Hindawi Publishing Corporation BioMed Research International Volume 2016, Article ID 1934518, 10 pages http://dx.doi.org/10.1155/2016/1934518

Review Article

Sunlight Effects on Immune System: Is There Something Else in addition to UV-Induced Immunosuppression?

D. H. González Maglio, M. L. Paz, and J. Leoni

Instituto de Estudios de la Inmunidad Humoral (IDEHU), CONICET, Universidad de Buenos Aires, Junín 956, C1113AAD Buenos Aires, Argentina

Correspondence should be addressed to D. H. González Maglio; danielgm@ffyb.uba.ar

Received 6 September 2016; Revised 2 November 2016; Accepted 6 November 2016

Academic Editor: Maxim E. Darvin

Copyright © 2016 D. H. González Maglio et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Sunlight, composed of different types of radiation, including ultraviolet wavelengths, is an essential source of light and warmth for life on earth but has strong negative effects on human health, such as promoting the malignant transformation of skin cells and suppressing the ability of the human immune system to efficiently detect and attack malignant cells. UV-induced immunosuppression has been extensively studied since it was first described by Dr. Kripke and Dr. Fisher in the late 1970s. However, skin exposure to sunlight has not only this and other unfavorable effects, for example, mutagenesis and carcinogenesis, but also a positive one: the induction of Vitamin D synthesis, which performs several roles within the immune system in addition to favoring bone homeostasis. The impact of low levels of UV exposure on the immune system has not been fully reported yet, but it bears interesting differences with the suppressive effect of high levels of UV radiation, as shown by some recent studies. The aim of this article is to put some ideas in perspective and pose some questions within the field of photoimmunology based on established and new information, which may lead to new experimental approaches and, eventually, to a better understanding of the effects of sunlight on the human immune system.

1. Introduction

Sunlight is composed of ultraviolet (UV), visible, and infrared radiations. It is essential for life on earth as a source of energy, light, and warmth and to maintain oxygen levels in our atmosphere, due to the role it plays in photosynthesis. However, it also causes profound changes in the human body.

The effects of sunlight, particularly UV radiation, on the skin cell biology as well as on the immune system have been described at length. One of its most important effects is UV-induced immunosuppression, a defective immune response triggered by UV radiation affecting the skin first, and then the whole body. Thousands of experimental papers have been published since the first descriptions of UV-induced immunosuppression and its role in the development of skin carcinogenesis [1–4]. In addition to causing immune cell alterations, UV radiation absorption produces molecular changes, many of which have been extensively reported (though it is impossible to know if all types have been covered). UV radiation is known to be directly absorbed

by DNA (in particular by adjacent pyrimidine bases) and by cis-urocanic acid in exposed cells [5-7] and to promote the production of reactive oxygen species (ROS), which in turn may cause DNA damage [8]. These alterations lead to changes in the production of different molecules related to the immune system, including interleukin-10 (IL-10), IL-4, and prostaglandin E₂ (PGE₂) [9-11]. These molecules, in turn, modulate systemic immune responses, promoting defects in cellular immunity [12-14]. In animal models, it has been demonstrated that UV-induced systemic immunosuppression is related to the development of antigenspecific regulatory T-cells (CD4+ CD25+ foxp3+ cells), which can be transferred into nonexposed animals [15, 16]. The development of these regulatory cells is associated with a particular environment of soluble molecules established after UV exposure, which include not only cytokines and PGE₂ but also Vitamin D (its role in UV-induced immunosuppression will be discussed below) [17]. It is known that this environment may condition skin dendritic cells in order to specifically promote the regulatory T-cell phenotype during

Table 1: Summary of irradiation protocols producing UV-induced immunosuppression. The experimental design of all the papers cited in the table included the irradiation before the sensitization stage of the immune responses. CHS: contact hypersensitivity; DTH: delayed type hypersensitivity; Oxa: oxazolone; DNFB: 2,4-dinitrofluorobenzene; Ova: ovalbumin.

Authors	UV source	UV dose employed	MED represented	Mice strain	Type of reaction (antigen)
Reeve et al. [43]	UVA/UVB	$3 \times 4500 \text{ mJ/cm}^2 \text{ (UVA)};$ $3 \times 252 \text{ mJ/cm}^2 \text{ (UVB)}$	1 MED	C57BL/6	CHS (Oxa)
Majewski et al. [44]	UVR	$4 \times 150 \mathrm{mJ/cm^2}$	Not expressed	C57BL/6	CHS (DNFB)
Wang et al. [45]	UVB	$3 \times 45 \text{ mJ/cm}^2$	Not expressed	C57BL/6	CHS (DNFB) OT1 T-cell proliferation (Ova)
Schwarz et al. [46]	UVB	$4 \times 150 \mathrm{mJ/cm}^2$	Not expressed	C57BL/6	CHS (DNFB)
Gorman et al. [47]	65% UVB	400 and 800 mJ/cm ²	>1 MED	Balb/c and C57BL/6	CHS (DNFB)
Zhang et al. [48]	45% UVB	$750 \mathrm{mJ/cm^2}$	Not expressed	C57BL/6	CHS (DNFB)
Guéniche et al. [49]	UVB + UVA	Not expressed	2.5 MED	SKH:HR1	CHS (DNFB)
Shreedhar et al. [9]	65% UVB	$1500 \mathrm{mJ/cm^2}$	Not expressed	C3H/HeN	DTH (alloantigen)
Li et al. [50]	UVB	$200\mathrm{mJ/cm}^2$	Not expressed	C57BL/6	CHS (DNFB)
Rana et al. [51]	UVB	$3 \times 150 \mathrm{mJ/cm}^2$	0.5 MED	C57BL/6	DTH (Ova)
Dixon et al. [52]	UVB and UVA	$3 \times 400 \mathrm{mJ/cm^2}$ (UVB)	3 MED	SKH:HR1	CHS (Oxa)

priming in regional lymph nodes [18, 19]. The induction of the tolerogenic phenotype in dendritic cells may be so intense that even bone marrow cells may develop it, leading to suppressive responses several days (and even months) after exposure [20]. However, regulatory T-cells and tolerogenic dendritic cells are not the only ones involved in UV-induced immunosuppression. Mast cells also have a role to play in the development of immunosuppression, since the number of such cells in the skin and their migration to draining lymph nodes have been correlated with the UV-induced suppressive response [21, 22]. Moreover, regulatory B-cells, capable of affecting dendritic cell-mediated T-cell activation, are also involved in this effect triggered by UV exposure. Their number and suppressive action in draining lymph nodes increase after UV exposure [23]. Molecular mechanisms involved in this effect include the production of IL-10 by regulatory B-cells after the interaction of the platelet-activation factor, a proinflammatory mediator, with its receptor in Bcells [24]. Finally, oxidative stress is also related to UVinduced immunosuppression, since the topical application of antioxidants before UV exposure may completely inhibit it [25].

