

# The FGF metabolic axis

Xiaokun Li (✉)

*School of Pharmaceutical Science, Wenzhou Medical University, Wenzhou 325035, China*

© The Author(s) 2019. This article is published with open access at [link.springer.com](http://link.springer.com) and [journal.hep.com.cn](http://journal.hep.com.cn)

**Abstract** Members of the fibroblast growth factor (FGF) family play pleiotropic roles in cellular and metabolic homeostasis. During evolution, the ancestor FGF expands into multiple members by acquiring divergent structural elements that enable functional divergence and specification. Heparan sulfate-binding FGFs, which play critical roles in embryonic development and adult tissue remodeling homeostasis, adapt to an autocrine/paracrine mode of action to promote cell proliferation and population growth. By contrast, FGF19, 21, and 23 coevolve through losing binding affinity for extracellular matrix heparan sulfate while acquiring affinity for transmembrane  $\alpha$ -Klotho (KL) or  $\beta$ -KL as a coreceptor, thereby adapting to an endocrine mode of action to drive interorgan crosstalk that regulates a broad spectrum of metabolic homeostasis. FGF19 metabolic axis from the ileum to liver negatively controls diurnal bile acid biosynthesis. FGF21 metabolic axes play multifaceted roles in controlling the homeostasis of lipid, glucose, and energy metabolism. FGF23 axes from the bone to kidney and parathyroid regulate metabolic homeostasis of phosphate, calcium, vitamin D, and parathyroid hormone that are important for bone health and systemic mineral balance. The significant divergence in structural elements and multiple functional specifications of FGF19, 21, and 23 in cellular and organismal metabolism instead of cell proliferation and growth sufficiently necessitate a new unified and specific term for these three endocrine FGFs. Thus, the term “FGF Metabolic Axis,” which distinguishes the unique pathways and functions of endocrine FGFs from other autocrine/paracrine mitogenic FGFs, is coined.

**Keywords** FGF19; FGF21; FGF23; FGFR; metabolism; endocrine; Klotho

## Introduction

Fibroblast growth factors (FGFs) are pleiotropic signal molecules for all types of cell and tissue systems in metazoans [1–3]. FGFs share a conserved core structure of  $\beta$ -trefoil fold consisting of 12-stranded  $\beta$ -sheets arranged in three similar lobes around a central axis, of which six strands form an antiparallel  $\beta$ -barrel [4,5]. Except for the four FGF-homologous intracrine factors that are functionally reminiscent of the ancestor FGF, the FGFs can be classified into mitogenic and metabolic FGFs, which overtly regulate cellular proliferation and substrate/energy metabolism, respectively, on the basis of their distinct functions and endpoint biological effects [6,7]. Both FGF classes signal through the same types of transmembrane receptor tyrosine kinases, that is, the FGF receptors (FGFRs) 1 to 4 with multiple splicing variants [8]. However, in physiology, these two types of regulatory

activities driven by the two FGF classes appear to be spatially and temporally segregated. At a physiological level, mitogenic FGFs appear to be incapable of traveling far to other tissues, including metabolic tissues, to promote cellular metabolism because of local trapping after secretion that is mediated by high affinity binding to the extracellular matrix heparan sulfate (HS). On the other hand, metabolic FGFs circulate but are inactive for nonmetabolic tissues or cells that often undergo active tissue remodeling via the renewed cycles of cell proliferation and population growth because of the lack of critical transmembrane accessory coreceptors. This divergence necessitates a distinction of the metabolic axis that is a term as we call hereafter, which the metabolic FGFs drive, from the mitogenic axis that the mitogenic FGFs drive. The metabolic axis still shares the major aspects of structural coevolution [9,10] while gaining unique structural and functional divergence with the mitogenic axis within each subfamily (Table 1), as our recent structural studies have revealed [2,5,11]. From the evolutionary standpoint, although the two axes largely parallel and drive differential effects via divergent intracellular mechanisms, they aim for a common goal of promoting the survival and homeostasis

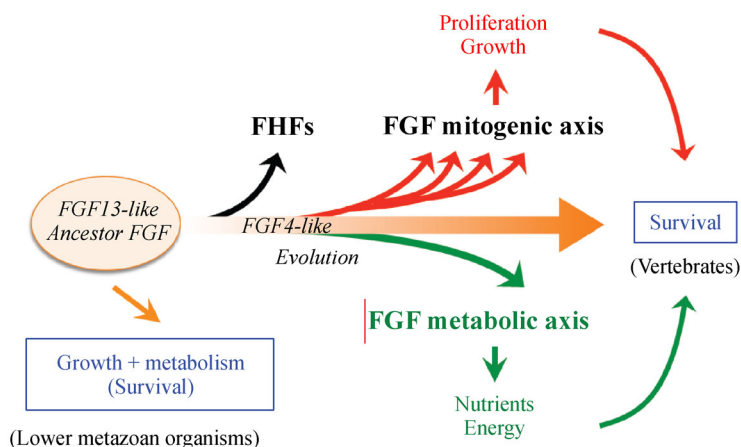
of each cell/tissue system and the organism as a whole (Fig. 1), as we have summarized in a previous review [7].

## The mitogenic FGF axis

The classic FGF family consists of 17 structurally related polypeptides, which are secreted and act as extracellular signaling molecules, in humans [1,3,7,12]. For the most part of FGF history beginning in the late 1970s [13,14], FGFs are known as short-range mitogens in a wide variety of cell types in the developing ectoderm, mesoderm, and endoderm. FGFs elicit a chemoattractant activity to promote cell migration and tissue remodeling and antiapoptotic effects to promote cell survival. FGF1 and 2 are the prototypes that are initially isolated based on potent mitogenic activity toward fibroblasts or fibroblast-like cells [13,14]. It was found early that the mitogenic FGFs bind tightly to the local extracellular matrix HS chains, do not circulate, and accordingly act in a paracrine or autocrine mode. This heparin/HS binding property renders their potent activity temporarily contained but timely released locally upon injury or demand of tissue remodeling [15]. These mitogenic FGFs include 14 members (Table 1), which strongly promote genomic DNA synthesis and subsequent cell division and population growth [12,16,17]. Therefore, mitogenic FGFs play critical roles in the development of multiple tissues/organs [18–20]. They initiate the mitogenic axis by binding to the Ig-like ectodomains of their cognate transmembrane

FGFRs in complex with HS motifs on diverse target cells and tissues in the first step [1,21,22]. The subsequent activation of the intracellular kinase domains of FGFRs results in downstream signal relay primarily through the PI3K-AKT, RAS-MAPK, and PLC $\gamma$ -PKC pathways [23–25], as we have summarized previously [3]. These HS and FGFR dependent activities driven by the mitogenic FGF axes contribute not only to the regulation of virtually all aspects of development and organogenesis but also to many natural processes of active post-developmental tissue repair, remodeling, and homeostasis [26].

Among mitogenic FGFs, FGF7, which is also known as keratinocyte growth factor (KGF), has the highest specificity for receptor isoforms [12,22,27]. FGF7 only activates the IIIb-type isoform of FGFR2. Given that FGF7 is produced in mesenchyme cells, while FGFR2IIIb resides on the epithelial or keratinocyte cells, FGF7 forms a unidirectional paracrine communication axis with FGFR2 from mesenchyme to epithelium compartment within a tissue or organ. On the other hand, epithelial cells secrete specific FGFs (e.g., FGF1 or 9), which then acts on mesenchymal cells that harbor FGFR1IIIc within two compartmental tissues. Therefore, these FGF1 and FGF7 driven mutual cell communication axes are poised to drive tissue remodeling and maintain tissue homeostasis [28]. The prolonged or abnormal activation of the FGFR-HS binary complexes by mitogenic FGF axes contributes to an array of cell/tissue-specific developmental diseases and multiple cancer types [3,29] (see a brief summary in Table 1). The proliferation- and survival-promoting



**Fig. 1** Scheme of FGF metabolic axis evolution. The FGF family originates from a common FGF13-like ancestor molecule in early metazoans that bifurcates into the so-called intracrine FGF-homologous factor (FHF) subgroup (black arrow), including FGF11, 12, 13, and 14 (not shown), and FGF4-like molecule, which continues to bifurcate into two major functional subgroups with diverging structural and functional specifications. The so-called mitogenic FGF subgroups, including the FGF5, 8, 9, and 10 subfamilies (red arrows, Table 1), bind extracellular matrix heparan sulfate and drive autocrine/paracrine mitogenic signal axes to promote cell proliferation and population growth. By contrast, the endocrine FGF subgroup members (green arrow, Table 2), including FGF19, 21, and 23, drive metabolic signal axes that elicit broad-spectrum functions in regulating the metabolic homeostasis of bile acid, lipids, glucose, energy, and minerals without direct proliferation-promoting activity. However, both the FGF mitogenic and FGF metabolic axes are designed to promote cell and organismal survival in the vertebrates (orange arrows and blue-colored font).

activities of the diverse mitogenic FGF axes have been a major focus of utilities as regenerative and repair agents in a range of medical settings [30–34]. In the past, we demonstrated the benefits of the application of mitogenic FGFs to tissue damage complications of diabetes mellitus, including diabetic cardiomyopathy, nephropathy, and neuropathy [35–37], as well as to wound healing and spinal cord injury repair [38–40]. On the other hand, the mitogenic FGF mediated cell miscommunications have also been on the menu for developing inhibitors to be used in cancer therapy [29,41,42].

It should be pointed out that, although at a physiological level mitogenic FGFs are not evolutionarily designed to circulate and target distal tissues or organs for an endocrine effect, at pharmacological or supraphysiological levels, mitogenic FGFs exert certain regulatory activities beyond promoting cell proliferation and growth possibly due to their accumulation sufficiently to achieve an effect in distal metabolic tissues, where a cognate FGFR isotype is expressed. It was shown in the early 1990s that a bolus intravenous injection of FGF1 or 2 could target vascular endothelium to decrease arterial blood pressure [43]. FGF16 is expressed in classical brown fat depots during the later stages of embryonic development, and recombinant FGF16 is a mitogen for adipocytes [44]. Mice overexpressing FGF16 delivered by adeno-associated virus display dramatic weight loss and uncoupling protein-1 (UCP1) upregulation in inguinal white adipose tissue (WAT), which is a common site for emergent active brown adipose tissue (BAT). These effects are likely a combined result of reduced food and water intake and abnormal feces replete with lipid and bile acid due to the brain, liver, and intestinal actions of overexpressed FGF16 [45]. Mice deficient in FGF1 exhibit insignificant phenotypes under standard dietary conditions; however, under a chronic high-fat diet, these mice develop an aggressive diabetic phenotype coupled with aberrant adipose phenotypes, including multiple histopathologies in the adipose vasculature network, accentuated inflammatory response, aberrant adipocyte size distribution and expansion, and ectopic expression of pancreatic lipases [46]. In particular, we show by structure-based mutagenesis that FGF1 can be designed to have full metabolic activity of wild-type FGF1 but with reduced proliferative potential both *in vitro* and *in vivo* [47]. These studies underscore the important role of FGF1 in maintaining local adipose tissue homeostasis, which upon significant tissue perturbations impinges on the metabolic functions that subsequently affect the systemic metabolic state. Thus, the metabolic effects of several mitogenic FGF axes may be due to either a local function in maintaining cellular homeostasis that is closely associated with local metabolic state at a physiological concentration or an induced metabolic response to a supraphysiological concentration from circulation in the

metabolic tissues or organs where FGFR resides. However, at pharmacological levels, few mitogenic FGFs may also be designed to elicit systemic metabolic effects.

## The metabolic FGF axis

In contrast to mitogenic FGFs, the metabolic FGF subfamily contains only three members, namely, FGF19 (mouse FGF15), 21, and 23 [2,7,48–51]. However, the metabolic axes of these three FGFs regulate a wide range of metabolic pathways, resulting in tissue and organismal metabolic homeostasis of bile acids, lipid, glucose, energy, and minerals. Although the metabolic FGF axes do not overtly promote DNA synthesis, thereby leading to cell proliferation [12,52,53], both metabolic and mitogenic FGF axes appear to enhance cell survival and promote an optimal state of homeostasis in the target tissues and organisms [7].

Based on current knowledge, the metabolic FGFs appear to originate from a common FGF13-like ancestor molecule as mitogenic FGFs and then bifurcate in early evolution through an FGF4-like molecule from all other mitogenic members by acquiring unique structural and mechanistic properties [5,10,11,54], thereby leading to specific activities in modulating metabolic states in specific cell and tissue types [2]. Instead of acting locally, metabolic FGFs take a hormonal or endocrine route of action by traveling through circulation from the originating tissue to other peripheral tissues/organs. This endocrine action can be attributed to the loss of the structurally conserved HS-binding domain characteristic of the mitogenic FGFs [5]. Both the expression and target tissues of the metabolic FGFs are relatively limited to the metabolically active endocrine organs, such as liver, intestine, adipose tissue, pancreas, muscle, bone, kidney, heart, parathyroid, and specific neurons in specific regions of the central nervous system (CNS) [55,56]. In expression tissues, metabolic FGF genes are subject to direct transcriptional control by several major metabolite-responsive nuclear receptors, including farnesoid X receptor (FXR), peroxisome proliferator-activated receptor  $\alpha$  (PPARA) and  $\gamma$  (PPARG), carbohydrate-response element-binding protein (ChREBP), sterol regulatory element-binding protein-1c (SREBP1c), retinoic acid-related orphan receptor  $\alpha$  (RORA), liver X receptor  $\beta$  (LXRB), vitamin D receptor (VDR) [48–50,57–65], and stress-sensing transcription factors, such as ATF4 [66], depending on the location of specific nutrition/energy-sensing cells in specific tissues. In target tissues, the biological effects of the metabolic FGF axes are still mediated by FGFRs but in a different binary complex with a new transmembrane nonkinase accessory coreceptor, the  $\alpha$ -Klotho (KL) or  $\beta$ -KL (KLB) [5,11,55,67] (Table 2), to which mitogenic FGFs do not bind.

