

Selective estrogen receptor modulators contribute to prostate cancer treatment by regulating the tumor immune microenvironment

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ABSTRACT

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Prostate cancer (PC) has previously been established as a cold tumor and develops in an inert immunosuppressive environment. Current research focuses on altering the immune microenvironment of PC from cold to hot; thus, in the present review, the diverse roles of estrogen and estrogen receptor (ER) signaling was examined in the tumor cell and tumor immune microenvironment (TIM). We hypothesized that ER α promotes PC progression and ER β impedes epithelial-mesenchymal transition in PC cells, while in the TIM, ER β mediates the immunosuppressive environment, and low levels of $ER\alpha$ is associated with disease development. Selective estrogen receptor modulators (SERMs) or selective ER degraders play diverse roles in the regulation of ER isoforms. Patients with PC may benefit from the use of SERMs, including raloxifene, in combination with anti-PD1/PD-L1 checkpoint immunotherapy, or TGF- β or Wnt antagonists. The present review demonstrated that immunotherapy-based strategies combined with SERMs may be an option for the future of PC-targeting therapy.

INTRODUCTION

Prostate cancer and tumor immune microenvironment

Prostate cancer (PC) remains a common health problem, and impacts both society and the economy. According to recent data, it ranks the most common cancer among men in Western countries and possesses the second highest rate of cancer-related death. At present, first-line treatment for PC includes androgen-deprivation therapy, which is used in patients with progressive, recurrent and metastatic PC. However, almost all cases of PC progress from hormone-sensitive PC to castration-resistant PC (CRPC) following treatment.^{1 2} Notably, numerous epidemiological studies, including meta-analyses, indicate that hormone induction and the tumor immune microenvironment (TIM) are associated with PC progression.³ The TIM of PC plays a critical role in endocrine therapy and immunotherapy targeting tumor

progression, metastasis and treatment resistance. Specifically, the prostatic stromal cells associated with immunological cells participate in prostatic pathologies, particularly in cancer, building a complex TIM. Additionally, increased accumulation of pro-inflammatory factors, cancer associated fibroblasts (CAFs) and myofibroblasts generate a progressive and tumor-promotive TIM, in which the stroma cell stimulates the proliferation and migration of PC cells. The diverse activation pathways in CAFs mainly include transforming growth factor- β (TGF- β), platelet-derived growth factor, hepatocyte growth factor, fibroblast growth factor, interleukins, metalloproteinases and reactive oxygen species, which take effect via activating heat shock factor 1 and nuclear factor kappa B (NF-kB).⁴ The TIM associated with PC is highly immunosuppressive, containing (1) cells, including tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs) and regulatory T cells (Tregs); (2) cytokine milieu, secreted by stromal cells and fibroblasts of the TIM; (3) adenosine, produced from prostatic acid phosphatase; and (4) molecular pathways, including TGF-B and Wnt/ β -catenin. Both adenosine and TGF- β may serve as potential immunosuppressive molecules or pathways, and act as therapeutic targets⁵ (figure 1). Moreover, in the TIM, TGF- β and Wnt/ β -catenin are frequently activated in advanced PC, and contribute to therapy resistance and metastasis, including resistance to docetaxel, enzalutamide and abiraterone acetate-prednisone.^{6–8}

Estrogen and ER in PC

Estrogen treatment for PC was verified to be effective in previous clinical trials.⁹ Estrogen is implicated in the initiation of PC.¹⁰ Estrogendependent signaling-induced TMPRSS2-ERG expression leads to TMPRSS2-ERG fusion



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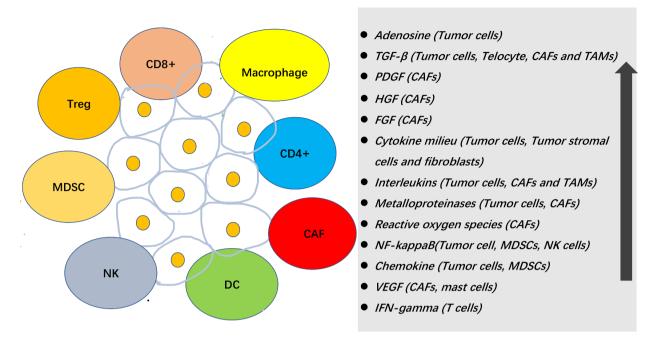


Figure 1 Communication between prostate cancer (PC) and stromal cells in the tumor immune microenvironment (TIM). These stromal cells include cancer associated fibroblasts (CAF), regulatory T cells (Tregs), tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs), CD8 +T cells, CD4 +T cells, dendritic cells (DCs) and natural killer (NK) cells. A series of immune mediators and their origins are presented in the gray box, containing adenosine, hepatocyte growth factor (HGF), platelet-derived growth factor (PDGF), transforming growth factor- β (TGF- β), fibroblast growth factor (FGF), interleukins, metalloproteinases, reactive oxygen species, cytokine milieu, heat shock factor 1 (HSF1) and nuclear factor kappa B (NF-kB).