2

Regardless of the cell types involved, an important biological consequence of the UV-induced immunosuppression is the loss of immunosurveillance on newly generated malignant cells. Skin cell DNA is affected by UV radiation either directly (dimerization of adjacent pyrimidines) or indirectly (oxidative damage induced by ROS), which may cause specific mutations that will eventually lead to the malignant transformation of these cells (mainly, melanocytes and keratinocytes) [26–28]. These malignant cells, under normal circumstances, can be identified and eliminated by the immune system in a process known as "immunosurveillance." However, after only a single exposure to UV radiation, this immune process can be severely affected, diminishing the ability of the body to fight skin tumors.

But sunlight exposure is not only associated with detrimental effects on human health. Sunlight exposure is essential to ensuring proper levels of circulating Vitamin D, since its synthesis is initiated in the skin with the photoconversion of 7-dehydrocholesterol to previtamin D [29]. Vitamin D is essential to maintain bone homeostasis, but it also has effects on the immune system [30, 31]. The role of Vitamin D in UV-induced effects will be discussed below, but it is worth noticing that this is one of the main benefits of sunlight exposure.

The above-mentioned effects of sunlight, especially UV radiation, on human health have been also widely reviewed. Plenty of excellent reviews covering the contrasting consequences of skin exposure to sunlight have been published [17, 32–42]. This article is intended to raise questions regarding the overall effects of skin exposure to sunlight that may lead to the use of different sources of radiation in the treatment of human diseases.

2. How Low Is a Low-Immunosuppressive-UV Dose?

The pioneering work of Dr. Kripke demonstrating UV-induced immunosuppression was based on chronic exposures to UV radiation (three times a week during three months) [1, 2]. Since then, a great number of irradiation protocols inducing immunosuppression were created. Table 1 briefly summarizes some of the doses used in such protocols with their respective correlation, in most of the cases, to a biological effect: the minimal erythema dose (MED).

It should be noted that the paradigm of UV-induced immunosuppression has changed over the last decades. While immunosuppression in Dr. Kripke's work was reached by chronic irradiation, later on it was proved that a single high irradiation (above the erythema dose) was also capable of producing the same effect. Regardless of the form

of irradiation involved, some of the papers mentioned in Table 1 refer to the source of irradiation as a "low-dose UV" [46, 51, 53-55] that can promote immunosuppression even under the MED. Consequently, the concept of UV dose causing immune suppression was then correlated to human exposures to sunlight, alerting about exposures even below the minimal erythema dose. Indeed, many works have demonstrated that immunosuppressive doses of UV radiation in humans are effectively below the MED. Wolf et al. published that the dose capable of producing 50% of the inhibition of CHS response to DNCB during sensitization phase ranged between 0.63 and 0.79 MED [56]. It is worth noticing that a major effect of sunlight on human health, as reviewed, is UV-induced immunosuppression, which is presented as having both a positive and a negative effect on human health. An example of this is the review by Dr. Schwarz [40]. Negative effects are related to defective immunosurveillance, allowing for tumor development, while the positive effects are related to the control of autoimmune diseases due to the generation of specific regulatory cells. In the latter case, the possible implications of the abovementioned "low UV doses" were reviewed a few years ago by Dr. Halliday et al. [32], which led to the design of clinical trials to assess the role of phototherapy in autoimmune diseases [57].

However, our question addresses the fact that half MED is not as low as a tenth (or even less) of the MED, which we classify as "very low doses." These "very low doses" of radiation are completely relevant if we think in photoprotectors not as blockers of radiation but as filters: radiation can be absorbed to a high degree (SPF 50 or more) but not entirely. Thus, SPF (Sun Protection Factor) is "a measure of how much solar energy (UV radiation) is required to produce sunburn on protected skin (i.e., in the presence of sunscreen) relative to the amount of solar energy required to produce sunburn on unprotected skin," according to the U.S. Food and Drug Administration [58]. However, it has been stated that the SPF of a given sunscreen may not be directly related to its Immune Protective Factor (IPF) [59], leading to the need of a standardized procedure to evaluate IPF. This topic is remarkably well treated in a publication of five groups of researchers from Australia, Austria, France, UK, and USA [60]. Based on SPF definition, we can estimate the approximate time of exposure to receive a tenth of the MED using SPF 50. For example, as it was published by Samanek et al., in Sydney (Australia) in summer time, people with skin phototype II required an exposure of 11 minutes to reach MED [61]. In that context and using a SPF 50 photoprotector, a tenth of the MED would be achieved after a 55-minute exposure. Moreover, these doses of radiation can also be obtained while walking normally outdoors in daylight in summer. One is likely to be frequently exposed to the above-mentioned "very low doses" of UV, but will such doses also cause an immunosuppressive effect? Or will they produce other effects on human health? These questions, which have been recently raised, will be discussed in another section.

3. Is Vitamin D a Soluble Mediator of UV Radiation Immune Effects or Just an Epiphenomenon?

It is very well known that UV radiation is essential for Vitamin D synthesis, in particular for the photoconversion of 7-dehydrocholesterol to cholecalciferol in the epidermis. The production of this vitamin represents one of the most important beneficial effects of sunlight exposure. Vitamin D synthesis and its impact on human health have been extensively reviewed in many of the cited papers [31, 36, 62–65] and it is not our purpose to analyze once again the relevance of this process.