**Table 1** The mitogenic FGF axis

Subfamily	Ligand member	Physiological function (knockout phenotypes)	Known pathologies	Receptor specificity						
				1b	1c	2b	2c	3b	3c	4
FGF1	FGF1	Adipose tissue homeostasis	Amplification — ovarian cancer	✓	✓	✓	✓	✓	✓	✓
	FGF2	Wound healing and angiogenesis	Overexpression — several cancer types	✓	✓	✓	✓	✓	✓	✓
	FGF4	Limb bud and heart development	Amplification — breast cancer	✓	✓	✓	✓	✓	✓	✓
	FGF5	Hair follicle growth and development	Overexpression — glioblastoma	✓	✓	✓	✓	✓	✓	✓
	FGF6	Muscle development and regeneration	Overexpression — prostate cancer	✓	✓	✓	✓	✓	✓	✓
	FGF3	Inner ear and skeleton development	1. Missense mutation — Michel aplasia, LAMM syndrome 2. Haploinsufficiency — otodental syndrome 3. Amplification — breast cancer	✓		✓				
FGF7	FGF7	Branching morphogenesis	1. Polymorphism — COPD 2. Overexpression — lung adenocarcinoma			✓				
	FGF10	1. Lung branching morphogenesis 2. Inner ear development 3. Hair follicle development 4. Fore and hind limb	1. Polymorphism — myopia 2. Nonsense mutation — LADD syndrome and ALSG 3. Overexpression — breast and prostate cancer	✓		✓				
	FGF22	Synaptogenesis	Undefined	✓		✓				
FGF8	FGF8	Brain, eye, ear, limb bud, kidney, and heart development	1. Missense mutation — cleft lip and palate, holoprosencephaly, craniofacial defects, and hypothalamo-pituitary dysfunction 2. Nonsense mutation — familial hypogonadotropic hypogonadism		✓		✓		✓	✓
	FGF17	Cerebellum and frontal cortex development	1. Missense mutation — familial hypogonadotropic hypogonadism 2. Overexpression — liver and prostate cancer		✓		✓		✓	✓
FGF18	FGF18	Lung alveolar and bone, CNS, skeletal, and palate development	1. Polymorphism — cleft lip and palate 2. Overexpression — liver cancer						✓	✓
	FGF9	Inner ear, gonad, and kidney development	1. Promoter mutation — sertoli cell-only syndrome 2. Missense mutation — multisynostosis syndrome 3. Mutations — colorectal and endometrial cancers 4. Overexpression — lung cancer		✓		✓		✓	✓
FGF16	FGF16	Heart development	1. Nonsense mutation — 4-5 metacarpal fusion 2. Overexpression — ovarian cancer				✓		✓	✓
	FGF20	Kidney, hair, teeth, cochlea, and central nervous development	1. Frame-shift mutation — bilateral renal agenesis 2. Polymorphism — risk of Parkinson's disease		✓		✓		✓	✓

Abbreviations: LAMM, labyrinthine aplasia, microtia, and microdontia; COPD, chronic obstructive pulmonary disease; LADD, lacrimo-auriculo-dento-digital syndrome; ALSG, aplasia of the lacrimal and salivary glands; and CNS, central nervous system.

Structurally, metabolic FGFs coevolve with coreceptor KL/KLB but also acquire new structural elements that direct specific contact interactions with KL/KLB and FGFRs, thereby leading to a tethered basic triad complex and subsequent activation of intracellular kinase domains of FGFRs [5,11]. The C-terminus of the metabolic FGFs mimics the interaction mode of a sugar chain that docks into the pseudo-glycolytic pocket of KL/KLB while interacting with FGFR ectodomains through the domains that are conserved across the FGF family [5,68]. Meanwhile, the interacting KL/KLB protrudes an “arm” from the membrane-proximal glycosidase domain gripping onto the FGFR ectodomain.

Although the FGFRs, in particular FGFR1, are broadly expressed, the highly restricted expression of KL/KLB and metabolic FGFs, and the new structural elements and mutual interaction modes, set the tone for tissue-specific functions of the metabolic FGF axes (Table 2). The different intracellular molecular constituents in different cells types, which are tailored to perform specific biological functions, may be also an important limiting

factor. For instance, the adult adipocytes are not poised in a normal context to increase population by direct proliferation due to the loss of several key proliferation-controlling pathways, thereby partly accounting for the inability of the activated FGFR1 by FGF21 to promote adipocyte proliferation. Overall, metabolic FGFs appear to be inducible stress factors in response to organismal metabolic perturbations [7,69] and signal distal peripheral tissues through the FGFR-KL/KLB complex to control due metabolic pathways. In this sense, the metabolic FGF acts as a key to ignite the FGF-FGFR-KLB/KLB triad complex, which functions similarly as an engine with an axis to drive effects in a tissue-specific manner, thereby leading to beneficial effects that offset the initial adverse metabolic changes and prevent metaflammation and tissue damage not only in the FGF-producing tissues but also systemically [2,7] (Table 2). Consequently, both the analogs of endocrine FGFs and the agonists of FGF-KL/KLB have been actively pursued clinically for the prevention and treatment of a wide range of metabolic diseases and comorbidities [2,70–75].

**Table 2** The metabolic FGF axis.

Subfamily	Members of ligands	Physiological function (knockout phenotypes)	Known pathologies	Receptor specificity								
				1b	1c	2b	2c	3b	3c	4	KL	KLB
FGF19	FGF19	1. Bile acid metabolism	1. Bile acid diarrhea, IBD		✓		✓		✓	✓		✓
		2. Gall bladder filling	2. Cholestasis									
	3. Lipid and energy metabolism	3. Overexpression — liver cancer										
FGF21	FGF21	1. Lipid metabolism — lipolysis, fatty acid oxidation, lipogenesis	1. Obesity		✓				✓			✓
		2. Energy metabolism — uncoupling thermogenesis	2. Diabetes									
		3. Macronutrient preference	3. NAFLD									
		4. Starvation response and associated physiology	4. Hyperlipidemia									
		5. Insulin sensitivity and glucose homeostasis	5. Metabolic syndrome									
FGF23	FGF23	Phosphate, calcium, sodium, and vitamin D homeostasis	1. Activation mutation — autosomal dominant hypophosphatemic rickets and tumor-induced osteomalacia		✓				✓	✓	✓	
			2. Inactivation mutation — familial tumoral calcinosis									
			3. Increase — X-linked dominant hypophosphatemia, CKD									
			4. Decrease — GALNT3-related familial tumoral calcinosis									

Abbreviations: IBD, inflammatory bowel disease; NAFLD, nonalcoholic fatty liver disease; CKD, chronic kidney disease; and GALNT3, polypeptide N-acetylglucosaminyltransferase 3.

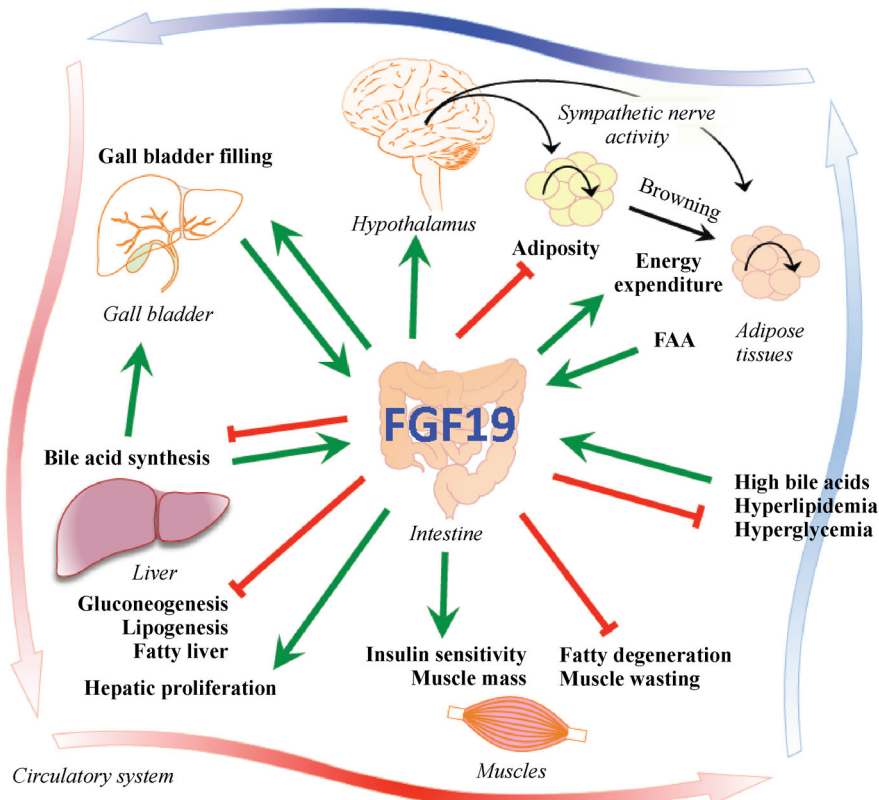
## FGF19 metabolic axis

FGF19 is the prime controller of diurnal bile acid flux, and the FGF19-driven metabolic axis is a temporal interorgan crosstalk from the ileum to the liver in response to the increase in the postprandial serum and transintestinal flux of bile acids [2,49] (Fig. 2). This axis serves to control the enterohepatic and systemic levels of bile acids negatively, which facilitate the uptake and absorption of dietary lipids after a meal but are toxic as biode detergent if the flux is prolonged at increased levels. The ileal initiation of the FGF19 signal is under the transcriptional control of FXR, which is stimulated by the reabsorbed enterocyte bile acids as a natural ligand that is originally released from gallbladder and mixed with food traveling down from the duodenum to jejunum and ileum. This enterocyte-derived FGF19 activates the remote FGFR4-KLB complex [67] residing across the membrane of hepatocytes in the liver, resulting in a major feedback termination of the transcription of the rate-limiting enzymes Cyp7A1 and Cyp8b1 in the bile acid biosynthesis pathways [49,76].

Therefore, the FGF19 axis triggers the shut-off of hepatic biosynthesis of new bile acids from cholesterol and the refilling of gallbladder approximately 2 h after the peak of serum bile acids is reached.

In experimental animals, FGF19 overexpression or administration elicits other metabolic effects [77,78]. Excessive FGF19 promotes lipolysis, metabolic rate, and energy expenditure and reduces body weight, serum glucose, and lipids. The FGFR1-KLB complex on adipose tissues, including WAT and BAT, was suggested in a large part to mediate these metabolic effects [79] (Fig. 2). However, the direct metabolic roles of bile acid fluctuation and bile acid-activated FXR and TGR5 cannot be excluded.

Although there is no evidence for any genetic mutation of FGF19 gene that may be involved in human metabolic diseases, its reduced synthesis and blood levels are suggestive of a causative factor of chronic bile acid diarrhea [80,81] and certain metabolic disorders, such as metabolic syndrome, nonalcoholic fatty liver disease (NAFLD), and insulin resistance. Experimentally, the



**Fig. 2** FGF19 metabolic axis. The major FGF19 metabolic axis drives a temporal interorgan crosstalk from the ileum to the liver in response to the increase in postprandial serum and transintestinal flux of bile acids to discontinue the biosynthesis of new bile acids after sufficient food digestion, thereby preventing the prolonged exposure of tissues to potential bile acid toxicity. Pharmacological FGF19 may also initiate multiple signal axes to drive effects on multiple tissues/organs, such as promoting (green arrow) energy expenditure in white and brown adipose tissues, increasing muscle mass and insulin sensitivity, and preventing (red long-tailed “T” sign) systemic hyperglycemia and hyperlipidemia. FAA: free fatty acids.

neutralization of FGF19 by specific anti-FGF19 antibodies causes severe diarrhea in monkeys accompanied by the increases in bile acid synthesis, serum and fecal total bile acids, specific bile acid transporters, and liver toxicity [82]. In obese patients who undergo Roux-en-Y gastric bypass bariatric surgery, FGF19 increases to normal values, which at least partially underlie the benefits of this approach [83]. On the other hand, high FGF19 expression levels are found in the livers of patients with extrahepatic cholestasis [84,85], suggesting FGF19 as a therapeutic target for this disease.

Recently, the FGF19 axis was shown to elicit hypertrophic and protective effects on the skeletal muscle presumably through a KLB-FGFR4-dependent mechanism by increasing myofiber size in the soleus, muscle mass, and grip strength [86]. Pharmacological FGF19 ameliorates skeletal muscle atrophy and prevents muscle wasting in mice with glucocorticoid treatment, obesity, or sarcopenia. These results highlight a potential treatment strategy for muscle wasting induced by glucocorticoid treatment, obesity, aging, and cachexia. However, whether the same treatment will have a similar adverse effect on the liver still has to be determined because muscle-specific transgenic mice developed prominent hepatocellular carcinoma (HCC) [87].

Despite the tumorigenic concern, the FGF19 analog NGM282 was tested in patients with nonalcoholic steatohepatitis (NASH). It markedly reduced liver fat content but with significant side effects [70]. In a phase 2 trial in patients with type 2 diabetes and chronic idiopathic constipation, NGM282 significantly improved bowel function by accelerating gastric emptying and colonic transit [81]. Furthermore, NGM282 was further tested in mouse models and human patients with cholestasis and primary biliary cholangitis, showing efficacy in significantly reducing bile acid levels and improving hepatic inflammatory injury and fibrosis [84,88,89].

