test (b-m). ns, not significant; **P*<0.05 and ***P*<0.01. Source data are provided in the Source Data file.

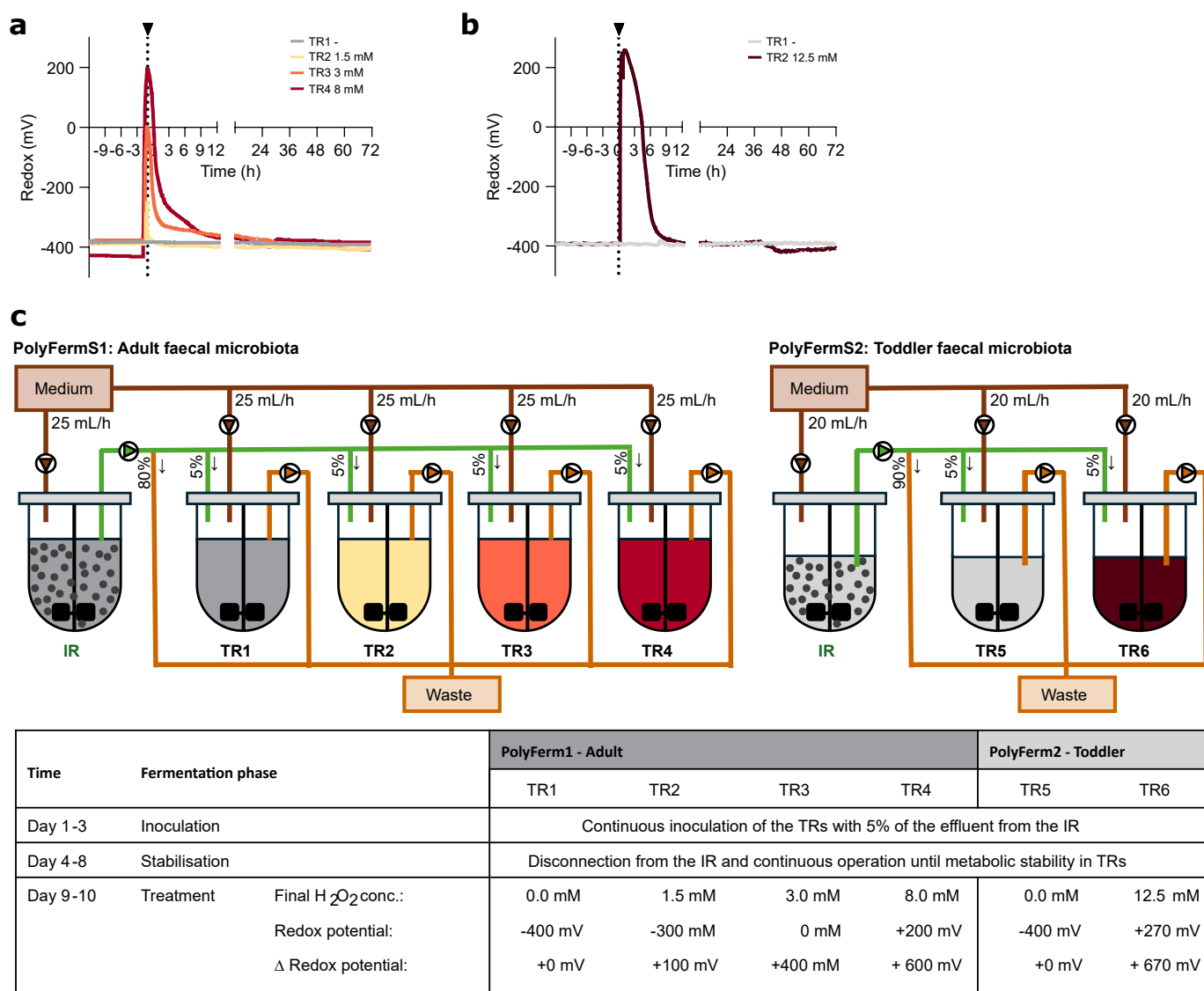


Supplementary Figure 3 Effect of systemic LPS exposure on the gut microbiota. (a) 16S rRNA sequencing strain-level microbiota composition at 0 and 6 h.p.inj. in Oligo-MM12 mice systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (b) 16S rRNA sequencing Shannon diversity index at 0 and 6 h.p.inj. in Oligo-MM12 mice systemically exposed to PBS or LPS (minimum mice n=5, at least two independent replicates). (c) Caecal microbiota metatranscriptome log₂ fold change of top 10 significantly differentially expressed genes involved in protein folding, oxidative stress, amino acid biosynthesis and ribosomes at 6 h.p.inj. in Oligo-MM12 mice systemically exposed to

