Check for updates

OPEN ACCESS

EDITED BY Anurag Kumar Singh, Alabama State University, United States

REVIEWED BY Prabhat Suman, Central University of Punjab, India Vivek K. Chaturvedi, Banaras Hindu University, India Ihtisham Bukhari, Henan Provincial People's Hospital, China Arun Chhokar, University of Delhi, India

*CORRESPONDENCE Md. Mominur Rhaman, mominur.ph@gmail.com Rohit Sharma, rohitsharma@bhu.ac.in

ronnandinna (a bina. ac. in

SPECIALTY SECTION This article was submitted to Molecular and Cellular Pathology, a section of the journal Frontiers in Cell and Developmental Biology

RECEIVED 08 July 2022 ACCEPTED 08 August 2022 PUBLISHED 02 September 2022

CITATION

Rhaman MM, Islam MR, Akash S, Mim M, Noor alam M, Nepovimova E, Valis M, Kuca K and Sharma R (2022), Exploring the role of nanomedicines for the therapeutic approach of central nervous system dysfunction: At a glance. *Front. Cell Dev. Biol.* 10:989471. doi: 10.3389/fcell.2022.989471

COPYRIGHT

© 2022 Rhaman, Islam, Akash, Mim, Noor alam, Nepovimova, Valis, Kuca and Sharma. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Exploring the role of nanomedicines for the therapeutic approach of central nervous system dysfunction: At a glance

Md. Mominur Rhaman¹*, Md. Rezaul Islam¹, Shopnil Akash¹, Mobasharah Mim¹, Md. Noor alam¹, Eugenie Nepovimova², Martin Valis³, Kamil Kuca^{2,4} and Rohit Sharma⁵*

¹Department of Pharmacy, Faculty of Allied Health Sciences, Daffodil International University, Dhaka, Bangladesh, ²Department of Chemistry, Faculty of Science, University of Hradec Králové, Hradec Králové, Czech Republic, ³Department of Neurology, Charles University in Prague, Faculty of Medicine in Hradec Králové and University Hospital, Hradec Králové, Czech Republic, ⁴Andalusian Research Institute in Data Science and Computational Intelligence (DaSCI), University of Granada, Granada, Spain, ⁵Department of Rasa Shastra and Bhaishajya Kalpana, Faculty of Ayurveda, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India

In recent decades, research scientists, molecular biologists, and pharmacologists have placed a strong emphasis on cutting-edge nanostructured materials technologies to increase medicine delivery to the central nervous system (CNS). The application of nanoscience for the treatment of neurodegenerative diseases (NDs) such as Alzheimer's disease (AD), Parkinson's disease (PD), multiple sclerosis (MS), Huntington's disease (HD), brain cancer, and hemorrhage has the potential to transform care. Multiple studies have indicated that nanomaterials can be used to successfully treat CNS disorders in the case of neurodegeneration. Nanomedicine development for the cure of degenerative and inflammatory diseases of the nervous system is critical. Nanoparticles may act as a drug transporter that can precisely target sick brain sub-regions, boosting therapy success. It is important to develop strategies that can penetrate the blood-brain barrier (BBB) and improve the effectiveness of medications. One of the probable tactics is the use of different nanoscale materials. These nano-based pharmaceuticals offer low toxicity, tailored delivery, high stability, and drug loading capacity. They may also increase therapeutic effectiveness. A few examples of the many different kinds and forms of nanomaterials that have been widely employed to treat neurological diseases include quantum dots, dendrimers, metallic nanoparticles, polymeric nanoparticles, carbon nanotubes, liposomes, and micelles. These unique qualities, including sensitivity, selectivity, and ability to traverse the BBB when employed in nano-sized particles, make these nanoparticles useful for imaging studies and treatment of NDs. Multifunctional nanoparticles carrying pharmacological medications serve two purposes: they improve medication distribution while also enabling cell dynamics imaging and pharmacokinetic study. However, because of the potential for wide-ranging clinical implications, safety concerns persist, limiting any potential for translation. The evidence for using nanotechnology to create drug delivery systems that could pass across the BBB and deliver therapeutic chemicals to CNS was examined in this study.

KEYWORDS

neurodegenerative diseases, blood-brain barrier, drug delivery, nanotechnology, nanomedicine and nanocarrier

Introduction

The World Health Organization categorizes central nervous system (CNS) malignancies based on their effective potential, likelihood of spread, and clinical prognosis (Louis et al., 2016; Singh et al., 2021a). Because the brain accounts for nearly all malignant CNS tumors, the focus will now be on the lookout for malignant brain tumors. Malignant brain tumors are a type of primary brain tumors; for instance, almost 2% of all cancers and are emblematically generated from glial cells (hence the name gliomas) (Miranda et al., 2017). Brain metastases affect 10%-30% of all cancer patients, with 70%-80% developing numerous injuries. The high rate of intracranial metastases is due to the fact that while new chemotherapeutic drugs have transformed prognosis for many forms of cancer, they have been able to deter neoplasms from spreading into the brain due to their limited blood-brain barrier (BBB) penetration. The invention of therapeutic approaches that can prevent the BBB and also increase their efficacy is required. To address this issue, researchers are actively developing therapeutic drug delivery techniques (Parveen et al., 2012a; Zhang et al., 2015; Singh et al., 2021b). Different approaches to solve this issue have been developed. The nano-based method is at the heart of recent advancements in therapeutic drug delivery (Sharma et al., 2019; Song et al., 2021). Different kinds of manufactured nanomaterial and nanoparticles with a size of 1-100 nm in at least one dimension are used in nanotechnology (Farokhzad and Langer, 2009; Sharma and Bhargava, 2013; Khan et al., 2016). As numerous nanoparticles have been used in brain studies and research, including quantum dots (QDs), polymeric nanoparticles, micelles, and metallic nanoparticles, nanotechnology and nanomaterials open new pathways in biomedical science (Chenthamara et al., 2019; Alshamrani, 2022; Waris et al., 2022). Because of their small size and capacity to interact with biological systems at the molecular level, these nanoscale materials also have special properties such as a high surfaceto-volume ratio that can be mono or diverse with surface modification and also high stability (Umut, 2013; Natarajan et al., 2019; Singh et al., 2020). This is a significant impediment to brain delivery (Au et al., 2017). In malignant brain tumors, the mainstay of care is maximal surgical resection (if possible), followed by radiation, chemotherapy, and symptomatic treatment (Alifieris and Trafalis, 2015). Nonetheless, since reappearance within the next few times is

typical, cancerous brains tumors are prone to relapse (brain metastases have quite a median survival of 8 months, even aggressive primary brain cancers have median survival of 14.2 months) and remain an unmet clinical challenge (Stupp et al., 2005; Au et al., 2017). Indeed, given the extraordinarily varied character of malignant brain tumors, it is no longer reasonable to expect a single treatment to be effective in all individuals. At most, each medication would be helpful only for specific target populations and disease stages. In this context, theragnostic, which is defined as the simultaneous delivery of imaging and therapeutic substances (Orive et al., 2010), has a lot of potential for the treatment of malignant brain tumors at various stages. A number of promising medications for the treatment of neurological diseases have been identified (Alguacil et al., 2003; Whiting, 2003; Langmead et al., 2008). Although these medications have shown therapeutic efficacy, the existence of the 1) BBB and the 2) blood-cerebrospinal fluid barrier (BCFB) continues to restrict and limit their effectiveness (Jain, 2007; Wong et al., 2012; Dominguez et al., 2014). A number of scientists are trying to develop an associative strategy using nanotechnology in order to get the better at these significant challenges toward the field of CNS therapy. In the future, nanoparticles and their complex mixture including therapeutic substances may be considered as an effective tool in brain medication delivery for the development of better medicines (Halberstadt et al., 2006; Jain, 2007; Singh et al., 2019a; Singh et al., 2019b). For the treatment of CNS diseases, nanoengineered materials are important and helpful for a number of reasons. The materials can first and foremost penetrate the BBB, which is a frequent barrier to CNS-targeted medicines (Srikanth and Kessler, 2012; Ai et al., 2016). In addition, nanomaterials can be created to interact with specific cellular subsets or chemicals, allowing for more targeted treatment. Also, the nanomaterial's incorporation of enzyme cleavage sequences permits modulation of activity in response to biological stimuli, such as pH-sensitive modification or cation-triggered self-assembly (Johnstone et al., 2016; Karamanos et al., 2018; Rai et al., 2021). For either transplanted cells, nanofibers endogenous or and nanoscaffolds can offer trophic support in addition to structural support. The fact that nanoengineered materials can include many properties into their structures to simultaneously provide targeting, bioactivity, gene transport, and imaging capabilities in a single material is significant (Srikanth and Kessler, 2012; Liu J. et al., 2015; Bhatia, 2016). We discussed the therapeutic significance of nanomedicine in CNS



dysfunction. In this review, we give an overview of the nanotechnologies that have been studied in relation to neurological disease, talk about the evidence for effectiveness and toxicity of nanomaterials in particular CNS disorders, and emphasize the potential for clinical application of nanotechnology.

Blood-brain barrier

The brain is thought to be the body's most safeguarded organ. The skull, peripheral nerves, cerebrospinal fluid, and BBB are all protective shields for the central nervous system. Many of these elements have the capacity to prevent the brain against internal and external traumas as well as disease prevention (Engelhardt and Sorokin, 2009; Nair et al., 2018). These protective barriers, however, reduces of entry therapeutic agents to the central nervous system in diseased states. The BBB is a physiological border made up of firmly bound cerebral vascular wall, hepatocytes, astrocytes, and basal membranes, with occasional endocytosis and transcytosis (Vorbrodt and Dobrogowska, 2003; Yazdani et al., 2019). Most medicines are rapidly effluxed due to the high expression of P glycoproteins on brain vascular endothelium (Andersson et al., 2002; Miller, 2010). The BBB blocks practically all polymers and a major proportion of tiny molecules (especially chemotherapy drugs) from reaching the CNS, especially the brain, because of relatively narrow openings in such connecting cells (Domínguez et al., 2013; Karim et al., 2016; Furtado et al., 2018). As a result, the BBB is blamed for the inadequacy of most treatment options. Epithelial lymphocytes and membranes structure such as vascular endothelium and

mucosal membranes have the capacity to transfer materials into the CNS because of their lipid-like composition (Figure 1) (Alam et al., 2010; Yu et al., 2016).

Drug targeting

Researchers are focusing on specific areas inside the target cell to help drugs accumulate at the target spot and reduce side effects. Most nanoparticles created to date have failed to deliver drug material to a particular sub-cellular target organelle. The sub-cellular localization of nano-particulate therapeutic compounds administrate applications is still under progress (D'Souza and Weissig, 2010). Organelle-specific targeting sites, such as TAT CPP (cellular permeating peptides), energy metabolism targeted identification, nanomaterial surfaces, target-specific cell organelles, and nuclear clustering signals are all used to their full capability for drug delivery (Flierl et al., 2003; De la Fuente and Berry, 2005; Kang et al., 2010). As a result, sub-cellular targeting could dramatically minimize the amount of therapeutic molecule required.

Endosomal targeting

Nanotherapeutics might be given to lysosomes or endosomes by receptor-specific endocytosis or by engaging to the target site on the endosomal cellular region; moderate lipid membranes, ferritin, and ventricular endothelium proliferation considerations are examples of these substances. The endosomes are also responsible for a fundamental function in

| Nanoparticles | Antigens | Shape | Size (nm) | Diseases | References |
|------------------------|---|------------------------------|--|---|--|
| Organic | | | | | |
| Polymeric | Hepatitis B surface antigen | Spherical | 474-940 | HBV | Thomas et al. (2011) |
| | VMP001 | Spherical | 290 | Malaria | Aikins et al. (2017) |
| Liposome | Hemagglutinin of IAV and IBVa | Spherical | 50-400 | IAV and IBV | (Glück et al., 1994; Patois et al., 2012) |
| | Membrane-proximal external region (MPER) peptide | Spherical | 150 | HIV | Hanson et al. (2015) |
| Virus-like particle | HPV16 L1 capsomeresa | Pentameric | 10 | HPV | Hassett et al. (2015) |
| Inorganic | | | | | |
| Gold | West Nile virus envelope protein | Spherical, rod, and cubic | 20-40 (spherical) 40×10 (rod) $40 \times 40 \times 40$ (cubic) | West Nile virus | Niikura et al. (2013) |
| Iron Oxide | M. tb fusion protein | Spherical | <20 | M. tb | Pusic et al. (2013) |
| | Mannose and HBsAg | Spherical | 60 | HBV | Rezaei et al. (2019) |
| Mesoporous Silica | Soluble worm antigenic preparation antigen | Spherical | 39 | Schistosoma mansoni | de Pádua Oliveira et al. (2016) |
| | Porcine circovirus type 2 opening reading frames (PCV2-ORF2) proteins | Spherical | 200 | Post-weaning multisystemic wasting syndrome | Guo et al. (2012) |
| | | | | | |

TABLE 1 Organic and inorganic vaccine nanoparticles against infectious diseases.

nanoparticle preservation by delaying depreciation so that they can continue to function in the cytoplasm (Sakhrani and Padh, 2013).

Endoplasmic reticulum-specified targeting

Golgi bodies are secretory cell organelles. They engage with endoplasmic reticulum (ER), a folded membrane network that extends from the nuclear membrane throughout the cytoplasm. The ER is associated within Ca²⁺ stockpiling, Ca²⁺ signaling, apoptosis regulation, and facilitating the folding of secretory proteins. Golgi bodies use a variety of enzymes to modify newly generated proteins after they have been translated. mTOR (mammals or mechanical mediator of starvation) is a proteolytic enzyme that plays a role in cell signaling that is mostly found in the ER and Golgi bodies. Cellular differentiation modulation, polypeptide hydrolysis, PCR amplification, protein interactions, phagocytosis, and cell function are all maintained by it, among other things (Lu et al., 2018; Sanati et al., 2019).

Mitochondrial targeting

A mitochondrion is known as a cell's powerhouse because it supplies ATP (Adenosine Triphosphate) for energy generation through oxidative phosphorylation process. It also removes ROS in a variety of ways. ROS are created as by-products of ATP synthesis, which can lead to mitochondrial malfunction. In the case of AD, this could interact with the peptides, causing amyloid buildup in the brain. The mitochondria have certain antioxidants, such as MitoQ and SS31, which are used to treat oxidative stress and synaptic dysfunction in AD (McManus et al., 2011).

Nuclear targeting

The nucleus, cell's control center, holds genetic material, making it vital because it must regulate expression of gene for protein creation, which is specifically dispersed within the nucleus and cytoplasm. Nuclear pore complexes (NPC) are pores in the bilayer membrane that divide the nucleus from the cytoplasm that allow for this translocation. NPC-targeting nanoparticles can be delivered into the nucleus (Xia et al., 2016; Popli et al., 2018).

Organic nanoparticle

Due to their biocompatibility, biodegradability, and overall non-toxicity, a broad range of distinct organic nanoparticles (Table 1) have been explored as vaccination platforms (Poon and Patel, 2020). In comparison to other nanoparticle platforms, organic nanoparticles have many benefits, such as the ability to self-assemble antigens and adjuvants under physiologically mild conditions, as well as chemical uniqueness to accommodate various methodologies, mixtures, dimensions, shapes, and surface modification (Poon et al., 2018; Poon and Patel, 2020; Zhang et al., 2021).

| Therapeutic class | Pharmacological Use | Limitation/Possible side effect | Examples | References |
|-------------------|--|---|--|--|
| Antidepressants | CNS stimulants; anticholinergic | Tricyclics may promote dry mouth, impaired vision, tachycardia, and cardiac arrythmias | Monoamine oxidase inhibitors and tricyclic antidepressants | (Walsh, 1979; Livingston and Livingston, 1996; Youdim et al., 2006) |
| Antipsychotics | Relieve anxiety and thought disturbances | These medications have the potential to induce drowsiness, hypothermia, hypotension, and lowering in seizure threshold. | Butyrophenones and phenothiazines | (Janssen, 1965; Martin et al., 1992) |
| Antiemetics | Relieve nausea and vomiting | Antihistamines are the only treatments that are recommended for divers to use in order to prevent motion sickness; nevertheless, the sleepiness generated by these medications might induce a reduction in cognitive ability. | Anticholinergics and antihistamines | Williams et al. (1988) |
| Anxiolytics | Relieve anxiety; depress CNS | Anxiolytic medications often produce sleepiness, lethargy, disorientation, and hypotension, all of which have the potential to be catastrophic in the water. | Benzodiazepines | (Groner-Strauss and Strauss, 1976; Greenblatt et al., 1983; Stewart, 2005) |
| CNS Stimulants | ncrease alertness; inhibit fatigue; suppress appetite; mood elevation | These may bring on symptoms such as exhilaration, increased perspiration, anxiety, and panic attacks. | Amphetamines | (Cheshire et al., 2001; Fleckenstein et al., 2007; Carvalho et al., 2012) |
| Hypnotics | Depress CNS and induce sleep | It may produce drowsiness, asthenia (weakness), headache, and aeuromuscular and skeletal weakness | Barbiturates | (Vermeeren, 2004; Charney et al., 2006) |

TABLE 2 Synthetic drugs acting on the CNS and their possible limitation.

Inorganic nanomaterials

Inorganic nanoparticles (Table 1), also known as INPs, have been the subject of research over the period of the several years for a broad range of different commercial applications. INPs have been put to use in the area of biomedicine for the goals of diagnosis as well as treatment (Lohse and Murphy, 2012; Giner-Casares et al., 2016). For instance, gold nanoparticles (AuNPs) have been the subject of a deep investigation along with their biomedical applications and relieve with which their particle sizes and shape can be controlled. Their structure can take on a variety of forms, including spheres, nanorods, and cubes, among others (Gurunathan et al., 2014; Singh et al., 2016). In addition, the surface chemistry of AuNPs is able to be readily changed by combining with a wide variety of polymers, antibodies, smallmolecule therapies, and molecular diagnostics (Lee et al., 2010; Zhang et al., 2020). This part will highlight new advancements in Inorganic and organic nanoparticle vaccine delivery platforms, such as, gold, mesoporous silica, iron oxide, polymeric nanoparticles, liposomes, and virus-like particles (VLPs).

Current FDA approved synthetic drugs for CNS dysfunction

The central nervous system, which comprises nerves located in both the brain and the spinal cord, is the part of our body that processes and regulates the vast majority of our biological activities (Elenkov et al., 2000; Rossini et al., 2015). Medicines that have an effect on the central nervous system or may trigger CNS are known as central nervous system medications (Misra et al., 2003). A variety of medications, such as sedatives, antidepressants, and anesthetics, are all examples of medications that may affect the central nervous system (Charney et al., 2006; Henschel et al., 2008; Attri et al., 2012). Details are given in the table (Table 2).