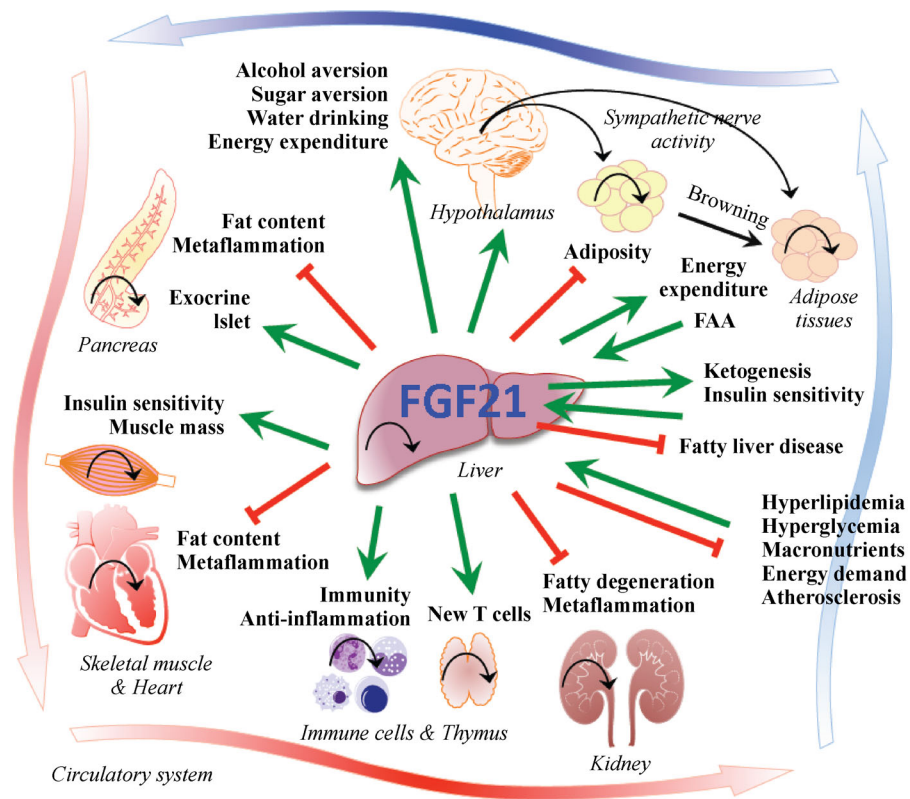
## FGF21 metabolic axis

FGF21 is a prime lipid catabolic factor that regulates energy balance. However, the physiological roles and pharmacological effects of FGF21-driven metabolic axes are multifaceted [2,7,90] (Fig. 3). FGF21 was discovered as a driver of glucose uptake in adipocytes and a PPAR $\alpha$ -dependent hepatic starvation hormone [48,50,51]. In mice, FGF21 levels are induced when calories are restricted or when glucose is low to allow fats to be burned for energy supply. The increasing levels of FGF21 drive diverse aspects of the adaptive starvation response, including stimulation of hepatic fatty acid oxidative for ketone body production during prolonged fasting and starvation. Whether this action of FGF21 is autocrine/paracrine in the liver or endocrine in adipose tissues through adipose lipolysis and fatty acid oxidation is a matter of debate. The

liver is a major contributor to the circulating FGF21 levels, which is associated with hepatic fat content and adiposity but inversely associated with serum glucose levels [91–93]. The hepatic expression of FGF21 is responsive not only to starvation but also to a broad spectrum of cellular, metabolic, or pathological changes in the liver as well as systemic metabolic perturbations [7,69,94]. As FGF21 is incapable of activating FGFR4-KLB complex [67], which is predominant in the liver that expresses FGFR1-KLB with lower levels, hepatic FGF21 acts mainly as an endocrine factor to drive the metabolic pathways in peripheral tissues, including WAT, BAT, muscle, heart, kidney, and CNS that express high levels of FGFR1/2/3-KLB, leading to the correction of metabolic derangements and amelioration of metaflammation and stress damage (Fig. 3) [7,94].

Although the liver is unlikely a major direct target of FGF21, the effects of FGF21 on the liver are prominent. In addition to its role as a regulator of integrated hepatic metabolism in multiple aspects [48,50,95–98], including fatty acid oxidation, ketogenesis, gluconeogenesis, and macronutrient preference, FGF21 counteracts hepatic pathologies in response to a number of nutritional and chemical insults, including ketogenic diet, high fat diet, high fructose diet, methionine and choline deficient diet, ethanol-supplemented diet, and diethylnitrosamine [99–103]. Under a chronic obesogenic diet, FGF21-deficient mice developed a spectrum of progressive fatty liver disease, including simple hepatosteatosis to NASH, fibrosis, and HCC, which is the most lethal complication of this disorder. These findings highlight the role of FGF21 metabolic axis as a defensive barrier for the deleterious stress damage caused by metabolic disorders in the liver [104]. Current clinical trials with FGF21 analogs show promising efficacy against NAFLD, NASH, and fibrosis without noticeable adverse side effects [73].

Acting on WAT and BAT, the FGF21 axis drives an array of catabolic effects, including insulin-independent glucose uptake, lipid droplet expansion inhibition, lipolysis, fatty acid oxidation, white adipocyte beigeing, and thermogenic dissipation of energy [79,105,106]. This route of action has been proposed as a major endocrine axis of FGF21 for insulin sensitization; lowering of systemic glucose, triacylglycerol, and LDL; fighting against obesity, diabetes, fatty liver diseases, hyperlipidemia, and associated comorbidities; and achieving metabolic health [2,73, 74,107]. Some of these effects are likely mediated by adipokines, such as CCL11 and adiponectin, as shown in mice [108,109]. In cold-induced nonshivering thermogenesis or exercise stress condition, BAT also becomes a source of endocrine FGF21 in a  $\beta$ -adrenergic- and cAMP-dependent manner, which in turn facilitates mitochondrial genesis, oxidative capacity, uncoupling, and heat generation, leading to adaptation to cold conditions and core body temperature maintenance [110–112].



**Fig. 3** FGF21 metabolic axis. The liver is the major organ of origin of endocrine FGF21 in response to a broad spectrum of stress conditions. The hepatic and pharmacological FGF21 drive multiple signal axes in multiple tissues/organs, resulting in multifaceted beneficiary metabolic effects, including promoting (green arrow) glucose, lipid, and energy homeostasis; offsetting metabolic derangements; and preventing (red long-tailed “T” sign) metaflammation, inflammatory tissue damage, and tissue-specific pathogenesis, including obesity, type 2 diabetes, fatty liver disease, metabolic syndrome, and associated comorbidities. FAA: free fatty acids. Black semicircular arrows indicate possibility of paracrine mode of FGF21 within local tissue environment.

In line with the beneficial effects of FGF21 on maintaining metabolic homeostasis during diverse adverse conditions, pharmacological FGF21 markedly extends the lifespan of mice by blunting the growth hormone/insulin-like growth factor-1 signaling pathway in the liver without reducing food intake or affecting longevity-associated markers of NAD<sup>+</sup> metabolism, AMP kinase, and mTOR signaling pathways [113]. The thymus functions in producing new T cells for the immune system, but with age, it becomes fatty and loses the ability to produce a sufficient amount of new T cells, which is an important cause of increased risks of infections, obesity, diabetes, and certain cancer types, leading to reduced lifespan in the elderly people. The FGF21 level in thymic epithelial cells is several folds higher than that in the liver. The high level of FGF21 is proposed to protect thymus from the age-related fatty degeneration and to increase the production of new T cells to bolster immune function, thereby lowering the incidence of diseases and promoting longevity [114].

The acinar cell compartment in the pancreas expresses

the highest levels of FGF21 constitutively among tissues, but contributes little to the circulation [56,115]. Acinar cells appear to be both the dominant source and target (via FGFR1-KLB complex) of pancreatic FGF21. The high levels of FGF21 is proposed to act as an exocrine pancreas secretagogue to stimulate pancreatic digestive enzyme secretion and pancreatic juice flow to the intestine, thereby relieving potential self-digestion caused proteostasis stress and protecting pancreas from pancreatitis, including but not limited to those caused by high-fat diet, pancreatic toxins, and alcoholism [116]. Although islets express significantly lower amounts of FGF21, acinar cell derived or endocrine FGF21 helps protect against fatty pancreas, high-fat diet induced islet hyperplasia, and inflammatory damage [117–119]. Demyelination in the CNS can cause severe neurological deficits, such as multiple sclerosis and neurological dysfunction. Pancreatic FGF21 acts on oligodendrocyte precursor cells to promote the remyelination process, leading to better recovery of neurological functions in mice [120].



Exposure to alcohol or sugar induces hepatic FGF21 through ChREBP, which then acts on the hypothalamus reward pathway to suppress the desire for sugar and alcohol in favor of drinking water in mice depending on the  $\beta$ -adrenergic circuit [97,98,121,122]. This finding may represent a new hydration pathway that is independent of the classical renin-angiotensin-aldosterone thirsty pathway in the kidney in response to nutritional stress, suggesting a previously underappreciated association of water intake to metabolism through the FGF21 metabolic axis. A human rs838133 allele in FGF21 is associated with higher alcohol and sugar intake and higher blood pressure and waist-hip ratio, with lower total body-fat percentage [123]. Comparison of the genomes of more than 105 000 light and heavy social drinkers also identifies a variation in the rs11940694 locus of the *KLB* gene in association with the aversion for alcohol [124]. Neuronal cell stress reaction, such as those caused by disturbances in the mitochondria and endoplasmic reticulum (ER), is an important factor in the development of neurodegenerative diseases. Studies found that the integrated stress response induces neuronal FGF21, which presumably serves to attenuate stress and neural damage [125].

In addition to the liver, pancreas, and adipose tissues, cardiac muscle produces FGF21 in response to cardiac stress, cardio exercise, and endurance training [126,127], which then speeds up glucose uptake, lipid catabolism, and energy metabolism, and protects against cardiovascular stress damage, apoptosis, and heart dysfunctions, such as cardiac hypertrophy, myopathy, steatosis, ischemic infarction, and atherosclerosis [128–132]. Through a multiorgan crosstalk, hepatic FGF21 drives the expression of angiotensin-converting enzyme 2 in adipocytes and renal cells, which hydrolyzes angiotensin II to active vasodilator angiotensin-(1-7) in the renin-angiotensin system, thereby alleviating angiotensin II-associated hypertension and reversing vascular damage [133]. Skeletal muscle under the bouts of exercise or stress, such as mitochondrial myopathies, also induces FGF21 expression [134–136]. In turn, FGF21 acts on muscle and adipose tissue to reduce lipid load by increasing lipolysis, fatty acid utilization, energy expenditure, and insulin sensitivity, thereby preventing diet-induced obesity and insulin resistance [137–140].

Hepatic FGF21 acts on the paraventricular nucleus in the hypothalamus to drive the release of corticotropin-releasing factor, which then stimulates the involuntary sympathetic nerve activity. This leads to the activation of brown adipose tissue by upregulating UCP1 and increases of glucose uptake, lipolysis, mitochondrial oxidation of fatty acids and glucose, body heat generation, and weight loss [141,142]. The increase in corticotropin-releasing factor levels may also stimulate the pituitary gland to release adrenocorticotrophic hormone and subsequent corticoster-

one production in adrenal cortex, leading to increased hepatic gluconeogenesis during prolonged fasting to prevent hypoglycemia [143]. Hepatic FGF21 acts on the suprachiasmatic nucleus (SCN) in the hypothalamus to suppress the vasopressin-kisspeptin and gonadotropin-releasing hormone signaling cascade, which then inhibits the proestrus surge in luteinizing hormone from anterior pituitary gland, thereby contributing to female infertility in response to nutritional challenge, such as prolonged starvation [144]. The SCN action of FGF21 may also alter circadian behavior [145]. By increasing neuropeptide Y levels and Y1 receptor activation, the hypothalamus action of FGF21 may decrease locomotive activity, metabolic rate, and body temperature, leading to torpor under nutrition limitation [146]. FGF21 may also act on the hippocampus to decrease reactive oxygen species and inflammatory damage, thus decreasing brain cell damage and improving cognition [147,148].

The endocrine FGF21 axes as well as the paracrine FGF21 axes within the local tissue compartments have been shown in many tissues and organs to counteract stress response and attenuate stress-ensued inflammation and inflammatory damage [7,104,117]. Therefore, FGF21 is not only a stress-responsive or -induced factor but also an anti-stress and anti-inflammatory factor. The stress-off-setting effects, in particular the anti-inflammatory activities, can be attributed to the metabolic effects of FGF21 axes that prevent fatty degeneration, gluco-lipototoxicity, oxidative and ER stress, and inflammatory and immune cell infiltration. These metabolic activities may be mediated in part through efficient and durable systemic and local glycemic and lipidemic control, improvement of insulin sensitivity, and promotion of lipid catabolism (lipolysis and fatty acid oxidation), adipose beigeing, and futile energy expenditure in adipose tissues, local adipocytes, and brain in both UCP1-dependent and adrenergic sympathetic nervous system-dependent mechanisms [79,105,106,141,149,150]. As a result, FGF21 effectively reverses hepatic steatosis in obese mice and clinical obese patients [73,105,151]. Furthermore, the pharmacological FGF21 analogs and FGFR1-KLB agonists have been shown to directly improve the spectrum of adverse components of metabolic syndrome, including central obesity, insulin resistance, fasting hyperglycemia, dyslipidemia, systemic hypertension, and fatty liver, which are the major risk factors for cardiovascular disease (CVD), type 2 diabetes mellitus, chronic kidney disease (CKD), and all-cause mortality [2,7,73,74,107]. The FGF21 axes suppress atherosclerotic plaque by reducing hypercholesterolemia, oxidative stress, and smooth muscle cell proliferation via adiponectin-dependent and adiponectin-independent mechanisms [129]. FGF21-deficient mice developed significant islet hyperplasia and periductal lymphocytic inflammation upon chronic challenge of an

obesogenic high-fat diet, indicating a protective role of FGF21 in compensatory islet hyperplasia and pancreatic inflammation associated with obesity [117,152]. FGF21 directly suppresses triglyceride levels and lipid accumulation in kidney tissues, thereby reducing lipotoxicity, oxidative stress, inflammation, glomerular abnormalities, fibrotic renal injury in diabetic nephropathy, while deficiency of FGF21 aggravates these conditions [153,154], indicating a defensive role of FGF21 against kidney pathogenesis associated with obesity and diabetes.