PC.¹¹ It has been demonstrated that estrogen and its active product, 17β-estradiol (E2), originate from androgen metabolism via steroidogenesis, which is associated with PC development.¹² In animal models and clinical samples, high levels of estrogen are positively associated with the aggressiveness of PC.¹³ Moreover, the modulation of estrogen in cancer progression is highlighted due to the activity of estrogen on its receptors ER α and ER β , which act as transcription factors, contributing to healthy prostate development.¹⁴ Results of a previous study demonstrated that treatment with E2 (1µM) for 24 hours may led to the downregulation of ERs, while treatment with physiological concentrations of E2 (0.1 nM) for 24 hours increased the levels of ERs. E2-induced ERK1/2 activation was not inhibited by ERs antagonists, indicating the involvement of alternative estrogen-regulated mechanisms. Therefore, the molecular mechanism underlying estrogen in the promotion of PC progression is complex. ER α activation using the agonist PPT, and ER β activation using the agonist DPN enhanced the phosphorylation of SRC in PC-3 cells of PC. ER enhanced the tumorigenic potential of PC by inducing rapid responses of SRC and PI3K/AKT.¹⁶

Results of a previous study demonstrated that ER α is mainly localized in prostate stroma cells, and estrogen plays regulatory effects on the prostate adenoepithelium using paracrine pathways.¹⁷ Results of a previous study further indicated that AR-positive human PC cell lines, including LNCaP, LAPC4 and 22Rv1 did not exhibit ER expression levels. AR-negative human PC cell lines, including DU145 and PC3 did not exhibit ER β expression levels, while in PC3 cells, weak expression of ER α was determined.¹⁸ ER α is crucial in the invasion and migration of PC cells.¹⁹ ER α , as the predominant subtype, is associated with high-grade PC.^{20 21} ER α has oncogenic functions in animal models, and its activation leads to cancer proliferation.^{22 23}

Compared with healthy prostate tissue, ER β is overexpressed in PC tissues.²⁴ Antiestrogens and selective estrogen receptor modulators (SERMs) are found to reduce the risk of PC development in PC metastases with high levels of ER β expression.^{25 26} ER β expression is associated with poor clinical outcomes in hormone-sensitive PC, and is increased in CRPC.²⁷ The ER isoforms β 2 and β 5 are associated with poor patient outcomes, and promote PC migration and invasion.²⁸ On the other hand, ER β is regarded as a tumor suppressor, and ER β inhibition promoted prostate hyperplasia and tumor development.^{29–31} ER β expression is decreased in the process of PC progression.^{32–34} Therefore, estrogen/ER pathways exhibit diverse and complex roles in PC development.¹⁸ The specific mechanisms underlying ER isoforms require further evaluation and exploration.

SERMs and PC

SERMs, including selective estrogen receptor degraders (SERDs), have previously been used for the treatment of breast cancer by targeting ERs.³⁵ These agents include tamoxifen, toremifene and raloxifene.¹⁸ SERMs competitively block the binding of estrogen with ERα. SERDs,

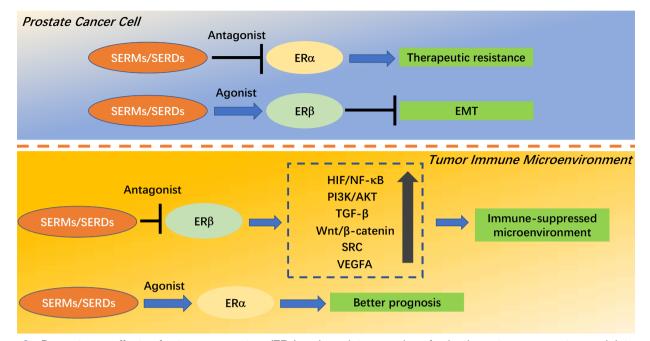


Figure 2 Downstream effects of estrogen receptors (ERs) and regulatory modes of selective estrogen receptor modulators (SERMs)/selective estrogen receptor degraders (SERDs) in prostate cancer (PC). In the top panel, SERM/SERDs which antagonize ER α would reverse resistance, while an agonist of ER β activity would reverse epithelial-mesenchymal transition (EMT) in PC cells. In the bottom panel, SERMs/SERDs which antagonize ER β would shift the immunosuppressed tumor immune microenvironment (TIM) to an immune-activated TIM, while an agonist of ER α activity would improve prognosis. VEGFA, vascular endothelial growth factor A.

such as fulvestrant, contribute to ER α downregulation and degradation. 36

Notably, $ER\beta$ may play a role as a response marker of tamoxifen.³⁷ The majority of clinical trials associated with tamoxifen yielded moderate rates of survival against cancer.³⁸ Targeting ERa with tamoxifen provides a novel therapeutic strategy for the castration-resistant ER-positive subtype of metastatic PC, which is associated with the suppression of phosphatidylinositol-4phosphate 5-kinase- α /AKT and matrix metalloproteinase (MMP)-9/vascular endothelial growth factor (VEGF) signaling pathways.³⁹ Thus, SERMs may exhibit potential in ER-progressed PC therapy. Results of a phase II clinical trial of CRPC demonstrated that treatment with raloxifene leads to a stable disease status, by inhibiting the growth of tumors in PC xenograft models.⁴⁰ However, previous studies have also demonstrated the antiproliferative role of ER β , and a loss of ER β expression in highgrade tumors. Therefore, treatment with single SERMs targeting ERs may not fully inhibit PC progression.

ESTROGEN/ER ON THE TIM

Estrogen can induce the activation of M2-type macrophages leading to tumor progression.⁴¹ Multiple cytokines secreted by these functional M1-type macrophages, including IL-6, MMP-9, IL-1 β and TNF α , are inhibited following activated estrogen/ER α signaling.⁴²⁻⁴⁴ Moreover, estrogen may directly or indirectly suppress the cytotoxic activity of NK cells.⁴⁵⁻⁴⁶ As an immune-suppressive regulator, estrogen supports tumor initiation and progression. Therefore, estrogen-targeted therapy may orchestrate an antitumor TIM and guide proimmune surveillance.