Modeling techniques for CNS disorders

Sickness modeling in the lab is essential for improving comprehension of pathophysiology and evaluation of efficiency of innovative pharmacological strategies. Cancerous cells can come from a variety of places: cancer tissues or tumor cell culture generated in the laboratories, which are mainly derived from transgenic mice or from patients (Liu F. et al., 2015; Humpel, 2015). Brain or systemic infusions of created cancer cells into recipient mice are used to generate multiple myeloma characteristics in desired experimental animals (Verreault et al., 2016). Submucosal or cerebral infusion of cancerous cells into rats is the most extensively used approach in laboratory animals. Transgenic models administered with a dermal injection continue to grow but are not well restricted within the intramuscular region, rendering this model distinct from the TME (Verreault et al., 2016). Intracerebral infusion for tumor transplantation is preferable because cancer cells are directly implanted into the brains rather than being degraded TABLE 3 Neuroendocrine tumor, sickness of Alzheimer, and disease of Parkinson animal models are summarized.

| Disease | Model | Characteristics | Reference | Plant name | Part used | Neuroprotective activity | Reference |
|---------|---|--|--|---|----------------|---|--|
| GMB | Xenograft/HT1080 (human cell line) | IDH1 mutant (MGG152) | Tateishi et al. (2015) | <i>Fumaria indica</i> (Hausskn.) Pugsley | Leaf | Significant activity of ethanolic extract on rat cognitive dysfunctions. Potential antianxiety activity of leaf | (Singh et al., 2013a; Singh et al., 2013b) |
| | Xenograft/LNT- 229 and LN-308 | IDH1 R132C mutant (HT1080) | Szabo et al. (2016) | | | extract; preclinical study | |
| | (human cell line) Xenograft/BT111 (TIC), BT116 (TIC) Xenograft/U251 (human cell line) | Unmethylated MGMT (BT111) and (BT116) N/A | Sharpe et al. (2015) Sharpe et al. (2015) | Alhagipseud alhagi (M. Bieb) Desv. ex B. Keller &Shap. | Whole plant | Traditionally used for neuroprotective disorders. Compounds having neuroprotective activity such as flavanone glycosides and alkaloids such as β - phenethylamine and tetrahydroisoquinoline have been reported | (Ghosal et al., 1974; Singh et al., 1999; Muhammad et al., 2015) |
| | Allograf GL261- Luc (mouse cell line) | N/A | Zeng et al. (2013) | | | | |
| AD | PDAPP | Unspecified microgliosis and resistant microglia are both related to diseases | Cohen et al. (2013) | Premna mucronata Roxb | Whole plant | Luteolin and apigenin are reported, and they are reported to be neuroprotective | Dave et al. (2015) |
| | Tg2576 | Plaques are related to neural stem cells that have been allocated a particular character | Elfenbein et al. (2007) | | | to be neuroprotective | |
| | APP23 | In specifically, the glial cell is associated in fibrils deposits, epidemic, and | Karch and Goate, (2015) | Semecarpus anacardium L.f | Fruits | Stress-induced neuroprotective activity | Shukla et al. (2000) |
| | J20 | also cytosis Microgliosis and astrogliosis are two forms of engraftment that may occur in the brain and have negative consequences | (Hartley et al., 2015) | Sida cordifolia L | Whole plant | Ameliorative effect in parkinsonism | SJ and Kumar B, (2020) |
| PD | MPTP Neurotoxin: inhibition of complex I | No time passes during the process of accumulation, and cholinergic neurotoxicity occurs quickly and severely, resulting in a major motor deficit | Dauer and Przedborski, (2003) | <i>Tinospora</i> <i>cordifolia</i> (Thunb.) Miers | Stems | Suppresses neuro- inflammation in Parkinsonian Mouse Model; potential neuro-regenerative candidate against glutamate induced excitotoxicity: | (Sharma et al., 2018; Birla et al., 2019) |
| | 6-OHDA Suppression of 6- OHDA suppression of enzyme I as well as antioxidant distress caused by neurotoxic | Heterogeneous sensory impairments, no synuclein aggregation, fast and severe cholinergic deterioration | Schober and research, (2004) | Trichosanthes dioica Roxb | Rhizome | Neuropharmacological properties of root | Bhattacharya and Haldar, (2013) |
| | UCH-L1 I93M mutation | In rodents, there were no synuclein aggregation, cholinergic neurotoxicity, or minor motor impairments | Setsuie et al. (2007) | | | | |

via subcutaneous administration (Hoffman, 2015). Other methods of mutant hereditary materials supply, such as chromosomal- and viral carrier (retroviral administration of genomic material)-based template matching delivery of materials (Bardella et al., 2016; Miyai et al., 2017; Oldrini et al., 2018), can also be used to create models. In addition to this, the desired mice should be produced by integrating mice that have mutated genes and the progression of neurological

06

disorders in humans should be triggered. Till now, the majority of AD lab animals have been genetic pigs in which transcription factor gene mutations were linked to the formation of β amyloidosis and/or cornerstones β peptide aggregates and filaments (Banik et al., 2015). Botulism toxins, which are utilized to cause the buildup of senile lesions in patients with AD, are employed to induce physiological features. Conversely, PD animals are classified into neurotoxicity or genetically caused, and they incorporate cholinergic neurons loss in the globus pallidus (Blesa and Przedborski, 2014). In particular, neurotoxicity treatments produce sudden and strong cell death in the ventral striatum, as well as developmental delays and abnormal movements. Heritable traits rodents, on the other hand, show varied point, which is an important pathogenesis, as well as certain obvious manifestations such cellular disintegration and behavioral abnormalities. a-Synuclein is the main component that causes PD and AD with laws clumps (Henderson et al., 2019). The accumulation of α -synuclein, as well as the depletion of dopamine, happens in geographical cells called synapses throughout the brains. Specific genes can be used to illustrate gene variations, or antiviral translation can be used to cause chromosomes variations. The instances of frequently utilized laboratory animals for CNS disorder are given in Table 3.

Challenges of regular drug delivery to the CNS

The medicine must be lipophilic in nature and have a low molecular mass (400-600 Daltons) in order to be successful in conventional chemotherapy (Pardridge, 1997). This transportation can be accomplished through percutaneous, passive, and other methods (Dominguez et al., 2013; Chauhan and Chauhan, 2015), although BBB provides for regulated entrance of prospective medicines (Crone, 1986; Jain, 2007; Stockwell et al., 2014; Yi et al., 2014). Slow drug activity, affiliation or transformation of the medication into non transporting forms, and poorer synaptic bioavailability are all main factors for pharmacological difficulties in the CNS (Tamai and Tsuji, 1996). Some enzymatic processes in the CNS also breakdown or keep dormant drugs in the brain that have a nonspecific function (Crone, 1986). The thin blood vessel wall, which makes up the majority of the physiological barrier, is made of endothelial cells. Glial cells, specifically astrocytes, firmly encircle the capillary endothelial cells, with neurons nearby, and control BBB functions (Figure 2) (Zlokovic, 2008; Neuwelt et al., 2011; Yamazaki and Kanekiyo, 2017). The neurovascular connections are important for a healthy brain as well as the efficient administration of drugs (Woodworth et al., 2014; Wang et al., 2021). Other leukocytes located in the periphery, such as pericytes, oligodendrocytes, and extracellular matrix, among others, play an essential role in maintaining homeostasis in the brain as well as a healthy BBB. In addition, owing to the presence of tight junction proteins such as occludins, claudins, and tight junction's adhesion molecule-A (Sweeney et al., 2019; Bhattacharya et al., 2020), large serum proteins and electrolytes are unable to pass across endothelial cells in a paracellular manner. Because of this, the BBB is responsible for establishing a distinct anatomical divide between the albuminal brain and the lumen of the blood vessel (Zeevi et al., 2010; Naik and Cucullo, 2012; Heidarzadeh et al., 2021). In addition to a physical barrier that is provided by the BBB, the endothelial cells of the brain possess an enzymatic activity that permits some compounds to be degraded before their passage over the BBB (Lorke et al., 2008; Upadhyay, 2014). Brain endothelial cells are known to express a number of important transport proteins, including organic anion carriers (OATs), P-glycoprotein/MDR1 (Pgp), multidrug 1 (MRPs), resistance-associated protein breast cancer-resistant protein (BCRP), and several other proteins (Englund, 2005; Löscher and Potschka, 2005; Sangha et al., 2022). These proteins control the pathway of several different xenobiotics and drugs. These proteins may inhibit the buildup of macromolecules in the brain by effluxing chemicals back into the blood supply. Nevertheless, the OATs have been discovered to be situated at almost all of the body's barrier epithelia and the endothelium, and they have shown to play some roles in the management of intercellular mobility of several diverse organic anionic molecules across these epithelial barriers and between body fluid compartments. In addition, the endothelium has been found to be a localization site for the OATs as well (such as blood and the central nervous system, blood and urine, intestine and blood, blood and bile, blood and the placenta, and others). Regardless of the fact that the prototype members of this transporter family are capable of the bidirectional movement of substrates, the vast majority of organic anion transporters (OATs) are usually thought of as influx carriers, which aid in the passage of organic anions into the epithelial and endothelial cells. Because of these factors, the neuroprotective actions of the dysfunctional BBB make it visibly challenging to transfer medications to the brain, particularly in circumstances in which efflux carriers are hyperactive (Klaassen et al., 2013; Sanchez-Covarrubias et al., 2014; Saunders et al., 2016). Thus, it may be difficult for drugs to reach their target tissue or to concentrate enough in the brain, a problem that has been seen in a variety of conditions that affect the CNS to this day (Thorne and Frey, 2001; Doran et al., 2005; Bae and Park, 2011). Because of its significance in the medical community and the pharmaceutical business, the study of drug delivery has become an increasingly important topic of research over the course of the last two decades. This review delves into deeper information regarding recent studies and research initiatives that have proven inventive approaches to circumvent the BBB in medication delivery (Nance, 2019). These investigations and projects are discussed in this review in greater depth.



Nanotechnology for drug delivery to the brain

Engineered adjustable devices with sizes in order of billionths of meters have been offered as an attractive tool, which is potentially able to answer the unmet problem of increasing transportation of drug over the BBB in recent years including the advent of nanomedicine (Re et al., 2012; Holmes, 2013). NP technology is quickly advancing across a variety of technologies. NPs are objects that range in size from 1 to 100 nm (Demetzos and Pippa, 2014) and function as a single unit in terms of transport and characteristics. The promise of NPs' multifunctionality, along with their ability to deliver therapeutic payloads, including BBB impermeant medicines, is the primary factors of this enthusiasm. The use of NPs for drug delivery in the brain is demonstrated by the fact that adequate surface functionalization may increase either their targeting of the BBB or their enhancement of its crossing at the same time. When BBB impermeant pharmaceuticals are vehicled by NPs, their ability to cross the barrier is defined by the physicochemical and biomimetic properties of the NPs, rather than the chemical structure of the drug, which is impeded inside the NPs. The ability to confer on NPs' features such as high chemical and biological stability, feasibility of incorporating both hydrophilic and hydrophobic pharmaceuticals, and the ability to be administered via a variety of routes (including oral, inhalational, and parenteral) (Petkar et al., 2011) makes them even more appealing for medical applications. Furthermore, NPs can be functionalized to target-specific tissues by covalently conjugating them to several ligands (such as antibodies, proteins, or aptamers). Multiple copies of a ligand can be added to NPs, also greatly increasing their binding affinity by multivalent functionalization (Montet et al., 2006), when developing NPs for therapeutic applications, by keeping in mind that their systemic delivery can cause significant changes. The nonspecific interaction between the shell of NPs, also several kinds of proteins circulating in the circulation, results in the adsorption of opsonin on the surface of proteins that generate the so-called corona. These proteins significantly alter the basic material qualities which govern NPs removal from circulation via the reticuloendothelial system, which is mostly found in the spleen, also in the liver. The most frequent methods for evading RES are constructing particles with a neutral surface charge, also covering them with different hydrophilic surfactants such as polysorbates and polyethylene glycol (PEG), and using small nanoparticles (e.g., 80 nm) (Provenzale and Silva, 2009). These "stealth" NPs are able to avoid the reticuloendothelial system which have a long circulation time including blood

stability and can be functionalized to successfully target and penetrate the BBB (Gabathuler, 2010). Finally, NPs must be nontoxic to blood cells, healthy bystander cells, as well as biodegradable and biocompatible, also noninflammatory and nonimmunogenic (Andersen et al., 2010). It is also worth noting that designing NPs to improve drug delivery toward the brain does not always suggest that they can cross the BBB on their own. NPs could play this role in at least two ways, as expected: by boosting drug concentration inside BBB cells or at the luminal surface of BBB cells. This results in a local high concentration gradient between blood and the brain that is higher than that obtained following systemic delivery of the free drug. The gradient should therefore favor toward increased drug passive diffusion. This aim, for example, could be accomplished by creating NPs which are functionalized to target brain capillary endothelial cells. This property can be confirmed or disproven by their subsequent uptake from specific cells (Haque et al., 2012) and by transporting themselves and their drug cargo into the CNS. This task can be accomplished, for example, by allowing NPs to target brain capillary endothelial cells and after that transcellular transit through the BBB (Beg et al., 2019). The key aspects of NPs routinely used for medical purposes and already used or potential candidates for brain medication delivery will be outlined in the next part for the sake of completeness of this review.

Inside the brain parenchyma: NPs diffusion

Although this feature is unlikely to be crucial when looking for delivery of therapeutic agents, it could be critical when looking for NPs to cross through the BBB, whereas the extracellular space (ECS) of the cerebral epithelium could hinder their dispersion or possibly prevent them from entering the brains. The opportunity to accomplish neural insertion with greater NPs is anticipated to help further homogeneousness which is longer lasting, as well as efficacious biomedical applications inside the nervous system. It also could be used to diagnose brain lesions, brain hemorrhage, neuronal injury, and other brain diseases where the BBB is undermined or municipal service methodologies which are conceivable (Masserini, 2013). It is now firmly understood that the ECS represents a volumetric proportion of between 15%, as well as 30% in healthy mature neural tissues, with a mean frequency of 20%, and this drops to 5% during worldwide hemorrhage (Syková and Nicholson, 2008; Wolak and Thorne, 2013). The exact magnitude of the intervals between the cell lines is seldom visible. Organic compounds such as inulin as well as sugar have a lower partition coefficient in the ECS than that in freshwater, implying that their transportation is restricted (Levin et al., 1970). 1) Cell membrane corridors; 2) entrapment of compounds in killed microspheres; 3) viscoelastic drag exacted by the biomolecules which assemble the collagen fibers or drag emerging from the wall surfaces of the networks while compounds are massive, such as in the situation of NPs; 4) temporary adhering to living cells or matrix proteins; and 5) nonspecific engagement with deleterious distribution on the surface of the NPs. Thorne and Nicholson (2006) offered an alternative type of framework to describe the greater compressibility empirically detected in the in vivo rat brain for larger molecules such as dextran and particularly huge synthesized nanoscale nanoparticles. It was hypothesized because nanocrystals with one hydraulic dimension of 35 nm were near to the median thickness of the ECS, as well as the predicted ECS breadth was 38-64 nm, regardless of whether a rectangular or cylindrical paradigm was used. Nance et al. (2012) claimed that NPs as massive as 114 nm in dimension disseminated inside the mammalian as well as rat brains only if they were thickly covered with PEG, in contrast to Thorne and Nicholson (2006). Researchers determined that actual normal brain cell ECS has some holes greater than 200 nm using those marginally sticky PEG-coated NPs. More than a fifth of all openings are less than 100 nm.

Impact of protein corona

Nanomaterials' capability to penetrate the brains may be determined by physical and chemical factors other than size. The creation of a "chromosphere" of macromolecules on the interfaces of nanomaterials, whose constitution varies relying on the pathway of access and the chemical and physical interface qualities of NPs, considerably complicates the scenario. The presence of a corona of macromolecules on the exterior of NPs could also influence whether they can pass from one capsule to the next and if these are efficiently picked up by cells (Pietroiusti et al., 2013). Depending on the location of distribution, nanomaterials are originally introduced to a physiological environment, such as bloodstream, in various situations where nanomaterials interface with living beings. For example, NPs implanted via IV infusion would indeed be introduced to bloodstream, which contains an abundance of enzymes and several other sophisticated bioactive molecules that compete with the nanomaterials' membrane. According to prevailing thinking, the "naked" nanomaterials do not reside in vivo even though individuals which are instantaneously adapted by the adsorbate of hemoglobin in red blood cells with elevated affiliations for the surface of nanoparticles. This results in the formation of a too much or little covalently bonded stack (the as such tough corona) as well as a tenuously affiliated interactive surface (the so-called soft corona). Despite the reality that bodily liquids comprise millions of protein, it was discovered that ordinary tequilas have a restricted quantity and also different kinds of compounds were obtained from the bloodstream when this

topic was researched (Mahon et al., 2012). It is also worth noting that perhaps the corona might play a significant role in those other negative consequences of NPs in biological processes, such as cytokine production and plasma coagulation, and is not always involved in intracellular ingestion. This should be noted that none of the in vitro studies looking into the mechanism of NPs transportation to the brains take into consideration the membrane functionalization of NPs within the bloodstream and whether it can influence BBB bridging. This is a subject that requires more research. Furthermore, after being ingested by endothelium BBB units, NPs may depart the brains wrapped in various macromolecules, also depending on whether they have completed endosomal escape, transcellular, or pinocytosis, and hence impose considerable impacts (e.g., cytotoxicity) on neurons. Moreover, that matter has still not been explored (Masserini, 2013).

BBB changes in neurological diseases

The BBB's unique pharmacological properties influence medication ductility and, for most situations, severely restrict the quantity of medications absorbed by the brains. It really should be noted since the physiology and structure of the BBB are modified in a variety of clinical states, and these alterations could have had an impact on physiologically circulating compounds, medicines, as well as NPs traversing the barriers. It also should be noted that these alterations are not uniform across all illnesses: variation is significant, making it difficult to anticipate the consequences of any given medicine. The BBB changes could affect the dosing, effectiveness, and adverse effects of regularly utilized medications in general (Masserini, 2013). The impact of hypoxic conditions on the barriers, for example, has been thoroughly studied. Oxygen starvation triggers a cascade of processes that enhance BBB vulnerability, perhaps due to limited connection breakdown and regulated by secondary messengers such cytokine and nitrous oxide (Yang and Rosenberg, 2011). Considering this scenario, biomolecules' transit through the BBB is anticipated to rise. In laboratory animals of hypoxic conditions, nanocrystalline antidiuretic hormone has been shown to preserve neurons (Chen et al., 2012). Albumin has been demonstrated to penetrate target cell from the bloodstream in animals (Deng et al., 1995), suggesting that the BBB is similarly disrupted in septic encephalopathy. Although there is no indication of a significant rupture of the BBB in AD in individuals or experimental animals, the AD brains do require less sugar and oxygen. It is uncertain whether the reduction is due to a fault in the circulation through the membrane or a reduction in CNS demand; nonetheless, it is compatible with the AD microenvironment encouraging barriers and cellular discharges which are deleterious to cognitive health (Sharma et al., 2012).

Size

One underappreciated question is when the diameter of NPs developed for CNS medication distribution creates a contribution. Various inquiries have been conducted in an endeavour to elucidate the situation. Sarin et al. (2008) discovered that NPs may pass through holes in the blood-brain tumor boundary of RG-2 aggressive glioblastoma after being systemically supplied with synthesized PAMAM microparticles with a diameter of about 12 nm. Sonavane et al. (2008) investigated the biodistribution of aqueous metal nanoparticles of various sizes (15, 50, 100, and 200 nm) administered intravenously to rodents. The number of NPs that penetrated the brains within 24 h was inversely related to their mass, with 15 nm gold NPs having a 500-fold amount than 100 nm NPs and 200 nm NPs having a relatively small quantity. Nevertheless, for 15 and 50 nm, the overall amount of currency, which really is proportional to the entire capacity of NPs penetrated to the burden of medication in the instance of drug-loaded NPs, was comparable, as well as approximately 30% less for 100 nm. Models are used to describe (Oberdörster et al., 2004) created crystalline nanoscale graphene nanoparticles with a diameter of roughly 36 nm that have been inhaled by animals. The researchers noted whether flying NPs of such magnitude can reach the CNS through the olfactory bulb from the olfactory mucosa. Emerging investigation uncovers whether the passage of NPs throughout the digestive tract has a significant impact on their future accessibility to the brains. Hillyer and Albrecht (2001) investigated the digestive assimilation and vasculature dissemination of magnetic dispersion of gold nanoparticle with diameters of 4, 10, 28, and 58 nm. The golden nanoparticles were administered orally, and golden concentrations in several tissues/organs, including the brain, were measured after 12 h. The overall quantity of metal effort to uphold in the brain was comparable for 4 nm and 58 nm particle diameter, according to the study. Schleh et al. (2012) looked into the effect of golden nanomaterials size and morphology charged on diffusion through gastrointestinal obstacles and accumulated in the brain following oral treatment.

Nanomethods for delivering CNS drugs

Nanotechnology is a novel, effective, and cutting-edge method of delivering neurotherapeutics across the blood-brain barrier. Nano-medicines have shown considerable promise in CNS drug delivery over the last few decades due to their nanosized range, certain physicochemical features, and capacity to leverage surface-tailored biocompatible and biodegradable nanomaterials (Kaur et al., 2008). Site-specific transport of medicines including other chemicals across the

BBB using nanotechnology might potentially be tailored, which can perform specific roles whenever needed. The medication, which is a component of the nanoengineered complex with other essential activities, is the pharmacologically active component to be given, for example, encapsulating the active drug, protecting it from enzymatic degradation, allowing it to release at certain pH, crossing the BBB, and targeting specific brain cells (Silva, 2008). Liposomes, PNPs, SLNs, micelles, dendrimers, and numerous other pharmacological nanocarriers have all been created (Rajadhyaksha et al., 2011; Wong et al., 2012; Ozkizilcik et al., 2017).

Micelles

Micelles are vesicles made of amphiphilic surfactants (nonpolymeric micelles) or amphiphilic copolymers (polymeric micelles), and they have lately attracted the attention of scientists because they may be used as a means of delivering drugs to the CNS (Aliabadi and Lavasanifar, 2006; Torchilin, 2007). Polymeric micelles have a longer duration of action and higher biodistribution than non-polymeric micelles (Ozkizilcik et al., 2017). They have a core-shell structural design with a size range of 10-100 nm that is made up of an outer hydrophilic environment primarily composed of polyethylene glycol (PEG) and as well as an inner hydrophobic core composed of molecules such as polycaprolactone, polypropylene glycols, phospholipids, and fatty acids, allowing hydrophobic drugs to be loaded (Torchilin, 2007). In an aquatic environment, the outer hydrophilic coating keeps micelles stable. In addition, it extends the time they spend circulating in the blood, shielding them from the reticuloendothelial system (RES) and increasing their accumulation in leaky vascular areas (Ozkizilcik et al., 2017). Because it may block drug efflux transporters, the pluronic copolymer class, commonly referred to as poloxamers, is of particular interest. Drug distribution to the CNS is made easier by the P-gp efflux carrier, which is widely expressed on the BBB (Park et al., 2008). In addition, they have been shown to increase drug solubility and stability in plasma as well as the distribution of low molecular mass drugs into the brain. Numerous efforts have been made to change micelles in a manner that enables a larger concentration of loaded medication to pass the BBB on the opposite side without much difficulty. One such modification is the binding of polyclonal antibodies to the receptor on the luminal side of the BBB against the brain-specific antigen, 2-glycoprotein, or insulin. When these modified micelles are administered intravenously to mice after being loaded with either a fluorescent dye or the neuroleptic drug haloperidol, both the fluorescent dye transport and the neuroleptic activity of the drug are significantly enhanced (Batrakova and Kabanov, 2008). Another variation of the micelle approach involves direct conjugation of the drug molecule and the targeting moiety to the amphiphilic area. For instance, we studied cyclo(Arg-GlyAsp-d-Phe-Lys) paclitaxel conjugate–loaded micelles modified with transferrin and discovered better absorption by brain microvascular endothelial cells *in vitro* as well as prolonged retention in glioma tumors *in vivo*, both without considerable toxicity (Naqvi et al., 2020). The interaction of chitosan oleate self-assembled polymeric micelle–based nano-systems and polylactic–glycolic acid (PLGA) NPs coated with chitosan oleate (CS-OA), which may be conferred as a positive surface charge, with Caco-2 and Hela cells was investigated. According to data from cell line interactions, TGA studies, and influence the way, PLGA-CS-OA was shown to be more stable than polymeric micelles when it was packed with the lipophilic drug carrier resveratrol (Zhang et al., 2012).