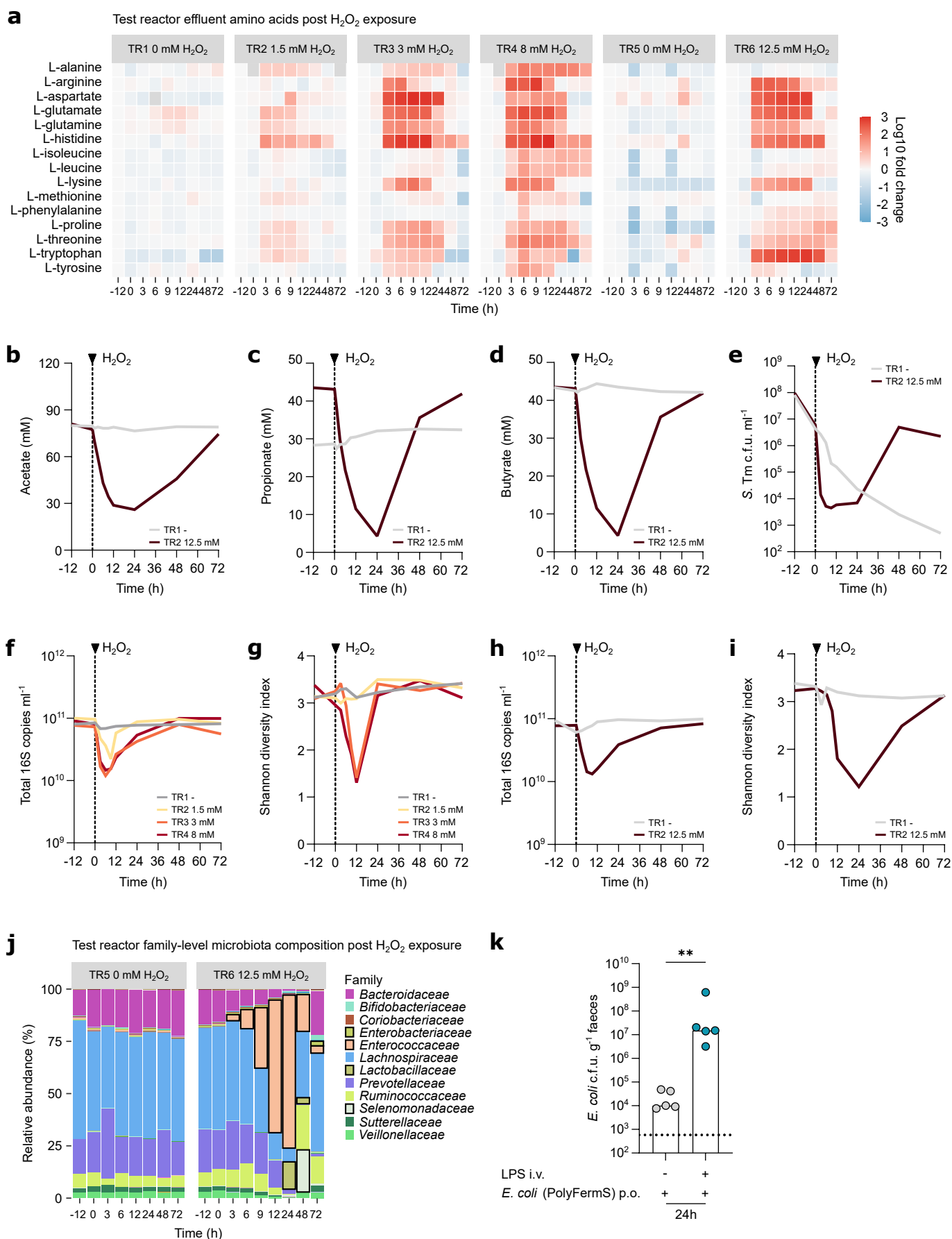
LPS compared to PBS, (mice n=6, at least two independent replicates). Bars indicate median values. *P* values were calculated using the two-sided Mann-Whitney U test (b) or two-sided Wald test with Benjamini-Hochberg multiple test correction (c). ns, not significant. Source data are provided in the Source Data file.