However, there is some puzzling evidence of the role of Vitamin D in UV-induced immunosuppression: (a) Vitamin D is a mediator of UV-induced immunosuppression and it mimics this effect [46]; (b) Vitamin D is not necessary to immunosuppress UV irradiated animals [46, 47]; (c) Vitamin D and a nongenomic analog are protective against UV-induced immunosuppression and carcinogenesis [52]. Do these differences lie on the concentration used and/or particular preactivated pathways? Schwarz et al. [46] topically applied 0.1 µg of 1,25(OH)₂VitD₃ (diluted in acetone/olive oil, 4:1), which represents 240 pmoles, in order to induce immunosuppression. On the other hand, Dixon et al. [52] used 159.6 and 44.8 pmoles (diluted in ethanol, propylene glycol, and water to a final solvent ratio of 2:1:1, resp.) of the vitamin in order to obtain significant protection against UV-induced immunosuppression. Even though the concentration of Vitamin D used in these experiences is different, an important question arises: which one best represents the concentration of vitamin in the skin after UV exposure? Gorman et al. measured Vitamin D in the ear skin of irradiated animals (on a Vitamin D replete or deficient diet) and they reported that "no change in 1.25(OH)₂D3 levels was detected in the ear skin of either male or female Vitamin D3-deficient mice with UV irradiation" [47]. It is difficult to find out the role of this vitamin in UV-induced immunosuppression in humans, but we would like to discuss two papers related to human exposure to radiation. In a study with volunteers subjected to a CHS protocol, a single exposure to 3 MED (an average dose of 420 mJ/cm², since MED depends on skin type) prior to sensitization significantly suppressed the immunological reaction [66]. On the other hand, measurements performed in a group of Danish people during a summer holiday revealed that with a total exposure of 10100 mJ/cm² (an average for 25 individuals) serum concentration of Vitamin D increased only 1.44 times [67]. It seems like biological Vitamin D increments after UV exposure are not sufficient to justify the suppression of specific immune responses, but certainly more evidence is needed.

Finally, to conclude this topic, there is a very good review by Dr. Byrne, which we completely agree with, that presents a conclusion about a clinical trial on the use of Vitamin D in autoimmune diseases suggesting that "boosting Vitamin D levels alone may not have the desired therapeutic effect and that there is something else about UV exposure that explains the protective properties of sunlight" [42].

4. What Determines Different Susceptibility to UV-Induced Immunosuppression in Mice? Is There a Correlation in Humans?

Susceptibility to UV-induced immunosuppression is very well known. Yoshikawa et al. used CHS reaction in humans to study this phenomenon [68]. They observed an absence of immunosuppression in 60% of the healthy volunteers analyzed. In contrast, patients with a history of nonmelanoma skin cancer showed marked susceptibility to immunosuppression. They posed the possibility that this increased susceptibility may be the cause of skin cancer development. Moreover, twenty years ago, Noonan and Hoffman performed an extensive analysis of susceptibility to CHS reactions in 16 strains of inbred mice [69]. They informed the UV radiation dose needed to produce a 50% inhibitory effect on a CHS protocol, which ranged from 70 mJ/cm² to 260 mJ/cm² in highly susceptible mice (C57 mice); from 470 mJ/cm² to 690 mJ/cm² in moderately susceptible mice (DBA/2 and A/J mice); and from 930 mJ/cm² to 1230 mJ/cm² in low susceptible mice (Balb/c mice). This almost 20-time difference in the UV doses causing immunosuppression reflects substantial differences in the genetic background involved in the response to UV light. Hair pigmentation does not seem to justify these results, since C57 (highly pigmented mice) are more susceptible than Balb/c (albino nonpigmented mice). What differences in what genes can condition UV radiation response? In a more recent work, Welsh et al. investigated the association of human polymorphisms in functional variants of 10 genes involved in the response to UV radiation (IL-10, IL-4, and TNF- α among others), with the risk of Basal Cell Carcinoma and Squamous Cell Carcinoma (both associated with UV-induced immunosuppression) [70]. Major effects were observed for skin type, lifetime severe sunburns, and IL-10 haplotypes in both BCC and SCC. The haplotypes studied were in the promoter region of the gene and may be correlated with an increased production of this cytokine [71]. Moreover, Nagano et al. published a study on a non-Caucasian population, where similar results were observed: patients who developed skin cancer in sun-exposed areas (more susceptible to immunosuppression) presented less frequency of low expression IL-10 promoter genotype [72]. The work by Welsh and collaborators reinforces the idea of the impact of UV exposure in the development of skin cancer and UV-induced immunosuppression, since skin type and burns were highly associated with the development of tumors, but it also incorporates the notion of genetic susceptibility, as it was demonstrated long ago by Yoshikawa, in humans, and Noonan and Hoffman, in mice.

We have recently studied the response of different mice strains to a single, 2 MED, UVB exposure and observed that C57BL/6 mice greatly differ in their inflammatory and oxidative responses from Balb/c and other mice strains. C57BL/6 mice produce a stronger inflammatory response (increased levels of serum and epidermal IL-6) and a weaker oxidative epidermal response (with less production of superoxide anion in epidermal cells) after irradiation [73].

Even though differences in the response to UV light between individuals with different genetic background exist, we cannot precisely determine their influence on the overall response to natural or therapeutic exposures. In the future, this issue may be unraveled, once more studies are performed in the area.

5. Is There Something Else (with Immune Effects) in Sunlight than UV Radiation?

Sunlight is a complex source of different types of radiation that includes UV radiation but it also exceeds it. The impact of a specific source of radiation on living organisms depends on the capacity of the cells to absorb it by producing molecules with a specific absorption spectrum. Once radiation is absorbed by these molecules, different cellular pathways can be activated. In this way, UV radiation is absorbed by DNA, cis-urocanic acid, and proteins, among other molecules, and promotes many of the very well-known effects mentioned before. But, are visible radiations absorbed by skin cells? Or in other words, do visible radiations directly induce any effect, either positive or negative, on skin health? The answer is yes. Laser therapies with visible wavelengths for cosmetic or therapeutic purposes are distributed worldwide, but do they only promote beneficial effects? What cellular pathway do they affect?