Liposomes

The first generation of novel colloidal nanocarriers, known as liposomes, were developed and tested as a medicine delivery system in the 1970s (Bawarski et al., 2008). These are tiny, spherical vesicles that include one or more phospholipid bilayers around a hydrophilic compartment in the core. This has morphological similarities with cell membranes, which are used to transfer medicines, proteins, and peptides (Martins et al., 2007; Wong et al., 2012; Ozkizilcik et al., 2017). The three varieties are classified according to their size and number of bilayers as small unilamellar (10-50 nm), gigantic unilamellar (50-1000 nm), and multilamellar (20-100 nm). In addition to hydrogen bonding between molecules, non-covalent interactions such as van der Wall forces also result from this. These buildings can be turned around (Ramos-Cabrer and Campos, 2013). Normal, unaltered liposomes circulate in the body for a brief period of time before being swiftly eliminated from the systemic circulation by RES cells. The creation of long-circulating and tailored liposomes has so been the subject of various efforts. It has been discovered that covering liposomes with polyethylene glycol (PEG) may successfully evade RES detection of nanocarrier systems (Doxil, 2012; Wong et al., 2012). The targeted delivery of PEG-modified liposomes to the brain may be improved with further modifications using ligands such monoclonal antibodies (mAbs) against glial fibrillary acidic proteins, transferrin receptors (TRs), or human insulin receptors (Pardridge, 1999; Miele et al., 2019). Because of receptor-mediated endocytosis, transferrin-conjugated liposomes have been shown to transfer payloads, such as 5fluorouracil to the brain, more effectively (Shah et al., 2013). Prednisolone-loaded liposomes and mAbs that are recognized by cell surface receptors in the targeted tissue are combined to form immunoliposomes, which have been shown to have improved liposome dispersion inside the brain and to be highly effective against experimental autoimmune encephalomyelitis (Soni et al., 2008). It has been shown that immunoliposomes, which are nanocarrier systems for brain medicine administration, may also

be employed in gene therapy by linking a TRsMAb-targeted liposome with a plasmid for tyrosine hydroxylase in the treatment of Parkinson's disease in a rat model. Small interfering RNA (siRNA) targeting the epidermal growth factor receptor (EGFR) has also been delivered via this technique, leading to EGFR knockdown and improved survival in mice implanted intracranially with brain tumors (Schmidt et al., 2003; Zou et al., 2010). Another method of enhancing liposomes' capacity to penetrate barriers and improve therapeutic efficacy is to modify them with cell penetrating peptides. For instance, doxorubicin-encapsulated liposomes may be used in combination with the specific ligand transferrin (T7) and non-specific cell penetrating peptide (TAT), which has been shown to have high availability across the BBB and specific cell targeting to brain glioma (Pardridge, 2007). When in contact with water, nimodipine PR liposomes form a liposomal structure. This has just been shown to increase the medication's oral absorption (Zong et al., 2014).

Polymeric nanoparticles

These polymeric nanoparticles are solid colloidal dispersions of biocompatible and biodegradable polymers with sizes ranging from 10 to 100 nm, such as PACA, PLA, PLGA, and natural proteins and polysaccharides (García, 2009). Polymeric nanoparticles can be used in a variety of applications, biodegradation, biocompatibility, including and biodegradation. Lipophilic medications are encapsulated in a thick polymer matrix core, while a hydrophilic corona provides steric stability to NPs. The NPs' surface might be used to encapsulate, adsorb, or chemically bind the drug to be delivered (Parveen et al., 2012b). PEG and other hydrophilic polymers have been shown to increase the NP's shelf-life in systemic circulation, which may be achieved by the addition of tissue-targeted polysaccharides or other hydrophilic polymers. When combined with traditional therapy, coating poly(n-butylcyanoacrilate) NPs with 1% polysorbate 80 (PS80) has been shown to enhance the concentration of rivastigmine or tacrine medication in the brain while decreasing hepatic or gastrointestinal adverse effects (Mohamed and van der Walle, 2008; Wilson et al., 2008). Dalargin nanoparticles coated with PS80 were reported to pass the blood-brain barrier and elicit antinociception upon oral distribution in another investigation (Das and Lin, 2005). It is possible that PS80-coated PACA nanoparticles injected intravenously may bind to ApoE and B in the blood, which can then be transcytosed across the BBB through low-density lipoprotein receptors (Kreuter, 2013). PLGA nanoparticles may transport medicine over the BBB without damaging it. C57/bl6 mice and hCMEC/D3 cells were used to study the effects of vanlafaxine-loaded PLGA nanoparticles on depression. P-gP pump efflux does not influence intranasal delivery of receptor-mediated PLGA

nanoparticles, which leads to improved biodistribution in the brain and does not alter P-gP pump concentration in the basolateral side after 24 h through receptor-mediated endocytosis (Cayero-Otero et al., 2019).

Drug delivery methods for the brain

The BBB operates as a microvascular endothelium connection, allowing important chemicals and ionic transfer to the brain (Tamai and Tsuji, 1996). The BBB is usually a major stumbling block for medicine administration systems. Water-soluble compounds have been found to migrate via particular carrier-mediated endocytic, transporters, and the extracellular channel. Dispersion and addition to introducing have been used to transfer liposomes compounds (Stenehjem et al., 2009). The following are examples of drug distribution pathways.

Invasive approach

This mechanically violated approach breaks through the BBB and inserts the medicine straight into the brains. It necessitates a brain surgery for intracranial hemorrhage medication delivery and intracerebroventricular (ICV) injection (Stenehjem et al., 2009; Gabathuler, 2010). Collapsing the tight junction of endothelial causes BBB disturbance for drug diffusion (Inamura and Black, 1994; Stenehjem et al., 2009). Osmolarity disruption (Neuwelt et al., 1991; Rapoport and neurobiology, 2000) or destructive plasma dissolved substances can be used to deliver this (Aird, 1984; Erdlenbruch et al., 2003). Because the medicine is carried in the periphery circulating blood rather than the different tissues, ICV delivery of drugs is regarded a poor technique (Lee et al., 2011). Instead of advancing high molecular weight drug absorption, ICV was confined to confined drug dealing and lack of intended CNS action following straight prescribed medication owing to excessive intraocular pressure (Soni et al., 2016a).

Pharmacological approach

The spontaneous inactive passage of medicines *via* the blood-brain barrier is the basis for this empirical methodology (Stenehjem et al., 2009; Gabathuler, 2010). Because of their tiny molecular size, poor hydrophobic interactions capability, and solubility in water, these compounds can traverse the blood-brain barrier without assistance (Hsiung et al., 2003). Physiological changes, such as a decline in the amount of polar functional groups, are also part of this strategy, which improves medication transport across the BBB (Pardridge, 1998). If the solubilization of the changed

particles increases by several folds, it may operate as a P-glycoprotein outflow pump (Stenehjem et al., 2009).

Physiological approach

Drug transport to the CNS *via* receptors and carriers is deemed a cutting-edge pharmacological approach (Stenehjem et al., 2009; Gabathuler, 2010). The BBB is frequently seen with associated protein and insulin channels (Gabathuler, 2010). As a result, the medicine may convey pharmaceuticals to the specified cerebral location if it interacts with the ligands of these channels. The medicine must be capable of mimicking the indigenous carrying substrates when using transporter-mediated distribution (Allen and Cullis, 2013). However, the pharmacokinetics and interaction capability of the carrier molecules constrain the administration of CNS drugs *via* a pharmacological methodology.

Treatment of CNS disorders with nanocarriers and BBB-bypassing techniques

The presence of the tightly selective BBB is one of the greatest roadblocks in the development of successful CNS therapies. As a result, the use of various nanocarriers in combination with various administration routes may be of tremendous interest in bypassing the BBB.

Parenteral drug delivery

Since IV injections are less invasive and allow precise control over bioavailability, thorough dosages of frequently toxic drugs, and continuous administration of several-hour injections, obtaining a relatively better drug concentration within the duration of treatment is the most effective mode of injecting drugs for CNS afflictions. Medicines given intravenously, engaging other organs or cell, should either have a greater possibility to passively disperse through the BBB (such as the two very hydrophobic medication including carmustine and temozolomide) or employ one of the aided transcytosis pathways, as shown graphically in Figure 2. Exogenous chemicals may utilize the BBB's most common facilitators and transporters to access the brain barrier, according to an assessment by Griffith et al. (2020) and Mulvihill et al. (2020). Nanotherapies may be implicated in a wide range of transcytosis pathways because of their heterogeneous physicochemical character (Figure 3). Furthermore, to stimulate a sufficient brain translocation, their tridimensional properties almost always necessitate the appearance of a certain specific side on their surface. In addition to the widely utilized intravenous

delivery, which requires a special design of nanoparticles to a specified site and circumvent the BBB, direct intracerebral drug infusion is another viable but quite invasive technique of administration. This surgically aided method of administration can be accomplished either by stereotactic injection in the diseased area (i.e., the intraparenchymal route) to build local depots or by intraventricular administration (Griffith et al., 2020). Because of their high danger, these invasive administration methods are only used in life-threatening situations and do not allow for repeated injections (Yi et al., 2014).

Intranasal drug delivery

Because it permits the formulation to circumvent the BBB, for brain targeting, intranasal, or nose-to-brain pharmaceutical delivery is a highly fascinating form of distribution that has just developed. The respiratory and olfactory parts of the nose can both be used to carry out this operation. Millions of olfactory neurons reside in the nasal epithelium, allowing substances to be transported directly to the brain *via* transcellular diffusion *via* the olfactory bulb (Griffith et al., 2020; Mulvihill et al., 2020). The second channel involves the respiratory epithelium, which allows drugs to directly reach brain tissue *via* the trigeminal nerve (Griffith et al., 2020). As recently reviewed by Yi et al. (2014), the intranasal route has been successfully tested in a range of neuroinflammation-related illnesses.

Intracarotid infusion

The intracarotid infusion technique involves injecting a medicine or fluid into the carotid artery, which is the main blood vessel that connects the heart and the brain (Joshi et al., 2008). Intracarotid drug delivery can provide extremely high local concentrations of the unique pharmacological agent in a matter of seconds, facilitating regional delivery. This technique has long been overlooked in the development of medications for CNS illnesses, and the kinetics of such a route of administration is still unknown. However, a few studies have indicated that it can be an effective delivery method, particularly for the treatment of brain tumors (Inamura et al., 1994).

Transmucosal drug delivery

Transmucosal drug administration is a novel and popular approach of direct CNS drug injecting that uses heterotopic mucosal grafts to circumvent first-pass metabolism and enables for fast drug uptake into the systemic circulation (Abhang et al., 2014; Pandey et al., 2016; Dong, 2018). This strategy is based on "cohesion", or attachment to the human



mucosa, which is extremely close to the blood circulation and may even transfer high molecular weight and polar substances to the CNS. This approach may be adapted to accommodate items delivered by nasal and oral routes by utilizing of droplets, creams, pills, or even suppositories as delivery systems (Bleier et al., 2013).

BBB disruption caused by pharmacological or physical activity

To allow the brief passage of therapeutic substances such as nanocarriers, the BBB's physical integrity can be partially and temporarily disturbed. These bioactive disturbances can be achieved by "physically" broadening and opening tight junctions, such as by utilizing an external ultrasonic (US) source to drive an electromagnetic differential to propel nanomaterials throughout the BBB or microscopic pores throughout the BBB (Teixeira et al., 2020): the nanomaterials can be osmotic (sorbitol, sugar, lactose moieties, ammonia, or glycerin) or form effective (cholecystokinin, antihistamine, tryptophan, and glutamine) or pro-inflammatory (particularly, prostaglandins and interleukins) or biologically active substances or tumor necrosis factor (Chenthamara et al., 2019; Xie et al., 2019).

The ECM as a barrier to nanopharmacology

Because of the fact that it slows down the diffusion of macromolecules, the extracellular matrix (ECM) might be considered a barrier to the administration of nanomedicines. Hyaluronidase, an enzyme that breaks down HA, was injected before the NPs; this resulted in a more even dispersion of the NPs throughout the tissues (Lee et al., 2021; Liu et al., 2022). Nevertheless, improved NP distribution is attainable even without altering the ECM design. Therefore, paclitaxel (PTX)loaded PEG-PLGA block copolymer NPs, which have a mean particle size of 70 nm, are able to diffuse through brain tissue one hundred times more quickly than identical particles that do not have PEG coverings. In accordance with this, the local administration of strongly PEGylated PTX-loaded nanoparticles dramatically slowed down the development of 9L liposarcoma as compared to PTX-loaded PLGA nanoparticles or unencapsulated PTX (Palei et al., 2018). Despite this, the diffusion of 100-nm PEG-modified particles was eight times slower in 9L liposarcomas compared to normal brain tissue. This was because the tumor tissue had a larger cell density and more extensive collagen content than normal brain tissue (Soni et al., 2016b; Henrich-Noack et al., 2019). These findings demonstrate that a combination of a vector targeting the ECM with a dense layer of PEG on the surface of NPs could improve the efficacy of the NPs by reducing their high emphasis with brain tissue and enhancing their percolation into the tumor (LeClaire, 2021). The perivascular space and lymphatic systems are also important factors that should be taken into consideration in relation to the diffusion of NPs in brain tissue (Khawar et al., 2015). It has been proven that the perivascular space offers a favorable permeable channel for the rapid convective transport of large NPs across tissue. This is true in particular for the transportation of bigger particles, the passage of which is impeded by the ECM (Voigt et al., 2014; Witten and Ribbeck, 2017). In point of fact, this might be the mechanism that lies behind the supposed presence of channels inside the ECM that make it easier for NPs to move about.

Nanosystems' possible side effects on the ECM

In addition to this, we need to take into account the possible impacts that NP have on the structure of the ECM. Therefore, silver nanoparticles may activate an inflammatory signaling pathway, leading to an upregulation of matrix metalloproteinases (MMPs) and an induction of ECM breakdown (Raja et al., 2020). In addition, it has been shown that the cytotoxicity of multiwalled carbon nanotubes may be altered *in vitro* depending on the kind of endothelial cells they are exposed to.

Brain tumor nanotheranostics

T1-weighted MRI is used to guide the treatment of malignant brain tumors using image-guided therapy. Gadolinium chelates are often employed in contrast-enhanced T1-weighted MRI imaging of malignant brain tumors to differentiate between white and gray tissue. Because the BBB is disrupted in the tumor location in order to accurately define the tumor borders, these contrast chemicals do not flow into healthy brain areas. In order to improve the contrast agent's sensitivity and efficacy, scientists are working on developing customized gadolinium-loaded nanocarriers (Fink et al., 2015; Aparicio-Blanco and Torres-Suárez, 2018). Thus, amphiphilic compounds containing gadolinium in the chelated form as a T1weighted MRI contrast agent for brain nanotheranostics have been created. Currently, radiation is the usual treatment for malignant brain tumors, and the T1-weighted MRI imaging function has been used for theragnostic reasons (Gløgård et al., 2000; Richard et al., 2008). It is hypothesized that gold nanoparticles coated with gadolinium chelates (zeta potential: -30 mV) can absorb high energy ionizing radiation to cause thermal ablation of tumors, and thus act as a radio-sensitizing agent, in rats orthotopically implanted with a 9L liposarcoma, which can be used for image-guided radiotherapy (Simonet, 2018). It is possible to monitor the distribution of gold nanoparticles after intravenous injection using T1-weighted MRI since gadolinium had been added to the solution (Jung et al., 2012). This helped establish the best time to administer the X-rays. Comparing treated to untreated rats, the median survival period following tumor implantation was days, a 473% improvement over the non-radiosensitive nanotheranostics radiosensitizing treatment (median survival time: 2.5 days after tumor implantation). X-rays penetrate deeper into the tissue than visible light, making this thermal treatment a viable option for brain tumor theragnostic. Gadolinium chelate (particle size: 2 nm)-doped polyciliate nanoparticles were then used to investigate gadolinium's radiosensitizing potential. After utilizing MRI-guided microbeam radiotherapy to target the tumor while sparing healthy tissue, the researchers were able to achieve a five-fold increase in median survival time, and half of all liposarcoma-bearing rats survived 100 days after tumor implantation using gadolinium-rich microbeam radiotherapy (Sancey et al., 2014; Bechet et al., 2015). Bechet et al. developed 3-nm-sized silica-based nanoparticles (zeta potential: 22 mV) functionalized with an anti-angiogenic heptapeptide (ATWLPPR) that targets neuropilin-1, an endothelial receptor found only on malignant brain tumors. This was done to improve the distribution of gadoliniumcontaining nanotheranostics to malignant brain tumors. MRI contrast agent and photosensitizer for PDT of brain malignancies guided by interventional MRI were contained in these particles. Using MRI guidance, the optical fiber was stereotactically implanted for PDT in rats with the orthotopic glioma model U87 after intravenous injection of neo vasculature-focused nanocarriers (Kopelman et al., 2005). Significantly reduced intratumorally blood perfusion in rats treated with targeted nanotheranostics was seen in this study (80%). A histological study of brain slices after MRI-guided PDT with specific carriers showed vascular damage and swelling (Hirschberg et al., 2008).

T2-weighted MRI

Because of the fact that they have a high magnetic susceptibility, superparamagnetic iron oxide nanoparticles are often utilized as T2 agents to produce contrast enhancement for the purpose of identifying watery tissue (Qiao et al., 2009; Yang et al., 2009). The degree of tissue contrast enhancement is dependent on vascular extravasation, much as it is with T1 agents. As a direct result of this, brain-targeted iron oxide nanoparticles have been produced for the purposes of glioma therapeutic research. Since superparamagnetic iron oxide nanoparticles are already in use as MRI contrast agents, and since they can be easily upgraded to nanotheranostics by encapsulating various therapeutic agents, there is a significant

possibility that they will be used in the clinical translation of brain diagnostics (Dehaini et al., 2016; Ag Seleci et al., 2021). For the purposes of theragnosis, various anticancer agents (drug substances, genes, and antibodies) or photosensitizers can be loaded into iron oxide nanoparticles. This allows for the possibility of combining MRI with chemotherapy or photodynamic therapy for the purpose of providing localized treatment (Aljiffry et al., 2009; Barreto et al., 2011). In addition, iron oxide nanoparticles themselves simultaneously exhibit therapeutic properties. These properties include the generation of heat under alternating magnetic fields, which can be used for the thermal ablation of tumors (Estelrich and Busquets, 2018; Tong et al., 2019). As a result, iron oxide nanoparticles are functioning independently capable of as brain nanotheranostics (Nelson et al., 2020; Lorkowski et al., 2021).

Other imaging techniques

Although alternative scanners have a great potential for use in brain imaging, their use as diagnostic tools for brain disorders have not yet been investigated (De Bie et al., 2010; Pereira et al., 2016). Unfortunately, the majority of research that describe the performance of brain diagnostic nanomedicines in vivo examine either the therapeutic or imaging function, but seldom examine both of these functions at the same time. Imaging and therapeutic chemicals are often loaded independently onto a particular nanocarrier so that each may be evaluated independently for their biodistribution and effectiveness (Duncan and Gaspar, 2011; Chen et al., 2013). In these independent investigations, a larger armamentarium of imaging methods (radionuclide-based imaging and optical imaging, respectively) and nanocarrier architectures (liposomes, polymer nanosphere, and solid lipid nanoparticles) have been employed (Sanson et al., 2011; Butler et al., 2016; Yao et al., 2016; Houston et al., 2020). The development of theragnostic nanomedicines with multiple imaging functions is an additional promising future area of research. These nanomedicines would improve the sensitivity of monitoring the response of tumors to treatment, as well as guide stimulus-responsive therapies and surgical resection of malignant brain tumors (Nurhidayah et al., 2019; Das et al., 2020). Even though various nanocarriers with numerous imaging functionalities have previously been produced and tested in orthotopic models of malignant brain tumors, the coupling of these nanocarriers with a treatment method has not yet been explored in vivo (Baumann et al., 2013; Dixit et al., 2015; Aparicio-Blanco and Torres-Suárez, 2018).

