

\*Correspondence to:

0000-0002-7164-4116

Accepted: 2017-03-22

(Christian Guerra-Araiza)

doi: 10.4103/1673-5374.205098

orcid:

Christian Guerra-Araiza,

christianguerra2001@gmail.com.

# Tibolone modulates neuronal plasticity through regulating Tau, GSK3β/Akt/PI3K pathway and CDK5 p35/p25 complexes in the hippocampus of aged male mice

Teresa Neri-Gómez<sup>1, 2</sup>, Judith Espinosa-Raya<sup>2</sup>, Sofía Díaz-Cintra<sup>3</sup>, Julia Segura-Uribe<sup>4</sup>, Sandra Orozco-Suárez<sup>4</sup>, Juan Manuel Gallardo<sup>5</sup>, Christian Guerra-Araiza<sup>1,\*</sup>

1 Unidad de Investigación Médica en Farmacología, Hospital de Especialidades, Centro Médico Nacional Siglo XXI, Instituto Mexicano del Seguro Social, Ciudad de México, México

2 Laboratorio de Farmacología Conductual, Escuela Superior de Medicina, Instituto Politécnico Nacional, Plan de San Luis y Díaz Mirón Col. Sto. Tomás, Ciudad de México, México

3 Departamento de Neurobiología del Desarrollo y Neurofisiología, Instituto de Neurobiología, Universidad Nacional Autónoma de México Campus Juriquilla, Querétaro, Querétaro, México

4 Enfermedades Neurológicas (Neurological Diseases), Hospital de Especialidades, Centro Médico Nacional Siglo XXI, Instituto Mexicano del Seguro Social, Ciudad de México, México

5 Enfermedades Nefrológicas (Kidney Diseases), Hospital de Especialidades, Centro Medico Nacional Siglo XXI, Instituto Mexicano del Seguro Social, Ciudad de Mexico, Mexico

*How to cite this article:* Neri-Gómez T, Espinosa-Raya J, Díaz-Cintra S, Segura-Uribe J, Orozco-Suárez S, Gallardo JM, Guerra-Araiza C (2017) Tibolone modulates neuronal plasticity through regulating Tau,  $GSK3\beta/Akt/PI3K$  pathway and CDK5 p35/p25 complexes in the hippocampus of aged male mice. Neural Regen Res 12(4):588-595.

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Funding: This study was supported by FIS/IMSS project No. FIS/IMSS/PROT/G13/1216, COFAA, SIP-IPN and by DGAPA-UNAM IN203616.

# Abstract

Aging is a key risk factor for cognitive decline and age-related neurodegenerative disorders. Also, an age-related decrease in sex steroid hormones may have a negative impact on the formation of neurofibrillary tangles (NFTs); these hormones can regulate Tau phosphorylation and the principal kinase GSK3β involved in this process. Hormone replacement therapy decreases NFTs, but it increases the risk of some types of cancer. However, other synthetic hormones such as tibolone (TIB) have been used for hormone replacement therapy. The aim of this work was to evaluate the long-term effects of TIB (0.01 mg/kg and 1 mg/kg, intragastrically for 12 weeks) on the content of total and hyperphosphorylated Tau (PHF-1) proteins and the regulation of GSK3β/Akt/PI3K pathway and CDK5/p35/p25 complexes in the hippocampus of aged male mice. We observed that the content of PHF-1 decreased with TIB administration. In contrast, no changes were observed in the active form of GSK3 $\beta$  or PI3K. TIB decreased the expression of the total and phosphorylated form of Akt while increased that of p110 and p85. The content of CDK5 was differentially modified with TIB: it was increased at low doses and decreased at high doses. When we analyzed the content of CDK5 activators, an increase was found on p35; however, the content of p25 decreased with administration of low dose of TIB. Our results suggest a possible mechanism of action of TIB in the hippocampus of aged male mice. Through the regulation of Tau and GSK3β/Akt/PI3K pathway, and CDK5/p35/ p25 complexes, TIB may modulate neuronal plasticity and regulate learning and memory processes.