In order to analyze some specific responses activated by visible light, there is an increasing number of reports describing different effects of these radiations on skin or its constituent cells. It has been published that low level light therapy with different wavelengths (470 nm, blue, and 629 nm, red) induces angiogenesis and improves wound healing in an ischemia disturbed rodent flap model, supporting an interesting application of artificial light sources on human health [74]. Another benefit is a specific bactericidal effect on Staphylococcus aureus and Pseudomona aeruginosa, which has been proved both in vitro (in bacterial culture techniques) and in vivo (in infection models) [75, 76]. The last reference is a work by Dai et al. which used blue light (415 nm) therapy to effectively treat a potentially fatal mice infection with Pseudomona, showing that the bactericidal effect could be very useful in skin infections, especially in those produced by multiresistant microorganisms. Therefore, the questions that arise are the following: are both (healing and bactericidal) effects simultaneously produced? Is it possible for blue light to modulate immune responses? These questions need to be addressed in new experimental studies in order to test, for example, blue light therapy in different skin conditions in which healing and antimicrobial barriers have to be rapidly improved, like in massive burns.

However, blue light has also shown some potentially harmful effects. As an example, Mamalis et al. described that this type of radiation promotes a decrease in skin fibroblasts proliferation as well as an increase in ROS production [77]. ROS and matrix degrading enzymes production increase is a very well-known harmful effect of UV radiation, but this is also caused by visible light [78]. In addition, a recently published article by Vandersee and collaborators demonstrated that blue light skin exposure affects the antioxidant

defense of the skin, by decreasing the cutaneous carotenoid concentration, in addition to its effects as a promoter of ROS, as previously described [79]. Does this blue light-induced oxidative damage also affect keratinocytes and Langerhans cells? If so, how does it affect the immune functions of those cells?

We would like to briefly comment on another skin cellular response to radiation: the induction of pigmentation. This important response of the skin to sunlight is thought to have a protective role against DNA damage [80]. It has been proved that blue light (415 nm), but not red light (630 nm), is able to induce pigmentation in type III and IV healthy subjects. Moreover, when compared to UVB irradiation, the blue light induces a significantly more pronounced hyperpigmentation that lasts up to 3 months [81]. This means that blue light is capable of affecting epidermal cells and, in particular, that melanocytes are very sensitive to this radiation. Again, it raises the question about blue light effects on skin immune cells. There are just a few reports about the effects of blue light on dendritic or Langerhans cells. On the one side, it has been shown that during a photodynamic therapy with blue light the number of epidermal Langerhans cells was not affected; neither was the oxidative damage to their DNA, by the radiation itself or the photosensitizer [82, 83]. But on the other side, in vitro irradiation of dendritic cells affects their capacity to respond to a LPS/IFN-y stimulus, thus decreasing the production of cytokines (IL-12, IL-6, and TNF- α) and the level of expression of costimulatory molecules (CD83 and CD80) in a dose dependent fashion [84, 85].

The exact role of blue light in immunosuppression and the possible involvement of visible radiations in the very well-known detrimental effects of sun exposures are yet to be explored. But sunlight includes much more than UV and visible light. Infrared- (IR-) A radiation (780–1400 nm), which accounts for more than 30% of sunlight, can penetrate deeply into the skin and promote the production of ROS and MMP-1 by skin cells [86-88]. Moreover, IR exposure may have other effects on skin cells, modulating their response to deleterious doses of UV radiation. Jantschitsch et al. showed that IR-exposed keratinocytes were protected from UV-induced apoptosis, through the reduction of DNA damage and modulation of the expression of apoptosisrelated proteins (upregulation of antiapoptotic FLIP_L and BCL-X_L and downregulation of proapoptotic BAX) [89]. These cellular alterations lead to a delay in the onset of skin cancer development in an in vivo model but promote a more aggressive phenotype of the tumors developed [90]. Regarding IR effects on the immune system, Lee et al. have recently published that IR exposure promoted an increase in the number of epidermal Langerhans cells, while lymph nodes cells stimulated with an anti-CD3 antibody led to the production of Th1 and Th2 cytokines, but not to regulatory ones [91]. These effects may be used to modulate immune responses locally and systemically. The IR/far red light mediated photobiomodulation has been described as an effective treatment for cutaneous infection by methicillin-resistant Staphylococcus aureus [92, 93], experimental autoimmune encephalitis (a mouse model of multiple sclerosis) [94], and brain disorders in humans [95]. The contribution of these

wavebands to the final effects of sunlight exposure remains to be elucidated. Figure 1 summarizes the effects triggered by different wavebands of sunlight on the skin and the immune system.

6. Are "Very Low UV Doses" Free of Immunomodulatory Effects?

To conclude this review, we would like to return to the discussion about the "very low UV doses" mentioned above. These doses of radiation we all tend to be exposed to—during occasional walks under the sun in summer time or during intentional sun exposures using sun protectors—are very relevant to study and we are working in this direction in our laboratory. We have recently published an article in which we worked with the idea of a model of daily "casual" exposures to sunlight, in particular to UV radiation. We performed UVB irradiation of hairless mice during four consecutive days with only 20 mJ/cm² (a tenth of the MED, described as repetitive low UV doses or rlUVd) and compared the effects on skin innate immunity with animals exposed to a single high dose irradiation (400 mJ/cm², 2 MED, described as single high UV dose or shUVd). We found a strong inflammatory response, as it has been largely described, in shUVd exposed animals which was completely absent in the rlUVd exposed ones. However, these "very low-dose" irradiations are far from producing no alterations at all, since a strong reinforcement of the epidermal barrier function was observed in mice irradiated with rlUVd. This reinforcement is based in a slight increment in epidermal thickness, without signs of histological alteration or metabolic dysfunction of epidermal cells, and a strong induction of antimicrobial peptides transcription [96]. The reinforcement of the barrier function was also described by Hong et al., who reported an increase in antimicrobial peptides' synthesis and permeability barrier reinforcement (measured after a tape-stripping insult) in hairless mice exposed to 40 mJ/cm² just once or three times in consecutive days [97].