The effect of nanobiotechnology in nanomedicines to treat CNS diseases

The term "nanotechnology" originates from the Greek word "nano," which translates to "dwarf". Nanotechnology refers to the

production and utilization of materials, equipment, and frameworks through the control of matter on the nanometer-length scale, or more specifically, at the level of atoms, molecules, and supramolecular structures (Fulekar, 2010; Ramanathan, 2012). It is the common term for the construction and utilization of multiple functions with at least one characteristic dimension measured in the nanometer scale. A nanometer (nm) is equal to one billionth of a meter (10-9 m), so this term refers to the construction and utilization of structures with at least one characteristic dimension evaluated in the nanometer scale (Nasrollahzadeh et al., 2019). This is about the same as having four times the diameter of a single atom. DNA molecules have a width of around 2.5 nm, whereas protein molecules range in size from 1 to 20 nm (Heng et al., 2006). It was inevitable that nanotechnology would be used for biotechnology, which gave origin to the name "nanobiotechnology". Given the intrinsic nanoscale functional components of live cells, it was unavoidable that nanotechnology would be applied to biotechnology (Ball, 2004; Boisseau et al., 2007). The use of nanobiotechnology to medical practice is known as nanomedicine. The field of healthcare is already beginning to feel the effects of nanobiotechnology's impact (Jain, 2008; Fakruddin et al., 2012). Over the course of the previous half century, early ideas about nanotechnology have given rise to a wide variety of technologies, and there are even a few treatments that are based on nanotechnology that are now on the market (Ratner and Ratner, 2003; Lemley, 2005). The phrase "nano bio-pharmaceuticals" may refer to a wide variety of applications within the pharmaceutical business. Some examples of these applications include drug discovery and drug delivery (Invernizzi, 2010). The development of the microscope ushered in a new era in medical practice by making it possible to identify microscopic organisms and investigate the histology of illness. Microsurgery was a significant improvement over crude microsurgery and opened the door to the possibility of surgeries that either had not been performed in the past or had a high mortality and morbidity rate. A comparable influence will be exerted on medical practice and surgical procedures by the advent of nanotechnologies, which will make it possible to see beyond the microscale (Morrow et al., 2007; Martel, 2016). This is due to the fact that both physiological and pathological processes that take place at the cellular level take place on a nanoscale. The incorporation of recent developments in genomes and proteomics into nanomedicine, which paves the way for the creation of customized medicine, is another way in which we may think of nanomedicine as an improvement on molecular medicine (Stratakis et al., 2009; Wei et al., 2017). A discussion on the connections between nanobiotechnology, nanomedicine, and other technological fields was carried out (Riehemann et al., 2009; Haseeb et al., 2020). This diagram provides an overview of how nanobiotechnology will influence the growth of nanomedicine both directly and indirectly via the enhancement of other fields, such as the distribution of nanomedicines and molecular diagnostics. Parallel to the development of nanomedicine, personalized medicine is made easier by the same technology that underpins nanomedicine.

Neuro-inflammatory cell targeting with nanoparticles and modules

Cell membrane coating strategies

Greater blood flow lengths and focusing capabilities have subsequently been demonstrated using biological membranes fragmentation as a nanomaterial interface covering element. Cellular membrane coating approaches offer immunological masquerade, lower neutrophil assimilation, and an appealing alternative for targeted non-immune cellular components including glial as well as synapses. Synthesis of cells plasma nanostructures (such as cellular barrier nanocrystals) and nanomaterials embellishment with biological membranes segments (membrane-coated nanomaterials or harnesses) are two innovative methods that permit development of novel therapeutic techniques for several diseases (Luk and Zhang, 2015). Cell-coated nanoparticles provide another option for avoiding immune detection, increasing prolonged circulation, and delivering targeted drug delivery. Various sorts of membrane surface could be employed for a range of specialized biomedical techniques without demanding substantial physiological modifications, which is clinically meaningful and can help the Food and Drug Administration to approve the product faster. Rapid innovations in the production of fibroblast nanomaterials that can discharge drugs for longer periods of time in responsiveness to the surroundings are paving the way for more complex treatment approaches, which could lead to distinct benefits for CNS drug delivery (Hu et al., 2011; Luk and Zhang, 2015; Liu et al., 2019).

Neuro-inflammatory treatment using stimuli-responsive nanoparticles

A current emphasis of investigation has been the production of nanostructures which exhibit medicinal benefits in accordance to biological responses or application of an extrinsic stimulus. These materials, also known as stimuli-responsive materials, are designed to replicate the behavior of living beings. Formulating nanocarriers that release medications selectively at the target site in response to preset illness signals could revolutionize the development of successful neuroinflammation therapeutics (d'Arcy and Tirelli, 2014).

Neurological disorders and diseases tested by nanomedicines

Nanomedicine in stroke

Understanding the pathophysiology of ischemic stroke, as well as how nanoparticles engage with ischemic cells, is essential. The basic issue of how nanoparticles carry medicines over the BBB is still unknown, but new breakthroughs in nanoparticle design and manufacture, as well as innovative *in vivo* imaging tools (such as intravital microscopy) may be helpful. The patients with ischemic stroke will benefit from the development of new medicines and innovative nanoparticle-based drug delivery approach (Dong et al., 2020).

Traumatic brain injury and nanotechnology

Traumatic brain injuries (TBIs) are injuries to the cerebrum tissue that cause temporary or permanent impairment of brain functions. TBIs cause 135,000 deaths and cases of lifelong disability in the United States each year. Acute respiratory distress syndrome (ARDS) is a life-threatening lung illness characterized by dyspnea, acute hypoxemia, reduced lung compliance, and diffused bilateral pulmonary infiltrates, among other symptoms. Patients with ARDS are severely unwell or hospitalized as a result of serious injuries, one of which is a TBI. Researchers have been looking for biomarkers in various biofluids for brain damage and its consequences. TBI biomarkers in urine xiv and serum that have been clinically validated still have low sensitivity and specificity (Vuong, 2018).

Application nanomedicine in the treatment of brain tumors

Invasive brain CNS tumors account for approximately about 2% of all cancers. These are, nevertheless, typically associated with significant incidence and fatality percentages. Although recent standard-of-care interventions such as operation, radiotherapy, and chemotherapeutic are already present, there has yet to be curative or non-toxic therapies for aggressive CNS tumors. Nanoscience has the potential to transform this situation. It offers new promise in the identification and treatment of patients. This new technique could establish a framework for combining diagnoses, therapies, and transport to the cancer, as well as ongoing performance surveillance by creating and producing materials employing atomic and molecular elements. This study examines current advances in cancer nanotechnology, with a focus on nanoparticle systems, which are significant tools for improving medication delivery in brain tumors (Figure 4) (Invernici et al., 2011).

Alzheimer's disease

Since the discovery and definition of AD, the most common dementing dementia in the world, a century has passed in research. However, there are currently no definitive diagnostic



procedures or viable treatments for AD. Furthermore, the currently available diagnostic methods are insufficient for early detection of AD so that preventive measures can be taken. Nanotechnology, a new field that has just emerged, has promised new approaches to address some of the issues associated with AD. Nanotechnology is the process of designing and fabricating nanoscale (1-100 nm) structures by manipulating the positioning and/or self-assembly of atoms and molecules in a controlled manner. In this study, we discuss the benefits of nanotechnology in AD detection and treatment research. They include the possibility for a greater knowledge of the molecular pathways that underlie AD, early detection of the disease, and successful therapy. The contributions of atomic force microscopy, single molecule fluorescence microscopy, and nano SIMS microscopy for AD research are discussed. In addition, the bio-barcode test, localized surface plasmon resonance nanosensor, quantum dot, and nanomechanical cantilever arrays, which were newly proposed uses of nanotechnology for the early diagnosis of AD, are examined. Nanotechnology applications in AD therapy are reviewed and studied, including neuroprotection toward oxidative stress and anti-amyloid therapies, neurodegeneration, and drug delivery across the BBB. All of these applications have the potential to improve treatment for AD and other NDs (Nazem and Mansoori, 2008).

Parkinson's disease

Screening imaging tools are being established and treatment instruments will rely heavily on nanotechnology. Nanostructures make use of manufactured nanomaterials with the tightest organizational structure on the nanoscale level in at least one parameter. Modification of certain substance qualities can lead to novel bioactive components. Nanomaterials could be utilized to create technologies which restrict as well as repair neurobiological disorder processes, cultivate as well as encourage efficient neuronal replacement, provide neuroprotective effects, and make drug and small molecule administration across the BBB easier. All of these are relevant for improving existing PD (Torres-Ortega et al., 2019).

Spinal cord injury and nanotechnology in regenerative medicine

Because of the significant nerve damage, spinal cord injury (SCI) can result in the loss of perceptual and athletic function. Yet, pieces of evidence describing the exact pathogenic pathways in SCI remain ambiguous. The necessity for systemic distribution, which has a major negative effect on the patient, makes it difficult for pharmaceutical treatment to adequately relieve SCI impairments. As a result, developing SCI treatment options is both necessary and

TABLE 4 Treatment of neurodegenerative disorders used various potential nanostructures.

| Neuro-degenerated disease | Nanomaterial | Outcome | References |
|---------------------------|--|--|----------------------------|
| AD | AGulX NPs | Enhancing the susceptibility of MRI, penetrating the BBB with minimal toxicity, addressing amyloid accumulations in the brain, and enabling excellent viewing of amyloid plaques without causing negative impacts | Pansieri et al. (2018) |
| | Boron-dipyrromethene biosensor | Might be utilized to analyze the accumulation of $A\beta$ A in Alzheimer's disease and identify its self-assembly | Quan et al. (2019) |
| PD | Micelles | Intravenous delivery/efficient drug delivery | Brynskikh et al. (2010) |
| | Rasagiline (Double emulsion/solvent evaporation) | Significantly affect MAO-B enzyme protect your neurons from the oxidative stress | Myers, (2004) |
| MS | Quantum dot complexed with MMP-9 siRNA and MMP-1–loaded PLGA nanoparticles | Interpretation of MMP9 was significantly suppressed in the brain, as well as in microvascular endothelial cells and leukocytes. inhibition | Singh et al. (2017) |
| HD | Peptides QBP1, NT17, and PGQ9P 2 nanoprecipitation | Significantly affect mHtt clumping bring back the functioning of the motor systems | Joshi et al. (2018) |

helpful. Nanopharmaceutical-based regenerative medicine will provide enormous potential space for clinical medicine as a result of the application of nanotechnology in pharmaceutical research. These nano pharmaceuticals (i.e., nanocrystalline medications and nanocarrier drugs) are made with various materials or bioactive compounds to increase therapeutic efficacy, lessen adverse effects, and distribute drugs quietly, among other things. Drug regulatory agencies are currently approving a growing number of nanopharmaceutical items, prompting more academics to focus on prospective SCI therapy options (Zhao et al., 2021).

Nano-formulation approaches

For an influential drug delivery strategy in CNS therapy, nanoparticles affect the pharmacokinetics of drug and boost drug loading capacity (Khanbabaie and Jahanshahi, 2012). Before being loaded into various nanomaterial-based vehicles and delivered to the brain, drugs must be chemically changed (C Dinda and Pattnaik, 2013). It was also able to reach the brain *via* transcytosis through the BBB (Stenehjem et al., 2009). Nanobiotechnology has made significant progress in the realm of medicine delivery. We reviewed the features, nanotechnology-based medication delivery, and drug release mechanism using a few examples of patent nanomedicine (Table 4) (Haque et al., 2011).

Nanoformulations of natural compounds and herbal medications

Polymeric nanoparticles, nanocapsules, and nanospheres

Drug-loading capabilities of polymers nanostructures allow the device to safeguard and maintain the included medication toward breakdown. As a result, there is a greater possibility of drug incorporation and cerebral accessibility. They can elude neutrophils because of their stable architectures and distinctive characteristics, which facilitates the drug transport to the CNS. Although nano emulsions are created by a uniform thin wrapper enclosing a hydrocarbon chamber, nanomaterials are a robust elastomeric matrix created using micro-emulsion polymer (Figure 5) (Vickers, 2017).

Polymeric nanogels and nanosuspensions

Extremely covalently bonded submicron gelatin processes made of charge density, or standard quantity polymers are referred to as nanoemulsion. The hydrogels range from 20 to 200 nm range in size. The capability of this mechanism to load drugs is between 40% and 60%. Recent research studies revealed that polynucleotide consumption by hepatic and the spleen might be reduced and increased by hydrogels Crystal dosage form stabilized frameworks. bv combinations of triglycerides or polymeric detergents make up matrix tablets nano emulsions. The ease of usage and impressive drug unloading and distribution capabilities of nano emulsion are only a few of their many benefits (Modi et al., 2010).

Carbon nanotubes and nanofibers

Mesoporous silicon nanomaterials, CNTs, stacked dual ions in aqueous solution, ferromagnetic nanoclusters, as well as calcium phosphate nanostructures are a few examples of matrix composite targeted distribution that have found curative use in a variety of illnesses, including NDs. While improving drug concentration, penetration, retention impact,



durability, and accessibility to desired places, properties of nano complexes can withstand extended blood vessels. This nanotechnology could also alter how a drug is released and make it easier to see and analyze a medication's action. In addition, the CNTs are a remarkable find for nanopharmacology due to their flexibility to a variety of stimuli, including heat, pH, compounds, pressures, and magneto electric forces. Many of the most notable methods for neuropsychiatric applicability is the use of carbon-based nanomaterials, such as CNTs. Carbon alloying elements with tubular nanomaterials are known as CNTs. To improve their electrical currents, CNTs are currently the subject of intensive research. Direct electrical stimulation is one of the most efficient methods for treating many psychological and neurological conditions, particularly PD. When these activating conductors are present, the immune response might sometimes respond negatively, making it difficult to use these fibers. Nanofibrous production is less dangerous than CNT production and poses less of a threat to the environment. It is fascinating that cerebral prostheses are created and produced using nanowires. In compared to nanostructured materials, alternative nano methods might not be capable of achieving the same functionalities (Modi et al., 2009; Naz et al., 2019).

Polymeric nano micelles

Most of the most intriguing methodologies in nanotechnology are polymers microspheres. This arrangement is comprised a conceptual model with a buildings of polyethylene glycol in the core and a liposomes interior. The inclusion of hydrophobic toxic compounds is this program's key benefit. The polymeric nanoparticles have sizes ranging between 10 and 100 nanometric scale (Vickers, 2017).

Polymeric nano liposomes

Lipoproteins called nano liposomes have a hydrophilic top and two hydrophobic cores. Their diameters range from 30 nm to a few meters. The phospholipid liposomes or the watery chambers of the liposomes can both hold a substantial quantity of medicines. Having an active component with changed interfaces can speed up bloodstream, minimize the likelihood that such lipid membranes will be eliminated by the hepatic, and diminish chemical peroxidation in serum. *In vitro* tests demonstrated their effectiveness for specific CNS dosage forms and demonstrated their extraordinary propensities for BBB drug translocation.

Exosomes: New promising nanocarrier

A bulk of eukaryotic cells, including T and B lymphocytes, progenitor cells, and monocytes, produce macrovesicles, which are phospholipid relating to or characteristic extrinsic packets with micrometer sizes greater than 30-150 nm. Liposomes are exceptional and different from other solid lipid nanoparticles due to a number of unique characteristics. High cytocompatibility, nanoscale size, institutional and municipal cell communication abilities, minimal business consists, phenomenal capacity to overcome biological membranes, and significant capacity for stroma trying to target, entrap, and transport of a variety of subgroups of instability chemotherapeutic drugs, including fatty acids, hormone levels, peptides, and genetic mutations, have identified extracellular vesicles as viable chemical delivery vehicles for treating a variety of diseases, including cancer and myocardial infarction (Aryani and Denecke, 2016; Sarko and McKinney, 20172017; Niu et al., 2019; Zhang et al., 2019).

Conclusion and future perspectives

CNS disorders are a big issue in today's industrialized world. Nanotechnology has shown to be a cutting-edge and promising way for delivering drugs to the brain with pinpoint accuracy. However, in order to assess their dynamic behavior in biomedical science, we still need to learn more about their qualities and characteristics. For various CNS disorders that may be caused by multiple metabolic pathways, we now need a multimodal medicine. Nanodrugs could be the answer to this problem. Neurological issues can be linked to a variety of illnesses, including diabetes, trauma, and different psychotic conditions. As a result, nanomedicine requires the total eradication of these co-morbidity variables while limiting negative consequences. Apart from that, because genetic alterations in neuronal cells are difficult to achieve, nanotechnology-based drug delivery could be a viable treatment option for CNS illnesses. Although clinical trials using polymer-based gold nanoparticles and CNT nanomedicines are limited because to their strong physical and mechanical robustness, they may be helpful in transporting medications whose transit is still unclear. Despite the numerous benefits of nanoparticle-based therapy, there are still a number of difficulties to be resolved. There is no established method for evaluating the level of toxicity and targeted pharmaceutical release in the CNS at this time. As a result, current nanotechnology applications must be substantially developed in order to be safe and focused. Gold nanoparticle (US2011262546, US2011111040), lipid nanoparticle (WO2008024753, WO2008018932), chitosan nanoparticle (US2010260686), and SLN (US2011208161) are some of the nanomedicines that have registered for patents in complex CNS treatment in recent years. A growing population and a rise in brain illnesses necessitate the development of new promising remedies. Nanotechnology's use in neuroscience will fill an unmet medical need and give sufferers hope. Nanomedicine of the next generation may be able to regulate prolonged and focused drug administration in a specific way. We still need to improve nanotechnological ways in pharmaceuticals for greater comprehension and improved living quality, rather than reducing side effects and increasing viability of nanodrugs. Nanomedicines' potential use cannot be overstated, but their opportunity and risk formula also point to potentially dangerous adverse effects. Nanotechnology's quick advancement in today's research makes it hard to dismiss it merely on the basis of its disadvantages. Specific instructions must be followed in order to avoid the most harmful effects of nanotechnology. Nanotechnology-based drug delivery is also expected to revolutionize the timeframe for traditional drug distribution, with personalized pharmaceuticals significantly more efficient than the current standard. NPs are well-suited for the diagnosis and treatment of brain disorders due to their physical, chemical, and biological characteristics. The possibility of applying this technical technology to treat and diagnose CNS diseases has been promising. For improved medicine delivery to patients with CNS illnesses, several nanosolutions based on polymerbased techniques and nanoparticles are being investigated. To achieve development and hasten the treatment of illnesses affecting the CNS, experts in biomedical sciences (neuroscience, immunology, pharmacology, molecular imaging), materials science, biomaterials, and pharmaceuticals (polymers, nanomaterials, medicine, and genetics) are necessary. In both in vitro and in vivo models of malignant brain tumors, targeted delivery of chemotherapy drugs and antisense gene therapy has been made possible using nanocarriers, which has produced significant disease progression inhibition. Sadly, very few, if any, of this encouraging preclinical research have effectively been transferred to the clinic to affect patient care. Insufficient research on the toxicity of nanomaterials, their immunological compatibility, and the relative scarcity of in vivo trials are obstacles to this successful translation. In the future, nanomaterials might be utilized to replace a niche for transplanted stem cells, offering structural support as well as a sustained release of signaling molecules. In addition, the mechanical features of the nanomaterials as well as the incorporation of bioactive signals allow them to be functionalized to interact with stem cells. Collaboration across disciplinary boundaries will be essential to the effective application of nanotechnology in medicine. Although the use of nanotechnology alone is unlikely to be able to complete the challenging process of CNS repair, it does have a significant potential to influence clinical neuroscience treatment options. The most successful uses of nanoparticles in the treatment of CNS disorders have paired the effectiveness of nanoscale interventions with growth factors or cells that boost the overall effect of the treatment, emphasizing the significance of a combined approach to nanotherapeutics. Furthermore, improvements in our biology understanding of the mechanisms behind these illnesses can only increase the usefulness of nanotechnology applications in CNS disease. The aspect that has the potential to radically alter the practice of neurology is the interaction between new knowledge of the molecular causes of neurological illnesses and the multipurpose capabilities of nanotechnology. The severity of this condition and the current difficulties in AD neuropharmacology research are the main causes of the failure of CNS medication development. As a result, the future for their synergistic combination effect in CNS disease will be paved by the expansion in research of combinations of new nanocarriers and therapeutic targets. Although the development of nanomedicine with strong biodegradability and biocompatibility as a therapeutic agent for the treatment of CNS illnesses is a possibility, each NP has advantages and disadvantages of its own. As was previously mentioned, there are some negatives to nanomedicine that cannot be disregarded since they may buildup in the liver, kidney, and spleen and cause oxidative damage in the brain, which may be harmful to long-term health. In addition, there have been relatively few studies on the pharmacodynamics and pharmacokinetics of this nanomedicine, and their side effects continue to be a significant clinical concern.

Author contributions

MR, MI, and MM: conceptualization and writing original manuscript. SA, MN, EN, and MV: editing and proofreading. KK and RS: editing, proofreading, and supervision.

References

Abhang, P., Momin, M., Inamdar, M., and Kar, S. J. D. D. L. (2014). Transmucosal drug delivery- an overview. *Drug Deliv. Lett.* 4, 26–37. doi:10.2174/22103031113039990011

Ag Seleci, D., Maurer, V., Barlas, F. B., Porsiel, J. C., Temel, B., Ceylan, E., et al. (2021). Transferrin-decorated niosomes with integrated InP/ZnS quantum dots and magnetic iron oxide nanoparticles: Dual targeting and imaging of glioma. *Int.* J. Mol. Sci. 22, 4556. doi:10.3390/ijms22094556

Ai, X.-L., Liang, R.-C., Wang, Y.-C., and Fang, F. (2016). Stem cells combined with nano materials-novel therapeutics for central nervous system diseases. *J. Nanosci. Nanotechnol.* 16, 8895–8908. doi:10.1166/jnn.2016.12743

Aikins, M. E., Bazzill, J., and Moon, J. J. J. N. (2017). Vaccine nanoparticles for protection against HIV infection. *Nanomedicine* 12, 673–682. doi:10.2217/nnm-2016-0381

Aird, R. B. J. E. n. (1984). A study of intrathecal, cerebrospinal fluid-to-brain exchange. *Exp. Neurol.* 86, 342–358. doi:10.1016/0014-4886(84)90192-4

Alam, M. I., Beg, S., Samad, A., Baboota, S., Kohli, K., Ali, J., et al. (2010). Strategy for effective brain drug delivery. *Eur. J. Pharm. Sci.* 40, 385–403. doi:10.1016/j.ejps. 2010.05.003

Alguacil, L., Pérez-García, C. J. C. D. T.-C., and Disorders, N. (2003). Histamine H3 receptor: A potential drug target for the treatment of central nervous system

Funding

It is supported by Research program of University of Granada, Excellence project PrF UHK 2217/2022-2023, Spain. This study was also partially supported by the Charles University in Prague, Czechia (PROGRES Q40/15) and Ministry of Health project MHCZ – DRO (UHHK, 00179906).