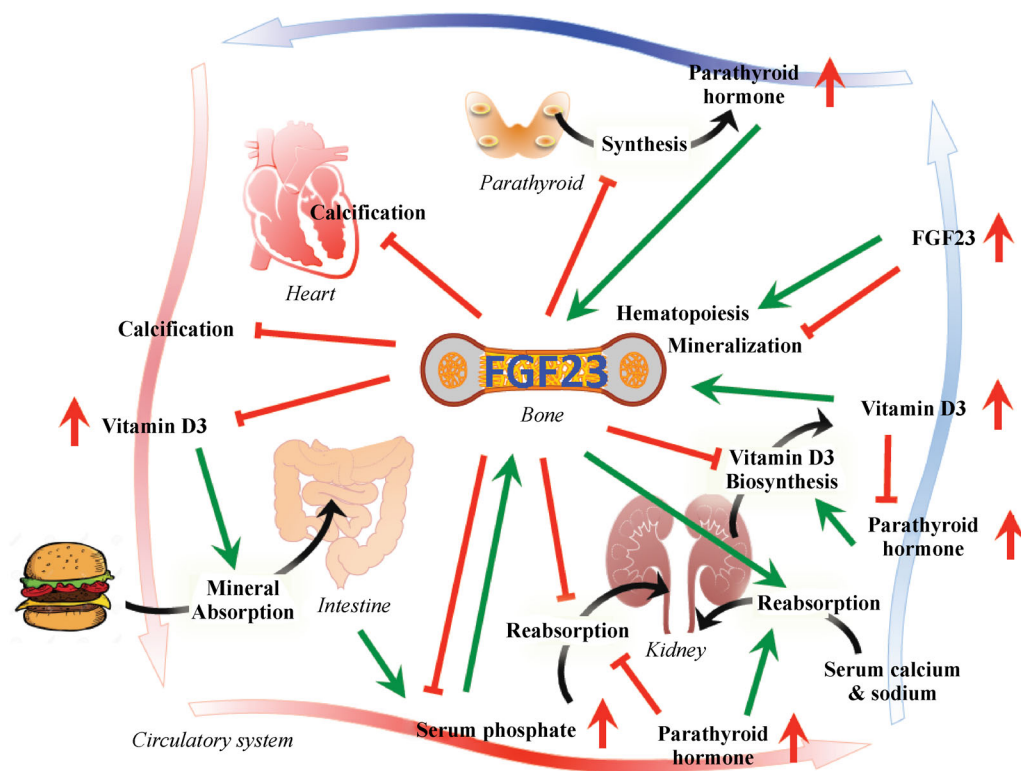
The anti-stress and anti-inflammatory effects of FGF21 may be also attributable to its direct action on non-metabolic cells and non-metabolic activities. FGF21 directly inhibits cardiomyocyte apoptosis, oxidative stress, myocardial injury, thereby reducing the risk of pathological cardiac remodeling and dysfunction, cardiac hypertrophy, myocardial ischemia, and heart failure in ischemic heart tissue and diabetic cardiomyopathy [130,155]. FGF21 protects the pancreas from caerulein- and L-arginine-induced pancreatitis, acinar cell injury, and fibrosis in mice [118,119,156]. FGF21 acts directly on renal mesangial cells to reduce glucose reabsorption and prevent hyperglycemia-induced fibrogenesis in *db/db* mice [157,158]. Interestingly, recent evidence supports that FGF21 can directly act on inflammatory and immune cells to attenuate inflammation and inflammatory damage. FGF21 activates THP-1-derived macrophages to promote cholesterol efflux, oxidized low-density lipoprotein (oxLDL) uptake, and foam cell formation and inhibits macrophage inflammatory capacity through the Nrf2 pathway [156,159,160]. Adipose tissue is an endocrine organ and plays an active role in the inflammation in obesity that can favor CVD and CKD progression by inducing a chronic and low-grade inflammation via secreted proinflammatory adipokines and cytokines. Studies in diet-induced obesity and pancreatitis models indicate that FGF21 promotes anti-inflammatory macrophage polarization in adipose depots and pancreas, WAT browning, and insulin sensitivity, thereby effectively preventing adipose tissues from adapting proinflammatory profiles and the pancreas from inflammatory fibrosis [109,156,159,161]. Interestingly, FGF21 was found highly expressed in neutrophils and monocytes among circulating leukocytes and stimulates phagocytosis, glucose uptake, and reactive oxygen species production in a NADPH oxidase-dependent manner in the neutrophil-like HL-60 and monocytic THP-1 cells [162–164]. In the type II collagen-induced arthritis mouse model, FGF21 acts on the spleen to reduce inflammatory IL-17, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-8, and MMP3 and the number of splenic TH17 cells, thereby alleviating arthritis severity [165]. These studies highlight the potential mediator role of FGF21 in innate immunity and inflammatory disorders. The direct impact of FGF21 on the function of inflammatory and immune cells and associated health consequences is yet to be validated.

## FGF23 metabolic axis

FGF23 is a key hormonal regulator of phosphate, vitamin D, and calcium metabolism, and its metabolic axes drive a complex interorgan crosstalk network for bone health and systemic mineral balance (Fig. 4) [2,61,166–168]. Osteoblastic cells in osseous tissue are the major source of FGF23 in response to elevated calcitriol, increased phosphate and calcium burdens, increased parathyroid hormone, iron and magnesium loss, and active bone remodeling in a vitamin D receptor dependent mechanism. Acting on kidneys that express the FGFR1-KL complex, the FGF23 signal axis represses the expression of NPT2a and NPT2c, the sodium-phosphate cotransporters in the proximal tubule, thereby decreasing reabsorption and increasing secretion of phosphate in renal brush border membrane vesicles. Another important function of this bone to kidney FGF23 signal axis is suppressing the expression of 25-hydroxyvitamin D3-1- $\alpha$ -hydroxylase and stimulating the expression of 1,25-dihydroxyvitamin D(3) 24-hydroxylase, thereby inhibiting the production of active calcitriol in renal proximal tubules, which subsequently inhibits the expression of NPT2b and phosphate absorption in the apical brush border of small intestine. The bone FGF23 acts on the basolateral FGFR1-KL complex in the renal distal tubules to increase the intracellular transport of fully glycosylated TRPV5 from the Golgi apparatus to the plasma membrane, thereby stimulating calcium reabsorption in distal renal tubules and preventing calcium loss [169]. These FGF23-associated axes also directly increase the membrane abundance of the Na<sup>+</sup>:Cl<sup>-</sup> cotransporter NCC in distal renal tubules, and thus, increase sodium reabsorption, plasma volume, and blood pressure [170]. This change may be a new cause of high blood pressure and heart disease under the modern processed phosphate-rich foods.

Bone FGF23 also acts on the parathyroid gland to inhibit the production and secretion of parathyroid hormone (Fig. 4) [171], which then reduces serum calcium through its effects on the bone, kidney, and intestine. High serum FGF23 levels in patients with CKD decrease calcitriol, thereby contributing to the development of secondary hyperparathyroidism, which has a crucial role in increasing the levels of FGF23 because the parathyroid hormone stimulates FGF23 expression.

Recent studies revealed the potential roles of the FGF23 axis in suppressing erythropoiesis in bone marrow. Erythroid progenitor cells highly express FGF23 and FGFR-KL, suggesting that they are both a source and a target of FGF23. The loss of FGF23 or injection of an FGF23-blocking peptide in mice results in increased erythropoiesis, reduced erythroid cell apoptosis, and increased renal and bone marrow erythropoietin (EPO) expression with increased circulating EPO levels. On the other hand, the increased EPO or acute blood loss increases



**Fig. 4** FGF23 metabolic axis. The bone-derived FGF23 drives signal axes to promote (green arrows) the metabolic homeostasis of phosphate, vitamin D, and calcium through a complex interorgan crosstalk network for bone health and systemic mineral balance. The bone to the kidney axis of FGF23 is central to the metabolic roles of FGF23, which inhibits (red long-tailed “T” sign) the reabsorption of phosphate and the production of active calcitriol in renal proximal tubules while increasing the calcium and sodium reabsorption in renal distal tubules. The bone to parathyroid axis of FGF23 inhibits the production and secretion of parathyroid hormone that also plays critical roles in mineral and vitamin D balance.

FGF23 expression in the bone marrow with a concomitant increase in serum FGF23 [172,173]. A recent study suggests that FGF23 is involved in the association between functional iron deficiency, increased EPO levels, and death. The further elucidation of the role of the EPO-FGF23 signaling axis in hereditary anemia and chronic hemolytic diseases, CKD, and mineralization disorders will add to the understanding of the pathophysiology of these diseases and life expectancy and will inform new treatment strategies for the diseases.

Current evidence indicates that FGF23 is more structurally unique than FGF19 and 21 [5]. FGF23 contains a conserved furin-sensitive  $^{176}\text{RHTR}^{179}$  cleavage site near the C-terminus, which inactivates the intact FGF23 upon cleavage, leading to signal attenuation. The biological importance of this activity control mechanism is demonstrated by point mutations (e.g., R176Q, R179Q, and R179W) of this site, which results in cleavage-resistant FGF23 and increased circulating levels of active FGF23, in autosomal dominant hypophosphatemic and vitamin D-deficient rickets characterized by renal phosphate wasting, hypophosphatemia, rickets, osteomalacia, leg deformities,

short stature, bone pain, and dental abscesses [168,174, 175]. FGF23 levels are increased and may play important roles in other hereditary and acquired phosphate wasting disorders, including X-linked dominant hypophosphatemic rickets, autosomal recessive hypophosphatemic rickets, hypophosphatemic rickets associated with McCune-Albright syndrome/fibrous dysplasia of bone, and linear sebaceous nevus syndrome [176,177]. The increased FGF23 levels are also found in acquired phosphate wasting disorders in some tumor types, such as the benign mesenchymal neoplasm phosphaturic mesenchymal tumor, causing tumor-induced osteomalacia, a paraneoplastic syndrome [168].

During post-translational modification, FGF23 is glycosylated at Thr-178 in the cleavage site by GalNT3, which facilitates its secretion and protects the protein from being broken down, suggesting a novel posttranslational regulatory model of FGF23 involving competing O-glycosylation and proteolytic processing to determine the level of secreted active FGF23 [178]. The importance of this glycosylation modification is demonstrated by inactivating GalNT3 mutations that render FGF23 susceptible to

proteolysis [179,180], thereby reducing circulating intact hormone levels and leading to autosomal recessive familial tumoral calcinosis that manifests with hyperphosphatemic and massive calcium deposits in the skin and subcutaneous tissues throughout the body. Consistently, at least seven mutations in the conserved backbone of FGF23, such as S71G, M96T, S129F, and F157L, destabilize the tertiary structure and render it susceptible to degradation, thereby resulting in autosomal recessive familial tumoral calcinosis with hyperphosphatemia [2,5,181–184].

Patients with CKD have increased serum levels of phosphate as well as FGF23, which lead to increased uptake of calcium by the kidneys, resulting in vascular calcification. This explains the CVD complications, such as cardiac hypertrophy and congestive heart failure, in patients with CKD [185,186]. The inhibition of FGF23 or its axis could be a strategy to bring CVD and vascular calcification under control. The FGF23 level in patients with CKD can even indicate their life expectancy. The dysregulation of calcium levels can have an array of serious health consequences. Chronic hypocalcemia can potentially lead to heart failure, nervous system and muscle disorders, and encephalopathy, while hypercalcemia can increase the risk of kidney stones, cause muscle weakness, and worsen psychological issues, such as dementia and depression. This may explain some current observations that people with high serum FGF23 can be at risk of dementia, and that mice lacking FGF23 exhibit defective learning and memory problems similar to those seen in KL-deficient mice [187,188].

## Conclusions and future perspectives

The three members of the metabolic FGFs, including FGF19, 21, and 23, share a conserved core structure of  $\beta$ -trefoil fold but diverge in functions from other mitogenic members of the FGF family during evolution (Fig. 1). These metabolic FGFs acquire specific structural elements that endow them with abilities to function via an endocrine mode and to bind new accessory receptors that have strict expression patterns in metabolic tissues. Although metabolic FGFs still signal through the transmembrane FGFR tyrosine kinases as the mitogenic FGFs, these new properties divert their functions to metabolic regulation. As such, FGF19, 21, and 23 drive a wide range of diverse metabolic axes that function in maintaining the homeostasis of bile acids, glucose, lipids, energy, and minerals; offsetting detrimental metabolic derangements; and achieving optimal metabolic health without an overt effect on cell proliferation and population growth. In this sense, each of the metabolic axes of FGF19, 21, and 23 stands alone as a driver of specific metabolic effects with important physiological functions and pathological consequences. Therefore, these axes together constitute the

“FGF Metabolic Axis,” which is a new term that we start to call hereafter, with broad-spectrum pathophysiological roles and consequences on the quality of survival (Fig. 1).

## Acknowledgements

I would like to acknowledge the long-term contributions of many members of my Wenzhou FGF team to the FGF field research as I cited in the text that made the idea of “The FGF Metabolic Axis” possible. I thank Dr. Yongde Luo for the expert assistance on the conceptual and practical aspects of the manuscript and Dr. Jin-San Zhang and Dr. Jian Xiao for their assistance. I apologize to those whose works have not been cited here due to the limited discussion scope on this evolving field. This work is supported by the National Key R&D Program of China (No. 2017YFA0506000, Xiaokun Li).

## Compliance with ethics guidelines

Xiaokun Li declares no conflict of interests. This manuscript is a review article and does not involve a research protocol requiring approval by the relevant institutional review board or ethics committee.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made.

The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.

To view a copy of this licence, visit <https://creativecommons.org/licenses/by/4.0/>.

## References

1. Beenken A, Mohammadi M. The FGF family: biology, pathophysiology and therapy. *Nat Rev Drug Discov* 2009; 8(3): 235–253
2. Luo Y, Ye S, Li X, Lu W. Emerging structure-function paradigm of endocrine FGFs in metabolic diseases. *Trends Pharmacol Sci* 2019; 40(2): 142–153
3. Li X, Wang C, Xiao J, McKeehan WL, Wang F. Fibroblast growth factors, old kids on the new block. *Semin Cell Dev Biol* 2016; 53: 155–167
4. Eriksson AE, Cousens LS, Weaver LH, Matthews BW. Three-dimensional structure of human basic fibroblast growth factor. *Proc Natl Acad Sci USA* 1991; 88(8): 3441–3445
5. Chen G, Liu Y, Goetz R, Fu L, Jayaraman S, Hu MC, Moe OW, Liang G, Li X, Mohammadi M.  $\alpha$ -Klotho is a non-enzymatic