Does this type of irradiation impact only innate skin immunity? It is difficult to find evidences of adaptive immunity-based reinforcement by repetitive low UV doses. However, more than a decade ago, Khaskely et al. published a very interesting work on cutaneous leishmaniasis in mice model (using Balb/c mice) [98], in which an experimental infection with parasite Leishmania amazonensis was introduced after a UVB irradiation procedure that consisted of daily exposures to 25 mJ/cm² during four consecutive days (very similar to the schedule used in our lab). Twenty-four hours after the last exposure, mice were challenged intradermally with the parasite. They found that the development of skin lesion produced by the infection was significantly reduced by a pretreatment with low-dose UV irradiation. The authors also reported an increase in serum IFN-y as a finding that could explain the control of the infection, since L. amazonensis is an intracellular parasite that is capable of surviving within the macrophages. This essential article raises new questions again: are repetitive low UV doses capable of predisposing adaptive immunity to a stronger response? Or does it only promote a better response of skin macrophages

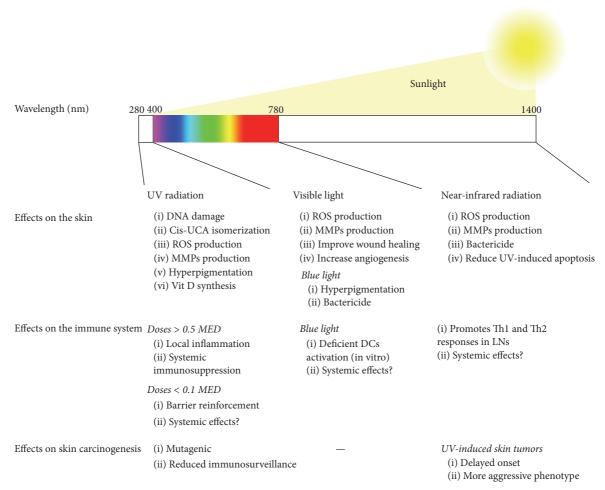


FIGURE 1: Sunlight effects on the skin and the immune system. The effects triggered by different wavebands of radiation are summarized, focusing on those effects described in the text. UCA: urocanic acid; ROS: reactive oxygen species; MMPs: matrix metalloproteases; MED: minimal erythema dose; DCs: dendritic cells; LNs: lymph nodes.

without T-cell activation? Is it possible to obtain similar responses in immune reinforcement with rlUVd once the infection has developed?

Unfortunately, neither the authors nor other researchers have published any new articles on this topic since then, but the questions remain there to be investigated and we ourselves are trying to answer some of them.

7. Conclusions

Sunlight exposure cannot be considered only as a carcinogen nowadays, though the highly relevant mechanisms leading to immunosuppression and consequently to skin cancer development have been and are still very well characterized and the most reported in the literature. There is also abundant evidence showing that sunlight effects, especially of "very low doses," are indeed beneficial and not only due to Vitamin D synthesis. We believe that plenty of work has yet to be done in the field of photoimmunology, which needs to cover the impact not only of "very low doses" of radiation, but also of exposure to non-UV light (focusing on the effects of various

doses) on the immune system. We as immunologists and particularly photoimmunologists have to move forward from the milestone of UV-induced immunosuppression towards a more comprehensive analysis of the interaction of human beings with the environment, leading to the possibility of establishing new therapies, which might be useful in different pathologies and not only in those that require a specific suppression of the immune response.

Competing Interests

The authors declare that they have no competing interests.

Acknowledgments

This study was funded by grants from Universidad de Buenos Aires (UBACyT 2011–2014 and 2013–2016), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET, PIP 2011–2013), and Agencia Nacional de Promoción Científica y Tecnológica (ANPCyT, PICT 2012). The authors are members of the CONICET Research Career Program.

References

- [1] M. S. Fisher and M. L. Kripke, "Systemic alteration induced in mice by ultraviolet light irradiation and its relationship to ultraviolet carcinogenesis," *Proceedings of the National Academy* of Sciences of the United States of America, vol. 74, no. 4, pp. 1688–1692, 1977.
- [2] M. S. Fisher and M. L. Kripke, "Further studies on the tumorspecific suppressor cells induced by ultraviolet radiation," *Journal of Immunology*, vol. 121, no. 3, pp. 1139–1144, 1978.
- [3] S. E. Ullrich and M. L. Kripke, "Mechanisms in the suppression of tumor rejection produced in mice by repeated UV irradiation," *Journal of Immunology*, vol. 133, no. 5, pp. 2786–2790, 1984.
- [4] M. L. Kripke, "Antigenicity of murine skin tumors induced by ultraviolet light," *Journal of the National Cancer Institute*, vol. 53, no. 5, pp. 1333–1336, 1974.
- [5] J. Garssen, F. De Gruijl, D. Mol, A. De Klerk, P. Roholl, and H. Van Loveren, "UVA exposure affects UVB and cis-Urocanic acid-induced systemic suppression of immune responses in Listeria monocytogenes-infected Balb/c mice," *Photochemistry* and *Photobiology*, vol. 73, no. 4, pp. 432–438, 2001.
- [6] N. K. Gibbs and M. Norval, "Urocanic acid in the skin: a mixed blessing?" *Journal of Investigative Dermatology*, vol. 131, no. 1, pp. 14–17, 2011.
- [7] A. R. Young, "Chromophores in human skin," *Physics in Medicine and Biology*, vol. 42, no. 5, pp. 789–802, 1997.
- [8] G. M. Halliday, "Inflammation, gene mutation and photoimmunosuppression in response to UVR-induced oxidative damage contributes to photocarcinogenesis," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 571, no. 1-2, pp. 107–120, 2005.
- [9] V. Shreedhar, T. Giese, V. W. Sung, and S. E. Ullrich, "A cytokine cascade including prostaglandin E2, IL-4, and IL-10 is responsible for UV-induced systemic immune suppression," *The Journal of Immunology*, vol. 160, no. 8, pp. 3783–3789, 1998.
- [10] C. C. Miller, P. Hale, and A. P. Pentland, "Ultraviolet B injury increases prostaglandin synthesis through a tyrosine kinasedependent pathway. Evidence for UVB-induced epidermal growth factor receptor activation," *The Journal of Biological Chemistry*, vol. 269, no. 5, pp. 3529–3533, 1994.
- [11] M. L. Paz, A. Ferrari, F. S. Weill, J. Leoni, and D. H. Gonzalez Maglio, "Time-course evaluation and treatment of skin inflammatory immune response after ultraviolet B irradiation," *Cytokine*, vol. 44, no. 1, pp. 70–77, 2008.
- [12] K. Loser, J. Apelt, M. Voskort et al., "IL-10 controls ultravioletinduced carcinogenesis in mice," *Journal of Immunology*, vol. 179, no. 1, pp. 365–371, 2007.
- [13] J. M. Rivas and S. E. Ullrich, "The role of IL-4, IL-10, and TNFα in the immune suppression induced by ultraviolet radiation," *Journal of Leukocyte Biology*, vol. 56, no. 6, pp. 769–775, 1994.
- [14] J. M. Rivas and S. E. Ullrich, "Systemic suppression of delayed-type hypersensitivity by supernatants from UV-irradiated keratinocytes: an essential role for keratinocyte-derived IL-10," *The Journal of Immunology*, vol. 149, no. 12, pp. 3865–3871, 1992.
- [15] A. Schwarz and T. Schwarz, "UVR-induced regulatory T cells switch antigen-presenting cells from a stimulatory to a regulatory phenotype," *Journal of Investigative Dermatology*, vol. 130, no. 7, pp. 1914–1921, 2010.
- [16] M. Ghoreishi and J. P. Dutz, "Tolerance induction by transcutaneous immunization through ultraviolet-irradiated skin