*Key Words:* nerve regeneration; Tibolone; hippocampus; aged mice; sex steroids; Akt; GSK3β; PI3K; neural plasticity; Tau; neurofibrillary tangles; neural regeneration

# Introduction

The association between low testosterone levels and increased risk for Alzheimer's disease (AD) in aged men may reflect the loss of beneficial androgen-mediated actions in the brain (Rosario et al., 2010). There are several androgen actions potentially relevant to AD, including promotion of neuron viability (Pike, 2001; Ramsden et al., 2003), synaptic plasticity (Leranth et al., 2003), selected aspects of cognition (Cherrier et al., 2005; Moffat, 2005; Janowsky, 2006), and reduction of Tau phosphorylation (Papasozomenos, 1997). Neurofibrillary tangles (NFTs) are predominantly composed of post-translationally modified forms of the microtubule-associated protein Tau and can become widespread in hippocampal, neocortical, and limbic neurons in AD (Yankner et al., 2008).

Tau modulates the extent and rate of microtubule assembly and plays an essential role in morphogenetic processes, such as axonal growth (Drechsel et al., 1992). It also increases the rate of tubulin polymerization, decreases the rate of transit into the shrinking phase, and inhibits the rate of depolymerization (Drechsel et al., 1992). In several diseases, hyperphosphorylation of Tau at multiple sites is related with its dissociation from microtubules and aggregation into oligomeric and fibrillar forms. This hyperphosphorylation may destabilize microtubules and impair anterograde axonal transport of essential macromolecules and organelles to synaptic endings (Skovronsky et al., 2006).

It has been shown that several kinases regulate Tau phosphorylation, but only two have been co-immunoprecipitated with microtubules, GSK3 $\beta$  and CDK5 (Ishiguro et al., 1992; Flaherty et al., 2000). CDK5 is a member of the cyclin-dependent protein kinase family. Whereas CDK5 does not have any role in the cell cycle, it participates in neuronal development and survival, phosphorylation of cytoskeletal proteins, and synaptic plasticity (Malumbres and Barbacid, 2005; Shukla et al., 2012).

Some evidence has suggested that GSK3 $\beta$  levels are increased in AD brains (Baum et al., 1996; Imahori and Uchida, 1997), and that the imbalance in GSK3 $\beta$  activation could compromise neurotrophin signaling. The abnormal Tau phosphorylation that results from increased activity of GSK3 $\beta$  could lead to sequestration of Tau, inhibition of microtubule formation, destabilization of the existing cytoskeleton, and promotion of assembly of large Tau-containing aggregates, and these would inhibit both anterograde and retrograde microtubule-based transport (Niewiadomska et al., 2006).

Some gonadal hormones such as estrogens, progestogens, and androgens can modulate the activation of Tau and GSK3 $\beta$  in female rats (Pinto-Almazán et al., 2012). In a mouse model of AD, dihydrotestosterone can prevent an increase of hyperphosphorylation of Tau as well (Rosario et al., 2010). In addition, different doses of estrogens have been shown to exert different effects (Espinosa-Raya et al., 2011).

Tibolone (TIB), a synthetic steroid, is widely prescribed to treat menopausal symptoms. TIB is metabolized into three biologically active metabolites: 3α-hydroxy and 3β-hydroxy (estrogenic), and  $\Delta$ -4 keto isomer, which displays progestogenic and androgenic effects (Kloosterboer, 2001; Campisi and Marengo, 2007). TIB metabolites circulate as inactive forms, and its conversion to bioactive forms depends on tissue-specific desulfation (Verheuland Kloosterboer, 2006). TIB also exhibited neuroprotective effects when prevented oxidative stress neurodegeneration in male rats (Pinto-Almazán et al., 2014). TIB also exhibits different effects on memory and learning in a dose-dependent manner (Espinosa-Raya et al., 2012; Farfán-García et al., 2014). However, the effects of TIB on the activation of Tau protein and its signaling cascade in the brain of aged males are unknown. The aim of this study was to evaluate the long-term effects of two (low and high) doses of TIB on Tau hyperphosphorylation and PI3K/Akt/ GSK3 $\beta$  signaling pathway, and CDK5/p35/p25 complexes.