Supplementary Figure 4 Effect of systemic LPS exposure on the gut microbiota. (a) Caecal amino acid levels at 6 h.p.inj. in germ-free and SPF mice systemically exposed to PBS or LPS. Area ratio calculated to ^{13}C -L-glutamic acid internal standard (minimum mice $n=6$, at least two independent replicates). (b) Caecal L-glutamate levels at 0, 3, 6 or 9 h.p.inj. in mice systemically exposed to LPS (minimum mice $n=6$, at least two independent replicates). (c) Area under the curve of hydrogen between 32 h and 56 h in mice systemically exposed to PBS or LPS at 32 h (minimum mice $n=4$, at least two independent replicates). (d) Cumulative food consumption of mice systemically exposed to PBS or LPS at 32 h, curves obtained by smoothing function of data obtained every 24 min per mouse (mice $n=4$, at least two independent replicates). (e) Correlation between cumulative food consumption and average hydrogen at food consumption levels calculated based on 3 h intervals. R^2 was calculated by linear regression (mice $n=4$, at least two independent replicates). Bars indicate median values. Dashed lines indicate time of injection. Grey rectangles indicate dark phase. P values were calculated using the two-sided Kruskal-Wallis test with Dunn's multiple comparisons adjustment (b) or two-sided Mann-Whitney U test (c,d). ns, not significant; *** $P<0.001$. Source data are provided in the Source Data file.