Results of a previous study demonstrated that TGF- β and ER signaling interact during cancer development.⁴⁷ ER α , downstream of TGF- β , plays a dual role in regulating TGF- β signaling and cancer development, via a novel non-genomic mechanism.^{48–50} The activated ER signaling pathway is associated with TGF- β in the progression of LNCaP PC models.⁵¹ Cell migration and invasion induced by ER α are associated with the upregulated activity of MMP-2 and MMP-9, accompanied with the upregulation of a series of epithelial-mesenchymal transition (EMT)related genes, such as TGF- β 1, zinc finger E-box-binding homeobox 1/2 and vimentin.¹⁹ In prostate stromal cells, estrogen induces MMP-2 expression via binding ER α to induct TGF- β , and stimulates PC cell invasion in a paracrine manner.⁵²

ERβ is mainly expressed in immune cells, particularly in the inflammatory and hypoxic TIM, compared with ERα,⁵³ which may differentially contribute to the response to anti-estrogen or treatment with SERMs/ SERDs.⁵⁴ Following E2 stimulation, ERβ promotes Treg differentiation, guides the secretion of IL-10 and TGF-β, downregulates CD8 +T cell activity and suppresses tumor immunocytotoxicity induced by T cells.^{55–57} E2 requires ERβ rather than ERα signaling for increasing the expression levels of Foxp3, GATA3 and CD25 in Treg cells.⁵⁸

SERMs or SERDs	Cancer stage	Primary purpose	Primary indication /outcome	Clincial trial phase	Clinical trial number	Combination drug/ therapy	Sponsor
Raloxifene	Metastatic or hormone- refractory prostate cancer	Treatment	PFS, AEs, QoL assessment, survival time 1 (completed)	I (completed)	NCT01050842	Bicalutamide	Mayo Clinic
Toremifene	Stage I or stage II prostate cancer followed by radical prostatectomy	Prevention (neoadjuvant therapy)	Percent of radical prostatectomy tissue volume	II (completed)	NCT00020735	Surgery	University of Pittsburgh
Tamoxifen	Locally advanced prostate cancer	Prevention	Incidence of gynecomastia and breast pain	III (completed)	NCT00233610	Bicalutamide	AstraZeneca
Zuclomiphene citrate	Advanced prostate cancer	Treatment	Change in frequency of moderate to severe hot flashes	II (completed)	NCT03646162	ADT	Veru Inc.
Toremifene citrate	Prostate cancer on androgen deprivation therapy	Prevention	Percentage of subjects at 24 months with III (completed) at least one new vertebral fracture	III (completed)	NCT00129142	ADT	GTx
Toremifene	Prostate cancer on androgen deprivation therapy	Prevention	The efficacy of toremifene in the reduction in the risk of new bone fracture occurrences	III (withdrawn)	NCT01214291	ADT	GTx
Raloxifene hydrochloride	Prostate cancer undergoing surgery	Treatment (neoadjuvant therapy)	Collection and interrogation of prostate cancer samples	II (withdrawn)	NCT03147196	Bicalutamide surgery	Mayo Clinic
Tamoxifen	Adenocarcinoma of the prostate gland but with no evidence of distant metastasis	Treatment	The extent of gynaecomastia and breast II (completed) pain by treatment group	II (completed)	NCT00637871	Casodex	AstraZeneca
Toremifene	High grade prostatic intraepithelial neoplasia (PIN)	Prevention	The efficacy of toremifene in the prevention of prostate cancer	III (completed)	NCT00106691		GTx
Tamoxifen	Prostate cancer with stage T1a, Treatment T1b T2a or T2b Nx Mx	Treatment	Total prostate volume	ll (unknown)	NCT00866554	Bicalutamide, dutasteride, brachytherapy	CHU de Quebec- Universite Laval
Toremifene citrate	High grade prostate intraepithelial neoplasia	Prevention (chemoprevention)	Investigational medication that reduce high grade PIN and prevent the occurrence of prostate cancer	II (completed)	NCT00028353		GTx
Fulvestrant	Recurrent prostate cancer	Treatment	Proportion of patients who respond to treatment	II (terminated)	NCT00217464		Roswell Park Cancer Institute
Fulvestrant	Hormone refractory prostate cancer	Treatment	PSA reduction ≥50%	II (completed)	NCT00476645		Stanford University
Fulvestrant	Advanced prostate cancer	Treatment	PSA objective response rate	II (completed)	NCT00244998		Roswell Park Cancer Institute

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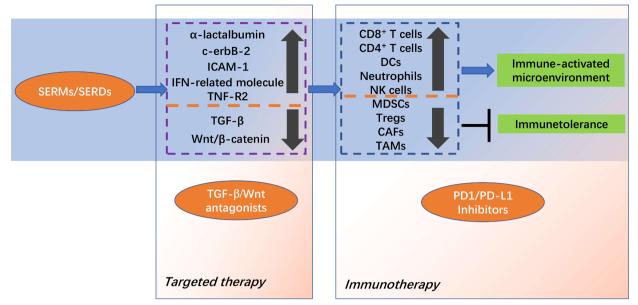


Figure 3 Schematic diagram of a novel immunotherapy-based comprehensive strategy combined with SERMs/SERDs, transforming growth factor- β (TGF- β) or Wnt antagonists for PC-targeting therapy. The SERMs/SERDs increase immune cell infiltrations, including natural killer (NK) cells, CD8 +T cells, CD4 +T cells, dendritic cells (DCs) and neutrophils, and decrease immunosuppressive cancer associated fibroblasts (ICAFs), tumor-associated macrophages (TAMs) and myeloid-derived suppressor cells (MDSCs) by altering the activity of downstream pathways and synergizing with other agents.