# Materials and Methods

### Animals

Adult male aged (18 months) 129/C57BL/6 mice, weighing  $25 \pm 5$  g, were housed five per cage, under a 12-hour light/ dark cycle (lights on at 9:00 p.m.) and provided water and food *ad libitum*. All procedures were performed by the Mexican Guidelines for Animal Care and Handling (NOM-

062-ZOO-1999). All efforts were made to minimize animal discomfort and reduce the number of animals used.

### Treatments

Aged intact male mice were randomly divided into three groups (n = 6 per group). According to previous reports, mice in these groups were intregastrically administered vehicle (Veh), 0.01 mg/kg (low dose), or 1.0 mg/kg (high dose) of TIB (Sigma-Aldrich, San Luis, MO, USA), respectively once a day for 12 successive weeks (Espinosa-Raya et al., 2012; Farfán-García et al., 2014).

# Western blot analysis

Twenty-four hours after the last TIB administration (at 10:00 a.m.), mice were decapitated and the brains were removed. The hippocampus was dissected according to the Atlas of Paxinos and Watson (1998) and then immediately homogenized in lysis buffer with protease inhibitors for protein extraction, as previously described (Espinosa-Raya et al., 2012). Protein samples were obtained by centrifugation and quantified using the Bradford method (Bio-Rad, Hercules, CA, USA). Proteins (50 µg) were separated by electrophoresis and transferred to Immobilon-P membranes (Millipore, Bedford, MA, USA). Membranes were blocked with Blocking Buffer (Bio-Rad) and incubated with primary antibodies (diluted 1:1,000). To determine the molecular weight of the proteins, broad range pertained protein markers were added.

The following antibodies were used: rabbit anti-tau (H150) polyclonal antibody, mouse anti-GSK3 $\beta$  monoclonal antibody, mouse anti-phosphorylated GSK3 $\beta$  (pSer9 GSK3 $\beta$ ) monoclonal antibody, rabbit Akt polyclonal antibody, rabbit pAkt1/2/3 (Ser473) polyclonal antibody, rabbit PI3-kinase (p110) antibody, rabbit pPI3 kinase p110g (Tyr485) polyclonal antibody, mouse PI3-kinase (p85 $\alpha$ ) monoclonal antibody, goat pPI3 kinase p85a (Tyr508) antibody and rabbit CDK5 (J3) polyclonal antibody were purchased from Santa Cruz Biotechnology (Dallas, TX, USA). Rabbit p35/25 (C64B10) polyclonal antibody was purchased from Cell Signaling (Danvers, MA, USA). Mouse anti-glyceraldehyde-3-phosphate-dehydrogenase (GAPDH) was purchased from Chemicon (Billerica, MA, USA). Rabbit anti-tau (PHF-1, phospho S396) monoclonal antibody was purchased from Abcam (Cambridge, UK).

The abovementioned membranes were incubated overnight at 4°C with the primary antibody, then washed and incubated with anti-mouse or anti-rabbit secondary antibody as appropriate for 2 hours at room temperature (Santa Cruz Biotechnology, diluted 1:15,000). Immunoreactive bands were detected using an enhanced chemiluminescence system (GE Healthcare, Piscataway, NJ, USA). Subsequently, the membranes were stripped with a commercial solution (Chemicon, Billerica, MA, USA) and reprobed with anti-GAPDH monoclonal antibody detected by the system mentioned above. The intensity of protein bands was quantified and analyzed by densitometry (KODAK 1D Image Analysis Software). The density of each band was normalized to its respective loading control (GAPDH). Samples from all animal groups were processed concurrently and under the same conditions in each experiment.



Figure 1 Changes in the content of total Tau and PHF-1 in the hippocampus of aged mice treated with tibolone (TIB). (A) Western blots for total Tau content in the hippocampus of aged mice. (B) PHF-1 levels in the hippocampus of aged mice treated with TIB. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. (C) The ratio of PHF-1 and Tau to their respective control (GAPDH) expression in the hippocampus of mice treated with low- and high-dose TIB. Results are expressed as the mean  $\pm$  SE of six individual experiments. \**P* < 0.05, *vs.* vehicle (Veh).