- is transferable through CD4⁺CD25⁺ T regulatory cells and is dependent on host-derived IL-10," *Journal of Immunology*, vol. 176, no. 4, pp. 2635–2644, 2006.
- [17] S. Gorman and P. H. Hart, "The current state of play of rodent models to study the role of vitamin D in UV-induced immunomodulation," *Photochemical & Photobiological Sciences*, vol. 11, no. 12, pp. 1788–1796, 2012.
- [18] S. Gorman, M. A. Judge, and P. H. Hart, "Topical 1,25-dihydroxyvitamin D3 subverts the priming ability of draining lymph node dendritic cells," *Immunology*, vol. 131, no. 3, pp. 415–425, 2010.
- [19] R. L. X. Ng, N. M. Scott, J. L. Bisley et al., "Characterization of regulatory dendritic cells differentiated from the bone marrow of UV-irradiated mice," *Immunology*, vol. 140, no. 4, pp. 399– 412, 2013.
- [20] R. L. X. Ng, J. L. Bisley, S. Gorman, M. Norval, and P. H. Hart, "Ultraviolet irradiation of mice reduces the competency of bone marrow-derived CD11c+ cells via an indomethacin-inhibitable pathway," *Journal of Immunology*, vol. 185, no. 12, pp. 7207–7215, 2010.
- [21] P. H. Hart, M. A. Grimbaldeston, G. J. Swift, A. Jaksic, F. P. Noonan, and J. J. Finlay-Jones, "Dermal mast cells determine susceptibility to ultraviolet B-induced systemic suppression of contact hypersensitivity responses in mice," *Journal of Experimental Medicine*, vol. 187, no. 12, pp. 2045–2053, 1998.
- [22] S. N. Byrne, A. Y. Limón-Flores, and S. E. Ullrich, "Mast cell migration from the skin to the draining lymph nodes upon ultraviolet irradiation represents a key step in the induction of immune suppression," *The Journal of Immunology*, vol. 180, no. 7, pp. 4648–4655, 2008.
- [23] S. N. Byrne and G. M. Halliday, "B cells activated in lymph nodes in response to ultraviolet irradiation or by interleukin-10 inhibit dendritic cell induction of immunity," *Journal of Investigative Dermatology*, vol. 124, no. 3, pp. 570–578, 2005.
- [24] Y. Matsumura, S. N. Byrne, D. X. Ngheim, Y. Miyahara, and S. E. Ullrich, "A role for inflammatory mediators in the induction of immunoregulatory B cells," *Journal of Immunology*, vol. 177, no. 7, pp. 4810–4817, 2006.
- [25] D. P. T. Steenvoorden and B. Van Henegouwen, "Protection against UV-induced systemic immunosuppression in mice by a single topical application of the antioxidant vitamins C and E," *International Journal of Radiation Biology*, vol. 75, no. 6, pp. 747–755, 1999.
- [26] B. K. Armstrong and A. Kricker, "The epidemiology of UV induced skin cancer," *Journal of Photochemistry and Photobiol*ogy B: Biology, vol. 63, no. 1-3, pp. 8–18, 2001.
- [27] S. J. Miller, M. Alam, J. Andersen et al., "Basal cell and squamous cell skin cancers," *Journal of the National Comprehensive Cancer Network*, vol. 8, no. 8, pp. 836–864, 2010.
- [28] G. Pennello, S. Devesa, and M. Gail, "Association of surface ultraviolet B radiation levels with melanoma and nonmelanoma skin cancer in United States blacks," *Cancer Epidemiology, Biomarkers and Prevention*, vol. 9, no. 3, pp. 291–297, 2000.
- [29] B. L. Diffey, "Solar ultraviolet radiation effects on biological systems," *Physics in Medicine and Biology*, vol. 36, no. 3, pp. 299– 328, 1991
- [30] M. Wacker and M. F. Holiack, "Vitamin D—effects on skeletal and extraskeletal health and the need for supplementation," *Nutrients*, vol. 5, no. 1, pp. 111–148, 2013.
- [31] N. Khazai, S. E. Judd, and V. Tangpricha, "Calcium and vitamin D: skeletal and extraskeletal health," *Current Rheumatology Reports*, vol. 10, no. 2, pp. 110–117, 2008.