However, results of previous studies have demonstrated that in CD8 +T cells, ER β signaling increases the activation of T cell receptors and antitumor immunity via a phosphotyrosine switch. ER β activation promotes IL-1 β secretion and enhances neutrophil infiltration in TIM, further suppressing tumor progression and metastasis.⁵⁹ Therefore, the specific role of ER β and the phosphotyrosine switch in cancer development is complex, and further studies are required. Recent clinical data indicated that in PC tumor associated stromal cells, a high expression level of ERa was associated with elevated clinical failure-free survival and PC death-free survival, while a high expression of $ER\beta$ was associated with decreased biochemical failure-free survival.⁶⁰ These studies suggested that $ER\beta$ may mediate antitumor immunity and play an opposite role to ER α in the TIM.

SERMS ON THE TIM

Increased evidence based on experimental and clinical research has demonstrated the diverse immunomodulatory roles of SERMs and SERDs.⁶¹ Previous studies have explored how SERMs and SERDs act on the TIM. By inhibiting antitumor immunity, Tamoxifen caused a phenotypic shift of CD4 +T cells from Th1-targeting cancer to Th2-tolerating cancer.⁶¹ ⁶² Moreover, TGF- β production induced by Tamoxifen and Fulvestrant inhibited the cytotoxic effects of CD8 +T cells, and promoted CD4 +T cell polarization into Foxp3 +Tregs. Tamoxifen and fulvestrant evoked programmed death-ligand 1 (PD-L1) upregulation by blocking Er α , contributing to the cytotoxic T cell evasion.⁶³ Notably, SERMs and SERDs

mediated immunosuppressive effects and increased the chance of tumors escaping an immune response.

On the other hand, SERMs also enhance the anticancer immunity and cancer immunogenicity. Tamoxifen and Fulvestrant inhibit cancer via targeted immunotherapy, without impacting healthy tissue.⁶⁴ Tamoxifen increased the cytotoxicity of NK cells, leading to tumor cell lysis.⁶⁵ Toremifene bridges the immunological synapses between cancer and NK cells by upregulating the expression levels of intercellular adhesion molecule-1 (ICAM-1).⁶⁶ Tamoxifen abolished estrogen-induced cancer-immune tolerance though suppressing FasL and blocking chemokine (C-C motif) ligand 2(CCL-2)/CCL-5.67 68 Depletion of TAMs in combination with tamoxifen increased the inhibitory effects of tamoxifen on aggressive PC.³⁹ Fulvestrant reduced MDSCs and Tregs, and increased the infiltration of DCs, CD8 +and CD4+T cells in a xenograft tumor model. The effects of SERDs on TIM markedly improved the efficiency of PD-L1-targeted immunotherapy.⁶⁹ SERMs/SERDs-mediated anticancer immunity along with immune check-point blockers (ICBs) increases the potential of PD-L1 inhibition. Moreover, tamoxifen and toremifene were also found to inhibit INK activation and enhance activated T cells.⁷⁰

PHARMACODYNAMIC EVALUATION OF SERMS

Certain SERMs exert the same pharmacodynamic effects on both ER α and ER β , including 4-hydroxytamoxifen (IC50; 3.3nM), raloxifene (IC50; 2.9–5.7nM) and toremifene (IC50; 1000nM), while bazedeoxifene, lasofoxifene and SERD fulvestrant exhibit differential effects on each. Notably, bazedeoxifene exerts differential effects on ER α (IC50; 26 nM) and ER β (IC50; 99 nM); lasofoxifene exerts differential effects on ER α (IC50; 1.08 nM) and ER β (IC50; 4.41 nM) and fulvestrant exerts differential effects on ER α (IC50; 0.47 nM) and ER β (IC50; 3.8 nM).¹⁸ Based on these pharmacodynamic analyses, we concluded that SERMs exert inhibitory activities on both ER α and ER β isoforms. Moreover, 4-hydroxytamoxifen, toremifene and raloxifene exert the same inhibitory activities on both isoforms. Compared with the effects on ER β , bazedeoxifene, lasofoxifene and fulvestrant (SERD) exert increased inhibitory activities on ER α . Moreover, in PC cells, raloxifene induced apoptosis via ER β activation and ER α inhibition.⁷¹

HYPOTHESES AND RESULTS

- 1. In PC cells, ER α activation and ER β inactivation may lead to therapeutic resistance and EMT, respectively. A SERM that antagonizes ER α may reverse resistance, while an agonist of ER β activity may reverse EMT.
- 2. In the TIM, ER β activation and ER α inactivation will result in immunosuppression and poor prognosis, respectively. A SERM that antagonizes ER β may cause a shift from an immunosuppressed TIM to an immuneactivated TIM, while an agonist of ER α activity may improve prognosis (figure 2).
- 3. ER isoforms are expressed in different locations in PC. ERs exert different roles in PC cells and the TIM, for example, an ER α -dominant phenotype in PC cells contributes to therapeutic resistance, while an ER β -dominant phenotype in the TIM induces the suppression of immunotherapies.
- 4. Patients with PC may benefit from the use of SERMs, including raloxifene, in combination with anti-PD1/ PD-L1 checkpoint immunotherapy, or TGF- β or Wnt antagonists.
- 5. Uncovering the optimal activity profile of SERMs should include (1) engineering a novel SERM that is easily attached to or highly selective to PC; (2) designing highly selective SERMs for ER α and ER β ; and (3) developing a targeted delivery system to exert downstream effects.