Figure 2 Changes in the content of total and phosphorylated GSK3 $\beta$  in the hippocampus of aged mice treated with tibolone (TIB). Western blots for total (A) and phosphorylated (Ser9) (B) GSK3 $\beta$  content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. (C) Ratio of phosphorylated (Ser9) GSK3 $\beta$  to GSK3 $\beta$ . Results are expressed as the mean  $\pm$  SE of six individual experiments. \**P* < 0.05, *vs.* vehicle (Veh).

## Statistical analysis

All data are presented as the mean  $\pm$  standard error (SE) and analyzed by one-way analysis of variance to compare group means for each variable. Tukey's *post-hoc* test was used to determine differences between means. Prism 5.0 program (GraphPad, La Jolla, CA, USA) was used to calculate probability values. *P* values < 0.05 were considered statistically significant.

# Results

# Effect of TIB on the content of Tau and GSK3 $\beta$

Western blot was used to analyze the effect of TIB on the content of PHF-1 and total Tau in hippocampal samples of aged mice. The expression of PHF-1 decreased with TIB treatment at both doses (**Figure 1A**); in contrast, the expression of total Tau was unaffected by TIB (**Figure 1B**). Furthermore, the ratio of hyperphosphorylated Tau *vs.* total Tau



Figure 3 Changes in the content of total and phosphorylated Akt in the hippocampus of aged mice treated with tibolone (TIB). Western blots for total (A) and phosphorylated (B) Akt content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. (C) Ratio of phosphorylated to total Akt. Results are expressed as the mean  $\pm$  SE of six individual experiments. \**P* < 0.05 *vs.* vehicle (Veh); #*P* < 0.05, *vs.* 1.0 mg/kg TIB.



#### Figure 4 Changes in the content of the catalytic subunit PI3K p110 in the hippocampus of aged mice treated with tibolone (TIB).

Representative western blots for PI3K p110 and phosphorylated p110 content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. Densitometric analysis of total (B) and phosphorylated (C) catalytic subunit PI3K p110. (D) Ratio of phosphorylated versus total PI3K p100. Results are expressed as the mean ± SE of six individual experiments.\**P* < 0.05, *vs*. vehicle (Veh).

was significantly increased (P < 0.05) after TIB treatment at either dose (**Figure 1C**).

Tau phosphorylation is associated with the modification of GSK3 $\beta$  phosphorylation (Ser9) (Haque et al., 1999). Therefore, we analyzed the expression of total GSK3 $\beta$ , and the levels of phosphorylated GSK3 $\beta$  (Ser9) were assessed to determine if the expression of total GSK3 $\beta$  (**Figure 2**) was decreased in the hippocampus with the low-dose TIB treatment. However, TIB did not change the expression of phosphorylated GSK3 $\beta$  (Ser9). Therefore, there was no difference in the ratio of phosphorylated *vs.* total GSK3 $\beta$ .

#### Effect of TIB on the content of PI3K/Akt pathway

PI3K/Akt pathway is involved in the activation of GSK3 $\beta$  (Woodgett, 1990), so we investigated if there were any changes in Akt content. A decrease in the total content of Akt after treatment with TIB at both doses was observed (**Figure 3A**). Phosphorylated Akt content decreased only with the low-dose TIB treatment (**Figure 3B**). **Figure 3C** shows that the ratio of phosphorylated Akt and total Akt increased with the low-dose TIB while it decreased with treatment with high-dose TIB.

The content of the catalytic subunit of PI3 kinase, p110, and its phosphorylated form (pPI3 kinase), were also ana-

lyzed (**Figure 4**). No changes were observed for total PI3 kinase p110 (**Figure 4B**), whereas TIB increased the expression of pPI3 kinase p110 (**Figure 4C**). This effect was obvious when phosphorylated PI3K/PI3K ratio was analyzed (**Figure 4D**). The levels of the regulatory subunit of PI3 kinase p85 decreased with low-dose TIB treatment and increased with high-dose TIB treatment (**Figure 5B**). TIB at both doses increased the expression of pPI3 kinase p85 (**Figure 5C**), and the pPI3Kp85/PI3Kp85 ratio was the highest after treatment with TIB at 0.01 mg/kg (**Figure 5D**).