- [32] G. M. Halliday, D. L. Damian, S. Rana, and S. N. Byrne, "The suppressive effects of ultraviolet radiation on immunity in the skin and internal organs: implications for autoimmunity," *Journal of Dermatological Science*, vol. 66, no. 3, pp. 176–182, 2012.
- [33] P. H. Hart, S. Gorman, and J. J. Finlay-Jones, "Modulation of the immune system by UV radiation: more than just the effects of vitamin D?" *Nature Reviews Immunology*, vol. 11, no. 9, pp. 584– 596, 2011.
- [34] M. Norval and G. M. Halliday, "The consequences of UVinduced immunosuppression for human health," *Photochem-istry and Photobiology*, vol. 87, no. 5, pp. 965–977, 2011.
- [35] K. M. Dixon, W. Tongkao-On, V. B. Sequeira et al., "Vitamin D and death by sunshine," *International Journal of Molecular Sciences*, vol. 14, no. 1, pp. 1964–1977, 2013.
- [36] R. M. Lucas and A.-L. Ponsonby, "Considering the potential benefits as well as adverse effects of sun exposure: can all the potential benefits be provided by oral vitamin D supplementation?," Progress in Biophysics and Molecular Biology, vol. 92, no. 1, pp. 140–149, 2006.
- [37] R. M. Lucas, M. Norval, R. E. Neale et al., "The consequences for human health of stratospheric ozone depletion in association with other environmental factors," *Photochemical and Photobi*ological Sciences, vol. 14, no. 1, pp. 53–87, 2015.
- [38] T. Schwarz and S. Beissert, "Milestones in photoimmunology," The Journal of Investigative Dermatology, vol. 133, no. 1, pp. E7– E10, 2013.
- [39] T. Schwarz, "Photoimmunosuppression," Photodermatology, Photoimmunology and Photomedicine, vol. 18, no. 3, pp. 141–145, 2002
- [40] T. Schwarz, "The dark and the sunny sides of UVR-induced immunosuppression: photoimmunology revisited," *Journal of Investigative Dermatology*, vol. 130, no. 1, pp. 49–54, 2010.
- [41] S. Kannan and H. W. Lim, "Photoprotection and vitamin D: a review," *Photodermatology, Photoimmunology & Photomedicine*, vol. 30, no. 2-3, pp. 137–145, 2013.
- [42] S. N. Byrne, "How much sunlight is enough?" *Photochemical & Photobiological Sciences*, vol. 13, no. 6, pp. 840–852, 2014.
- [43] V. E. Reeve, R. M. Tyrrell, M. Allanson, D. Domanski, and L. Blyth, "The role of interleukin-6 in UVA protection against UVB-induced immunosuppression," *Journal of Investigative Dermatology*, vol. 129, no. 6, pp. 1539–1546, 2009.
- [44] S. Majewski, C. Jantschitsch, A. Maeda, T. Schwarz, and A. Schwarz, "IL-23 antagonizes UVR-induced immunosuppression through two mechanisms: reduction of UVR-induced DNA damage and inhibition of UVR-induced regulatory T cells," *The Journal of Investigative Dermatology*, vol. 130, no. 2, pp. 554–562, 2010.
- [45] L. Wang, S. C. Jameson, and K. A. Hogquist, "Epidermal langerhans cells are not required for UV-induced immunosuppression," *Journal of Immunology*, vol. 183, no. 9, pp. 5548–5553, 2009.
- [46] A. Schwarz, F. Navid, T. Sparwasser, B. E. Clausen, and T. Schwarz, "1,25-Dihydroxyvitamin D exerts similar immuno-suppressive effects as UVR but is dispensable for local UVR-induced immunosuppression," *The Journal of Investigative Dermatology*, vol. 132, no. 12, pp. 2762–2769, 2012.
- [47] S. Gorman, N. M. Scott, D. H. W. Tan et al., "Acute erythemal ultraviolet radiation causes systemic immunosuppression in the absence of increased 25-hydroxyvitamin D₃ levels in male mice," *PLoS ONE*, vol. 7, no. 9, Article ID e46006, 2012.