DISCUSSION

ERs promote PC development/progression

ERs may promote PC development and progression, whereas exogenous estrogen is a treatment option currently used to inhibit the development of PC. Multiple signaling pathways are involved in the process. Both ER α and ER β activation promotes migration, invasion and anchorage-independent growth of PC-3 cells. Their activation upregulates β -catenin expression and contributes to cell proliferation. These effects are blocked by PKF 118–310, a compound disrupting the complex β -catenin/TCF/LEF. PKF 118–310 also inhibited the upregulation of ERs-induced VEGF A.⁷² ERs activate both the SRC and PI3K/AKT signaling pathways, enhancing the tumorigenic potential, cell proliferation, migration, invasion and tumor formation of PC.¹⁶ Moreover, ERs can also intersect with the HIF/NF-kB signaling pathway in PC.⁷³ ERs-mediated long non-coding RNAs, including NEAT1, H19, MALAT1 and HOTAIR to promote PC development and the acquisition of the CRPC phenotype.⁷⁴ The progressive emergence of the ER α combined with a series of genes that are regulated by ER α during CRPC progression, including progesterone receptor, PS2, TMPRSS2-ERG fusion and NEAT1 indicates that ER signaling can bypass the AR for tumor growth.¹⁴

Immunogenic potential of SERMs/SERDs in PC

PC is considered a cold tumor due to the limited number of tumor-associated antigens, a suppressive TIM and resistance to ICBs.⁷⁵ PC grows slowly compared with other types of cancers and becomes a potential target for immunotherapy. However, previous clinical trials associated with various types of immunotherapies achieved minimal effects in metastatic CRPC. These limitations included an immunosuppressive TIM leading to limited efficacy of immunotherapy. The TIM associated with PC decreases the number of tumorinfiltrating immune cells and increases the number of immunosuppressive cells, including TAM, Tregs and MDSCs. Moreover, a low tumor mutation burden, PD-L1 expression and less somatic mutations induce fewer responses to ICBs or PD-1/PD-L1 inhibitors. The upregulated expression of TGF- β is also a limitation, as blocking TGF-β combined with ICBs increases Th1 and CD8 T cells, and promotes the regression of metastatic CRPC.⁷⁶ Thus, shifting from an immunosuppressed TIM to an immune-activated TIM-based immunotherapy is key for the successful treatment of PC. Immunotherapies may also be combined with ADT, radiotherapy, chemotherapy and immunogenic agents, highlighting that personalized and combination therapy may exhibit potential for the treatment of PC.

As previously described, SERMs and SERDs have been used to enhance the immunogenicity of BC and improve the antitumor immunity. SERMs and SERDs effectively promote the expression of α-lactalbumin, c-erbB-2 and ICAM-1, which boost the cytotoxicity of NK cells, bridge and form the immunological synapses, reduce the number of MDSCs and Tregs, and increase the infiltration of DCs, and CD8 +and CD4+T cells.^{64-66 69} Tamoxifen and toremifene upregulate the expression levels of IFN-related genes and TNF-R2, which inhibit the activation of JNK and promote T cell proliferation.^{70 77} SERMs and SERDs abolish estrogeninduced tumor immunetolerance and improve the efficiency of anti-PD-L1 therapy.^{67 68} Numerous clinical trials evaluating the efficiency of ICBs in combination with SERMs and SERDs in BCs are ongoing.⁵⁴ These agents, in combination with immunotherapeutics, are the current focus of research for the successful treatment of PC. Moreover, SERMs and SERDs may enhance immunogenicity by improving the expression of immunotherapeutic targets; SERMs and SERDs may increase the infiltration of NK cells, neutrophils, DCs, CD8 + and CD4+T cells, and reduce the counts of MDSCs and Tregs; and SERMs and SERDs may upregulate the expression levels of ICAM-1, IFN and TNF-R2, and inhibit the activation of JNK to promote cellular immune responses. Moreover, SERMs and SERDs may abolish tumor immunetolerance in combination with anti-PD-L1 therapy and ICBs. Results of a previous study demonstrated that fulvestrant downregulated AR expression and diminished androgenic responses in PC LNCaP cells, by the direct repression of AR gene transcription.⁷⁸

CONCLUSION AND PERSPECTIVE

Androgen-targeted endocrine therapyand chemotherapy-based drug treatment models is not largely enough for impeding PC progression and could not bring overall benefits for patients. An increasing number of ER-targeted therapies and immunotherapies targeting PC and adverse effects followed by firstline therapies have entered clinical trials. A summary of 14 clinical trials on SERMs and SERDs focusing on PC prevention and treatment are summarized in table 1. A total of 192 clinical trials focusing on the use of immunotherapy for the treatment of PC were found in a keyword search on https://clinicaltrialsgov. According to our hypotheses, the ER signaling pathway may regulate the TIM, and immunotherapy-based strategies combined with selective SERMs may be the future direction for adjunctive therapy of PC. Further clinical trials based on immunotherapies and SERMs are required.

Changing the immune microenvironment of PC from cold to hot remains an ongoing problem. The majority of clinical trials that aim to develop a single immunosuppressive target result in limited therapy efficacy.⁷⁹ According to the stromal status of the TIM, PC may be divided into immune-activated and immunosuppressive subtypes. Moreover, the WNT/TGF- β signaling pathway and the extracellular matrix were highly concentrated in the immunosuppressive group. Therefore, immunosuppressive patients may benefit from ICB therapy in combination with TGF- β inhibitors.⁸⁰ The present review indicated that a combination of immunotherapy-based strategies with SERMs, TGF-β or Wnt antagonists requires further investigation for the development of future treatment options for PC (figure 3).

Contributors DT wrote and reviewed the MS.