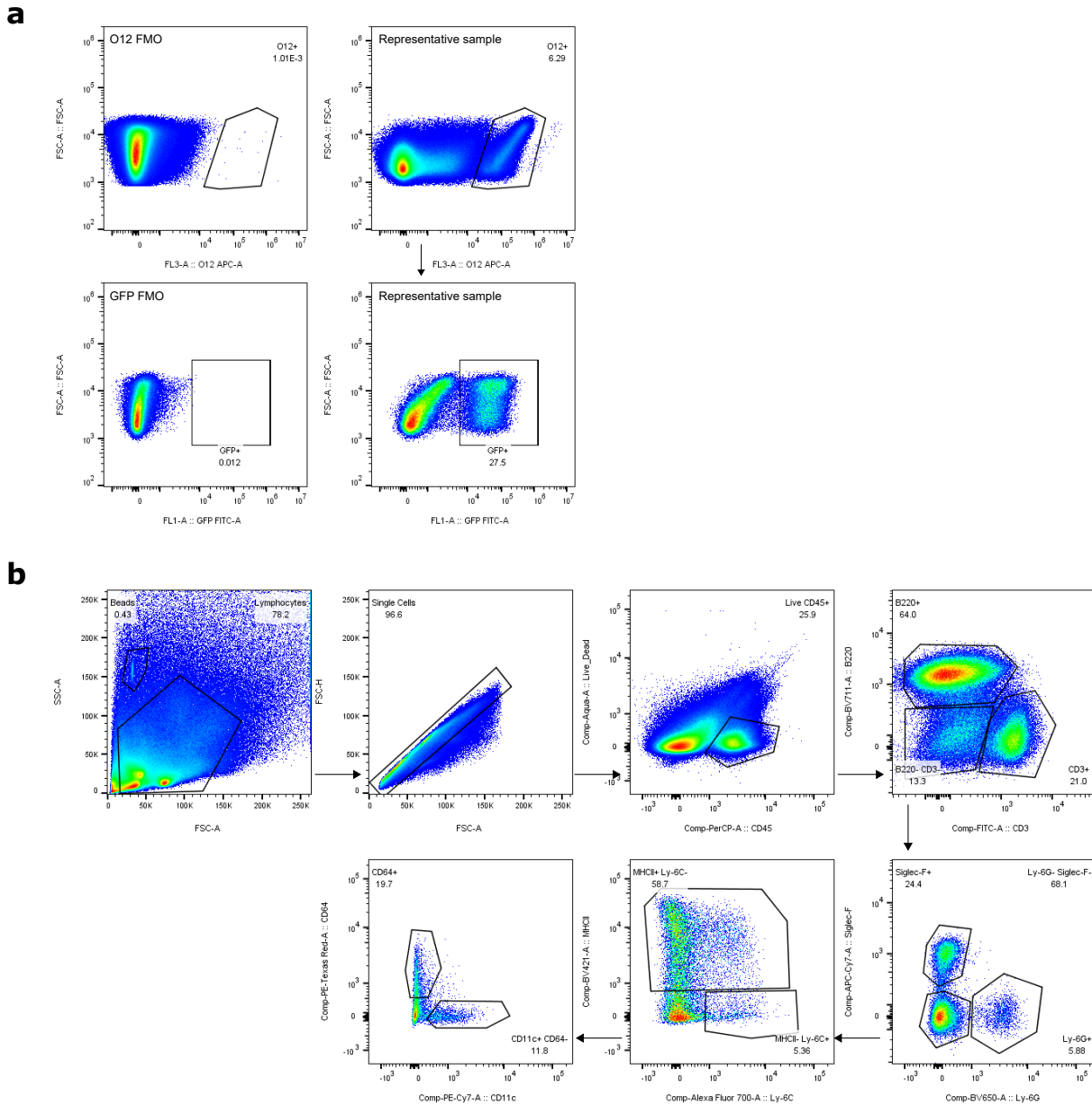


Supplementary Figure 5 PolyFermS set-up. (a,b) Redox potential in test reactors (TR) post-H₂O₂ exposure (reactors n=4, n=2). (c) Two independent PolyFermS experiments were conducted under the proximal colon conditions of a healthy adult and a healthy toddler. Each experimental set-up included an inoculum reactor (IR), containing immobilised human faecal microbiota gel beads and consecutive TRs, which were continuously inoculated with 5% of the fermented effluent. The IR of PolyFermS1 contained immobilised faecal microbiota of an adult donor and inoculated TR1-4 while the IR of PolyFermS2 harboured immobilised faecal microbiota from a toddler and inoculated TR5 and TR6. After a 3-day-period of continuous fermentation, the TRs were disconnected from the IR and further stabilised for another 4 days to reach metabolic stability before treatment initiation. TRs were inoculated with *S. Tm* at -12h to reach a density of 1×10^8 c.f.u. ml⁻¹. 12h post *S. Tm* inoculation, TRs were exposed to varying H₂O₂ concentrations 0 mM (TR1,TR5), 1.5 mM (TR2), 3 mM (TR3), 8mM (TR4) and 12.5 mM (TR6) to increase the redox potential, a common measurement of the degree of anaerobiosis (reactors n=6). Dashed lines indicate time of H₂O₂ exposure. Source data are provided in the Source Data file.



Supplementary Figure 6 Oxidative stress inhibits human gut microbiota fermentation and promotes facultative anaerobic pathogens blooms. (a) An inoculation reactor (IR) containing immobilised faecal microbiota of an adult donor inoculated TR1-4, and an IR harbouring immobilised faecal microbiota from a toddler inoculated TR5 and TR6. TRs were inoculated with *S. Tm* at -12h to reach a

density of 1×10^8 c.f.u. ml⁻¹. 12h post *S. Tm* inoculation, TRs were exposed to varying H₂O₂ concentrations 0 mM (TR1/5), 1.5 mM (TR2), 3 mM (TR3), 8mM (TR4) and 12.5 mM (TR6) (reactors n=6). Log10 fold change amino acids in the effluent post-H₂O₂ exposure compared to respective mock TR (TR1/TR5) (reactors n=6). (b-d) Acetate, propionate, and butyrate concentrations in TR effluent post-H₂O₂ exposure (reactors n=2). (e) *S. Tm* loads in TR effluent post-H₂O₂ exposure (reactors n=2). (f-i) 16S rRNA sequencing copy number and Shannon diversity index in TR effluent post-H₂O₂ exposure (reactors n=6). (j) 16S rRNA sequencing family-level microbiota composition in TR effluent post-H₂O₂ exposure (reactors n=2). (k) Faecal *E. coli* (PolyFermS) loads at 24 h.p.i. in mice systemically exposed to PBS or LPS (mice n=5, at least two independent replicates). Dashed lines indicate time of H₂O₂ exposure. Bars indicate median values. Dotted lines indicate conservative average limit of detection. *P* values were calculated using the two-sided Mann-Whitney U test (k). ***P*<0.01. Source data are provided in the Source Data file.