Acknowledgments

The authors are grateful to the Department of Pharmacy, Daffodil International University for giving the opportunity to conduct this research.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer VC declared a shared affiliation with the author RS to the handling editor at the time of review

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

disorders. Curr. Drug Targets. CNS Neurol. Disord. 2, 303-313. doi:10.2174/ 1568007033482760

Aliabadi, H. M., and Lavasanifar, A. J. E. o. o. d. d. (2006). Polymeric micelles for drug delivery. *Expert Opin. Drug Deliv.* 3, 139–162. doi:10.1517/17425247.3.1.139

Alifieris, C., and Trafalis, D. T. J. P. (2015). Glioblastoma multiforme: Pathogenesis and treatment. *Pharmacol. Ther.* 152, 63–82. doi:10.1016/j. pharmthera.2015.05.005

Aljiffry, M., Walsh, M. J., and Molinari, M. J. W. j. o. g. W. (2009). Advances in diagnosis, treatment and palliation of cholangiocarcinoma. *World J. Gastroenterol.* 15, 4240. doi:10.3748/wjg.15.4240

Allen, T. M., and Cullis, P. R. J. A. d. d. r. (2013). Liposomal drug delivery systems: From concept to clinical applications. *Adv. Drug Deliv. Rev.* 65, 36–48. doi:10.1016/ j.addr.2012.09.037

Alshamrani, M. J. P. (2022). Broad-spectrum theranostics and biomedical application of functionalized nanomaterials. *Polym. (Basel).* 14, 1221. doi:10. 3390/polym14061221

Andersen, A. J., Hashemi, S. H., Galimberti, G., Re, F., Masserini, M., and Moghimi, S. M. J. J. o. B. (2010). The interaction of complement system with abetabinding liposomes: Towards engineering of safer vesicles for the management of alzheimer's disease. J. Biotechnol. 150, 97–98. doi:10.1016/j.jbiotec.2010.08.252 Andersson, U., Grankvist, K., Bergenheim, A. T., Behnam-Motlagh, P., Hedman, H., and Henriksson, R. J. M. O. (2002). Rapid induction of long-lasting drug efflux activity in brain vascular endothelial cells but not malignant glioma following irradiation. *Med. Oncol.* 19, 1–9. doi:10.1385/MO:19:1:1

Aparicio-Blanco, J., and Torres-Suárez, A.-I. J. A. B. (2018). Towards tailored management of malignant brain tumors with nanotheranostics. *Acta Biomater*. 73, 52–63. doi:10.1016/j.actbio.2018.04.029

Aryani, A., and Denecke, B. J. M. n. (2016). Exosomes as a nanodelivery system: A key to the future of neuromedicine? *Mol. Neurobiol.* 53, 818–834. doi:10.1007/s12035-014-9054-5

Attri, J. P., Bala, N., and Chatrath, V. J. I. j. o. a. (2012). Psychiatric patient and anaesthesia. *Indian J. Anaesth.* 56, 8–13. doi:10.4103/0019-5049.93337

Au, K., Meng, Y., Suppiah, S., Nater, A., Jalali, R., Zadeh, G. J. N. A. t. t. M. o. P., et al. (2017). Current management of brain metastases: Overview and teaching cases. *New Approaches to the Management of Primary and Secondary CNS Tumors*. doi:10.5772/66310

Bae, Y. H., and Park, K. J. J. o. c. r. (2011). Targeted drug delivery to tumors: Myths, reality and possibility. *J. Control. Release* 153, 198–205. doi:10.1016/j.jconrel. 2011.06.001

Ball, P. J. N. (2004). Synthetic biology for nanotechnology. Nanotechnology 16, R1–R8. doi:10.1088/0957-4484/16/1/r01

Banik, A., Brown, R. E., Bamburg, J., Lahiri, D. K., Khurana, D., Friedland, R. P., et al. (2015). Translation of pre-clinical studies into successful clinical trials for alzheimer's disease: What are the roadblocks and how can they be overcome? *J. Alzheimers Dis.* 47, 815–843. doi:10.3233/JAD-150136

Bardella, C., Al-Dalahmah, O., Krell, D., Brazauskas, P., Al-Qahtani, K., Tomkova, M., et al. (2016). Expression of Idh1R132H in the murine subventricular zone stem cell niche recapitulates features of early gliomagenesis. *Cancer Cell* 30, 578–594. doi:10.1016/j.ccell.2016.08.017

Barreto, J. A., O'Malley, W., Kubeil, M., Graham, B., Stephan, H., and Spiccia, L. J. A. m. (2011). Nanomaterials: Applications in cancer imaging and therapy. *Adv. Mat.* 23, H18–H40. doi:10.1002/adma.201100140

Batrakova, E. V., and Kabanov, A. V. J. J. o. c. r. (2008). Pluronic block copolymers: Evolution of drug delivery concept from inert nanocarriers to biological response modifiers. *J. Control. Release* 130, 98–106. doi:10.1016/j. jconrel.2008.04.013

Baumann, B. C., Kao, G. D., Mahmud, A., Harada, T., Swift, J., Chapman, C., et al. (2013). Enhancing the efficacy of drug-loaded nanocarriers against brain tumors by targeted radiation therapy. *Oncotarget* 4, 64–79. doi:10.18632/oncotarget.777

Bawarski, W., Chidlowsky, E., Bharali, D., and Mousa, S. J. B. M. (2008). Emerging nanopharmaceuticals. *Nanomedicine* 4, 273–282. doi:10.1016/j.nano. 2008.06.002

Bechet, D., Auger, F., Couleaud, P., Marty, E., Ravasi, L., Durieux, N., et al. (2015). Multifunctional ultrasmall nanoplatforms for vascular-targeted interstitial photodynamic therapy of brain tumors guided by real-time MRI. *Nanomedicine* 11, 657–670. doi:10.1016/j.nano.2014.12.007

Beg, S., Rahman, M., Barkat, M. A., and Ahmad, F. J. (2019). Nanomedicine for the treatment of disease: From concept to application. New York: CRC Press.

Bhatia, S. (2016). Nanoparticles types, classification, characterization, fabrication methods and drug delivery applications, Natural polymer drug delivery systems. Springer, 33–93.

Bhattacharya, A., Kaushik, D. K., Lozinski, B. M., and Yong, V. W. J. J. o. N. R. (2020). Beyond barrier functions: Roles of pericytes in homeostasis and regulation of neuroinflammation. *J. Neurosci. Res.* 98, 2390–2405. doi:10.1002/jnr.24715

Bhattacharya, S., and Haldar, P. K. J. C. j. o. n. m. (2013). Neuropharmacological properties of Trichosanthes dioica root. *Chin. J. Nat. Med.* 11, 158–163. doi:10. 1016/S1875-5364(13)60043-6

Birla, H., Rai, S. N., Singh, S. S., Zahra, W., Rawat, A., Tiwari, N., et al. (2019). Tinospora cordifolia suppresses neuroinflammation in parkinsonian mouse model. *Neuromolecular Med.* 21, 42–53. doi:10.1007/s12017-018-08521-7

Bleier, B. S., Kohman, R. E., Feldman, R. E., Ramanlal, S., and Han, X. J. P. o. (2013). Permeabilization of the blood-brain barrier via mucosal engrafting: Implications for drug delivery to the brain. *PLoS One* 8, e61694. doi:10.1371/journal.pone.0061694

Blesa, J., and Przedborski, S. J. F. i. n. (2014). Parkinson's disease: Animal models and dopaminergic cell vulnerability. *Front. Neuroanat.* 8, 155. doi:10.3389/fnana. 2014.00155

Boisseau, P., Houdy, P., and Lahmani, M. (2007). Nanoscience. Springer.

Brynskikh, A. M., Zhao, Y., Mosley, R. L., Li, S., Boska, M. D., Klyachko, N. L., et al. (2010). Macrophage delivery of therapeutic nanozymes in a murine model of Parkinson's disease. *Nanomedicine* 5, 379–396. doi:10.2217/nnm.10.7 Butler, K. S., Durfee, P. N., Theron, C., Ashley, C. E., Carnes, E. C., and Brinker, C. J. J. s. (2016). Protocells: Modular mesoporous silica nanoparticle-supported lipid bilayers for drug delivery. *Small* 12, 2173–2185. doi:10.1002/smll.201502119

C Dinda, S., and Pattnaik, G. J. C. P. B. (2013). Nanobiotechnology-based drug delivery in brain targeting. *Curr. Pharm. Biotechnol.* 14, 1264–1274. doi:10.2174/1389201015666140608143719

Carvalho, M., Carmo, H., Costa, V. M., Capela, J. P., Pontes, H., Remião, F., et al. (2012). Toxicity of amphetamines: An update. *Arch. Toxicol.* 86, 1167–1231. doi:10. 1007/s00204-012-0815-5

Cayero-Otero, M., Gomes, M. J., Martins, C., Álvarez-Fuentes, J., Fernández-Arévalo, M., Sarmento, B., et al. (2019). *In vivo* biodistribution of venlafaxine-PLGA nanoparticles for brain delivery: Plain vs. *Funct. nanoparticles* 16, 1413–1427.

Charney, D. S., Mihic, S. J., Harris, R. A. J. T. P. B. o. T. t. e. B. L., Lazo, J. S., and Parker, K. L. (2006). *Hypnotics and sedatives*. New York: McGraw-Hill, 401-427.

Chauhan, M. B., and Chauhan, N. B. (2015). Brain uptake of neurotherapeutics after intranasal versus intraperitoneal delivery in mice. J. Neurol. Neurosurg. 2 (1), 009.

Chen, H., Spagnoli, F., Burris, M., Rolland, W. B., Fajilan, A., Dou, H., et al. (2012). Nanoerythropoietin is 10-times more effective than regular erythropoietin in neuroprotection in a neonatal rat model of hypoxia and ischemia. *Stroke* 43, 884–887. doi:10.1161/STROKEAHA.111.637090

Chen, Y., Chen, H., and Shi, J. J. A. M. (2013). *In vivo* bio-safety evaluations and diagnostic/therapeutic applications of chemically designed mesoporous silica nanoparticles. *Adv. Mat.* 25, 3144–3176. doi:10.1002/adma.201205292

Chenthamara, D., Subramaniam, S., Ramakrishnan, S. G., Krishnaswamy, S., Essa, M. M., Lin, F.-H., et al. (2019). Therapeutic efficacy of nanoparticles and routes of administration. *Biomater. Res.* 23, 20–29. doi:10.1186/s40824-019-0166-x

Cheshire, W. P., Jr, Ott, M. C. J. H. T. J. o. H., and Pain, F. (2001). Headache in divers. *Headache* 41, 235–247. doi:10.1046/j.1526-4610.2001.111006235.x

Cohen, R. M., Rezai-Zadeh, K., Weitz, T. M., Rentsendorj, A., Gate, D., Spivak, I., et al. (2013). A transgenic Alzheimer rat with plaques, tau pathology, behavioral impairment, oligomeric aβ, and frank neuronal loss. *J. Neurosci.* 33, 6245–6256. doi:10.1523/JNEUROSCI.3672-12.2013

Crone, C. (1986). The blood-brain barrier: A modified tight epithelium. Chichester: Ellis Horwood.

d'Arcy, R., and Tirelli, N. J. P. f. a. t. (2014). Fishing for fire: Strategies for biological targeting and criteria for material design in anti-inflammatory therapies. *Biodegrad. Polym.* 25, 478–498. 10.1002/pat.3264.

Das, D., and Lin, S. J. J. o. p. s. (2005). Double-coated poly (butylcynanoacrylate) nanoparticulate delivery systems for brain targeting of dalargin via oral administration. *J. Pharm. Sci.* 94, 1343–1353. doi:10.1002/jps.20357

Das, S. S., Bharadwaj, P., Bilal, M., Barani, M., Rahdar, A., Taboada, P., et al. (2020). Stimuli-responsive polymeric nanocarriers for drug delivery, imaging, and theragnosis. *Polymers* 12, 1397. doi:10.3390/polym12061397

Dauer, W., and Przedborski, S. J. N. (2003). Parkinson's disease: Mechanisms and models. *Neuron* 39, 889–909. doi:10.1016/s0896-6273(03)00568-3

Dave, N., Cetiner, U., Arroyo, D., Fonbuena, J., Tiwari, M., Barrera, P., et al. (2015). Validated HPTLC method for quantification of luteolin and apigenin in Premna mucronata Roxb. *Adv. Pharmacol. Sci.* 10, 2015. doi:10.1155/2015/682365

De Bie, H., Boersma, M., Wattjes, M. P., Adriaanse, S., Vermeulen, R. J., Oostrom, K. J., et al. (2010). Preparing children with a mock scanner training protocol results in high quality structural and functional MRI scans. *Eur. J. Pediatr.* 169, 1079–1085. doi:10.1007/s00431-010-1181-z

De la Fuente, J. M., and Berry, C. C. J. B. c. (2005). Tat peptide as an efficient molecule to translocate gold nanoparticles into the cell nucleus. *Bioconjug. Chem.* 16, 1176–1180. doi:10.1021/bc050033+

de Pádua Oliveira, D. C., de Barros, A. L. B., Belardi, R. M., de Goes, A. M., de Oliveira Souza, B. K., and Soares, D. C. F. J. J. o. D. D. S. (2016). Mesoporous silica nanoparticles as a potential vaccine adjuvant against Schistosoma mansoni. *J. Drug Deliv. Sci. Technol.* 35, 234–240. doi:10.1016/j.jddst.2016.07.002

Dehaini, D., Fang, R. H., Zhang, L. J. B., and medicine, t. (2016). Biomimetic strategies for targeted nanoparticle delivery. *Bioeng. Transl. Med.* 1, 30–46. doi:10. 1002/btm2.10004

Demetzos, C., and Pippa, N. J. D. d. (2014). Advanced drug delivery nanosystems (aDDnSs): A mini-review. *Drug Deliv.* 21, 250–257. doi:10.3109/10717544.2013.844745

Deng, X., Wang, X., and Andersson, R. J. J. o. A. P. (1995). Endothelial barrier resistance in multiple organs after septic and nonseptic challenges in the rat. *J. Appl. Physiol.* 78, 2052–2061. doi:10.1152/jappl.1995.78.6.2052

Dixit, S., Novak, T., Miller, K., Zhu, Y., Kenney, M. E., and Broome, A.-M. J. N. (2015). Transferrin receptor-targeted theranostic gold nanoparticles for photosensitizer delivery in brain tumors. *Nanoscale* 7, 1782–1790. doi:10.1039/ c4nr04853a

Dominguez, A., Álvarez, A., Hilario, E., Suarez-Merino, B., and Goni-de-Cerio, F. (2013). Central nervous system diseases and the role of the blood-brain barrier in their treatment. *Neurosci. Discov.* 1, 3. doi:10.7243/2052-6946-1-3

Domínguez, A., Álvarez, A., Hilario, E., Suarez-Merino, B., and Goñi-de-Cerio, F. J. N. D. (2013). Central nervous system diseases and the role of the blood-brain barrier in their treatment. *Neurosci. Discov.* 1, 3. doi:10.7243/2052-6946-1-3

Dominguez, A., Alvarez, A., Suarez-Merino, B., and Goni-de-Cerio, F. J. R. d. N. (2014). Neurological disorders and the blood-brain barrier. Strategies and limitations for drug delivery to the brain. *Rev. Neurol.* 58, 213–224.

Dong, X., Gao, J., Su, Y., and Wang, Z. J. I. J. o. M. S. (2020). Nanomedicine for ischemic stroke. *Int. J. Mol. Sci.* 21, 7600. doi:10.3390/ijms21207600

Dong, X. J. T. (2018). Current strategies for brain drug delivery. *Theranostics* 8, 1481–1493. doi:10.7150/thno.21254

Doran, A., Obach, R. S., Smith, B. J., Hosea, N. A., Becker, S., Callegari, E., et al. (2005). The impact of P-glycoprotein on the disposition of drugs targeted for indications of the central nervous system: Evaluation using the mdr1a/1B knockout mouse model. *Drug Metab. Dispos.* 33, 165–174. doi:10.1124/dmd.104.001230

Doxil, B. J. J. C. R. (2012). The first FDA-approved nanodrug: lessons learned. J. Control Release 160, 117–134. doi:10.1016/j.jconrel.2012.03.020

D'Souza, G. G., and Weissig, V. J. O.-S. P. N. (2010). An introduction to subcellular and nanomedicine: Current trends and future developments, 1–13.

Duncan, R., and Gaspar, R. J. M. p. (2011). Nanomedicine(s) under the microscope. *Mol. Pharm.* 8, 2101–2141. doi:10.1021/mp200394t

Elenkov, I. J., Wilder, R. L., Chrousos, G. P., and Vizi, E. S. J. P. r. (2000). The sympathetic nerve—an integrative interface between two supersystems: The brain and the immune system. *Pharmacol. Rev.* 52, 595–638.

Elfenbein, H., Rosen, R., Stephens, S., Switzer, R., Smith, Y., Pare, J., et al. (2007). Cerebral ß-amyloid angiopathy in aged squirrel monkeys. *Histol. Histopathol.* 22 (2), 155–167. doi:10.14670/HH-22.155

Engelhardt, B., and Sorokin, L. (2009). The blood-brain and the blood-cerebrospinal fluid barriers: Function and dysfunction, seminars in immunopathology. Springer, 497–511.

Englund, G. (2005). Interindividual variability of drug transport proteins: Focus on intestinal pgp (ABCB1) and BCRP (ABCG2). *Acta Univ. Ups.*.

Erdlenbruch, B., Schinkhof, C., Kugler, W., Heinemann, D. E., Herms, J., Eibl, H., et al. (2003). Intracarotid administration of short-chain alkylglycerols for increased delivery of methotrexate to the rat brain. *Br. J. Pharmacol.* 139, 685–694. doi:10. 1038/sj.bjp.0705302

Estelrich, J., and Busquets, M. A. J. M. (2018). Iron oxide nanoparticles in photothermal therapy. *Molecules* 23, 1567. doi:10.3390/molecules23071567

Fakruddin, M., Hossain, Z., and Afroz, H. J. J. o. n. (2012). Prospects and applications of nanobiotechnology: A medical perspective. *J. Nanobiotechnology* 10, 1–8. doi:10.1186/1477-3155-10-31

Farokhzad, O. C., and Langer, R. J. A. n. (2009). Impact of nanotechnology on drug delivery. ACS Nano 3, 16–20. doi:10.1021/nn900002m

Fink, J. R., Muzi, M., Peck, M., and Krohn, K. A. J. J. o. N. M. (2015). Multimodality brain tumor imaging: MR imaging, PET, and PET/MR imaging. J. Nucl. Med. 56, 1554–1561. doi:10.2967/jnumed.113.131516

Fleckenstein, A. E., Volz, T. J., Riddle, E. L., Gibb, J. W., and Hanson, G. R. J. A. R. P. T. (2007). New insights into the mechanism of action of amphetamines. *Annu. Rev. Pharmacol. Toxicol.* 47, 681–698. doi:10.1146/annurev.pharmtox.47.120505. 105140

Flierl, A., Jackson, C., Cottrell, B., Murdock, D., Seibel, P., and Wallace, D. J. M. T. (2003). Targeted delivery of DNA to the mitochondrial compartment via import sequence-conjugated peptide nucleic acid. *Mol. Ther.* 7, 550–557. doi:10.1016/s1525-0016(03)00037-6

Fulekar, M. (2010). Nanotechnology: Importance and applications. New Delhi: IK International Pvt Ltd.

Furtado, D., Björnmalm, M., Ayton, S., Bush, A. I., Kempe, K., and Caruso, F. J. A. m. (2018). Overcoming the blood-brain barrier: The role of nanomaterials in treating neurological diseases. *Adv. Mat.* 30, 1801362. doi:10.1002/adma.201801362

Gabathuler, R. J. N. o. d. (2010). Approaches to transport therapeutic drugs across the blood-brain barrier to treat brain diseases. *Neurobiol. Dis.* 37, 48–57. doi:10. 1016/j.nbd.2009.07.028

García, J. J. S. S. U. d. E. S. (2009). Evaluación del Rendimiento de Extracción de Pectina en Aguas Mieles del Beneficiado de Café Procedentes del Desmusilaginado Mecánico.

Ghosal, S., Srivastava, R., Bhattacharya, S., and Debnath, P. J. P. m. (1974). The active principles of alhagi pseudalhagi: $\beta\text{-phenethylamine}$ and

tetrahydroisoquinoline bases. Planta Med. 26, 318-326. doi:10.1055/s-0028-1099394

Giner-Casares, J. J., Henriksen-Lacey, M., Coronado-Puchau, M., and Liz-Marzán, L. M. J. M. T. (2016). Inorganic nanoparticles for biomedicine: Where materials scientists meet medical research. *Mat. TodayKidlingt.* 19, 19–28. doi:10. 1016/j.mattod.2015.07.004

Gløgård, C., Hovland, R., Fossheim, S. L., Aasen, A. J., and Klaveness, J. J. J. o. t. C. S. (2000). Perkin Transactions 2, Synthesis and physicochemical characterisation of new amphiphilic gadolinium DO3A complexes as contrast agents for MRI, 1047–1052.