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REFERENCES

- 1 Tong D. The role of JMJD6/U2AF65/AR-V7 axis in castrationresistant prostate cancer progression. *Cancer Cell Int* 2021;21:45.
- 2 Tong D. Unravelling the molecular mechanisms of prostate cancer evolution from genotype to phenotype. *Crit Rev Oncol Hematol* 2021;163:103370.
- 3 Liu Q, Tong D, Liu G, et al. Metformin reverses prostate cancer resistance to enzalutamide by targeting TGF-β1/STAT3 axisregulated EMT. Cell Death Dis 2017;8:e3007.
- 4 Sanches BDA, Maldarine JS, Vilamaior PSL, et al. Stromal cell interplay in prostate development, physiology, and pathological conditions. *Prostate* 2021;81:926–37.
- 5 Stultz J, Fong L. How to turn up the heat on the cold immune microenvironment of metastatic prostate cancer. *Prostate Cancer Prostatic Dis* 2021;24:697–717.
- 6 Li Y, Zhang B, Xiang L, et al. TGF-β causes docetaxel resistance in prostate cancer via the induction of Bcl-2 by acetylated KLF5 and protein stabilization. *Theranostics* 2020;10:7656–70.
- 7 Paller C, Pu H, Begemann DE, et al. TGF-β receptor I inhibitor enhances response to enzalutamide in a pre-clinical model of advanced prostate cancer. Prostate 2019;79:31–43.
- 8 Wang L, Dehm SM, Hillman DW, et al. A prospective genomewide study of prostate cancer metastases reveals association of wnt pathway activation and increased cell cycle proliferation with primary resistance to abiraterone acetate-prednisone. *Ann Oncol* 2018;29:352–60.
- 9 Bailar JC, Byar DP, The veterans administration cooperative urological research group. Estrogen treatment for cancer of the prostate.Early results with 3 doses of diethylstilbestrol and placebo. *Cancer* 1970;26:257–61.
- 10 Cavalieri EL, Devanesan P, Bosland MC, et al. Catechol estrogen metabolites and conjugates in different regions of the prostate of noble rats treated with 4-hydroxyestradiol: implications for estrogen-induced initiation of prostate cancer. *Carcinogenesis* 2002;23:329–33.
- 11 Setlur SR, Mertz KD, Hoshida Y, et al. Estrogen-Dependent signaling in a molecularly distinct subclass of aggressive prostate cancer. J Natl Cancer Inst 2008;100:815–25.
- 12 Dobbs RW, Malhotra NR, Greenwald DT, et al. Estrogens and prostate cancer. *Prostate Cancer Prostatic Dis* 2019;22:185–94.
- 13 Ozten N, Vega K, Liehr J, et al. Role of estrogen in androgen-induced prostate carcinogenesis in nbl rats. Horm Cancer 2019;10:77–88.
- 14 Bonkhoff H. Estrogen receptor signaling in prostate cancer: implications for carcinogenesis and tumor progression. *Prostate* 2018;78:2–10.
- 15 Pisolato R, Lombardi APG, Vicente CM, et al. Expression and regulation of the estrogen receptors in PC-3 human prostate cancer cells. Steroids 2016;107:74–86.
- 16 Lombardi APG, Cavalheiro RP, Porto CS, et al. Estrogen receptor signaling pathways involved in invasion and colony formation of androgen-independent prostate cancer cells PC-3. *Int J Mol Sci* 2021;22. doi:10.3390/ijms22031153. [Epub ahead of print: 25 Jan 2021].
- 17 Aronica SM, Katzenellenbogen BS. Stimulation of estrogen receptormediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-I. *Mol Endocrinol* 1993;7:743–52.
- 18 Lafront C, Germain L, Weidmann C, et al. A systematic study of the impact of estrogens and selective estrogen receptor modulators on prostate cancer cell proliferation. Sci Rep 2020;10:4024.
- 19 Kowalska K, Habrowska-Górczyńska D, Urbanek K, et al. Estrogen receptor α is crucial in Zearalenone-Induced invasion and migration of prostate cancer cells. *Toxins* 2018;10:98.