#### Effect of TIB on the content of CDK5/p35/p25 complexes

Another kinase involved in the regulation of Tau hyperphosphorylation is CDK5. We analyzed the expression of CDK5 and its activators p35 and p25. The content of CDK5 was differentially modified with TIB treatment: it increased with treatment with low dose TIB, and with treatment with high dose TIB, decreased CDK5 content was observed (**Figure 6**). When we analyzed the content of the activators, an increase was observed in the content of p35. In contrast, the content of p25 decreased with treatment with low dose TIB (**Figure 7**).

# Discussion

Our results suggest that TIB can modulate the content of total and phosphorylated microtubule-associated protein Tau through the PI3K/Akt/GSK3 $\beta$  pathway and CDK5/p35/p25 complexes in the hippocampus of aged male mice.

Our results showed that aging increased the phosphorylation of Tau and TIB administration decreased the phosphorylation of Tau in the hippocampus of the aged mice. These results were in accordance with previous studies showing that aging increased the phosphorylation of Tau in the brain tissue of monkeys and rats (Niewiadomska et al., 2006; Carlyle et al., 2014) and that TIB decreased the phosphorylation of Tau in the hippocampus of ovariectomized rats (Pinto-Almazán et al., 2012)

Tau phosphorylation reduces the affinity for microtubules and its ability to promote microtubule assembly in vitro (Lindwall and Cole, 1984; Biernat et al., 1993; Lu and Wood, 1993). Many of the phosphorylated residues in Tau during the development of paired helical filaments in Alzheimer's disease are serines and threonines preceding a proline (Mietelska-Porowska et al., 2014). GSK3β, a proline-directed kinase, induces phosphorylation of Tau at least on some of these residues. Co-transfection of Tau with GSK3β elevates GSK3β activity in fibroblast cells, so as to phosphorylate Tau (Lovestone et al., 1994; Sperber et al., 1995). Androgen depletion has been shown to significantly accelerate the development of Alzheimer's disease-like neuropathology in the triple-transgenic mouse model of AD (3xTg-AD). This effect is prevented by androgen treatment. It was observed that the levels of Tau hyperphosphorylation in sham gonadectomy (GDX) 3xTg-AD male mice were modest and only increased slightly by GDX. In GDX male mice, treatment with testosterone (T) or 17β-estradiol (E2) but not with dihydrotestosterone (DHT) reduced Tau hyperphosphorylation to levels lower than observed in sham animals. Because T is metabolized into both the androgen DHT and the estrogen E2 in the brain, T

has been shown to mediate its effects through androgen or estrogen metabolic pathways (Rosario et al., 2010).

The most abundant free TIB metabolite in the hippocampus and cerebellum of cynomolgus monkeys is the 3-hydroxy tibolone, which has estrogenic activity (Verheul and Kloosterboer, 2006). These data suggest that TIB modulates Tau phosphorylation through this metabolite in both the hippocampus and cerebellum. Our results suggest that the decrease in the phosphorylation of Tau in the hippocampus of aged male mice may involve estrogen-regulated pathways. Nevertheless, further research needs to be undertaken in this area.

Considering that GSK3 $\beta$  plays a significant role in the regulation of Tau phosphorylation *in vivo*, the understanding of signaling pathways that modulate the activity of GSK3 $\beta$  is of critical importance. It is well known that phosphorylation on the Ser<sup>9</sup> residue of GSK3 $\beta$  inhibits its activity, while phosphorylation on Tyr<sup>216</sup> increases it (Jope and Johnson, 2004). GSK3 $\beta$  regulates Tau hyperphosphorylation at Ser<sup>198</sup>/Ser<sup>199</sup>/Ser<sup>202</sup> sites and Ser<sup>396</sup>/Ser<sup>404</sup> sites (Haque et al., 1999). Furthermore, as a down-regulator of insulin signaling, GSK3 $\beta$  is regulated by Akt (Beurel et al., 2015). In addition, long-term treatment with insulin or IGF-1 results in Akt activation, which in turn phosphorylates GSK3 $\beta$  on Ser<sup>9</sup> (Hong and Lee, 1997; Lesort and Johnson, 2000).