Glück, R., Mischler, R., Finkel, B., Que, J., Cryz, S., Jr, and Scarpa, B. J. T. L. (1994). Immunogenicity of new virosome influenza vaccine in elderly people. *Lancet* 344, 160–163. doi:10.1016/s0140-6736(94)92758-8

Greenblatt, D. J., Shader, R. I., and Abernethy, D. R. J. N. E. J. o. M. (1983). Drug therapy. Current status of benzodiazepines. *N. Engl. J. Med.* 309, 410–416. doi:10. 1056/NEJM198308183090705

Griffith, J. I., Rathi, S., Zhang, W., Zhang, W., Drewes, L. R., Sarkaria, J. N., et al. (2020). Addressing BBB heterogeneity: A new paradigm for drug delivery to brain tumors. *Pharmaceutics* 12, 1205. doi:10.3390/pharmaceutics12121205

Groner-Strauss, W., and Strauss, M. J. P. S. M., Divers face special peril in use/ abuse of drugs. 4 (1976) 30-36.

Guo, H.-C., Feng, X.-M., Sun, S.-Q., Wei, Y.-Q., Sun, D.-H., Liu, X.-T., et al. (2012). Immunization of mice by hollow mesoporous silica nanoparticles as carriers of porcine circovirus type 2 ORF2 protein. *Virol. J.* 9, 108–110. doi:10.1186/1743-422X-9-108

Gurunathan, S., Han, J., Park, J. H., and Kim, J.-H. J. N. r. l. (2014). A green chemistry approach for synthesizing biocompatible gold nanoparticles. *Nanoscale Res. Lett.* 9, 1–11. doi:10.1186/1556-276X-9-248

Halberstadt, C., Emerich, D. F., and Gonsalves, K. J. E. O. o. B. T. (2006). Combining cell therapy and nanotechnology. *Expert Opin. Biol. Ther.* 6, 971–981. doi:10.1517/14712598.6.10.971

Hanson, M. C., Abraham, W., Crespo, M. P., Chen, S. H., Liu, H., Szeto, G. L., et al. (2015). Liposomal vaccines incorporating molecular adjuvants and intrastructural T-cell help promote the immunogenicity of HIV membrane-proximal external region peptides. *Vaccine* 33, 861–868. doi:10.1016/j.vaccine. 2014.12.045

Haque, S., Sahni, S. M., Alam, M. I., Sahni, J. K., Ali, J., Baboota, S. J. D. D., et al. (2012). Nanostructure-based drug delivery systems for brain targeting. *Drug Dev. Ind. Pharm.* 38, 387–411. doi:10.3109/03639045.2011.608191

Haque, S., Kaur, S. M., Intekhab Alam, M., Kaur Sahni, J., Ali, J., and Baboota, S. J. R. P. O. N. (2011). Nanomedicines for brain targeting: A patent review. *Nanomedicines Brain Target. A Pat. Rev.* 1, 149–161. doi:10.2174/1877912311101020149

Hartley, D., Blumenthal, T., Carrillo, M., DiPaolo, G., Esralew, L., Gardiner, K., et al. (2015). Down syndrome and Alzheimer's disease: Common pathways, common goals. *Alzheimers Dement.* 11, 700–709. doi:10.1016/j.jalz.2014.10.007

Haseeb, Q., Hamdani, S. D. A., Akram, A., Khan, D. A., Rajput, T. A., and Babar, M. M. J. N. (2020). *Nanobiotechnology: Paving the way to personalized medicine*, 17–32.

Hassett, K. J., Meinerz, N. M., Semmelmann, F., Cousins, M. C., Garcea, R. L., Randolph, T. W., et al. (2015). Development of a highly thermostable, adjuvanted human papillomavirus vaccine. *Eur. J. Pharm. Biopharm.* 94, 220–228. doi:10.1016/ j.ejpb.2015.05.009

Heidarzadeh, M., Gürsoy-Özdemir, Y., Kaya, M., Eslami Abriz, A., Zarebkohan, A., Rahbarghazi, R., et al. (2021). Exosomal delivery of therapeutic modulators through the blood-brain barrier; promise and pitfalls. *Cell Biosci.* 11, 1–28. doi:10. 1186/s13578-021-00650-0

Henderson, M. X., Cornblath, E. J., Darwich, A., Zhang, B., Brown, H., Gathagan, R. J., et al. (2019). Spread of α-synuclein pathology through the brain connectome is modulated by selective vulnerability and predicted by network analysis. *Nat. Neurosci.* 22, 1248–1257. doi:10.1038/s41593-019-0457-5

Heng, J. B., Aksimentiev, A., Ho, C., Marks, P., Grinkova, Y. V., Sligar, S., et al. (2006). The electromechanics of DNA in a synthetic nanopore. *Biophys. J.* 90, 1098–1106. doi:10.1529/biophysj.105.070672

Henrich-Noack, P., Nikitovic, D., Neagu, M., Docea, A. O., Engin, A. B., Gelperina, S., et al. (2019). Biology, and Medicine, the blood-brain barrier and beyond: Nano-based neuropharmacology and the role of extracellular matrix. *Nanomedicine* 17, 359–379.

Henschel, O., Gipson, K. E., Bordey, A. J. C., and Targets, N. D.-D. (2008). GABAA receptors, anesthetics and anticonvulsants in brain development. CNS Neurol. Disord. Drug Targets 7, 211–224. doi:10.2174/187152708784083812 Hillyer, J. F., and Albrecht, R. M. J. J. o. p. s. (2001). Gastrointestinal persorption and tissue distribution of differently sized colloidal gold nanoparticles. *J. Pharm. Sci.* 90, 1927–1936. doi:10.1002/jps.1143

Hirschberg, H., Uzal, F. A., Chighvinadze, D., Zhang, M. J., Peng, Q., and Madsen, S. J. I. L. i. S. (2008). Medicine, and Surgery, Disruption of the blood-brain barrier following ALA-mediated photodynamic therapy. *Lasers Surg. Med.* 40, 535–542. doi:10.1002/lsm.20670

Hoffman, R. M. J. N. R. C. (2015). Patient-derived orthotopic xenografts: Better mimic of metastasis than subcutaneous xenografts. *Nat. Rev. Cancer* 15, 451–452. doi:10.1038/nrc3972

Holmes, D. J. T. L. N. (2013). The next big things are tiny. Lancet. Neurol. 12, 31-32. doi:10.1016/S1474-4422(12)70313-7

Houston, Z. H., Bunt, J., Chen, K.-S., Puttick, S., Howard, C. B., Fletcher, N. L., et al. (2020). Understanding the uptake of nanomedicines at different stages of brain cancer using a modular nanocarrier platform and precision bispecific antibodies. *ACS Cent. Sci.* 6, 727–738. doi:10.1021/acscentsci.9b01299

Hsiung, S. c., Adlersberg, M., Arango, V., Mann, J. J., Tamir, H., and Liu, K. p. J. J. o. n. (2003). Attenuated 5-ht1a receptor signaling in brains of suicide victims: Involvement of adenylyl cyclase, phosphatidylinositol 3-kinase, akt and mitogenactivated protein kinase. *J. Neurochem.* 87, 182–194. doi:10.1046/j.1471-4159.2003. 01987.x

Hu, C.-M. J., Zhang, L., Aryal, S., Cheung, C., Fang, R. H., and Zhang, L. J. P. o. t. N. A. o. S. (2011). Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. *Proc. Natl. Acad. Sci. U. S. A.* 108, 10980–10985. doi:10.1073/pnas.1106634108

Humpel, C. J. N. (2015). Organotypic brain slice cultures: A review. *Neuroscience* 305, 86–98. doi:10.1016/j.neuroscience.2015.07.086

Inamura, T., and Black, K. L. (1994). Bradykinin selectively opens blood-tumor barrier in experimental brain tumors. *J. Cereb. Blood Flow. Metab.* 14, 862–870. doi:10.1038/jcbfm.1994.108

Inamura, T., Nomura, T., Bartus, R. T., and Black, K. L. (1994). Intracarotid infusion of RMP-7, a bradykinin analog: A method for selective drug delivery to brain tumors. *J. Neurosurg.* 81, 752–758. doi:10.3171/jns.1994.81.5.0752

Invernici, G., Cristini, S., Alessandri, G., E Navone, S., Canzi, L., Tavian, D., et al. (2011). Nanotechnology advances in brain tumors: The state of the art. *Recent Pat. anticancer. Drug Discov.* 6, 58–69. doi:10.2174/157489211793979990

Invernizzi, N. (2010). Science policy and social inclusion: Advances and limits of brazilian nanotechnology policy, nanotechnology and the challenges of equity, equality and development. Springer, 291–307.

Jain, K. J. N. D. (2007). Nanobiotechnology-based drug delivery to the central nervous system. *Neurodegener. Dis.* 4, 287–291. doi:10.1159/000101884

Jain, K. K. (2008). Nanomedicine: Application of nanobiotechnology in medical practice. *Med. Princ. Pract.* 17, 89–101. doi:10.1159/000112961

Janssen, P. A. J. I. R. o. N. (1965). The evolution of the butyrophenones, haloperidol and trifluperidol, from meperidine-like 4-phenylpiperidines. *Int. Rev. Neurobiol.* 8, 221–263. doi:10.1016/s0074-7742(08)60759-x

Johnstone, T. C., Suntharalingam, K., and Lippard, S. J. J. C. r. (2016). The next generation of platinum drugs: Targeted Pt (II) agents, nanoparticle delivery, and Pt (IV) prodrugs. *Chem. Rev.* 116, 3436–3486. doi:10.1021/acs. chemrev.5b00597

Joshi, A. S., Singh, V., Gahane, A., and Thakur, A. K. J. A. C. N. (2018). Biodegradable nanoparticles containing mechanism based peptide inhibitors reduce polyglutamine aggregation in cell models and alleviate motor symptoms in a Drosophila model of Huntington's disease. *ACS Chem. Neurosci.* 10, 1603–1614. doi:10.1021/acschemneuro.8b00545

Joshi, S., Meyers, P. M., and Ornstein, E. J. T. J. o. t. A. S. o. A. (2008). Intracarotid delivery of drugs: The potential and the pitfalls. *Anesthesiology* 109, 543–564. doi:10. 1097/ALN.0b013e318182c81b

Jung, J., Kim, M. A., Cho, J.-H., Lee, S. J., Yang, I., Cho, J., et al. (2012). Europiumdoped gadolinium sulfide nanoparticles as a dual-mode imaging agent for T1weighted MR and photoluminescence imaging. *Biomaterials* 33, 5865–5874. doi:10. 1016/j.biomaterials.2012.04.059

Kang, B., Mackey, M. A., and El-Sayed, M. A. J. J. o. t. A. C. S. (2010). Nuclear targeting of gold nanoparticles in cancer cells induces DNA damage, causing cytokinesis arrest and apoptosis. *J. Am. Chem. Soc.* 132, 1517–1519. doi:10.1021/ja9102698

Karamanos, N. K., Piperigkou, Z., Theocharis, A. D., Watanabe, H., Franchi, M., Baud, S., et al. (2018). Proteoglycan chemical diversity drives multifunctional cell regulation and therapeutics. *Chem. Rev.* 118, 9152–9232. doi:10.1021/acs.chemrev.8b00354

Karch, C. M., and Goate, A. M. J. B. p. (2015). Alzheimer's disease risk genes and mechanisms of disease pathogenesis. *Biol. Psychiatry* 77, 43–51. doi:10.1016/j. biopsych.2014.05.006

Karim, R., Palazzo, C., Evrard, B., and Piel, G. J. J. o. c. r. (2016). Nanocarriers for the treatment of glioblastoma multiforme: Current state-of-the-art. *J. Control. Release* 227, 23–37. doi:10.1016/j.jconrel.2016.02.026

Kaur, I. P., Bhandari, R., Bhandari, S., and Kakkar, V. J. J. o. C. r. (2008). Potential of solid lipid nanoparticles in brain targeting. *J. Control. Release* 127, 97–109. doi:10. 1016/j.jconrel.2007.12.018

Khan, W. S., Hamadneh, N. N., Khan, W. A. J. S., and Nanostructures, a. o. T. (2016). Polymer nanocomposites-synthesis techniques, classification and properties, 50.

Khanbabaie, R., and Jahanshahi, M. J. C. n. (2012). Revolutionary impact of nanodrug delivery on neuroscience. *Curr. Neuropharmacol.* 10, 370–392. doi:10. 2174/157015912804143513

Khawar, I. A., Kim, J. H., and Kuh, H.-J. J. J. o. C. R. (2015). Improving drug delivery to solid tumors: Priming the tumor microenvironment. *J. Control. Release* 201, 78–89. doi:10.1016/j.jconrel.2014.12.018

Klaassen, I., Van Noorden, C. J., Schlingemann, R. O. J. P. i. r., and research, e. (2013). Molecular basis of the inner blood-retinal barrier and its breakdown in diabetic macular edema and other pathological conditions. *Prog. Retin. Eye Res.* 34, 19–48. doi:10.1016/j.preteyeres.2013.02.001

Kopelman, R., Koo, Y.-E. L., Philbert, M., Moffat, B. A., Reddy, G. R., McConville, P., et al. (2005). Multifunctional nanoparticle platforms for *in vivo* MRI enhancement and photodynamic therapy of a rat brain cancer. *J. Magn. Magn. Mat.* 293, 404–410. doi:10.1016/j.jmmm.2005.02.061

Kreuter, J. J. J. o. m. (2013). Mechanism of polymeric nanoparticle-based drug transport across the blood-brain barrier (BBB). *J. Microencapsul.* 30, 49–54. doi:10. 3109/02652048.2012.692491

Langmead, C. J., Watson, J., and Reavill, C. J. P. (2008). Muscarinic acetylcholine receptors as CNS drug targets. *Pharmacol. Ther.* 117, 232–243. doi:10.1016/j. pharmthera.2007.09.009

LeClaire, M. J. (2021). Biophysical characterization of cancer-derived cells and extracellular vesicles. Los Angeles: University of California.

Lee, H., Park, J., Yoon, O., Kim, H. J. N. N., Lee do, Y., Kim do, H., et al. (2011). Amine-modified single-walled carbon nanotubes protect neurons from injury in a rat stroke model. *Nat. Nanotechnol.* 6, 121–125. doi:10.1038/nnano.2010.281

Lee, S., Xie, J., and Chen, X. J. C. t. i. m. c. (2010). Activatable molecular probes for cancer imaging. *Curr. Top. Med. Chem.* 10, 1135–1144. doi:10.2174/156802610791384270

Lee, W. T., Lee, J., Kim, H., Nguyen, N. T., Lee, E. S., Oh, K. T., et al. (2021). Photoreactive-proton-generating hyaluronidase/albumin nanoparticles-loaded PEG-hydrogel enhances antitumor efficacy and disruption of the hyaluronic acid extracellular matrix in AsPC-1 tumors. *Mat. Today. Bio* 12, 100164. doi:10. 1016/j.mtbio.2021.100164

Lemley, M. A. J. S. L. R. (2005). Patenting nanotechnology. Stanf. Law Rev. 58, 601-630.

Levin, V. A., Fenstermacher, J. D., and Patlak, C. S. J. A. J. o. P.-L. C. (1970). Sucrose and inulin space measurements of cerebral cortex in four mammalian species. *Am. J. Physiol.* 219, 1528–1533. doi:10.1152/ajplegacy. 1970.219.5.1528

Liu, F., Hon, G. C., Villa, G. R., Turner, K. M., Ikegami, S., Yang, H., et al. (2015). EGFR mutation promotes glioblastoma through epigenome and transcription factor network remodeling. *Mol. Cell* 60, 307–318. doi:10.1016/j. molcel.2015.09.002

Liu, J., Wen, J., Zhang, Z., Liu, H., and Sun, Y. (2015). Voyage inside the cell: Microsystems and nanoengineering for intracellular measurement and manipulation. *Microsyst. Nanoeng.* 1, 861–868. doi:10.1038/micronano. 2015.20

Liu, Y., Luo, J., Chen, X., Liu, W., and Chen, T. J. N.-M. L. (2019). Cell membrane coating technology: A promising strategy for biomedical applications. *Nanomicro. Lett.* 11, 100–146. doi:10.1007/s40820-019-0330-9

Liu, Y., Xu, D., Liu, Y., Zheng, X., Zang, J., Ye, W., et al. (2022). Remotely boosting hyaluronidase activity to normalize the hypoxic immunosuppressive tumor microenvironment for photothermal immunotherapy. *Biomaterials* 284, 121516. doi:10.1016/j.biomaterials.2022.121516

Livingston, M. G., and Livingston, H. M. J. D. s. (1996). Monoamine oxidase inhibitors. An update on drug interactions. *Drug Saf.* 14, 219–227. doi:10.2165/00002018-199614040-00002

Lohse, S. E., and Murphy, C. J. J. J. o. t. A. C. S. (2012). Applications of colloidal inorganic nanoparticles: From medicine to energy. J. Am. Chem. Soc. 134, 15607–15620. doi:10.1021/ja307589n

Lorke, D., Kalasz, H., Petroianu, G., and Tekes, K. J. C. m. c. (2008). Entry of oximes into the brain: A review. *Curr. Med. Chem.* 15, 743–753. doi:10.2174/092986708783955563

Lorkowski, M. E., Atukorale, P. U., Ghaghada, K. B., and Karathanasis, E. J. A. h. m. (2021). Stimuli-responsive iron oxide nanotheranostics: A versatile and powerful approach for cancer therapy. *Adv. Healthc. Mater* 10, 2001044. doi:10.1002/adhm.202001044

Löscher, W., and Potschka, H. J. P. i. n. (2005). Role of drug efflux transporters in the brain for drug disposition and treatment of brain diseases. *Prog. Neurobiol.* 76, 22–76. doi:10.1016/j.pneurobio.2005.04.006

Louis, D. N., Perry, A., Reifenberger, G., Von Deimling, A., Figarella-Branger, D., Cavenee, W. K., et al. (2016). The 2016 world health organization classification of tumors of the central nervous system: A summary. *Acta Neuropathol.* 131, 803–820. doi:10.1007/s00401-016-1545-1

Lu, H., Wu, L., Wang, J., Wang, Z., Yi, X., Wang, J., et al. (2018). Voltammetric determination of the Alzheimer's disease-related ApoE 4 gene from unamplified genomic DNA extracts by ferrocene-capped gold nanoparticles. *Mikrochim. Acta* 185, 1–7. doi:10.1007/s00604-018-3087-9

Luk, B. T., and Zhang, L. J. J. o. C. R. (2015). Cell membrane-camouflaged nanoparticles for drug delivery. *J. Control. Release* 220, 600–607. doi:10.1016/j. jconrel.2015.07.019

Mahon, E., Salvati, A., Bombelli, F. B., Lynch, I., and Dawson, K. A. J. J. o. C. R. (2012). Designing the nanoparticle-biomolecule interface for "targeting and therapeutic delivery". *J. Control. Release* 161, 164–174. doi:10.1016/j.jconrel. 2012.04.009

Martel, S. J. B. (2016). Swimming microorganisms acting as nanorobots versus artificial nanorobotic agents: A perspective view from an historical retrospective on the future of medical nanorobotics in the largest known three-dimensional biomicrofluidic networks. *Biomicrofluidics* 10, 021301. doi:10.1063/1.4945734

Martin, L., Smith, T., Weiler-Ravell, D., Mcdonough, J., Barutt, J., and Saffron, J. J. T. N. E. j. o. m. (1992). The medical problems of underwater diving. *N. Engl. J. Med.* 326, 1497–1498.

Martins, S., Sarmento, B., Ferreira, D. C., and Souto, E. B. J. I. j. o. n. (2007). Lipidbased colloidal carriers for peptide and protein delivery–liposomes versus lipid nanoparticles. *Int. J. Nanomedicine* 2, 595–607.

Masserini, M. J. I. S. R. N. (2013). Nanoparticles for brain drug delivery, 2013.

McManus, M. J., Murphy, M. P., and Franklin, J. L. J. J. o. N. (2011). The mitochondria-targeted antioxidant MitoQ prevents loss of spatial memory retention and early neuropathology in a transgenic mouse model of Alzheimer's disease. J. Neurosci. 31, 15703–15715. doi:10.1523/JNEUROSCI.0552-11.2011

Miele, D., Catenacci, L., Sorrenti, M., Rossi, S., Sandri, G., Malavasi, L., et al. (2019). Chitosan oleate coated polylactic-glycolic acid (PLGA) nanoparticles versus chitosan oleate self-assembled polymeric micelles, loaded with resveratrol. *Mar. Drugs* 17, 515. doi:10.3390/md17090515

Miller, D. S. J. T. i. p. s. (2010). Regulation of P-glycoprotein and other ABC drug transporters at the blood-brain barrier. *Trends Pharmacol. Sci.* 31, 246–254. doi:10. 1016/j.tips.2010.03.003

Miranda, A., Blanco-Prieto, M., Sousa, J., Pais, A., and Vitorino, C. J. I. j. o. p. (2017). Breaching barriers in glioblastoma. Part I: Molecular pathways and novel treatment approaches. *Int. J. Pharm.* 531, 372–388. doi:10.1016/j.ijpharm.2017. 07.056

Misra, A., Ganesh, S., Shahiwala, A., and Shah, S. P. J. J. P. P. S. (2003). Drug delivery to the central nervous system: A review. J. Pharm. Pharm. Sci. 6, 252–273.