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- 20 Ehara H, Koji T, Deguchi T, et al. Expression of estrogen receptor in diseased human prostate assessed by non-radioactive in situ hybridization and immunohistochemistry. *Prostate* 1995;27:304–13.
- 21 Bonkhoff H, Fixemer T, Hunsicker I, et al. Estrogen receptor expression in prostate cancer and premalignant prostatic lesions. Am J Pathol 1999;155:641–7.
- 22 Takizawa I, Lawrence MG, Balanathan P, et al. Estrogen receptor alpha drives proliferation in PTEN-deficient prostate carcinoma by stimulating survival signaling, Myc expression and altering glucose sensitivity. *Oncotarget* 2015;6:604–16.
- 23 Chakravarty D, Sboner A, Nair SS, et al. The oestrogen receptor alpha-regulated IncRNA NEAT1 is a critical modulator of prostate cancer. *Nat Commun* 2014;5:5383.
- 24 Choi JH, Choi SM, Lee SW, et al. Expression patterns of ERα, ERβ, Ar, SIRT1, SIRT2, and SIRT3 in prostate cancer tissue and normal prostate tissue. Anticancer Res 2021;41:1377–86.
- 25 Lai JS, Brown LG, True LD, *et al*. Metastases of prostate cancer express estrogen receptor-beta. *Urology* 2004;64:814–20.
- 26 Steiner MS, Raghow S. Antiestrogens and selective estrogen receptor modulators reduce prostate cancer risk. *World J Urol* 2003;21:31–6.
- 27 Zellweger T, Stürm S, Rey S, *et al.* Estrogen receptor β expression and androgen receptor phosphorylation correlate with a poor clinical outcome in hormone-naive prostate cancer and are elevated in castration-resistant disease. *Endocr Relat Cancer* 2013;20:403–13.
- 28 Leung Y-K, Lam H-M, Wu S, et al. Estrogen receptor β2 and β5 are associated with poor prognosis in prostate cancer, and promote cancer cell migration and invasion. *Endocr Relat Cancer* 2010;17:675–89.
- 29 Mak P, Li J, Samanta S, *et al*. Prostate tumorigenesis induced by PTEN deletion involves estrogen receptor β repression. *Cell Rep* 2015;10:1982–91.
- 30 Wu W-F, Maneix L, Insunza J, et al. Estrogen receptor β, a regulator of androgen receptor signaling in the mouse ventral prostate. Proc Natl Acad Sci U S A 2017;114:E3816–22.
- 31 McPherson SJ, Hussain S, Balanathan P, et al. Estrogen receptorbeta activated apoptosis in benign hyperplasia and cancer of the prostate is androgen independent and TNFalpha mediated. Proc Natl Acad Sci U S A 2010;107:3123–8.
- 32 Horvath LG, Henshall SM, Lee CS, et al. Frequent loss of estrogen receptor-beta expression in prostate cancer. Cancer Res 2001;61:5331–5.
- 33 Bonkhoff H, Berges R. The evolving role of oestrogens and their receptors in the development and progression of prostate cancer. *Eur Urol* 2009;55:533–42.
- 34 Shaw GL, Whitaker H, Corcoran M, et al. The early effects of rapid androgen deprivation on human prostate cancer. Eur Urol 2016;70:214–8.
- 35 Bhatnagar S, Soni A, Kaushik S, et al. Nonsteroidal estrogen receptor isoform-selective biphenyls. Chem Biol Drug Des 2018;91:620–30.
- 36 Traboulsi T, El Ezzy M, Gleason JL, et al. Antiestrogens: structureactivity relationships and use in breast cancer treatment. J Mol Endocrinol 2017;58:R15–31.
- 37 Seeliger H, Pozios I, Assmann G, et al. Expression of estrogen receptor beta correlates with adverse prognosis in resected pancreatic adenocarcinoma. BMC Cancer 2018;18:1049.
- 38 Tomao S, Romiti A, Massidda B, et al. A phase II study of gemcitabine and tamoxifen in advanced pancreatic cancer. Anticancer Res 2002;22:2361–4.
- 39 Semenas J, Wang T, Sajid Syed Khaja A, et al. Targeted inhibition of ERα signaling and PIP5K1α/Akt pathways in castration-resistant prostate cancer. *Mol Oncol* 2021;15:968–86.
- 40 Ho TH, Nunez-Nateras R, Hou Y-X, *et al.* A study of combination Bicalutamide and raloxifene for patients with castration-resistant prostate cancer. *Clin Genitourin Cancer* 2017;15:196–202.
- 41 Segovia-Mendoza M, Morales-Montor J. Immune tumor microenvironment in breast cancer and the participation of estrogen and its receptors in cancer physiopathology. *Front Immunol* 2019:10:348.
- 42 Vegeto E, Ghisletti S, Meda C, et al. Regulation of the lipopolysaccharide signal transduction pathway by 17beta-estradiol in macrophage cells. J Steroid Biochem Mol Biol 2004;91:59–66.
- Kramer PR, Kramer SF, Guan G. 17 beta-estradiol regulates cytokine release through modulation of CD16 expression in monocytes and monocyte-derived macrophages. *Arthritis Rheum* 2004;50:1967–75.
 Rogers A, Eastell R. The effect of 17beta-estradiol on production of
- 44 Rogers A, Eastell R. The effect of 17beta-estradiol on production of cytokines in cultures of peripheral blood. *Bone* 2001;29:30–4.
- 45 Nilsson N, Carlsten H. Estrogen induces suppression of natural killer cell cytotoxicity and augmentation of polyclonal B cell activation. *Cell Immunol* 1994;158:131–9.