Previous results have shown that TIB treatment increases only the phosphorylation of GSK3 $\beta$ , the inactive form of this protein, in the hippocampus. These results correlate with the decrease of Tau hyperphosphorylation in female rats (Pinto-Almazán et al., 2012), and suggest that TIB may decrease Tau phosphorylation through the inactivation of GSK3 $\beta$  in females. In contrast, our results indicate that, despite a decrease in the total GSK3 $\beta$  content with the low dose of TIB, this hormone does not modify the content of the active form of GSK3 $\beta$ in the hippocampus of aged male mice. Unlike the previous investigation, where a model with young, ovariectomized female rats was used (Pinto-Almazán et al., 2012), we used aged male mice as a model in the present investigation. The differences in the observed results may be because ovariectomy in female rats modifies the hypothalamic-pituitary-gonadal axis.

Furthermore, we investigated Akt activation, which can be achieved by the binding of neurotransmitters or growth factors on many specific cell-surface receptors, which in turn initiate a cascade of second messengers related to the PI3K pathway (Cardona-Gomez et al., 2001). Moreover, these two signaling pathways converge downstream to GSK3<sup>β</sup>, which is inhibited by Akt (Woodgett, 1990). PI3K signaling results in the activation of Akt following its phosphorylation on the Thr<sup>308</sup> and Ser<sup>473</sup> residues (Beaulieu, 2010). A previous report showed that in the rat cerebellum, E2 increased the phosphorylation of Akt at 6 and 12 hours after its administration (Morissette et al., 2008). However, in this study, we observed that TIB decreased the content and phosphorylation of Akt in the hippocampus of aged males. These results, together with those observed for GSK3 $\beta$ , indicate that the GSK3 $\beta$ / Akt signaling pathway is likely not involved in the regulation of TIB-mediated Tau phosphorylation in the hippocampus of the old mice. The negative regulation of TIB on Akt may have an effect on the decrease in Tau phosphorylation

through inactivating another kinase involved in this process such as CDK5. Overall, these results indicate the need for the analysis of the effect of TIB on the CDK5 signaling pathway in the old mouse hippocampus.

It has been observed that the activation of PI3K by the phosphorylation of tyrosine residues of its regulatory subunit p85 leads to the activation of the downstream kinase 3-phosphoinositide-dependent protein kinase-1 (PDK1) by phosphorylating PDK1 at Ser<sup>241</sup> (Fyffe and Falasca, 2013). Activated PDK1 then activates Akt by its phosphorylation at Thr<sup>308</sup>. Full activation of Akt also requires its phosphorylation at Ser<sup>473</sup>. A major target of Akt is GSK3. The activity of GSK3 is inhibited when it is phosphorylated at Ser<sup>21</sup> in GSK3 $\alpha$  or at Ser<sup>9</sup> in GSK3 $\beta$  by Akt, which results in glycogen synthesis. GSK3 $\beta$  is also a major Tau kinase (Takashima, 2006; Avila and Hernández, 2007). We observed that TIB increased the expression of the catalytic p110 and regulatory p85 subunits. Therefore, these modifications may lead to a down-regulation of the Tau phosphorylation pathway.

These differences may be because the regulation of Tau phosphorylation is subject to a balance between the action of kinases and phosphatases. Mitogen-activated protein kinases (MAPKs) and CDK5 are involved in the regulation of Tau phosphorylation in the hippocampus (Hyman et al., 1994; Crespo-Biel et al., 2007), and steroid hormones can modulate its activation (Guerra-Araiza et al., 2009; Harburger et al., 2009). In 2010, Amorim et al. reported that the phosphorylation of protein phosphatase 2A (PP2A) was regulated by hormones. PP2A is also capable of dephosphorylating Akt at Thr308 (Millward et al., 1999), and it regulates Tau phosphorylation directly or *via* GSK3 $\beta$  (Qian et al., 2010).

Another kinase involved in the hyperphosphorylation of Tau is CDK5. This kinase works with its co-activators, named p39/p35 and p29/p25. Without these activators, the monomeric form of CDK5 is enzymatically inactive (Gong and Iqbal, 2008).