Miyai, M., Tomita, H., Soeda, A., Yano, H., Iwama, T., and Hara, A. J. J. o. n.-o. (2017). Current trends in mouse models of glioblastoma. *J. Neurooncol.* 135, 423-432. doi:10.1007/s11060-017-2626-2

Modi, G., Pillay, V., and Choonara, Y. E. J. A. o. t. N. Y. A. o. S. (2010). Advances in the treatment of neurodegenerative disorders employing nanotechnology. *Ann. N. Y. Acad. Sci.* 1184, 154–172. doi:10.1111/j.1749-6632.2009.05108.x

Modi, G., Pillay, V., Choonara, Y. E., Ndesendo, V. M., du Toit, L. C., and Naidoo, D. J. P. i. N. (2009). Nanotechnological applications for the treatment of neurodegenerative disorders. *Prog. Neurobiol.* 88, 272–285. doi:10.1016/j. pneurobio.2009.05.002

Mohamed, F., and van der Walle, C. F. J. J. o. p. s. (2008). Engineering biodegradable polyester particles with specific drug targeting and drug release properties. J. Pharm. Sci. 97, 71–87. doi:10.1002/jps.21082

Montet, X., Funovics, M., Montet-Abou, K., Weissleder, R., and Josephson, L. J. J. o. m. c. (2006). Multivalent effects of RGD peptides obtained by nanoparticle display. *J. Med. Chem.* 49, 6087–6093. doi:10.1021/jm060515m

Morrow, K. J., Jr, Bawa, R., and Wei, C. J. M. C. o. N. A. (2007). Recent advances in basic and clinical nanomedicine. *Med. Clin. North Am.* 91, 805–843. doi:10.1016/ j.mcna.2007.05.009

Muhammad, G., Hussain, M. A., Anwar, F., Ashraf, M., and Gilani, A. H. J. P. r. (2015). Alhagi: A plant genus rich in bioactives for pharmaceuticals. *Phytother. Res.* 29, 1–13. doi:10.1002/ptr.5222 Mulvihill, J. J., Cunnane, E. M., Ross, A. M., Duskey, J. T., Tosi, G., and Grabrucker, A. M. J. N. (2020). Drug delivery across the blood-brain barrier: Recent advances in the use of nanocarriers. *Nanomedicine* 15, 205–214. doi:10. 2217/nnm-2019-0367

Myers, R. H. J. N. (2004). Huntington's disease genetics. *NeuroRx* 1, 255–262. doi:10.1602/neurorx.1.2.255

Naik, P., and Cucullo, L. J. J. o. p. s. (2012). *In vitro* blood-brain barrier models: Current and perspective technologies. *J. Pharm. Sci.* 101, 1337–1354. doi:10.1002/ jps.23022

Nair, K. G., Ramaiyan, V., and Sukumaran, S. K. J. I. (2018). Enhancement of drug permeability across blood brain barrier using nanoparticles in meningitis. *Inflammopharmacology* 26, 675–684. doi:10.1007/s10787-018-0468-y

Nance, E. A., Woodworth, G. F., Sailor, K. A., Shih, T.-Y., Xu, Q., Swaminathan, G., et al. (2012). A dense poly (ethylene glycol) coating improves penetration of large polymeric nanoparticles within brain tissue. *Sci. Transl. Med.* 4, 149ra119.

Nance, E. J. A. d. d. r. (2019). Careers in nanomedicine and drug delivery. *Adv. Drug Deliv. Rev.* 144, 180–189. doi:10.1016/j.addr.2019.06.009

Naqvi, S., Panghal, A., and Flora, S. J. F. i. n. (2020). Nanotechnology: A promising approach for delivery of neuroprotective drugs. *Front. Neurosci.* 14, 494. doi:10.3389/fnins.2020.00494

Nasrollahzadeh, M., Sajadi, S. M., Sajjadi, M., and Issaabadi, Z. (2019). An introduction to nanotechnology, Interface science and technology. Elsevier, 1–27.

Natarajan, S., Harini, K., Gajula, G. P., Sarmento, B., Neves-Petersen, M. T., and Thiagarajan, V. J. B. M. (2019). Multifunctional magnetic iron oxide nanoparticles: Diverse synthetic approaches, surface modifications. *Cytotox. towards Biomed. industrial Appl.* 1, 1–22.

Naz, S., Shamoon, M., Wang, R., Zhang, L., Zhou, J., and Chen, J. J. I. J. o. M. S. (2019). Advances in therapeutic implications of inorganic drug delivery nanoplatforms for cancer. *Int. J. Mol. Sci.* 20, 965. doi:10.3390/ijms20040965

Nazem, A., and Mansoori, G. A. J. J. o. A. s. d. (2008). Nanotechnology solutions for alzheimer's disease: Advances in research tools, diagnostic methods and therapeutic agents. *J. Alzheimers Dis.* 13, 199–223. doi:10.3233/jad-2008-13210

Nelson, N. R., Port, J. D., and Pandey, M. K. J. J. o. N. (2020). Use of superparamagnetic iron oxide nanoparticles (SPIONs) via multiple imaging modalities and modifications to reduce cytotoxicity: An educational review. *J. Nanotheranostics* 1, 105–135. doi:10.3390/jnt1010008

Neuwelt, E. A., Bauer, B., Fahlke, C., Fricker, G., Iadecola, C., Janigro, D., et al. (2011). Engaging neuroscience to advance translational research in brain barrier biology. *Nat. Rev. Neurosci.* 12, 169–182. doi:10.1038/nrn2995

Neuwelt, E. A., Goldman, D. L., Dahlborg, S. A., Crossen, J., Ramsey, F., Roman-Goldstein, S., et al. (1991). Primary CNS lymphoma treated with osmotic bloodbrain barrier disruption: Prolonged survival and preservation of cognitive function. *J. Clin. Oncol.* 9, 1580–1590. doi:10.1200/JCO.1991.9.9.1580

Niikura, K., Matsunaga, T., Suzuki, T., Kobayashi, S., Yamaguchi, H., Orba, Y., et al. (2013). Gold nanoparticles as a vaccine platform: Influence of size and shape on immunological responses *in vitro* and *in vivo*. ACS Nano 7, 3926–3938. doi:10. 1021/nn3057005

Niu, X., Chen, J., and Gao, J. J. A. j. o. p. s. (2019). Nanocarriers as a powerful vehicle to overcome blood-brain barrier in treating neurodegenerative diseases: Focus on recent advances. *Asian J. Pharm. Sci.* 14, 480–496. doi:10.1016/j.ajps.2018. 09.005

Nurhidayah, D., Maruf, A., Zhang, X., Liao, X., Wu, W., and Wang, G. J. N. (2019). Advanced drug-delivery systems: Mechanoresponsive nanoplatforms applicable in atherosclerosis management. *Nanomedicine* 14, 3105–3122. doi:10. 2217/nnm-2019-0172

Oberdörster, G., Sharp, Z., Atudorei, V., Elder, A., Gelein, R., Kreyling, W., et al. (2004). Translocation of inhaled ultrafine particles to the brain. *Inhal. Toxicol.* 16, 437–445. doi:10.1080/08958370490439597

Oldrini, B., Curiel-García, Á., Marques, C., Matia, V., Uluçkan, Ö., Graña-Castro, O., et al. (2018). Somatic genome editing with the RCAS-TVA-CRISPR-Cas9 system for precision tumor modeling. *Nat. Commun.* 9, 1–16. doi:10.1038/s41467-018-03731-w

Orive, G., Ali, O., Anitua, E., Pedraz, J., and Emerich, D. J. B. e. B. A.-R. o. C. (2010). Biomaterial-based technologies for brain anti-cancer therapeutics and imaging. *Biochim. Biophys. Acta* 1806, 96–107. doi:10.1016/j.bbcan.2010.04.001

Ozkizilcik, A., Davidson, P., Turgut, H., Sharma, H. S., Sharma, A., and Tian, Z. R. (2017). Nanocarriers as cns drug delivery systems for enhanced neuroprotection, drug and gene delivery to the central nervous system for neuroprotection. Springer, 33–55.

Palei, N. N., Mohanta, B. C., Sabapathi, M. L., and Das, M. K. (2018). Organic materials as smart nanocarriers for drug delivery. Elsevier, 415–470.Lipid-based nanoparticles for cancer diagnosis and therapy

Pandey, P. K., Sharma, A. K., and Gupta, U. J. T. b. (2016). Blood brain barrier: An overview on strategies in drug delivery, realistic *in vitro* modeling and *in vivo* live tracking, *Tissue Barriers* 4, e1129476. doi:10.1080/21688370.2015.1129476

Pansieri, J., Gerstenmayer, M., Lux, F., Mériaux, S., Tillement, O., Forge, V., et al. (2018). Magnetic nanoparticles applications for amyloidosis study and detection: A review. *Nanomaterials* 8, 740. doi:10.3390/nano8090740

Pardridge, W. M. (1997). Drug delivery to the brain. J. Cereb. Blood Flow. Metab. 17, 713-731. doi:10.1097/00004647-199707000-00001

Pardridge, W. M. J. J. o. n. (1998). CNS drug design based on principles of bloodbrain barrier transport. J. Neurochem. 70, 1781–1792. doi:10.1046/j.1471-4159. 1998.70051781.x

Pardridge, W. M. J. A. d. d. r. (1999). Vector-mediated drug delivery to the brain. *Adv. Drug Deliv. Rev.* 36, 299-321. doi:10.1016/s0169-409x(98)00087-8

Pardridge, W. M. J. A. d. d. r. (2007). shRNA and siRNA delivery to the brain. *Adv. Drug Deliv. Rev.* 59, 141-152. doi:10.1016/j.addr.2007.03.008

Park, J. H., Lee, S., Kim, J.-H., Park, K., Kim, K., and Kwon, I. C. J. P. i. p. s. (2008). Polymeric nanomedicine for cancer therapy. *Prog. Polym. Sci.* 33, 113–137. doi:10. 1016/j.progpolymsci.2007.09.003

Parveen, S., Misra, R., and Sahoo, S. J. B. (2012). Nanoparticles: A boon to drug delivery, therapeutics, diagnostics and imaging. *Nanomedicine* 8, 147–166. doi:10. 1016/j.nano.2011.05.016

Parveen, S., Misra, R., and Sahoo, S. K. J. N. N. (2012). Nanoparticles: A boon to drug delivery, therapeutics, diagnostics and imaging. *Nanomedicine* 8, 147–166. doi:10.1016/j.nano.2011.05.016

Patois, E., Capelle, M., Palais, C., Gurny, R., and Arvinte, T. J. J. o. d. d. s. (2012). Evaluation of nanoparticle tracking analysis (NTA) in the characterization of therapeutic antibodies and seasonal influenza vaccines: Pros and cons. J. Drug Deliv. Sci. Technol. 22, 427–433. doi:10.1016/s1773-2247(12)50069-9

Pereira, S., Pinto, A., Alves, V., and Silva, C. A. J. I. t. o. m. i. (2016). Brain tumor segmentation using convolutional neural networks in MRI images. *IEEE Trans. Med. Imaging* 35, 1240–1251. doi:10.1109/tmi.2016.2538465

Petkar, K. C., Chavhan, S. S., Agatonovik-Kustrin, S., and Sawant, K. J. C. R. i. T. D. C. S. (2011). Nanostructured materials in drug and gene delivery: A review of the state of the art. *Crit. Rev. Ther. Drug Carr. Syst.* 28. doi:10.1615/ critrevtherdrugcarriersyst.v28.i2.10

Pietroiusti, A., Campagnolo, L., and Fadeel, B. J. S. (2013). Interactions of engineered nanoparticles with organs protected by internal biological barriers. *Small* 9, 1557–1572. doi:10.1002/smll.201201463

Poon, C., Gallo, J., Joo, J., Chang, T., Bañobre-López, M., and Chung, E. J. J. J. o. n. (2018). Hybrid, metal oxide-peptide amphiphile micelles for molecular magnetic resonance imaging of atherosclerosis. *J. Nanobiotechnology* 16, 92–11. doi:10.1186/s12951-018-0420-8

Poon, C., and Patel, A. A. J. N. E. (2020). Organic and inorganic nanoparticle vaccines for prevention of infectious diseases. *Nano Express* 1, 012001.

Popli, D., Anil, V., Subramanyam, A. B., Mn, N., Vr, R., Rao, S. N., et al. (2018). Endophyte fungi, Cladosporium species-mediated synthesis of silver nanoparticles possessing *in vitro* antioxidant, anti-diabetic and anti-Alzheimer activity. *Artif. Cells Nanomed. Biotechnol.* 46, 676–683. doi:10.1080/21691401.2018.1434188

Provenzale, J., and Silva, G. J. A. j. o. n. (2009). Uses of nanoparticles for central nervous system imaging and therapy. *AJNR. Am. J. Neuroradiol.* 30, 1293–1301. doi:10.3174/ajnr.A1590

Pusic, K., Aguilar, Z., McLoughlin, J., Kobuch, S., Xu, H., Tsang, M., et al. (2013). Iron oxide nanoparticles as a clinically acceptable delivery platform for a recombinant blood-stage human malaria vaccine. *FASEB J.* 27, 1153–1166. doi:10.1096/fj.12-218362

Qiao, R., Yang, C., and Gao, M. J. J. o. M. C. (2009). Superparamagnetic iron oxide nanoparticles: From preparations to *in vivo* MRI applications. *J. Mat. Chem.* 19, 6274–6293. doi:10.1039/b902394a

Quan, L., Gu, J., Lin, W., Wei, Y., Lin, Y., Liu, L., et al. (2019). A BODIPY biosensor to detect and drive self-assembly of diphenylalanine. *Chem. Commun.* 55, 8564–8566. doi:10.1039/c9cc03810h

Rai, S. N., Tiwari, N., Singh, P., Mishra, D., Singh, A. K., Hooshmandi, E., et al. (2021). Therapeutic potential of vital transcription factors in alzheimer's and Parkinson's disease with particular emphasis on transcription factor eb mediated autophagy. *Front. Neurosci.* 15, 777347. doi:10.3389/fnins.2021.777347

Raja, G., Jang, Y.-K., Suh, J.-S., Kim, H.-S., Ahn, S. H., and Kim, T.-J. J. C. (2020). Microcellular environmental regulation of silver nanoparticles in cancer therapy: A critical review. *Cancers* 12, 664. doi:10.3390/cancers12030664

Rajadhyaksha, M., Boyden, T., Liras, J., El-Kattan, A., and Brodfuehrer, J. J. C. d. d. t. (2011). Current advances in delivery of biotherapeutics across the blood-brain barrier. *Curr. Drug Discov. Technol.* 8, 87–101. doi:10.2174/157016311795563866

Ramanathan, R. (2012). Towards understanding the influence of physio-chemical environment on biological synthesis of inorganic materials. RMIT University.

Ramos-Cabrer, P., and Campos, F. J. I. (2013). Liposomes and nanotechnology in drug development: Focus on neurological targets. *Int. J. Nanomedicine* 8, 951. doi:10.2147/IJN.S30721

Rapoport, S. I. J. C., and neurobiology, m. (2000). Osmotic opening of the bloodbrain barrier: Principles, mechanism, and therapeutic applications. *Cell. Mol. Neurobiol.* 20, 217–230. doi:10.1023/a:1007049806660

Ratner, M. A., and Ratner, D. (2003). Nanotechnology: A gentle introduction to the next big idea. London: Prentice Hall Professional.

Re, F., Gregori, M., and Masserini, M. J. M. (2012). Nanotechnology for neurodegenerative disorders. *Maturitas* 73, 45–51. doi:10.1016/j.maturitas.2011. 12.015

Rezaei, M., Hosseini, S. N., Khavari-Nejad, R. A., Najafi, F., and Mahdavi, M. J. A. C. (2019). Nanomedicine, and Biotechnology, HBs antigen and mannose loading on the surface of iron oxide nanoparticles in order to immuno-targeting: Fabrication, characterization. *Cell. humoral Immunoass.* 47, 1543–1558.

Richard, C., Doan, B.-T., Beloeil, J.-C., Bessodes, M., Tóth, É., and Scherman, D. J. N. L. (2008). Noncovalent functionalization of carbon nanotubes with amphiphilic Gd3+ chelates: Toward powerful T1 and T2 MRI contrast agents. *Nano Lett.* 8, 232–236. doi:10.1021/nl072509z

Riehemann, K., Schneider, S. W., Luger, T. A., Godin, B., Ferrari, M., and Fuchs, H. J. A. C. I. E. (2009). Nanomedicine--challenge and perspectives. *Angew. Chem. Int. Ed. Engl.* 48, 872–897. doi:10.1002/anie.200802585

Rossini, P. M., Burke, D., Chen, R., Cohen, L., Daskalakis, Z., Di Iorio, R., et al. (2015). Non-invasive electrical and magnetic stimulation of the brain, spinal cord, roots and peripheral nerves: Basic principles and procedures for routine clinical and research application. An updated report from an I.F.C.N. Committee. *Clin. Neurophysiol.* 126, 1071–1107. doi:10.1016/j.clinph.2015.02.001

Sakhrani, N. M., and Padh, H. J. D. d. (2013). Organelle targeting: Third level of drug targeting. *Drug Des. devel. Ther.* 7, 585–599. doi:10.2147/DDDT.S45614

Sanati, M., Khodagholi, F., Aminyavari, S., Ghasemi, F., Gholami, M., Kebriaeezadeh, A., et al. (2019). Impact of gold nanoparticles on amyloid β -induced alzheimer's disease in a rat animal model: Involvement of STIM proteins. *ACS Chem. Neurosci.* 10, 2299–2309. doi:10.1021/acschemneuro.8b00622

Sancey, L., Lux, F., Kotb, S., Roux, S., Dufort, S., Bianchi, A., et al. (2014). The use of theranostic gadolinium-based nanoprobes to improve radiotherapy efficacy. *Br. J. Radiol.* 87, 20140134. doi:10.1259/bjr.20140134

Sanchez-Covarrubias, L., Slosky, L. M., Thompson, B. J., Davis, T. P., and Ronaldson, P. T. J. C. p. d. (2014). Transporters at CNS barrier sites: Obstacles or opportunities for drug delivery? *Curr. Pharm. Des.* 20, 1422–1449. doi:10.2174/ 13816128113199990463

Sangha, V., Williams, E. I., Ronaldson, P. T., Bendayan, R. J. D. T. M. C., and Disposition, R. i. D. (2022). Drug transport in the brain, 283-317.

Sanson, C., Diou, O., Thevenot, J., Ibarboure, E., Soum, A., Brûlet, A., et al. (2011). Doxorubicin loaded magnetic polymersomes: Theranostic nanocarriers for MR imaging and magneto-chemotherapy. *ACS Nano* 5, 1122–1140. doi:10.1021/nn102762f

Sarin, H., Kanevsky, A. S., Wu, H., Brimacombe, K. R., Fung, S. H., Sousa, A. A., et al. (2008). Effective transvascular delivery of nanoparticles across the blood-brain tumor barrier into malignant glioma cells. *J. Transl. Med.* 6, 80–15. doi:10.1186/1479-5876-6-80

Sarko, D., and McKinney, C. (2017). Exosomes: Origins and therapeutic potential for neurodegenerative disease. *Front. Neurosci.* 11, 82. doi:10.3389/fnins.2017. 00082

Saunders, N. R., Habgood, M. D., Møllgård, K., and Dziegielewska, K. M. J. F. (2016). The biological significance of brain barrier mechanisms: Help or hindrance in drug delivery to the central nervous system? *F1000Res*, 5. doi:10.12688/f1000research.7378.1

Schleh, C., Semmler-Behnke, M., Lipka, J., Wenk, A., Hirn, S., Schäffler, M., et al. (2012). Size and surface charge of gold nanoparticles determine absorption across intestinal barriers and accumulation in secondary target organs after oral administration. *Nanotoxicology* 6, 36–46. doi:10.3109/17435390.2011.552811

Schmidt, J., Metselaar, J. M., Wauben, M. H., Toyka, K. V., Storm, G., and Gold, R. J. B. (2003). Drug targeting by long-circulating liposomal glucocorticosteroids increases therapeutic efficacy in a model of multiple sclerosis. *Brain* 126, 1895–1904. doi:10.1093/brain/awg176

Schober, A. J. C., and research, t. (2004). Classic toxin-induced animal models of Parkinson's disease: 6-OHDA and MPTP. *Cell Tissue Res.* 318, 215–224. doi:10. 1007/s00441-004-0938-y

Setsuie, R., Wang, Y.-L., Mochizuki, H., Osaka, H., Hayakawa, H., Ichihara, N., et al. (2007). Dopaminergic neuronal loss in transgenic mice expressing the

Parkinson's disease-associated UCH-L1 I93M mutant. Neurochem. Int. 50, 119-129. doi:10.1016/j.neuint.2006.07.015

Shah, L., Yadav, S., Amiji, M. J. D. d., and research, t. (2013). Nanotechnology for CNS delivery of bio-therapeutic agents. *Drug Deliv. Transl. Res.* 3, 336–351. doi:10. 1007/s13346-013-0133-3

Sharma, A., Kaur, G. J. B. c., and medicine, a. (2018). Tinospora cordifolia as a potential neuroregenerative candidate against glutamate induced excitotoxicity: An *in vitro* perspective. *BMC Complement. Altern. Med.* 18, 1–17. doi:10.1186/s12906-018-2330-6

Sharma, H. S., Castellani, R. J., Smith, M. A., and Sharma, A. J. I. R. N. (2012). The blood-brain barrier in alzheimer's disease: Novel therapeutic targets and nanodrug delivery. *Int. Rev. Neurobiol.* 102, 47–90. doi:10.1016/B978-0-12-386986-9.00003-X

Sharma, P., and Bhargava, M. J. R. D. (2013). Applications and characteristics of nanomaterials in industrial environment. *Nanotechnol. Environ.* 3, 63–72. doi:10. 5772/intechopen.91438

Sharma, P., Mehta, M., Dhanjal, D. S., Kaur, S., Gupta, G., Singh, H., et al. (2019). Emerging trends in the novel drug delivery approaches for the treatment of lung cancer. *Chem. Biol. Interact.* 309, 108720. doi:10.1016/j.cbi.2019.06.033

Sharpe, M. A., Livingston, A. D., Gist, T. L., Ghosh, P., Han, J., and Baskin, D. S. J. E. (2015). Successful treatment of intracranial glioblastoma xenografts with a monoamine oxidase B-activated pro-drug. *EBioMedicine* 2, 1122–1132. doi:10.1016/j.ebiom.2015.08.013

Shukla, S. D., Jain, S., Sharma, K., and Bhatnagar, M. (2000). Stress induced neuron degeneration and protective effects of semecarpus anacardium linn. And withania somnifera dunn. In hippocampus of albino rats: An ultrastructural study.