- molecular scaffold for FGF23 hormone signalling. *Nature* 2018; 553(7689): 461–466
6. Degirolamo C, Sabbà C, Moschetta A. Therapeutic potential of the endocrine fibroblast growth factors FGF19, FGF21 and FGF23. *Nat Rev Drug Discov* 2016; 15(1): 51–69
  7. Luo Y, Ye S, Chen X, Gong F, Lu W, Li X. Rush to the fire: FGF21 extinguishes metabolic stress, metaflammation and tissue damage. *Cytokine Growth Factor Rev* 2017; 38: 59–65
  8. McKeehan WL, Wang F, Kan M. The heparan sulfate-fibroblast growth factor family: diversity of structure and function. *Prog Nucleic Acid Res Mol Biol* 1998; 59: 135–176
  9. Itoh N, Ornitz DM. Evolution of the Fgf and Fgfr gene families. *Trends Genet* 2004; 20(11): 563–569
  10. Itoh N, Ornitz DM. Fibroblast growth factors: from molecular evolution to roles in development, metabolism and disease. *J Biochem* 2011; 149(2): 121–130
  11. Luo Y, Lu W, Li X. Unraveling endocrine FGF signaling complex to combat metabolic diseases. *Trends Biochem Sci* 2018; 43(8): 563–566
  12. Zhang X, Ibrahim OA, Olsen SK, Umemori H, Mohammadi M, Ornitz DM. Receptor specificity of the fibroblast growth factor family. The complete mammalian FGF family. *J Biol Chem* 2006; 281(23): 15694–15700
  13. Armelin HA. Pituitary extracts and steroid hormones in the control of 3T3 cell growth. *Proc Natl Acad Sci USA* 1973; 70(9): 2702–2706
  14. Gospodarowicz D. Localisation of a fibroblast growth factor and its effect alone and with hydrocortisone on 3T3 cell growth. *Nature* 1974; 249(453): 123–127
  15. Burgess WH, Maciag T. The heparin-binding (fibroblast) growth factor family of proteins. *Annu Rev Biochem* 1989; 58(1): 575–606
  16. Luo Y, Ye S, Kan M, McKeehan WL. Control of fibroblast growth factor (FGF) 7- and FGF1-induced mitogenesis and downstream signaling by distinct heparin octasaccharide motifs. *J Biol Chem* 2006; 281(30): 21052–21061
  17. Gospodarowicz D, Ill CR, Hornsby PJ, Gill GN. Control of bovine adrenal cortical cell proliferation by fibroblast growth factor. Lack of effect of epidermal growth factor. *Endocrinology* 1977; 100(4): 1080–1089
  18. Mansour SL, Goddard JM, Capocchi MR. Mice homozygous for a targeted disruption of the proto-oncogene int-2 have developmental defects in the tail and inner ear. *Development* 1993; 117(1): 13–28
  19. Guo C, Sun Y, Zhou B, Adam RM, Li X, Pu WT, Morrow BE, Moon A, Li X. A Tbx1-Six1/Eya1-Fgf8 genetic pathway controls mammalian cardiovascular and craniofacial morphogenesis. *J Clin Invest* 2011; 121(4): 1585–1595
  20. Ornitz DM, Marie PJ. Fibroblast growth factor signaling in skeletal development and disease. *Genes Dev* 2015; 29(14): 1463–1486
  21. Kan M, Wang F, Xu J, Crabb JW, Hou J, McKeehan WL. An essential heparin-binding domain in the fibroblast growth factor receptor kinase. *Science* 1993; 259(5103): 1918–1921
  22. Ye S, Luo Y, Lu W, Jones RB, Linhardt RJ, Capila I, Toida T, Kan M, Pelletier H, McKeehan WL. Structural basis for interaction of FGF-1, FGF-2, and FGF-7 with different heparan sulfate motifs. *Biochemistry* 2001; 40(48): 14429–14439
  23. Goetz R, Mohammadi M. Exploring mechanisms of FGF signalling through the lens of structural biology. *Nat Rev Mol Cell Biol* 2013; 14(3): 166–180
  24. Kouhara H, Hadari YR, Spivak-Kroizman T, Schilling J, Bar-Sagi D, Lax I, Schlessinger J. A lipid-anchored Grb2-binding protein that links FGF-receptor activation to the Ras/MAPK signaling pathway. *Cell* 1997; 89(5): 693–702
  25. Huang Z, Marsiglia WM, Basu Roy U, Rahimi N, Ilghari D, Wang H, Chen H, Gai W, Blais S, Neubert TA, Mansukhani A, Traaseth NJ, Li X, Mohammadi M. Two FGF receptor kinase molecules act in concert to recruit and transphosphorylate phospholipase C $\gamma$ . *Mol Cell* 2016; 61(1): 98–110
  26. Dorey K, Amaya E. FGF signalling: diverse roles during early vertebrate embryogenesis. *Development* 2010; 137(22): 3731–3742
  27. Lu W, Luo Y, Kan M, McKeehan WL. Fibroblast growth factor-10. A second candidate stromal to epithelial cell andromedin in prostate. *J Biol Chem* 1999; 274(18): 12827–12834
  28. Jin C, Wang F, Wu X, Yu C, Luo Y, McKeehan WL. Directionally specific paracrine communication mediated by epithelial FGF9 to stromal FGFR3 in two-compartment premalignant prostate tumors. *Cancer Res* 2004; 64(13): 4555–4562
  29. Carter EP, Fearon AE, Grose RP. Careless talk costs lives: fibroblast growth factor receptor signalling and the consequences of pathway malfunction. *Trends Cell Biol* 2015; 25(4): 221–233
  30. Goldberg JD, Zheng J, Castro-Malaspina H, Jakubowski AA, Heller G, van den Brink MR, Perales MA. Palifermin is efficacious in recipients of TBI-based but not chemotherapy-based allogeneic hematopoietic stem cell transplants. *Bone Marrow Transplant* 2013; 48(1): 99–104
  31. Uchi H, Igarashi A, Urabe K, Koga T, Nakayama J, Kawamori R, Tamaki K, Hirakata H, Ohura T, Furue M. Clinical efficacy of basic fibroblast growth factor (bFGF) for diabetic ulcer. *Eur J Dermatol* 2009; 19(5): 461–468
  32. Akita S, Akino K, Imaizumi T, Hirano A. Basic fibroblast growth factor accelerates and improves second-degree burn wound healing. *Wound Repair Regen* 2008; 16(5): 635–641
  33. Fu X, Shen Z, Chen Y, Xie J, Guo Z, Zhang M, Sheng Z. Randomised placebo-controlled trial of use of topical recombinant bovine basic fibroblast growth factor for second-degree burns. *Lancet* 1998; 352(9141): 1661–1664
  34. Maddaluno L, Urwyler C, Werner S. Fibroblast growth factors: key players in regeneration and tissue repair. *Development* 2017; 144(22): 4047–4060
  35. Zhao YZ, Zhang M, Wong HL, Tian XQ, Zheng L, Yu XC, Tian FR, Mao KL, Fan ZL, Chen PP, Li XK, Lu CT. Prevent diabetic cardiomyopathy in diabetic rats by combined therapy of aFGF-loaded nanoparticles and ultrasound-targeted microbubble destruction technique. *J Control Release* 2016; 223: 11–21
  36. Liang G, Song L, Chen Z, Qian Y, Xie J, Zhao L, Lin Q, Zhu G, Tan Y, Li X, Mohammadi M, Huang Z. Fibroblast growth factor 1 ameliorates diabetic nephropathy by an anti-inflammatory mechanism. *Kidney Int* 2018; 93(1): 95–109
  37. Li R, Li Y, Wu Y, Zhao Y, Chen H, Yuan Y, Xu K, Zhang H, Lu Y, Wang J, Li X, Jia X, Xiao J. Heparin-polyoxamer thermosensitive hydrogel loaded with bFGF and NGF enhances peripheral nerve regeneration in diabetic rats. *Biomaterials* 2018; 168: 24–37

38. Wu J, Zhu J, He C, Xiao Z, Ye J, Li Y, Chen A, Zhang H, Li X, Lin L, Zhao Y, Zheng J, Xiao J. Comparative study of heparin-polyoxamer hydrogel modified bFGF and aFGF for *in vivo* wound healing efficiency. *ACS Appl Mater Interfaces* 2016; 8(29): 18710–18721
39. Wu J, Ye J, Zhu J, Xiao Z, He C, Shi H, Wang Y, Lin C, Zhang H, Zhao Y, Fu X, Chen H, Li X, Li L, Zheng J, Xiao J. Heparin-based coacervate of FGF2 improves dermal regeneration by asserting a synergistic role with cell proliferation and endogenous facilitated VEGF for cutaneous wound healing. *Biomacromolecules* 2016; 17(6): 2168–2177
40. Wang Q, He Y, Zhao Y, Xie H, Lin Q, He Z, Wang X, Li J, Zhang H, Wang C, Gong F, Li X, Xu H, Ye Q, Xiao J. A thermosensitive heparin-polyoxamer hydrogel bridges aFGF to treat spinal cord injury. *ACS Appl Mater Interfaces* 2017; 9(8): 6725–6745
41. Katoh M. Therapeutics targeting FGF signaling network in human diseases. *Trends Pharmacol Sci* 2016; 37(12): 1081–1096
42. Liang G, Liu Z, Wu J, Cai Y, Li X. Anticancer molecules targeting fibroblast growth factor receptors. *Trends Pharmacol Sci* 2012; 33(10): 531–541
43. Cuevas P, Carceller F, Ortega S, Zazo M, Nieto I, Giménez-Gallego G. Hypotensive activity of fibroblast growth factor. *Science* 1991; 254(5035): 1208–1210
44. Konishi M, Mikami T, Yamasaki M, Miyake A, Itoh N. Fibroblast growth factor-16 is a growth factor for embryonic brown adipocytes. *J Biol Chem* 2000; 275(16): 12119–12122
45. Rulifson IC, Collins P, Miao L, Nojima D, Lee KJ, Hardy M, Gupte J, Hensley K, Samayoa K, Cam C, Rottman JB, Ollmann M, Richards WG, Li Y. *In vitro* and *in vivo* analyses reveal profound effects of fibroblast growth factor 16 as a metabolic regulator. *J Biol Chem* 2017; 292(5): 1951–1969
46. Jonker JW, Suh JM, Atkins AR, Ahmadian M, Li P, Whyte J, He M, Juguilon H, Yin YQ, Phillips CT, Yu RT, Olefsky JM, Henry RR, Downes M, Evans RMA. A PPAR $\gamma$ -FGF1 axis is required for adaptive adipose remodelling and metabolic homeostasis. *Nature* 2012; 485(7398): 391–394
47. Huang Z, Tan Y, Gu J, Liu Y, Song L, Niu J, Zhao L, Srinivasan L, Lin Q, Deng J, Li Y, Conklin DJ, Neubert TA, Cai L, Li X, Mohammadi M. Uncoupling the mitogenic and metabolic functions of FGF1 by tuning FGF1-FGF receptor dimer stability. *Cell Reports* 2017; 20(7): 1717–1728
48. Badman MK, Pissios P, Kennedy AR, Koukos G, Flier JS, Maratos-Flier E. Hepatic fibroblast growth factor 21 is regulated by PPAR $\alpha$  and is a key mediator of hepatic lipid metabolism in ketotic states. *Cell Metab* 2007; 5(6): 426–437
49. Inagaki T, Choi M, Moschetta A, Peng L, Cummins CL, McDonald JG, Luo G, Jones SA, Goodwin B, Richardson JA, Gerard RD, Repa JJ, Mangelsdorf DJ, Kliewer SA. Fibroblast growth factor 15 functions as an enterohepatic signal to regulate bile acid homeostasis. *Cell Metab* 2005; 2(4): 217–225
50. Inagaki T, Dutchak P, Zhao G, Ding X, Gautron L, Parameswara V, Li Y, Goetz R, Mohammadi M, Esser V, Elmquist JK, Gerard RD, Burgess SC, Hammer RE, Mangelsdorf DJ, Kliewer SA. Endocrine regulation of the fasting response by PPAR $\alpha$ -mediated induction of fibroblast growth factor 21. *Cell Metab* 2007; 5(6): 415–425
51. Kharitonov A, Shiyanova TL, Koester A, Ford AM, Micanovic R, Galbreath EJ, Sandusky GE, Hammond LJ, Moyers JS, Owens RA, Gromada J, Brozinick JT, Hawkins ED, Wroblewski VJ, Li DS, Mehrbod F, Jaskunas SR, Shanafelt AB. FGF-21 as a novel metabolic regulator. *J Clin Invest* 2005; 115(6): 1627–1635
52. Goetz R, Ohnishi M, Ding X, Kurosu H, Wang L, Akiyoshi J, Ma J, Gai W, Sidis Y, Pitteloud N, Kuro OM, Razzaque MS, Mohammadi M. Klotho co-receptors inhibit signaling by paracrine FGF8 subfamily ligands. *Mol Cell Biol* 32(10):1944–1954
53. Luo Y, Yang C, Lu W, Xie R, Jin C, Huang P, Wang F, McKeenan WL. Metabolic regulator  $\beta$ Klotho interacts with fibroblast growth factor receptor 4 (FGFR4) to induce apoptosis and inhibit tumor cell proliferation. *J Biol Chem* 2010; 285(39): 30069–30078
54. Itoh M, Nacher JC, Kuma K, Goto S, Kanehisa M. Evolutionary history and functional implications of protein domains and their combinations in eukaryotes. *Genome Biol* 2007; 8(6): R121
55. Kurosu H, Choi M, Ogawa Y, Dickson AS, Goetz R, Eliseenkova AV, Mohammadi M, Rosenblatt KP, Kliewer SA, Kuro-o M. Tissue-specific expression of  $\beta$ Klotho and fibroblast growth factor (FGF) receptor isoforms determines metabolic activity of FGF19 and FGF21. *J Biol Chem* 2007; 282(37): 26687–26695
56. Fon Tacer K, Bookout AL, Ding X, Kurosu H, John GB, Wang L, Goetz R, Mohammadi M, Kuro-o M, Mangelsdorf DJ, Kliewer SA. Research resource: comprehensive expression atlas of the fibroblast growth factor system in adult mouse. *Mol Endocrinol* 2010; 24(10): 2050–2064
57. Wang H, Qiang L, Farmer SR. Identification of a domain within peroxisome proliferator-activated receptor  $\gamma$  regulating expression of a group of genes containing fibroblast growth factor 21 that are selectively repressed by SIRT1 in adipocytes. *Mol Cell Biol* 2008; 28(1): 188–200
58. Iizuka K, Takeda J, Horikawa Y. Glucose induces FGF21 mRNA expression through ChREBP activation in rat hepatocytes. *FEBS Lett* 2009; 583(17): 2882–2886
59. Wang Y, Solt LA, Burris TP. Regulation of FGF21 expression and secretion by retinoic acid receptor-related orphan receptor  $\alpha$ . *J Biol Chem* 2010; 285(21): 15668–15673
60. Uebanso T, Taketani Y, Yamamoto H, Amo K, Tanaka S, Arai H, Takei Y, Masuda M, Yamanaka-Okumura H, Takeda E. Liver X receptor negatively regulates fibroblast growth factor 21 in the fatty liver induced by cholesterol-enriched diet. *J Nutr Biochem* 2012; 23(7): 785–790
61. Masuyama R, Stockmans I, Torrekens S, Van Looveren R, Maes C, Carmeliet P, Bouillon R, Carmeliet G. Vitamin D receptor in chondrocytes promotes osteoclastogenesis and regulates FGF23 production in osteoblasts. *J Clin Invest* 2006; 116(12): 3150–3159
62. Kolek OI, Hines ER, Jones MD, LeSueur LK, Lipko MA, Kiela PR, Collins JF, Haussler MR, Ghishan FK. 1 $\alpha$ ,25-Dihydroxyvitamin D3 upregulates FGF23 gene expression in bone: the final link in a renal-gastrointestinal-skeletal axis that controls phosphate transport. *Am J Physiol Gastrointest Liver Physiol* 2005; 289(6): G1036–G1042
63. Zhang Y, Lei T, Huang JF, Wang SB, Zhou LL, Yang ZQ, Chen XD. The link between fibroblast growth factor 21 and sterol regulatory element binding protein 1c during lipogenesis in hepatocytes. *Mol Cell Endocrinol* 2011; 342(1-2): 41–47
64. Liu TF, Tang JJ, Li PS, Shen Y, Li JG, Miao HH, Li BL, Song BL. Ablation of gp78 in liver improves hyperlipidemia and insulin