Supplementary Figure 7 Gating strategies. (a) Gating strategy for *S. Tm PsicA-gfp* using primary antibody human anti-*S. Tm* O12 and secondary antibody goat anti-human AF647. Including a control where the primary antibody was not added and a control *S. Tm* strain that does not express GFP. (b) Lamina propria gating strategy, initial gate was set based on size and granularity, live CD45⁺ cells, CD3⁺ T cells, B220⁺ B cells, Ly-6G⁺ neutrophils, Siglec-F⁺ eosinophils, Ly-6G⁻ Ly-6C⁺ monocytes, CD64⁺ macrophages and CD11c⁺ dendritic cells.

Supplementary Table 1 Bacterial strains

Relevant genotype	Strain number	Reference
Wild type <i>S. Tm</i> SL1344	SB300	Hoiseth & Stocker 1981 ⁸⁸
SB300 Δ sopB Δ sipA Δ sopE Δ sopE2	M566	Ehrbar 2003 ¹²¹
SB300 Δ cyxA:cat WITS17:aphT	Z6860	This study
SB300 Δ cydB:cat WITS19:aphT	Z6889	This study
SB300 Δ cyoA:cat WITS17:ahpT	T1728	This study
SB300 Δ cyxA:cat Δ cydB WITS19:ahpT	Z6859	This study
SB300 Δ gltA:cat WISH1:bla	Z8390	Nguyen 2024 ³⁷
SB300 Δ acnA:aphT Δ acnB:cat WISH3:bla	Z8382	Nguyen 2024 ³⁷
SB300 Δ icdA:aphT WISH4:bla	Z8384	Nguyen 2024 ³⁷
SB300 Δ sucB:aphT WISH5:bla	Z8386	Nguyen 2024 ³⁷
SB300 Δ sucD:cat WITS2:aphT	T1703	This study
SB300 Δ sdh:cat WITS13:aphT	Z3436	Nguyen 2020 ⁶⁸
SB300 Δ dcuA Δ dcuB Δ dcuC Δ frd WITS2:aphT	Z3444	Nguyen 2020 ⁶⁸
SB300 Δ fumAC Δ fumB WITS17:aphT	Z3460	Nguyen 2020 ⁶⁸
SB300 Δ mdh:cat WITS19:aphT	Z3468	This study
<i>E. coli</i>	CFT073	Mobley 1990 ¹²²
<i>K. pneumoniae</i>	T821	This study
<i>E. faecium</i>	T1749	This study
<i>E. coli</i> (PolyFermS)	T1740	This study
<i>Agathobacter rectalis</i>	A1-86	DSM No.: 17629
<i>Bacteroides fragilis</i>	EN-2	DSM No.: 2151
<i>Bacteroides thetaiotaomicron</i>	EP50	DSM No.: 2079
<i>Bacteroides uniformis</i>	VPI 0061	DSM No.: 6597
<i>Clostridium perfringens</i>	C36	DSM No.: 11782
<i>Collinsella aerofaciens</i>	VPI 1003	DSM No.: 3979
<i>Coprococcus comes</i>	VPI CI-38	ATCC No.: 27758
<i>Dorea formicigenerans</i>	VPI C8-13	DSM No.: 3992
<i>Enterocloster bolteae</i>	WAL 16351	DSM No.: 15670
<i>Escherichia coli</i>	BW25113	PMID:10829079
<i>Fusobacterium nucleatum</i>	VPI 4355	DSM No.: 15643
<i>Lacrimispora saccharolytica</i>	WM1	DSM No.: 2544
<i>Parabacteroides merdae</i>	VPI T4-1	DSM No.: 19495
<i>Phocaeicola vulgatus</i>	LRA 049 07 85	DSM No.: 1447
<i>Roseburia intestinalis</i>	L1-82	DSM No.: 14610
<i>Ruminococcus gnavus</i>	VPI C7-9	ATCC No.: 29149
<i>Streptococcus parasanguinis</i>	SS 898	DSM No.: 6778
<i>Streptococcus salivarius</i>	275	DSM No.: 20219
<i>Thomasclavelia ramosa</i>	113-I	DSM No.: 1402
<i>Bacteroides caecimuris</i>	I48	Brugiroux 2016 ⁵⁵
<i>Bifidobacterium animalis</i>	YL2	Brugiroux 2016 ⁵⁵
<i>Blautia pseudococcoides</i>	YL58	Brugiroux 2016 ⁵⁵
<i>Clostridium innocuum</i>	I46	Brugiroux 2016 ⁵⁵
<i>Enterocloster clostridioformis</i>	YL32	Brugiroux 2016 ⁵⁵
<i>Enterococcus faecalis</i>	KB1	Brugiroux 2016 ⁵⁵
<i>Flavonifractor plautii</i>	YL31	Brugiroux 2016 ⁵⁵
<i>Limosilactobacillus reuteri</i>	I49	Brugiroux 2016 ⁵⁵