Silva, G. A. J. B. (2008). Nanotechnology approaches to crossing the blood-brain barrier and drug delivery to the CNS. *BMC Neurosci.* 9, S4–S4. doi:10.1186/1471-2202-9-S3-S4

Simonet, S. p. (2018). Radiosensitizing effect of AGuIX[®] in head and neck squamous cell carcinoma (HNSCC): From cellular uptake to subcellular damage. Lyon: Université de Lyon.

Singh, A. K., Gothwal, A., Rani, S., Rana, M., Sharma, A. K., Yadav, A. K., et al. (2019). Dendrimer donepezil conjugates for improved brain delivery and better *in vivo* pharmacokinetics. *ACS Omega* 4, 4519–4529. doi:10.1021/acsomega.8b03445

Singh, A. K., Mishra, S. K., Mishra, G., Maurya, A., Awasthi, R., Yadav, M. K., et al. (2020). Inorganic clay nanocomposite system for improved cholinesterase inhibition and brain pharmacokinetics of donepezil. *Drug Dev. Ind. Pharm.* 46, 8–19. doi:10.1080/03639045.2019.1698594

Singh, A. K., Rai, S. N., Maurya, A., Mishra, G., Awasthi, R., Shakya, A., et al. (2021). Therapeutic potential of phytoconstituents in management of Alzheimer's disease. *Evid. Based Complement. Altern. Med.* 2021. doi:10.1155/2021/5578574

Singh, A. K., Singh, S. K., Nandi, M. K., Mishra, G., Maurya, A., Rai, A., et al. (2019). Berberine: A plant-derived alkaloid with therapeutic potential to combat alzheimer's disease. *Cent. Nerv. Syst. Agents Med. Chem.* 19, 154–170. doi:10.2174/1871524919666190820160053

Singh, A. K., Singh, S. S., Rathore, A. S., Singh, S. P., Mishra, G., Awasthi, R., et al. (2021). Lipid-coated MCM-41 mesoporous silica nanoparticles loaded with berberine improved inhibition of acetylcholine esterase and amyloid formation. ACS Biomater. Sci. Eng. 7, 3737–3753. doi:10.1021/acsbiomaterials.1c00514

Singh, A. V., Hosseinidoust, Z., Park, B.-W., Yasa, O., and Sitti, M. J. A. n. (2017). Microemulsion-based soft bacteria-driven microswimmers for active cargo delivery. *ACS Nano* 11, 9759–9769. doi:10.1021/acsnano.7b02082

Singh, G. K., Chauhan, S. K., Rai, G., Chatterjee, S. S., and Kumar, V. J. P. M. (2013). Potential antianxiety activity of fumaria indica: A preclinical study. *Pharmacogn. Mag.* 9, 14–22. doi:10.4103/0973-1296.108129

Singh, G. K., Rai, G., Chatterjee, S. S., and Kumar, V. J. A. (2013). Effects of ethanolic extract of Fumaria indica L. on rat cognitive dysfunctions. *Ayu* 34, 421–429. doi:10.4103/0974-8520.127727

Singh, P., Kim, Y.-J., Zhang, D., and Yang, D.-C. J. T. i. b. (2016). Biological synthesis of nanoparticles from plants and microorganisms. *Trends Biotechnol.* 34, 588–599. doi:10.1016/j.tibtech.2016.02.006

Singh, V., Yadav, B., and Pandey, V. J. P. (1999). Flavanone glycosides from Alhagi pseudalhagi. *Phytochemistry* 51, 587–590. doi:10.1016/s0031-9422(99)00010-2

Sj, R. D., and Kumar B, P. J. C. C.-A. D. D. (2020). *In silico* screening for antiinflammatory bioactive molecules from ayurvedic decoction, balaguluchyadi kashayam. *Curr. Comput. Aided. Drug Des.* 16, 435–450. doi:10.2174/ 1573409915666191015113753

Sonavane, G., Tomoda, K., Makino, K. J. C., and Biointerfaces, S. B. (2008). Biodistribution of colloidal gold nanoparticles after intravenous administration: Effect of particle size. *Colloids Surf. B Biointerfaces* 66, 274–280. doi:10.1016/j. colsurfb.2008.07.004 Song, Y., Jing, H., Vong, L. B., Wang, J., and Li, N. J. C. C. L. (2021). Recent advances in targeted stimuli-responsive nano-based drug delivery systems combating atherosclerosis. *Chin. Chem. Lett.* 33. doi:10.1016/j.cclet.2021.10.055

Soni, K. S., Desale, S. S., and Bronich, T. K. J. J. o. C. R. (2016). Nanogels: An overview of properties, biomedical applications and obstacles to clinical translation. *J. Control. Release* 240, 109–126. doi:10.1016/j.jconrel.2015.11.009

Soni, S., Ruhela, R. K., and Medhi, B. J. A. p. b. (2016). Nanomedicine in Central Nervous System (CNS) Disorders: A Present and Future Prospective. *Adv Pharm Bull* 6, 319. doi:10.15171/apb.2016.044

Soni, V., Kohli, D., and Jain, S. J. J. o. d. t. (2008). Transferrin-conjugated liposomal system for improved delivery of 5-fluorouracil to brain. *J. Drug Target.* 16, 73–78. doi:10.1080/10611860701725381

Srikanth, M., and Kessler, J. A. J. N. r. n. (2012). Nanotechnology-novel therapeutics for CNS disorders. *Nat. Rev. Neurol.* 8, 307–318. doi:10.1038/nrneurol.2012.76

Stenehjem, D. D., Hartz, A. M., Bauer, B., and Anderson, G. W. J. F. M. C. (2009). Novel and emerging strategies in drug delivery for overcoming the blood-brain barrier. *Future Med. Chem.* 1, 1623–1641. doi:10.4155/fmc.09.137

Stewart, S. A. J. J. o. C. P. (2005). The effects of benzodiazepines on cognition. J. Clin. Psychiatry 66, 9–13.

Stockwell, J., Abdi, N., Lu, X., Maheshwari, O., Taghibiglou, C. J. C. b., and design, d. (2014). Novel central nervous system drug delivery systems. *Chem. Biol. Drug Des.* 83, 507–520. doi:10.1111/cbdd.12268

Stratakis, E., Ranella, A., Farsari, M., and Fotakis, C. J. P. i. Q. E. (2009). Laserbased micro/nanoengineering for biological applications. *Prog. Quantum Electron.* 33, 127–163. doi:10.1016/j.pquantelec.2009.06.001

Stupp, R., Mason, W. P., Van Den Bent, M. J., Weller, M., Fisher, B., Taphoorn, M. J., et al. (2005). Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N. Engl. J. Med.* 352, 987–996. doi:10.1056/NEJMoa043330

Sweeney, M. D., Zhao, Z., Montagne, A., Nelson, A. R., and Zlokovic, B. V. J. P. r. (2019). Blood-brain barrier: From physiology to disease and back. *Physiol. Rev.* 99, 21–78. doi:10.1152/physrev.00050.2017

Syková, E., and Nicholson, C. J. P. r. (2008). Diffusion in brain extracellular space. *Physiol. Rev.* 88, 1277–1340. doi:10.1152/physrev.00027.2007

Szabo, E., Schneider, H., Seystahl, K., Rushing, E. J., Herting, F., Weidner, K. M., et al. (2016). Autocrine VEGFR1 and VEGFR2 signaling promotes survival in human glioblastoma models *in vitro* and *in vivo*. *Neuro*. *Oncol*. 18, 1242–1252. doi:10.1093/neuonc/now043

Tamai, I., and Tsuji, A. J. A. d. d. r. (1996). Drug delivery through the blood-brain barrier. Adv. Drug Deliv. Rev. 19, 401-424. doi:10.1016/0169-409x(96)00011-7

Tateishi, K., Wakimoto, H., Iafrate, A. J., Tanaka, S., Loebel, F., Lelic, N., et al. (2015). Extreme vulnerability of IDH1 mutant cancers to NAD+ depletion. *Cancer Cell* 28, 773–784. doi:10.1016/j.ccell.2015.11.006

Teixeira, M. I., Lopes, C. M., Amaral, M. H., and Costa, P. C. (2020). Current insights on lipid nanocarrier-assisted drug delivery in the treatment of neurodegenerative diseases. *Eur. J. Pharm. Biopharm.* 149, 192–217. doi:10.1016/j.ejpb.2020.01.005

Thomas, C., Rawat, A., Hope-Weeks, L., and Ahsan, F. J. M. p. (2011). Aerosolized PLA and PLGA nanoparticles enhance humoral, mucosal and cytokine responses to. *Hepat. B vaccine* 8, 405–415.

Thorne, R. G., and Frey, W. H. J. C. p. (2001). Delivery of neurotrophic factors to the central nervous system: Pharmacokinetic considerations. *Clin. Pharmacokinet.* 40, 907–946. doi:10.2165/00003088-200140120-00003

Thorne, R. G., and Nicholson, C. J. P. o. t. N. A. o. S. (2006). *In vivo* diffusion analysis with quantum dots and dextrans predicts the width of brain extracellular space. *Proc. Natl. Acad. Sci. U. S. A.* 103, 5567–5572. doi:10. 1073/pnas.0509425103

Tong, S., Zhu, H., and Bao, G. J. M. T. (2019). Magnetic iron oxide nanoparticles for disease detection and therapy. *Mat. Today* 31, 86–99. doi:10.1016/j.mattod.2019. 06.003

Torchilin, V. P. J. P. R. (2007). Micellar nanocarriers: Pharmaceutical perspectives. *Pharm. Res.* 24, 1–16. doi:10.1007/s11095-006-9132-0

Torres-Ortega, P. V., Saludas, L., Hanafy, A. S., Garbayo, E., and Blanco-Prieto, M. J. J. J. o. C. R. (2019). Micro-and nanotechnology approaches to improve Parkinson's disease therapy. *J. Control. Release* 295, 201–213. doi:10.1016/j.jconrel. 2018.12.036

Umut, E. J. M. S. E. T. (2013). Surface modification of nanoparticles used in biomedical applications. *Nanobiotechnology* 20, 185–208.

Upadhyay, R. K. J. B. r. i. (2014). Drug delivery systems, CNS protection, and the blood brain barrier, 2014.

Vermeeren, A. J. C. d. (2004). Residual effects of hypnotics: Epidemiology and clinical implications. CNS Drugs 18, 297–328. doi:10.2165/00023210-200418050-00003

Verreault, M., Schmitt, C., Goldwirt, L., Pelton, K., Haidar, S., Levasseur, C., et al. (2016). Preclinical efficacy of the MDM2 inhibitor RG7112 in MDM2-amplified and TP53 wild-type glioblastomas. *Clin. Cancer Res.* 22, 1185–1196. doi:10.1158/1078-0432.CCR-15-1015

Vickers, N. J. J. C. b. (2017). Animal communication: When i'm calling you, will you answer too? *Curr. Biol.* 27, R713–R715. doi:10.1016/j.cub.2017.05.064

Voigt, N., Henrich-Noack, P., Kockentiedt, S., Hintz, W., Tomas, J., and Sabel, B. A. (2014). Surfactants, not size or zeta-potential influence blood-brain barrier passage of polymeric nanoparticles. *Eur. J. Pharm. Biopharm.* 87, 19–29. doi:10. 1016/j.ejpb.2014.02.013

Vorbrodt, A. W., and Dobrogowska, D. H. J. B. R. R. (2003). Molecular anatomy of intercellular junctions in brain endothelial and epithelial barriers: Electron microscopist's view. *Brain Res. Brain Res. Rev.* 42, 221–242. doi:10.1016/s0165-0173(03)00177-2

Vuong, N. (2018). Nanotechnology-enhanced blood and urine analysis for the identification of biomarkers related to severe traumatic brain injury and acute respiratory distress Syndrome.

Walsh, J. (1979). "Behavioural effects of drugs in the hyperbaric environment, the twenty-first undersea medical society workshop," in *Interaction of drugs in the hyperbaric environment* (Bethesda: Undersea Medi Soc).

Wang, L., Xiong, X., Zhang, L., and Shen, J. J. C. N. (2021). Neurovascular unit: A critical role in ischemic stroke. CNS Neurosci. Ther. 27, 7–16. doi:10.1111/cns.13561

Waris, A., Ali, A., Khan, A. U., Asim, M., Zamel, D., Fatima, K., et al. (2022). Applications of various types of nanomaterials for the treatment of neurological disorders. *Nanomaterials* 12, 2140. doi:10.3390/nano12132140

Wei, G., Su, Z., Reynolds, N. P., Arosio, P., Hamley, I. W., Gazit, E., et al. (2017). Self-assembling peptide and protein amyloids: From structure to tailored function in nanotechnology. *Chem. Soc. Rev.* 46, 4661–4708. doi:10.1039/c6cs00542j

Whiting, P. J. J. D. d. t. (2003). GABA-A receptor subtypes in the brain: A paradigm for CNS drug discovery? *Drug Discov. Today* 8, 445–450. doi:10.1016/s1359-6446(03)02703-x

Williams, T., Wilkinson, A., Davis, F., and Frampton, C. J. U. b. r. (1988). Effects of transcutaneous scopolamine and depth on diver performance. *Undersea Biomed. Res.* 15, 89–98.

Wilson, B., Samanta, M. K., Santhi, K., Kumar, K. P. S., Paramakrishnan, N., and Suresh, B. J. B. r. (2008). Poly (n-butylcyanoacrylate) nanoparticles coated with polysorbate 80 for the targeted delivery of rivastigmine into the brain to treat Alzheimer's disease. *Brain Res.* 1200, 159–168. doi:10.1016/j.brainres.2008.01.039

Witten, J., and Ribbeck, K. J. N. (2017). The particle in the spider's web: Transport through biological hydrogels. *Nanoscale* 9, 8080–8095. doi:10.1039/c6nr09736g

Wolak, D. J., and Thorne, R. G. J. M. p. (2013). Diffusion of macromolecules in the brain: Implications for drug delivery. *Mol. Pharm.* 10, 1492–1504. doi:10.1021/mp300495e

Wong, H. L., Wu, X. Y., and Bendayan, R. J. A. d. d. r. (2012). Nanotechnological advances for the delivery of CNS therapeutics. *Adv. Drug Deliv. Rev.* 64, 686–700. doi:10.1016/j.addr.2011.10.007

Woodworth, G. F., Dunn, G. P., Nance, E. A., Hanes, J., and Brem, H. J. F. i. o. (2014). Emerging insights into barriers to effective brain tumor therapeutics. *Front. Oncol.* 4, 126. doi:10.3389/fonc.2014.00126

Xia, N., Wang, X., Zhou, B., Wu, Y., Mao, W., Liu, L. J. A. a. m., et al. (2016). Electrochemical detection of amyloid-β oligomers based on the signal amplification of a network of silver nanoparticles. *ACS Appl. Mat. Interfaces* 8, 19303–19311. doi:10.1021/acsami.6b05423

Xie, J., Shen, Z., Anraku, Y., Kataoka, K., and Chen, X. J. B. (2019). Nanomaterialbased blood-brain-barrier (BBB) crossing strategies. *Biomaterials* 224, 119491. doi:10.1016/j.biomaterials.2019.119491

Yamazaki, Y., and Kanekiyo, T. J. I. j. o. m. s. (2017). Blood-brain barrier dysfunction and the pathogenesis of Alzheimer's disease. *Int. J. Mol. Sci.* 18, 1965. doi:10.3390/ijms18091965

Yang, F., Li, Y., Chen, Z., Zhang, Y., Wu, J., and Gu, N. J. B. (2009). Superparamagnetic iron oxide nanoparticle-embedded encapsulated microbubbles as dual contrast agents of magnetic resonance and ultrasound imaging, *Biomaterials* 30, 3882–3890. doi:10.1016/j.biomaterials.2009.03.051

Yang, Y., and Rosenberg, G. A. J. S. (2011). Blood-brain barrier breakdown in acute and chronic cerebrovascular disease. *Stroke* 42, 3323–3328. doi:10.1161/STROKEAHA.110.608257

Yao, V. J., D'Angelo, S., Butler, K. S., Theron, C., Smith, T. L., Marchiò, S., et al. (2016). Ligand-targeted theranostic nanomedicines against cancer. *J. Control. Release* 240, 267–286. doi:10.1016/j.jconrel.2016.01.002

Yazdani, S., Jaldin-Fincati, J. R., Pereira, R. V., and Klip, A. J. T. (2019). Endothelial cell barriers: Transport of molecules between blood and tissues. *Traffic* 20, 390-403. doi:10.1111/tra.12645

Yi, X., Manickam, D. S., Brynskikh, A., and Kabanov, A. V. J. J. o. C. R. (2014). Agile delivery of protein therapeutics to CNS. *J. Control. Release* 190, 637–663. doi:10.1016/j.jconrel.2014.06.017

Youdim, M. B., Edmondson, D., and Tipton, K. F. J. N. R. N. (2006). The therapeutic potential of monoamine oxidase inhibitors. *Nat. Rev. Neurosci.* 7, 295–309. doi:10.1038/nrn1883

Yu, M., Wu, J., Shi, J., and Farokhzad, O. C. J. J. o. c. r. (2016). Nanotechnology for protein delivery: Overview and perspectives. *J. Control. Release* 240, 24–37. doi:10. 1016/j.jconrel.2015.10.012

Zeevi, N., Pachter, J., McCullough, L. D., Wolfson, L., and Kuchel, G. A. J. J. O. T. A. G. S. (2010). The blood-brain barrier: Geriatric relevance of a critical brain-body interface. *J. Am. Geriatr. Soc.* 58, 1749–1757. doi:10.1111/j.1532-5415.2010.03011.x

Zeng, J., See, A. P., Phallen, J., Jackson, C. M., Belcaid, Z., Ruzevick, J., et al. (2013). Anti-PD-1 blockade and stereotactic radiation produce long-term survival in mice with intracranial gliomas. *Int. J. Radiat. Oncol. Biol. Phys.* 86, 343–349. doi:10.1016/ j.ijrobp.2012.12.025

Zhang, F., Xu, C.-L., and Liu, C.-M. J. D. D. (2015). Drug delivery strategies to enhance the permeability of the blood-brain barrier for treatment of glioma. *Drug Des. devel. Ther.* 9, 2089–2100. doi:10.2147/DDDT.S79592

Zhang, J., Mou, L., and Jiang, X. J. C. S. (2020). Surface chemistry of gold nanoparticles for health-related applications. *Chem. Sci.* 11, 923–936. doi:10.1039/ c9sc06497d

Zhang, M., Zang, X., Wang, M., Li, Z., Qiao, M., Hu, H., et al. (2019). Exosome-based nanocarriers as bio-inspired and versatile vehicles for drug delivery: Recent advances and challenges. J. Mat. Chem. B 7, 2421–2433. doi:10. 1039/c9tb00170k

Zhang, P., Hu, L., Yin, Q., Feng, L., and Li, Y. J. M. p. (2012). Transferrin-modified c [RGDfK]-paclitaxel loaded hybrid micelle for sequential blood-brain barrier penetration and glioma targeting therapy. *Mol. Pharm.* 9, 1590–1598. doi:10. 1021/mp200600t

Zhang, Q., Zhang, J., Song, J., Liu, Y., Ren, X., and Zhao, Y. J. A. n. (2021). Proteinbased nanomedicine for therapeutic benefits of cancer. *ACS Nano* 15, 8001–8038. doi:10.1021/acsnano.1c00476

Zhao, C., Xing, Z., Zhang, C., Fan, Y., and Liu, H. J. J. o. M. C. B. (2021). Nanopharmaceutical-based regenerative medicine: A promising therapeutic strategy for spinal cord injury. *J. Mat. Chem. B* 9, 2367–2383. doi:10.1039/ d0tb02740e

Zlokovic, B. V. J. N. (2008). The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron* 57, 178–201. doi:10.1016/j.neuron.2008. 01.003

Zong, T., Mei, L., Gao, H., Cai, W., Zhu, P., Shi, K., et al. (2014). Synergistic dualligand doxorubicin liposomes improve targeting and therapeutic efficacy of brain glioma in animals. *Mol. Pharm.* 11, 2346–2357. doi:10.1021/mp500057n

Zou, Z., Jiang, X., Zhang, W., Zhou, Y., Ke, Y., Zhang, S., et al. (2010). Efficacy of Tyrosine Hydroxylase gene modified neural stem cells derived from bone marrow on Parkinson's disease–a rat model study. *Brain Res.* 1346, 279–286. doi:10.1016/j. brainres.2010.05.071