- resistance by inhibiting SREBP to decrease lipid biosynthesis. *Cell Metab* 2012; 16(2): 213–225
65. Muise ES, Azzolina B, Kuo DW, El-Sherbeini M, Tan Y, Yuan X, Mu J, Thompson JR, Berger JP, Wong KK. Adipose fibroblast growth factor 21 is up-regulated by peroxisome proliferator-activated receptor  $\gamma$  and altered metabolic states. *Mol Pharmacol* 2008; 74(2): 403–412
66. De Sousa-Coelho AL, Marrero PF, Haro D. Activating transcription factor 4-dependent induction of FGF21 during amino acid deprivation. *Biochem J* 2012; 443(1): 165–171
67. Yang C, Jin C, Li X, Wang F, McKeehan WL, Luo Y. Differential specificity of endocrine FGF19 and FGF21 to FGFR1 and FGFR4 in complex with KLB. *PLoS One* 2012; 7(3): e33870
68. Lee S, Choi J, Mohanty J, Sousa LP, Tome F, Pardon E, Steyaert J, Lemmon MA, Lax I, Schlessinger J. Structures of  $\beta$ -klotho reveal a ‘zip code’-like mechanism for endocrine FGF signalling. *Nature* 2018; 553(7689): 501–505
69. Luo Y, McKeehan WL. Stressed liver and muscle call on adipocytes with FGF21. *Front Endocrinol (Lausanne)* 2013; 4: 194
70. Harrison SA, Rinella ME, Abdelmalek MF, Trotter JF, Paredes AH, Arnold HL, Kugelmas M, Bashir MR, Jaros MJ, Ling L, Rossi SJ, DePaoli AM, Loomba R. NGM282 for treatment of non-alcoholic steatohepatitis: a multicentre, randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet* 2018; 391(10126): 1174–1185
71. Hirschfield GM, Chazouillères O, Drenth JP, Thorburn D, Harrison SA, Landis CS, Mayo MJ, Muir AJ, Trotter JF, Leeming DJ, Karsdal MA, Jaros MJ, Ling L, Kim KH, Rossi SJ, Somaratne RM, DePaoli AM, Beuers U. Effect of NGM282, an FGF19 analogue, in primary sclerosing cholangitis: a multicenter, randomized, double-blind, placebo-controlled phase II trial. *J Hepatol* 2019; 70(3): 483–493
72. Harrison SA, Rossi SJ, Paredes AH, Trotter JF, Bashir MR, Guy CD, Banerjee R, Jaros MJ, Owers S, Baxter BA, Ling L, DePaoli AM. NGM282 improves liver fibrosis and histology in 12 weeks in patients with nonalcoholic steatohepatitis. *Hepatology* 2019 Feb 25. [Epub ahead of print] doi: 10.1002/hep.30590
73. Sanyal A, Charles ED, Neuschwander-Tetri BA, Loomba R, Harrison SA, Abdelmalek MF, Lawitz EJ, Halegoua-DeMarzio D, Kundu S, Noviello S, Luo Y, Christian R. Pegbelfermin (BMS-986036), a PEGylated fibroblast growth factor 21 analogue, in patients with non-alcoholic steatohepatitis: a randomised, double-blind, placebo-controlled, phase 2a trial. *Lancet* 2018; 392(10165): 2705–2717
74. Talukdar S, Zhou Y, Li D, Rossulek M, Dong J, Somayaji V, Weng Y, Clark R, Lanba A, Owen BM, Brenner MB, Trimmer JK, Gropp KE, Chabot JR, Erion DM, Rolph TP, Goodwin B, Calle RA. A long-acting FGF21 molecule, PF-05231023, decreases body weight and improves lipid profile in non-human primates and type 2 diabetic subjects. *Cell Metab* 2016; 23(3): 427–440
75. Carpenter TO, Imel EA, Ruppe MD, Weber TJ, Klausner MA, Wooddell MM, Kawakami T, Ito T, Zhang X, Humphrey J, Insogna KL, Peacock M. Randomized trial of the anti-FGF23 antibody KRN23 in X-linked hypophosphatemia. *J Clin Invest* 2014; 124(4): 1587–1597
76. Yu C, Wang F, Kan M, Jin C, Jones RB, Weinstein M, Deng CX, McKeehan WL. Elevated cholesterol metabolism and bile acid synthesis in mice lacking membrane tyrosine kinase receptor FGFR4. *J Biol Chem* 2000; 275(20): 15482–15489
77. Fu L, John LM, Adams SH, Yu XX, Tomlinson E, Renz M, Williams PM, Soriano R, Corpuz R, Moffat B, Vandlen R, Simmons L, Foster J, Stephan JP, Tsai SP, Stewart TA. Fibroblast growth factor 19 increases metabolic rate and reverses dietary and leptin-deficient diabetes. *Endocrinology* 2004; 145(6): 2594–2603
78. Tomlinson E, Fu L, John L, Hultgren B, Huang X, Renz M, Stephan JP, Tsai SP, Powell-Braxton L, French D, Stewart TA. Transgenic mice expressing human fibroblast growth factor-19 display increased metabolic rate and decreased adiposity. *Endocrinology* 2002; 143(5): 1741–1747
79. Adams AC, Yang C, Coskun T, Cheng CC, Gimeno RE, Luo Y, Kharitonov A. The breadth of FGF21’s metabolic actions are governed by FGFR1 in adipose tissue. *Mol Metab* 2013; 2(1): 31–37
80. Walters JR, Tasleem AM, Omer OS, Brydon WG, Dew T, le Roux CW. A new mechanism for bile acid diarrhea: defective feedback inhibition of bile acid biosynthesis. *Clin Gastroenterol Hepatol* 2009; 7(11):1189–1194
81. Oduyebo I, Camilleri M, Nelson AD, Khemani D, Nord SL, Busciglio I, Burton D, Rhoten D, Ryks M, Carlson P, Donato L, Lueke A, Kim K, Rossi SJ, Zinsmeister AR. Effects of NGM282, an FGF19 variant, on colonic transit and bowel function in functional constipation: a randomized phase 2 trial. *Am J Gastroenterol* 2018; 113(5): 725–734
82. Pai R, French D, Ma N, Hotzel K, Plise E, Salphati L, Setchell KD, Ware J, Lauriault V, Schutt L, Hartley D, Dambach D. Antibody-mediated inhibition of fibroblast growth factor 19 results in increased bile acids synthesis and ileal malabsorption of bile acids in cynomolgus monkeys. *Toxicol Sci* 2012; 126(2): 446–456
83. Gerhard GS, Styer AM, Wood GC, Roesch SL, Petrick AT, Gabrielsen J, Strodel WE, Still CD, Argyropoulos G. A role for fibroblast growth factor 19 and bile acids in diabetes remission after Roux-en-Y gastric bypass. *Diabetes Care* 2013; 36(7): 1859–1864
84. Luo J, Ko B, Elliott M, Zhou M, Lindhout DA, Phung V, To C, Learned RM, Tian H, DePaoli AM, Ling L. A nontumorigenic variant of FGF19 treats cholestatic liver diseases. *Sci Transl Med* 2014; 6(247): 247ra100
85. Schaap FG, van der Gaag NA, Gouma DJ, Jansen PL. High expression of the bile salt-homeostatic hormone fibroblast growth factor 19 in the liver of patients with extrahepatic cholestasis. *Hepatology* 2009; 49(4): 1228–1235
86. Benoit B, Meugnier E, Castelli M, Chanon S, Vieille-Marchiset A, Durand C, Bendridi N, Pesenti S, Monternier PA, Durieux AC, Freyssenet D, Rieusset J, Lefai E, Vidal H, Ruzzin J. Fibroblast growth factor 19 regulates skeletal muscle mass and ameliorates muscle wasting in mice. *Nat Med* 2017; 23(8): 990–996
87. Nicholes K, Guillet S, Tomlinson E, Hillan K, Wright B, Frantz GD, Pham TA, Dillard-Telm L, Tsai SP, Stephan JP, Stinson J, Stewart T, French DM. A mouse model of hepatocellular carcinoma: ectopic expression of fibroblast growth factor 19 in skeletal muscle of transgenic mice. *Am J Pathol* 2002; 160(6): 2295–2307
88. Zhou M, Learned RM, Rossi SJ, DePaoli AM, Tian H, Ling L. Engineered fibroblast growth factor 19 reduces liver injury and resolves sclerosing cholangitis in Mdr2-deficient mice. *Hepatology* 2016; 63(3): 914–929

89. Mayo MJ, Wigg AJ, Leggett BA, Arnold H, Thompson AJ, Weltman M, Carey EJ, Muir AJ, Ling L, Rossi SJ, DePaoli AM. NGM282 for treatment of patients with primary biliary cholangitis: a multicenter, randomized, double-blind, placebo-controlled trial. *Hepatol Commun* 2018; 2(9): 1037–1050
90. BonDurant LD, Potthoff MJ. Fibroblast growth factor 21: a versatile regulator of metabolic homeostasis. *Annu Rev Nutr* 2018; 38(1): 173–196
91. Giannini C, Feldstein AE, Santoro N, Kim G, Kursawe R, Pierpont B, Caprio S. Circulating levels of FGF-21 in obese youth: associations with liver fat content and markers of liver damage. *J Clin Endocrinol Metab* 2013; 98(7): 2993–3000
92. Lin Z, Gong Q, Wu C, Yu J, Lu T, Pan X, Lin S, Li X. Dynamic change of serum FGF21 levels in response to glucose challenge in human. *J Clin Endocrinol Metab* 2012; 97(7): E1224–E1228
93. Yilmaz Y, Eren F, Yonal O, Kurt R, Aktas B, Celikel CA, Ozdogan O, Imeryuz N, Kalayci C, Avsar E. Increased serum FGF21 levels in patients with nonalcoholic fatty liver disease. *Eur J Clin Invest* 2010; 40(10): 887–892
94. Kliewer SA, Mangelsdorf DJ. A dozen years of discovery: insights into the physiology and pharmacology of FGF21. *Cell Metab* 2019; 29(2): 246–253
95. Laeger T, Henagan TM, Albarado DC, Redman LM, Bray GA, Noland RC, Münzberg H, Hutson SM, Gettys TW, Schwartz MW, Morrison CD. FGF21 is an endocrine signal of protein restriction. *J Clin Invest* 2014; 124(9): 3913–3922
96. Fisher FM, Kim M, Doridot L, Cunniff JC, Parker TS, Levine DM, Hellerstein MK, Hudgins LC, Maratos-Flier E, Herman MA. A critical role for ChREBP-mediated FGF21 secretion in hepatic fructose metabolism. *Mol Metab* 2017; 6(1): 14–21
97. von Holstein-Rathlou S, BonDurant LD, Peltekian L, Naber MC, Yin TC, Claflin KE, Urizar AI, Madsen AN, Ratner C, Holst B, Karstoft K, Vandenbeuch A, Anderson CB, Cassell MD, Thompson AP, Solomon TP, Rahmouni K, Kinnamon SC, Pieper AA, Gillum MP, Potthoff MJ. FGF21 mediates endocrine control of simple sugar intake and sweet taste preference by the liver. *Cell Metab* 2016; 23(2): 335–343
98. Talukdar S, Owen BM, Song P, Hernandez G, Zhang Y, Zhou Y, Scott WT, Paratala B, Turner T, Smith A, Bernardo B, Müller CP, Tang H, Mangelsdorf DJ, Goodwin B, Kliewer SA. FGF21 regulates sweet and alcohol preference. *Cell Metab* 2016; 23(2): 344–349
99. Fisher FM, Chui PC, Nasser IA, Popov Y, Cunniff JC, Lundasen T, Kharitonov A, Schuppan D, Flier JS and Maratos-Flier E. Fibroblast growth factor 21 limits lipotoxicity by promoting hepatic fatty acid activation in mice on methionine and choline-deficient diets. *Gastroenterology* 2014; 147(5): 1073–1083.e6
100. Huang X, Yu C, Jin C, Yang C, Xie R, Cao D, Wang F, McKeehan WL. Forced expression of hepatocyte-specific fibroblast growth factor 21 delays initiation of chemically induced hepatocarcinogenesis. *Mol Carcinog* 2006; 45(12): 934–942
101. Tanaka N, Takahashi S, Zhang Y, Krausz KW, Smith PB, Patterson AD, Gonzalez FJ. Role of fibroblast growth factor 21 in the early stage of NASH induced by methionine- and choline-deficient diet. *Biochim Biophys Acta* 2015; 1852(7): 1242–1252
102. Desai BN, Singhal G, Watanabe M, Stevanovic D, Lundasen T, Fisher FM, Mather ML, Vardeh HG, Douris N, Adams AC, Nasser IA, FitzGerald GA, Flier JS, Skarke C, Maratos-Flier E. Fibroblast growth factor 21 (FGF21) is robustly induced by ethanol and has a protective role in ethanol associated liver injury. *Mol Metab* 2017; 6(11): 1395–1406
103. Ye D, Wang Y, Li H, Jia W, Man K, Lo CM, Wang Y, Lam KS, Xu A. Fibroblast growth factor 21 protects against acetaminophen-induced hepatotoxicity by potentiating peroxisome proliferator-activated receptor coactivator protein-1 $\alpha$ -mediated antioxidant capacity in mice. *Hepatology* 2014; 60(3): 977–989
104. Singhal G, Kumar G, Chan S, Fisher FM, Ma Y, Vardeh HG, Nasser IA, Flier JS, Maratos-Flier E. Deficiency of fibroblast growth factor 21 (FGF21) promotes hepatocellular carcinoma (HCC) in mice on a long term obesogenic diet. *Mol Metab* 2018; 13: 56–66
105. Ye M, Lu W, Wang X, Wang C, Abbruzzese JL, Liang G, Li X, Luo Y. FGF21-FGFR1 coordinates phospholipid homeostasis, lipid droplet function, and ER stress in obesity. *Endocrinology* 2016; 157(12): 4754–4769
106. Foltz IN, Hu S, King C, Wu X, Yang C, Wang W, Weiszmann J, Stevens J, Chen JS, Nuanmanee N, Gupte J, Komorowski R, Sekirov L, Hager T, Arora T, Ge H, Baribault H, Wang F, Sheng J, Karow M, Wang M, Luo Y, McKeehan W, Wang Z, Véniant MM, Li Y. Treating diabetes and obesity with an FGF21-mimetic antibody activating the  $\beta$ Klotho/FGFR1c receptor complex. *Sci Transl Med* 2012; 4(162): 162ra153
107. Gaich G, Chien JY, Fu H, Glass LC, Deeg MA, Holland WL, Kharitonov A, Bumol T, Schilske HK, Moller DE. The effects of LY2405319, an FGF21 analog, in obese human subjects with type 2 diabetes. *Cell Metab* 2013; 18(3): 333–340
108. Lin Z, Tian H, Lam KS, Lin S, Hoo RC, Konishi M, Itoh N, Wang Y, Bornstein SR, Xu A, Li X. Adiponectin mediates the metabolic effects of FGF21 on glucose homeostasis and insulin sensitivity in mice. *Cell Metab* 2013; 17(5): 779–789
109. Huang Z, Zhong L, Lee JTH, Zhang J, Wu D, Geng L, Wang Y, Wong CM, Xu A. The FGF21–CCL11 axis mediates being of white adipose tissues by coupling sympathetic nervous system to type 2 immunity. *Cell Metab* 2017; 26(3): 493–508.e4
110. Lee P, Linderman JD, Smith S, Brychta RJ, Wang J, Idelson C, Perron RM, Werner CD, Phan GQ, Kammula US, Kebebew E, Pacak K, Chen KY, Celi FS. Irisin and FGF21 are cold-induced endocrine activators of brown fat function in humans. *Cell Metab* 2014; 19(2): 302–309
111. Hondares E, Iglesias R, Giralt A, Gonzalez FJ, Giralt M, Mampel T, Villarroya F. Thermogenic activation induces FGF21 expression and release in brown adipose tissue. *J Biol Chem* 2011; 286(15): 12983–12990
112. Ameka M, Markan KR, Morgan DA, BonDurant LD, Idiga SO, Naber MC, Zhu Z, Zingman LV, Grobe JL, Rahmouni K, Potthoff MJ. Liver derived FGF21 maintains core body temperature during acute cold exposure. *Sci Rep* 2019; 9(1): 630
113. Zhang Y, Xie Y, Berglund ED, Coate KC, He TT, Katafuchi T, Xiao G, Potthoff MJ, Wei W, Wan Y, Yu RT, Evans RM, Kliewer SA, Mangelsdorf DJ. The starvation hormone, fibroblast growth factor-21, extends lifespan in mice. *eLife* 2012; 1: e00065
114. Youm YH, Horvath TL, Mangelsdorf DJ, Kliewer SA, Dixit VD. Prolongevity hormone FGF21 protects against immune senescence by delaying age-related thymic involution. *Proc Natl Acad Sci*