- 46 Jiang X, Ellison SJ, Alarid ET, et al. Interplay between the levels of estrogen and estrogen receptor controls the level of the granzyme inhibitor, proteinase inhibitor 9 and susceptibility to immune surveillance by natural killer cells. Oncogene 2007;26:4106–14.
- 47 Band AM, Laiho M. Crosstalk of TGF-β and estrogen receptor signaling in breast cancer. *J Mammary Gland Biol Neoplasia* 2011;16:109–15.
- 48 Ito I, Hanyu A, Wayama M, et al. Estrogen inhibits transforming growth factor beta signaling by promoting Smad2/3 degradation. J Biol Chem 2010;285:14747–55.
- 49 Ren Y, Wu L, Frost AR, et al. Dual effects of TGF-beta on ERalphamediated estrogenic transcriptional activity in breast cancer. Mol Cancer 2009;8:111.
- 50 Goto N, Hiyoshi H, Ito I, *et al.* Estrogen and antiestrogens alter breast cancer invasiveness by modulating the transforming growth factor-β signaling pathway. *Cancer Sci* 2011;102:1501–8.
- 51 Lee H-R, Hwang K-A, Choi K-C. The estrogen receptor signaling pathway activated by phthalates is linked with transforming growth factor-β in the progression of LNCaP prostate cancer models. *Int J* Oncol 2014;45:595–602.
- 52 Yu L, Wang C-Y, Shi J, et al. Estrogens promote invasion of prostate cancer cells in a paracrine manner through up-regulation of matrix metalloproteinase 2 in prostatic stromal cells. *Endocrinology* 2011;152:773–81.
- 53 Straub RH. The complex role of estrogens in inflammation. *Endocr Rev* 2007;28:521–74.
- 54 Huang H, Zhou J, Chen H, et al. The immunomodulatory effects of endocrine therapy in breast cancer. J Exp Clin Cancer Res 2021;40:19.
- 55 Guo D, Liu X, Zeng C, et al. Estrogen receptor β activation ameliorates DSS-induced chronic colitis by inhibiting inflammation and promoting Treg differentiation. Int Immunopharmacol 2019;77:105971.
- 56 Jarnicki AG, Lysaght J, Todryk S, et al. Suppression of antitumor immunity by IL-10 and TGF-beta-producing T cells infiltrating the growing tumor: influence of tumor environment on the induction of CD4⁺ and CD8+ regulatory T cells. J Immunol 2006;177:896–904.
- 57 Wang T, Jin J, Qian C, et al. Estrogen/ER in anti-tumor immunity regulation to tumor cell and tumor microenvironment. Cancer Cell Int 2021;21:295.
- 58 Xiong Y, Zhong Q, Palmer T, *et al.* Estradiol resolves pneumonia via ERβ in regulatory T cells. *JCl Insight* 2021;6. doi:10.1172/jci. insight.133251. [Epub ahead of print: 08 02 2021].
- 59 Zhao L, Huang S, Mei S, et al. Pharmacological activation of estrogen receptor beta augments innate immunity to suppress cancer metastasis. Proc Natl Acad Sci U S A 2018;115:E3673–81.
- 60 Grindstad T, Skjefstad K, Andersen S, et al. Estrogen receptors α and β and aromatase as independent predictors for prostate cancer outcome. Sci Rep 2016;6:33114.
- 61 Behjati S, Frank M. The effects of tamoxifen on immunity. *Curr Med Chem* 2009;16:3076–80.
- 62 Bebo BF, Dehghani B, Foster S, *et al.* Treatment with selective estrogen receptor modulators regulates myelin specific T-cells and suppresses experimental autoimmune encephalomyelitis. *Glia* 2009;57:777–90.
- 63 Hühn D, Martí-Rodrigo P, Mouron S, et al. Prolonged estrogen deprivation triggers a broad immunosuppressive phenotype in breast cancer cells. *Mol Oncol* 2022;16:148–65.
- 64 Jaini R, Loya MG, Eng C. Immunotherapeutic target expression on breast tumors can be amplified by hormone receptor antagonism: a novel strategy for enhancing efficacy of targeted immunotherapy. Oncotarget 2017;8:32536–49.
- 65 Richards JO, Albers AJ, Smith TS, et al. Nk cell-mediated antibodydependent cellular cytotoxicity is enhanced by tamoxifen in HER2/ neu non-amplified, but not HER2/neu-amplified, breast cancer cells. Cancer Immunol Immunother 2016;65:1325–35.
- 66 Komi J, Lassila O. Toremifene increases the expression of intercellular adhesion molecule-1 (ICAM-1) on MCF-7 breast cancer cells and Jurkat cells. *Scand J Immunol* 2000;51:73–8.
- 67 Mor G, Kohen F, Garcia-Velasco J, et al. Regulation of Fas ligand expression in breast cancer cells by estrogen: functional differences between estradiol and tamoxifen. J Steroid Biochem Mol Biol 2000;73:185–94.
- 68 Svensson S, Abrahamsson A, Rodriguez GV, *et al.* Ccl2 and CCL5 are novel therapeutic targets for estrogen-dependent breast cancer. *Clin Cancer Res* 2015;21:3794–805.
- 69 Márquez-Garbán DC, Deng G, Comin-Anduix B, *et al.* Antiestrogens in combination with immune checkpoint inhibitors in breast cancer immunotherapy. *J Steroid Biochem Mol Biol* 2019;193:105415.

- 70 Komi J, Lassila O. Antioestrogens enhance tumour necrosis factor receptor 2 (TNF-R2) expression and TNF-R2-mediated proliferation in activated T cells. *Scand J Immunol* 1998;48:254–60.
- 71 Rossi V, Bellastella G, De Rosa C, *et al*. Raloxifene induces cell death and inhibits proliferation through multiple signaling pathways in prostate cancer cells expressing different levels of estrogen receptor α and β . *J Cell Physiol* 2011;226:1334–9.
- 72 Lombardi APG, Vicente CM, Porto CS. Estrogen receptors promote migration, invasion and colony formation of the androgenindependent prostate cancer cells PC-3 through β-catenin pathway. *Front Endocrinol* 2020;11:184.
- 73 Russo M, Ravenna L, Pellegrini L, et al. Hypoxia and inflammation in prostate cancer progression. cross-talk with androgen and estrogen receptors and cancer stem cells. *Endocr Metab Immune Disord Drug Targets* 2017;16:235–48.
- 74 Nanni S, Bacci L, Aiello A, et al. Signaling through estrogen receptors modulates long non-coding RNAs in prostate cancer. Mol Cell Endocrinol 2020;511:110864.

- 75 Cha H-R, Lee JH, Ponnazhagan S. Revisiting immunotherapy: a focus on prostate cancer. *Cancer Res* 2020;80:1615–23.
- 76 Jiao S, Subudhi SK, Aparicio A, et al. Differences in tumor microenvironment dictate T helper lineage polarization and response to immune checkpoint therapy. *Cell* 2019;179:1177–90.
- 77 Schild-Hay LJ, Leil TA, Divi RL, et al. Tamoxifen induces expression of immune response-related genes in cultured normal human mammary epithelial cells. *Cancer Res* 2009;69:1150–5.
- 78 Bhattacharyya RS, Krishnan AV, Swami S, et al. Fulvestrant (ICI 182,780) down-regulates androgen receptor expression and diminishes androgenic responses in LNCaP human prostate cancer cells. *Mol Cancer Ther* 2006;5:1539–49.
- 79 Stultz J, Fong L. How to turn up the heat on the cold immune microenvironment of metastatic prostate cancer. *Prostate Cancer Prostatic Dis* 2021;24:697–717.
- 80 Meng J, Zhou Y, Lu X, et al. Immune response drives outcomes in prostate cancer: implications for immunotherapy. *Mol Oncol* 2021;15:1358-1375.