Under pathological conditions, there is a cleavage of the activator p35 that leads to p25. The complex CDK5/p25 has been shown to induce the hyperphosphorylation of Tau, and also causes neurodegeneration. This complex was found in patients with Alzheimer's disease and co-localized with Tau aggregates (Imahori and Uchida, 1997). Moreover, the accumulation of this complex is located in several areas, including the frontal cortex, inferior parietal cortex, and hippocampus. Hyperphosphorylation of Tau and neurofilaments have been observed in mice overexpressing human p25, an activator of CDK5 (Tseng et al., 2002; Shelton and Johnson, 2004; Martina et al., 2013).

Our results showed that at a low dose, TIB increased the content of CDK5. On the contrary, at a high dose, the content of CDK5 decreased. When we analyzed the content of the p35 and p25 activators, it was observed that low doses of TIB increased the content of p35 and decreased the content of p25. As mentioned above, in mice overexpressing human p25, the hyperphosphorylation of Tau and neurofilaments have been observed in some areas, such as frontal cortex, cerebellum, and amygdala (Ahlijanian et al., 2000).

The effects of E2 on the regulation of CDK5 were demonstrated in SH-SY5Y cells using okadaic acid to induce Tau phosphorylation. Zhang and Simpkins (2010) reported that the hyperphosphorylation of Tau and a decrease in the upregulation of CDK5 were prevented by E2 in a dose-dependent manner, and this effect was blocked when the antagonist of estrogen receptor, ICI 182 780, was added.

Our results provide a possible mechanism of action of TIB in the aged hippocampus. Through the regulation of Tau, GSK3 $\beta$ /Akt/PI3K pathway and CDK5 p35/p25 complexes, TIB can modulate neuronal plasticity and participate in the regulation of learning and memory processes. Further studies are needed to determine whether TIB could be used as a neuroprotective agent for the prevention of tauopathies or other neurodegenerative diseases in aged males.

Acknowledgments: This work was submitted in partial fulfillment of the requirements for the Ph.D. degree of Teresa Neri-Gómez at Doctorado en Investigación en Medicina (ESM/IPN). Christian Guerra-Araiza received Beca de Excelencia en Investigación by Fundación IMSS, A. C.

**Author contributions:** TNG and JSU performed western blot analysis to detect protein expression levels and wrote the paper. SDC established aging mouse models. JER, SOS, JMG and CGA were responsible for data analysis. CGA designed this study and contributed to paper writing. All authors approved the final version of this paper.

Conflicts of interest: None declared.

**Plagiarism check:** This paper was screened twice using CrossCheck to verify originality before publication.

**Peer review:** This paper was double-blinded and stringently reviewed by international expert reviewers.

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#### Figure 5 Changes in the content of regulatory subunit PI3K p85 in the hippocampus of aged mice treated with tibolone (TIB).

Representative western blots for total PI3K p85 and phosphorylated p85 content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAP-DH protein content data. Densitometric analysis of total (B) and (C) phosphorylated catalytic subunit PI3K p85. (D) Ratio of phosphorylated versus total PI3K p85. Results are expressed as the mean  $\pm$  SE of six individual experiments.\*P < 0.05, vs. vehicle (Veh); #P < 0.05, vs. 1.0 mg/kg TIB.

#### Figure 6 Changes in the content of CDK5 in the hippocampus of aged mice treated with tibolone (TIB).

Western blots for CDK5 content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. Results are expressed as the mean  $\pm$  SE of six individual experiments. \**P* < 0.05, *vs.* vehicle (Veh); #*P* < 0.05, *vs.* 1.0 mg/kg TIB.

# Figure 7 Changes in the content of p35 and p25 in the hippocampus of aged mice treated with tibolone (TIB).

(A) Representative western blots for p35 and p25 content in the hippocampus of aged mice. GAPDH was used to correct differences of total loaded protein. Proteins detected by western blot analysis from the hippocampus of mice treated with TIB were quantified by densitometric analysis and corrected using GAPDH protein content data. (B-D) The densitometric analysis of p35 (B) and p25 (C) and the ratio of p35 versus p25 (D). Results are expressed as the mean  $\pm$  SE of six individual experiments. \*P < 0.05, vs. vehicle (Veh).

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