- USA 2016; 113(4): 1026–1031
115. Adams AC, Coskun T, Cheng CC, O'Farrell LS, Dubois SL, Kharitononkov A. Fibroblast growth factor 21 is not required for the antidiabetic actions of the thiazolidinediones. *Mol Metab* 2013; 2(3): 205–214
  116. Coate KC, Hernandez G, Thorne CA, Sun S, Le TDV, Vale K, Kliewer SA, Mangelsdorf DJ. FGF21 is an exocrine pancreas secretagogue. *Cell Metab* 2017; 25(2): 472–480
  117. Singhal G, Fisher FM, Chee MJ, Tan TG, El Ouaamari A, Adams AC, Najarian R, Kulkarni RN, Benoist C, Flier JS, Maratos-Flier E. Fibroblast growth factor 21 (FGF21) protects against high fat diet induced inflammation and islet hyperplasia in pancreas. *PLoS One* 2016; 11(2): e0148252
  118. Johnson CL, Mehmood R, Laing SW, Stepniak CV, Kharitononkov A, Pin CL. Silencing of the fibroblast growth factor 21 gene is an underlying cause of acinar cell injury in mice lacking MIST1. *Am J Physiol Endocrinol Metab* 2014; 306(8): E916–E928
  119. Johnson CL, Weston JY, Chadi SA, Fazio EN, Huff MW, Kharitononkov A, Köester A, Pin CL. Fibroblast growth factor 21 reduces the severity of cerulein-induced pancreatitis in mice. *Gastroenterology* 2009; 137(5): 1795–1804
  120. Kuroda M, Muramatsu R, Maedera N, Koyama Y, Hamaguchi M, Fujimura H, Yoshida M, Konishi M, Itoh N, Mochizuki H, Yamashita T. Peripherally derived FGF21 promotes remyelination in the central nervous system. *J Clin Invest* 2017; 127(9): 3496–3509
  121. Soberg S, Sandholt CH, Jespersen NZ, Toft U, Madsen AL, von Holstein-Rathlou S, Grevenkoed TJ, Christensen KB, Bredie WLP, Potthoff MJ, Solomon TPJ, Scheele C, Linneberg A, Jorgensen T, Pedersen O, Hansen T, Gillum MP, Grarup N. FGF21 is a sugar-induced hormone associated with sweet intake and preference in humans. *Cell Metab* 2017; 25(5): 1045–1053.e6
  122. Song P, Zechner C, Hernandez G, Canovas J, Xie Y, Sondhi V, Wagner M, Stadlbauer V, Horvath A, Leber B, Hu MC, Moe OW, Mangelsdorf DJ, Kliewer SA. The hormone FGF21 stimulates water drinking in response to ketogenic diet and alcohol. *Cell Metab* 2018; 27(6): 1338–1347.e4
  123. Frayling TM, Beaumont RN, Jones SE, Yaghootkar H, Tuke MA, Ruth KS, Casanova F, West B, Locke J, Sharp S, Ji Y, Thompson W, Harrison J, Etheridge AS, Gallins PJ, Jima D, Wright F, Zhou Y, Innocenti F, Lindgren CM, Grarup N, Murray A, Freathy RM, Weedon MN, Tyrrell J, Wood AR. A common allele in FGF21 associated with sugar intake is associated with body shape, lower total body-fat percentage, and higher blood pressure. *Cell Reports* 2018; 23(2): 327–336
  124. Schumann G, Liu C, O'Reilly P, Gao H, Song P, Xu B, Ruggeri B, Amin N, Jia T, Preis S, Segura Lepe M, Akira S, Barbieri C, Baumeister S, Cauchi S, Clarke TK, Enroth S, Fischer K, Hällfors J, Harris SE, Hieber S, Hofer E, Hottenga JJ, Johansson Å, Joshi PK, Kaartinen N, Laitinen J, Lemaitre R, Loukola A, Luan J, Lyytikäinen LP, Mangino M, Manichaikul A, Mbarek H, Milaneschi Y, Moayyeri A, Mukamal K, Nelson C, Nettleton J, Partinen E, Rawal R, Robino A, Rose L, Sala C, Satoh T, Schmidt R, Schraut K, Scott R, Smith AV, Starr JM, Teumer A, Trompet S, Uitterlinden AG, Venturini C, Vergnaud AC, Verweij N, Vitart V, Vuckovic D, Wedenoja J, Yengo L, Yu B, Zhang W, Zhao JH, Boomsma DI, Chambers J, Chasman DI, Daniela T, de Geus E, Deary I, Eriksson JG, Esko T, Eulenburt V, Franco OH, Froguel P, Gieger C, Grabe HJ, Gudnason V, Gyllensten U, Harris TB, Hartikainen AL, Heath AC, Hocking L, Hofman A, Huth C, Jarvelin MR, Jukema JW, Kaprio J, Kooner JS, Kutalik Z, Lahti J, Langenberg C, Lehtimäki T, Liu Y, Madden PA, Martin N, Morrison A, Penninx B, Pirastu N, Psaty B, Raitakari O, Ridker P, Rose R, Rotter JL, Samani NJ, Schmidt H, Spector TD, Stott D, Strachan D, Tzoulaki I, van der Harst P, van Duijn CM, Marques-Vidal P, Vollenweider P, Wareham NJ, Whitfield JB, Wilson J, Wolfenbittel B, Bakalkin G, Evangelou E, Liu Y, Rice KM, Desrivieres S, Kliewer SA, Mangelsdorf DJ, Müller CP, Levy D, Elliott P. KLB is associated with alcohol drinking, and its gene product  $\beta$ -Klotho is necessary for FGF21 regulation of alcohol preference. *Proc Natl Acad Sci USA* 2016; 113(50): 14372–14377
  125. Restelli LM, Oettinghaus B, Halliday M, Agca C, Licci M, Sironi L, Savoia C, Hench J, Tolnay M, Neutzner A, Schmidt A, Eckert A, Mallucci G, Scorrano L, Frank S. Neuronal mitochondrial dysfunction activates the integrated stress response to induce fibroblast growth factor 21. *Cell Reports* 2018; 24(6): 1407–1414
  126. Planavila A, Redondo I, Hondares E, Vinciguerra M, Munts C, Iglesias R, Gabrielli LA, Sitges M, Giralt M, van Bilsen M, Villarroya F. Fibroblast growth factor 21 protects against cardiac hypertrophy in mice. *Nat Commun* 2013; 4(1): 2019
  127. Morville T, Sahl RE, Trammell SA, Svenningsen JS, Gillum MP, Helge JW, Clemmensen C. Divergent effects of resistance and endurance exercise on plasma bile acids, FGF19, and FGF21 in humans. *JCI Insight* 2018; 3(15): 122737
  128. Brahma MK, Adam RC, Pollak NM, Jaeger D, Zierler KA, Pöcher N, Schreiber R, Romauch M, Moustafa T, Eder S, Ruelicke T, Preiss-Landl K, Lass A, Zechner R, Haemmerle G. Fibroblast growth factor 21 is induced upon cardiac stress and alters cardiac lipid homeostasis. *J Lipid Res* 2014; 55(11): 2229–2241
  129. Lin Z, Pan X, Wu F, Ye D, Zhang Y, Wang Y, Jin L, Lian Q, Huang Y, Ding H, Triggle C, Wang K, Li X, Xu A. Fibroblast growth factor 21 prevents atherosclerosis by suppression of hepatic sterol regulatory element-binding protein-2 and induction of adiponectin in mice. *Circulation* 2015; 131(21): 1861–1871
  130. Liu SQ, Roberts D, Kharitononkov A, Zhang B, Hanson SM, Li YC, Zhang LQ, Wu YH. Endocrine protection of ischemic myocardium by FGF21 from the liver and adipose tissue. *Sci Rep* 2013; 3(1): 2767
  131. Yang H, Feng A, Lin S, Yu L, Lin X, Yan X, Lu X, Zhang C. Fibroblast growth factor-21 prevents diabetic cardiomyopathy via AMPK-mediated antioxidation and lipid-lowering effects in the heart. *Cell Death Dis* 2018; 9(2): 227
  132. Zhang C, Huang Z, Gu J, Yan X, Lu X, Zhou S, Wang S, Shao M, Zhang F, Cheng P, Feng W, Tan Y, Li X. Fibroblast growth factor 21 protects the heart from apoptosis in a diabetic mouse model via extracellular signal-regulated kinase 1/2-dependent signalling pathway. *Diabetologia* 2015; 58(8): 1937–1948
  133. Pan X, Shao Y, Wu F, Wang Y, Xiong R, Zheng J, Tian H, Wang B, Wang Y, Zhang Y, Han Z, Qu A, Xu H, Lu A, Yang T, Li X, Xu A, Du J, Lin Z. FGF21 prevents angiotensin II-induced hypertension and vascular dysfunction by activation of ACE2/angiotensin-(1–7) axis in mice. *Cell Metab* 2018; 27(6): 1323–1337.e5
  134. Kim KH, Jeong YT, Oh H, Kim SH, Cho JM, Kim YN, Kim SS, Kim DH, Hur KY, Kim HK, Ko T, Han J, Kim HL, Kim J, Back SH, Komatsu M, Chen H, Chan DC, Konishi M, Itoh N, Choi CS, Lee MS. Autophagy deficiency leads to protection from obesity