Supplementary Table 2 RT² qPCR primers

Gene	Species	Ref.Seq.	Cat.Nr.
<i>Actb</i>	Mouse	NM_007393	PPM02945B-200
<i>Cxcl10</i>	Mouse	NM_021274	PPM02978E-200
<i>Il22</i>	Mouse	NM_016971	PPM05481A-200
<i>Ifnb1</i>	Mouse	NM_010510	PPM03594C-200
<i>Lcn2</i>	Mouse	NM_008491.1	PPM03770A-200
<i>Nos2</i>	Mouse	NM_010927	PPM02928B-200
<i>Il1b</i>	Mouse	NM_008361	PPM03109F-200
<i>Ifng</i>	Mouse	NM_008337	PPM03121A-200
<i>Il33</i>	Mouse	NM_133775	PPM32527A-200
<i>Duox2</i>	Mouse	NM_177610	PPM40846A-200
<i>Duoxa2</i>	Mouse	NM_025777	PPM32328A-200
<i>Nox1</i>	Mouse	NM_172203	PPM34199A-200
<i>Noxo1</i>	Mouse	NM_027988	PPM36220A-200
<i>Cybb</i>	Mouse	NM_007807	PPM32951A-200

Supplementary Table 3 Antibodies

Antigen	Fluorophore	Company	Clone	Cat #	Lot #	Dilution
Fixable live/dead dye	Aqua	Thermo Fisher Scientific		L34957	2335578	1:1000
CD45	PerCP	Biolegend	30-F11	103130	B236192	1:100
CD3	FITC	Biolegend	17A2	100203	B388790	1:100
B220	BV711	Biolegend	RA3-6B2	103255	B305860	1:200
Ly-6G	BV650	Biolegend	1A8	127641	B314454	1:100
Siglec-F	APC-Cy7	BD Bioscience	E50-2440	565527	1062707	1:200
MHCII	BV421	Biolegend	M5/114.15.2	107632	B335578	1:100
Ly-6C	AF700	Biolegend	HK1.4	128024	B318988	1:200
CD64	PE/Dazzle	Biolegend	X54-5/7.1	139320	B304964	1:100
CD11c	PE-Cy7	Biolegend	N418	117318	B264758	1:200

References

121. Ehrbar, K., Friebel, A., Miller, S. I. & Hardt, W. D. Role of the Salmonella Pathogenicity Island 1 (SPI-1) Protein InvB in Type III Secretion of SopE and SopE2, Two Salmonella Effector Proteins Encoded Outside of SPI-1. *J. Bacteriol.* **185**, 6950–6967 (2003). doi:10.1128/JB.185.23.6950-6967.2003.
122. Mobley, H. L. *et al.* Pyelonephritogenic *Escherichia coli* and killing of cultured human renal proximal tubular epithelial cells: Role of hemolysin in some strains. *Infect. Immun.* **58**, 1281–1289 (1990). doi:10.1128/iai.58.5.1281-1289.1990.