- and insulin resistance by inducing Fgf21 as a mitokine. *Nat Med* 2013; 19(1): 83–92
135. Suomalainen A, Elo JM, Pietiläinen KH, Hakonen AH, Sevastianova K, Korpela M, Isohanni P, Marjavaara SK, Tyni T, Kiuru-Enari S, Pihko H, Darin N, Öunap K, Kluijtmans LA, Paetau A, Buzkova J, Bindoff LA, Annunen-Rasila J, Uusimaa J, Rissanen A, Yki-Järvinen H, Hirano M, Tulinius M, Smeitink J, Tynismaa H. FGF-21 as a biomarker for muscle-manifesting mitochondrial respiratory chain deficiencies: a diagnostic study. *Lancet Neurol* 2011; 10(9): 806–818
  136. Geng L, Liao B, Jin L, Huang Z, Triggler CR, Ding H, Zhang J, Huang Y, Lin Z, Xu A. Exercise alleviates obesity-induced metabolic dysfunction via enhancing FGF21 sensitivity in adipose tissues. *Cell Rep* 2019; 26(10): 2738–2752.e4
  137. Pereira RO, Tadinada SM, Zasadny FM, Oliveira KJ, Pires KMP, Olvera A, Jeffers J, Souvenir R, Mcglauffin R, Seei A, Funari T, Sesaki H, Potthoff MJ, Adams CM, Anderson EJ, Abel ED. OPA1 deficiency promotes secretion of FGF21 from muscle that prevents obesity and insulin resistance. *EMBO J* 2017; 36(14): 2126–2145
  138. Tanimura Y, Aoi W, Takanami Y, Kawai Y, Mizushima K, Naito Y, Yoshikawa T. Acute exercise increases fibroblast growth factor 21 in metabolic organs and circulation. *Physiol Rep* 2016; 4(12): e12828
  139. Lee MS, Choi SE, Ha ES, An SY, Kim TH, Han SJ, Kim HJ, Kim DJ, Kang Y, Lee KW. Fibroblast growth factor-21 protects human skeletal muscle myotubes from palmitate-induced insulin resistance by inhibiting stress kinase and NF- $\kappa$ B. *Metabolism* 2012; 61(8): 1142–1151
  140. Izumiya Y, Bina HA, Ouchi N, Akasaki Y, Kharitonov A, Walsh K. FGF21 is an Akt-regulated myokine. *FEBS Lett* 2008; 582(27): 3805–3810
  141. Owen BM, Ding X, Morgan DA, Coate KC, Bookout AL, Rahmouni K, Kliewer SA, Mangelsdorf DJ. FGF21 acts centrally to induce sympathetic nerve activity, energy expenditure, and weight loss. *Cell Metab* 2014; 20(4): 670–677
  142. Douris N, Stevanovic DM, Fisher FM, Cisu TI, Chee MJ, Nguyen NL, Zarebidaki E, Adams AC, Kharitonov A, Flier JS, Bartness TJ, Maratos-Flier E. Central fibroblast growth factor 21 browns white fat via sympathetic action in male mice. *Endocrinology* 2015; 156(7): 2470–2481
  143. Liang Q, Zhong L, Zhang J, Wang Y, Bornstein SR, Triggler CR, Ding H, Lam KS, Xu A. FGF21 maintains glucose homeostasis by mediating the cross talk between liver and brain during prolonged fasting. *Diabetes* 2014; 63(12): 4064–4075
  144. Owen BM, Bookout AL, Ding X, Lin VY, Atkin SD, Gautron L, Kliewer SA, Mangelsdorf DJ. FGF21 contributes to neuroendocrine control of female reproduction. *Nat Med* 2013; 19(9): 1153–1156
  145. Bookout AL, de Groot MH, Owen BM, Lee S, Gautron L, Lawrence HL, Ding X, Elmquist JK, Takahashi JS, Mangelsdorf DJ, Kliewer SA. FGF21 regulates metabolism and circadian behavior by acting on the nervous system. *Nat Med* 2013; 19(9): 1147–1152
  146. Ishida N. Role of PPAR $\alpha$  in the control of torpor through FGF21-NPY pathway: from circadian clock to seasonal change in mammals. *PPAR Res* 2009; 2009: 412949
  147. Wang Q, Yuan J, Yu Z, Lin L, Jiang Y, Cao Z, Zhuang P, Whalen MJ, Song B, Wang XJ, Li X, Lo EH, Xu Y, Wang X. FGF21 attenuates high-fat diet-induced cognitive impairment via metabolic regulation and anti-inflammation of obese mice. *Mol Neurobiol* 2018; 55(6): 4702–4717
  148. Yu Y, Bai F, Wang W, Liu Y, Yuan Q, Qu S, Zhang T, Tian G, Li S, Li D, Ren G. Fibroblast growth factor 21 protects mouse brain against D-galactose induced aging via suppression of oxidative stress response and advanced glycation end products formation. *Pharmacol Biochem Behav* 2015; 133: 122–131
  149. Sarruf DA, Thaler JP, Morton GJ, German J, Fischer JD, Ogimoto K, Schwartz MW. Fibroblast growth factor 21 action in the brain increases energy expenditure and insulin sensitivity in obese rats. *Diabetes* 2010; 59(7): 1817–1824
  150. Véniant MM, Hale C, Helmering J, Chen MM, Stanislaus S, Busby J, Vonderfecht S, Xu J, Lloyd DJ. FGF21 promotes metabolic homeostasis via white adipose and leptin in mice. *PLoS One* 2012; 7(7): e40164
  151. Xu J, Lloyd DJ, Hale C, Stanislaus S, Chen M, Sivits G, Vonderfecht S, Hecht R, Li YS, Lindberg RA, Chen JL, Jung DY, Zhang Z, Ko HJ, Kim JK, Véniant MM. Fibroblast growth factor 21 reverses hepatic steatosis, increases energy expenditure, and improves insulin sensitivity in diet-induced obese mice. *Diabetes* 2009; 58(1): 250–259
  152. So WY, Cheng Q, Xu A, Lam KS, Leung PS. Loss of fibroblast growth factor 21 action induces insulin resistance, pancreatic islet hyperplasia and dysfunction in mice. *Cell Death Dis* 2015; 6(3): e1707
  153. Zhang C, Shao M, Yang H, Chen L, Yu L, Cong W, Tian H, Zhang F, Cheng P, Jin L, Tan Y, Li X, Cai L, Lu X. Attenuation of hyperlipidemia- and diabetes-induced early-stage apoptosis and late-stage renal dysfunction via administration of fibroblast growth factor-21 is associated with suppression of renal inflammation. *PLoS One* 2013; 8(12): e82275
  154. Kim HW, Lee JE, Cha JJ, Hyun YY, Kim JE, Lee MH, Song HK, Nam DH, Han JY, Han SY, Han KH, Kang YS, Cha DR. Fibroblast growth factor 21 improves insulin resistance and ameliorates renal injury in *db/db* mice. *Endocrinology* 2013; 154(9): 3366–3376
  155. Tang TT, Li YY, Li JJ, Wang K, Han Y, Dong WY, Zhu ZF, Xia N, Nie SF, Zhang M, Zeng ZP, Lv BJ, Jiao J, Liu H, Xian ZS, Yang XP, Hu Y, Liao YH, Wang Q, Tu X, Mallat Z, Huang Y, Shi GP, Cheng X. Liver-heart crosstalk controls IL-22 activity in cardiac protection after myocardial infarction. *Theranostics* 2018; 8(16): 4552–4562
  156. Wang N, Zhao TT, Li SM, Li YH, Wang YJ, Li DS, Wang WF. Fibroblast growth factor 21 ameliorates pancreatic fibrogenesis via regulating polarization of macrophages. *Exp Cell Res* 2019; 382(1): 111457
  157. Li S, Guo X, Zhang T, Wang N, Li J, Xu P, Zhang S, Ren G, Li D. Fibroblast growth factor 21 ameliorates high glucose-induced fibrogenesis in mesangial cells through inhibiting STAT5 signaling pathway. *Biomed Pharmacother* 2017; 93: 695–704
  158. Li S, Wang N, Guo X, Li J, Zhang T, Ren G, Li D. Fibroblast growth factor 21 regulates glucose metabolism in part by reducing renal glucose reabsorption. *Biomed Pharmacother* 2018; 108: 355–366
  159. Lin XL, He XL, Zeng JF, Zhang H, Zhao Y, Tan JK, Wang Z.

- FGF21 increases cholesterol efflux by upregulating ABCA1 through the ERK1/2-PPAR $\gamma$ -LXR $\alpha$  pathway in THP1 macrophage-derived foam cells. *DNA Cell Biol* 2014; 33(8): 514–521
160. Yu Y, He J, Li S, Song L, Guo X, Yao W, Zou D, Gao X, Liu Y, Bai F, Ren G, Li D. Fibroblast growth factor 21 (FGF21) inhibits macrophage-mediated inflammation by activating Nrf2 and suppressing the NF- $\kappa$ B signaling pathway. *Int Immunopharmacol* 2016; 38: 144–152
161. Li H, Wu G, Fang Q, Zhang M, Hui X, Sheng B, Wu L, Bao Y, Li P, Xu A, Jia W. Fibroblast growth factor 21 increases insulin sensitivity through specific expansion of subcutaneous fat. *Nat Commun* 2018; 9(1): 272
162. Li SM, Wang WF, Zhou LH, Ma L, An Y, Xu WJ, Li TH, Yu YH, Li DS, Liu Y. Fibroblast growth factor 21 expressions in white blood cells and sera of patients with gestational diabetes mellitus during gestation and postpartum. *Endocrine* 2015; 48(2): 519–527
163. Li JY, Wang N, Khoso MH, Shen CB, Guo MZ, Pang XX, Li DS, Wang WF. FGF-21 elevated IL-10 production to correct LPS-induced inflammation. *Inflammation* 2018; 41(3): 751–759
164. Wang WF, Ma L, Liu MY, Zhao TT, Zhang T, Yang YB, Cao HX, Han XH, Li DS. A novel function for fibroblast growth factor 21: stimulation of NADPH oxidase-dependent ROS generation. *Endocrine* 2015; 49(2): 385–395
165. Li SM, Yu YH, Li L, Wang WF, Li DS. Treatment of CIA mice with FGF21 down-regulates TH17-IL-17 axis. *Inflammation* 2016; 39(1): 309–319
166. Saito H, Kusano K, Kinoshita M, Ito H, Hirata M, Segawa H, Miyamoto K, Fukushima N. Human fibroblast growth factor-23 mutants suppress Na<sup>+</sup>-dependent phosphate co-transport activity and 1 $\alpha$ ,25-dihydroxyvitamin D3 production. *J Biol Chem* 2003; 278(4): 2206–2211
167. Shimada T, Kakitani M, Yamazaki Y, Hasegawa H, Takeuchi Y, Fujita T, Fukumoto S, Tomizuka K, Yamashita T. Targeted ablation of Fgf23 demonstrates an essential physiological role of FGF23 in phosphate and vitamin D metabolism. *J Clin Invest* 2004; 113(4): 561–568
168. Shimada T, Mizutani S, Muto T, Yoneya T, Hino R, Takeda S, Takeuchi Y, Fujita T, Fukumoto S, Yamashita T. Cloning and characterization of FGF23 as a causative factor of tumor-induced osteomalacia. *Proc Natl Acad Sci USA* 2001; 98(11): 6500–6505
169. Andrukhova O, Smorodchenko A, Egerbacher M, Streicher C, Zeitz U, Goetz R, Shalhoub V, Mohammadi M, Pohl EE, Lanske B, Erben RG. FGF23 promotes renal calcium reabsorption through the TRPV5 channel. *EMBO J* 2014; 33(3): 229–246
170. Andrukhova O, Slavic S, Smorodchenko A, Zeitz U, Shalhoub V, Lanske B, Pohl EE, Erben RG. FGF23 regulates renal sodium handling and blood pressure. *EMBO Mol Med* 2014; 6(6): 744–759
171. Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, Sirkis R, Naveh-Manly T, Silver J. The parathyroid is a target organ for FGF23 in rats. *J Clin Invest* 2007; 117(12): 4003–4008
172. Toro L, Barrientos V, León P, Rojas M, Gonzalez M, González-Ibáñez A, Illanes S, Sugikawa K, Abarzúa N, Bascuñán C, Arcos K, Fuentealba C, Tong AM, Elorza AA, Pinto ME, Alzamora R, Romero C, Michea L. Erythropoietin induces bone marrow and plasma fibroblast growth factor 23 during acute kidney injury. *Kidney Int* 2018; 93(5): 1131–1141
173. Rabadi S, Udo I, Leaf DE, Waikar SS, Christov M. Acute blood loss stimulates fibroblast growth factor 23 production. *Am J Physiol Renal Physiol* 2018; 314(1): F132–F139
174. ADHR Consortium. Autosomal dominant hypophosphataemic rickets is associated with mutations in FGF23. *Nat Genet* 2000; 26(3): 345–348
175. Bowe AE, Finnegan R, Jan de Beur SM, Cho J, Levine MA, Kumar R, Schiavi SC. FGF-23 inhibits renal tubular phosphate transport and is a PHEX substrate. *Biochem Biophys Res Commun* 2001; 284(4): 977–981
176. Riminucci M, Collins MT, Fedarko NS, Cherman N, Corsi A, White KE, Waguespack S, Gupta A, Hannon T, Econs MJ, Bianco P, Gehron Robey P. FGF-23 in fibrous dysplasia of bone and its relationship to renal phosphate wasting. *J Clin Invest* 2003; 112(5): 683–692
177. Hoffman WH, Jueppner HW, Deyoung BR, O'dorisio MS, Given KS. Elevated fibroblast growth factor-23 in hypophosphatemic linear nevus sebaceous syndrome. *Am J Med Genet A* 2005; 134(3): 233–236
178. Kato K, Jeanneau C, Tarp MA, Benet-Pagès A, Lorenz-Depiereux B, Bennett EP, Mandel U, Strom TM, Clausen H. Polypeptide GalNAc-transferase T3 and familial tumoral calcinosis. Secretion of fibroblast growth factor 23 requires O-glycosylation. *J Biol Chem* 2006; 281(27): 18370–18377
179. Ichikawa S, Imel EA, Sorenson AH, Severe R, Knudson P, Harris GJ, Shaker JL, Econs MJ. Tumoral calcinosis presenting with eyelid calcifications due to novel missense mutations in the glycosyl transferase domain of the GALNT3 gene. *J Clin Endocrinol Metab* 2006; 91(11): 4472–4475
180. Garringer HJ, Fisher C, Larsson TE, Davis SI, Koller DL, Cullen MJ, Draman MS, Conlon N, Jain A, Fedarko NS, Dasgupta B, White KE. The role of mutant UDP-N-acetyl- $\alpha$ -D-galactosamine-polypeptide N-acetylgalactosaminyltransferase 3 in regulating serum intact fibroblast growth factor 23 and matrix extracellular phosphoglycoprotein in heritable tumoral calcinosis. *J Clin Endocrinol Metab* 2006; 91(10): 4037–4042
181. Benet-Pagès A, Orlik P, Strom TM, Lorenz-Depiereux B. An FGF23 missense mutation causes familial tumoral calcinosis with hyperphosphatemia. *Hum Mol Genet* 2005; 14(3): 385–390
182. Chefetz I, Heller R, Galli-Tsinopoulou A, Richard G, Wollnik B, Indelman M, Koerber F, Topaz O, Bergman R, Sprecher E, Schoenau E. A novel homozygous missense mutation in FGF23 causes Familial Tumoral Calcinosis associated with disseminated visceral calcification. *Hum Genet* 2005; 118(2): 261–266
183. Araya K, Fukumoto S, Backenroth R, Takeuchi Y, Nakayama K, Ito N, Yoshii N, Yamazaki Y, Yamashita T, Silver J, Igarashi T, Fujita T. A novel mutation in fibroblast growth factor 23 gene as a cause of tumoral calcinosis. *J Clin Endocrinol Metab* 2005; 90(10): 5523–5527
184. Abbasi F, Ghafouri-Fard S, Javaheri M, Dideban A, Ebrahimi A, Ebrahim-Habibi A. A new missense mutation in FGF23 gene in a male with hyperostosis-hyperphosphatemia syndrome (HHS). *Gene* 2014; 542(2): 269–271
185. Faul C, Amaral AP, Oskouei B, Hu MC, Sloan A, Isakova T, Gutiérrez OM, Aguillon-Prada R, Lincoln J, Hare JM, Mundel P, Morales A, Scialla J, Fischer M, Soliman EZ, Chen J, Go AS,

- Rosas SE, Nessel L, Townsend RR, Feldman HI, St John Sutton M, Ojo A, Gadegbeku C, Di Marco GS, Reuter S, Kentrup D, Tiemann K, Brand M, Hill JA, Moe OW, Kuro-O M, Kusek JW, Keane MG, Wolf M. FGF23 induces left ventricular hypertrophy. *J Clin Invest* 2011; 121(11): 4393–4408
186. Gutiérrez OM, Januzzi JL, Isakova T, Laliberte K, Smith K, Collerone G, Sarwar A, Hoffmann U, Coglianese E, Christenson R, Wang TJ, deFilippi C, Wolf M. Fibroblast growth factor 23 and left ventricular hypertrophy in chronic kidney disease. *Circulation* 2009; 119(19): 2545–2552
187. McGrath ER, Himali JJ, Levy D, Conner SC, Pase MP, Abraham CR, Courchesne P, Satizabal CL, Vasani RS, Beiser AS, Seshadri S. Circulating fibroblast growth factor 23 levels and incident dementia: The Framingham heart study. *PLoS One* 2019; 14(3): e0213321
188. Liu P, Chen L, Bai X, Karaplis A, Miao D, Gu N. Impairment of spatial learning and memory in transgenic mice overexpressing human fibroblast growth factor-23. *Brain Res* 2011; 1412: 9–17