- [48] Q. Zhang, Y. Yao, R. L. Konger et al., "UVB radiation-mediated inhibition of contact hypersensitivity reactions is dependent on the platelet-activating factor system," *Journal of Investigative Dermatology*, vol. 128, no. 7, pp. 1780–1787, 2008.
- [49] A. Guéniche, J. Benyacoub, T. M. Buetler, H. Smola, and S. Blum, "Supplementation with oral probiotic bacteria maintains cutaneous immune homeostasis after UV exposure," *European Journal of Dermatology*, vol. 16, no. 5, pp. 511–517, 2006.
- [50] H. Li, R. Prasad, S. K. Katiyar, N. Yusuf, C. A. Elmets, and H. Xu, "Interleukin-17 mediated inflammatory responses are required for ultraviolet radiation-induced immune suppression," *Photo-chemistry and Photobiology*, vol. 91, no. 1, pp. 235–241, 2015.
- [51] S. Rana, L. J. Rogers, and G. M. Halliday, "Systemic low-dose UVB inhibits CD8 T cells and skin inflammation by alternative and novel mechanisms," *The American Journal of Pathology*, vol. 178, no. 6, pp. 2783–2791, 2011.
- [52] K. M. Dixon, A. W. Norman, V. B. Sequeira et al., "1α,25(OH)₂-vitamin D and a nongenomic vitamin D analogue inhibit ultraviolet radiation-induced skin carcinogenesis," *Cancer Prevention Research*, vol. 4, no. 9, pp. 1485–1494, 2011.
- [53] A. Schwarz, A. Maeda, M. K. Wild et al., "Ultraviolet radiation-induced regulatory T cells not only inhibit the induction but can suppress the effector phase of contact hypersensitivity," *Journal of Immunology*, vol. 172, no. 2, pp. 1036–1043, 2004.
- [54] S. Gorman, J. W.-Y. Tan, J. A. Thomas et al., "Primary defect in UVB-induced systemic immunomodulation does not relate to immature or functionally impaired APCs in regional lymph nodes," *The Journal of Immunology*, vol. 174, no. 11, pp. 6677– 6685, 2005.
- [55] L. Wang, K. Saito, M. Toda et al., "UV irradiation after immunization induces type 1 regulatory T cells that suppress Th2-type immune responses via secretion of IL-10," *Immunobiology*, vol. 215, no. 2, pp. 124–132, 2010.
- [56] P. Wolf, C. Hoffmann, F. Quehenberger, S. Grinschgl, and H. Kerl, "Immune protection factors of chemical sunscreens measured in the local contact hypersensitivity model in humans," *Journal of Investigative Dermatology*, vol. 121, no. 5, pp. 1080–1087, 2003.
- [57] J. Breuer, N. Schwab, T. Schneider-Hohendorf et al., "Ultraviolet B light attenuates the systemic immune response in central nervous system autoimmunity," *Annals of Neurology*, vol. 75, no. 5, pp. 739–758, 2014.
- [58] Center for Drug Evaluation and Research, About the Center for Drug Evaluation and Research—Sunburn Protection Factor (SPF), Center for Drug Evaluation and Research, http://www .fda.gov/AboutFDA/CentersOffices/OfficeofMedicalProductsandTobacco/CDER/ucm106351.htm.
- [59] D. A. Kelly, P. T. Seed, A. R. Young, and S. L. Walker, "A commercial sunscreen's protection against ultraviolet radiation-induced immunosuppression is more than 50% lower than protection against sunburn in humans," *Journal of Investigative Dermatology*, vol. 120, no. 1, pp. 65–71, 2003.
- [60] A. Fourtanier, D. Moyal, J. Maccario et al., "Measurement of sunscreen immune protection factors in humans: a consensus paper," *Journal of Investigative Dermatology*, vol. 125, no. 3, pp. 403–409, 2005.
- [61] A. J. Samanek, E. J. Croager, P. Gies et al., "Estimates of beneficial and harmful sun exposure times during the year for major Australian population centres," *Medical Journal of Australia*, vol. 184, no. 7, pp. 338–341, 2006.

[62] J. Reichrath, "Vitamin D and the skin: an ancient friend, revisited," Experimental Dermatology, vol. 16, no. 7, pp. 618–625, 2007.

- [63] A. R. Webb, R. Kift, M. T. Durkin et al., "The role of sunlight exposure in determining the vitamin D status of the U.K. white adult population," *British Journal of Dermatology*, vol. 163, no. 5, pp. 1050–1055, 2010.
- [64] A. R. Ralph, R. M. Lucas, and M. Norval, "Vitamin D and solar ultraviolet radiation in the risk and treatment of tuberculosis," *The Lancet Infectious Diseases*, vol. 13, no. 1, pp. 77–88, 2013.
- [65] P. Autier, M. Boniol, C. Pizot, and P. Mullie, "Vitamin D status and ill health: a systematic review," *The Lancet Diabetes & Endocrinology*, vol. 2, no. 1, pp. 76–89, 2014.
- [66] J. Narbutt, A. Lesiak, A. Sysa-Jedrzejowska et al., "Repeated low-dose ultraviolet (UV) B exposures of humans induce limited photoprotection against the immune effects of erythemal UVB radiation," *British Journal of Dermatology*, vol. 156, no. 3, pp. 539–547, 2007.
- [67] B. Petersen, H. C. Wulf, M. Triguero-Mas et al., "Sun and ski holidays improve vitamin D status, but are associated with high levels of DNA damage," *The Journal of Investigative Dermatology*, vol. 134, no. 11, pp. 2806–2813, 2014.
- [68] T. Yoshikawa, V. Rae, W. Bruins-Slot, J.-W. Van den Berg, J. R. Taylor, and J. W. Streilein, "Susceptibility to effects of UVB radiation on induction of contact hypersensitivity as a risk factor for skin cancer in humans," *The Journal of Investigative Dermatology*, vol. 95, no. 5, pp. 530–536, 1990.
- [69] F. P. Noonan and H. A. Hoffman, "Susceptibility to immunosuppression by ultraviolet B radiation in the mouse," *Immuno*genetics, vol. 39, no. 1, pp. 29–39, 1994.
- [70] M. M. Welsh, M. R. Karagas, J. K. Kuriger et al., "Genetic determinants of UV-susceptibility in non-melanoma skin cancer," *PLoS ONE*, vol. 6, no. 7, Article ID e20019, 9 pages, 2011.
- [71] E. Alamartine, P. Berthoux, C. Mariat, F. Cambazard, and F. Berthoux, "Interleukin-10 promoter polymorphisms and susceptibility to skin squamous cell carcinoma after renal transplantation," *The Journal of Investigative Dermatology*, vol. 120, no. 1, pp. 99–103, 2003.
- [72] T. Nagano, M. Kunisada, X. Yu, T. Masaki, and C. Nishigori, "Involvement of interleukin-10 promoter polymorphisms in nonmelanoma skin cancers—a case study in non-Caucasian skin cancer patients," *Photochemistry & Photobiology*, vol. 84, no. 1, pp. 63–66, 2008.
- [73] A. Friedrich, M. Paz, E. Cela, J. Leoni, and D. González Maglio, "Mitochondrial dysfunction and tissue alterations of ultraviolet-irradiated skin in five different mice strains," *Global Journal of Dermatology and Venereology*, vol. 2, no. 1, pp. 4–12, 2014.
- [74] P. Dungel, J. Hartinger, S. Chaudary et al., "Low level light therapy by LED of different wavelength induces angiogenesis and improves ischemic wound healing," *Lasers in Surgery and Medicine*, vol. 46, no. 10, pp. 773–780, 2014.
- [75] J. S. Guffey and J. Wilborn, "Effects of combined 405-nm and 880-nm light on *Staphylococcus aureus* and *Pseudomonas aeruginosa in vitro*," *Photomedicine and Laser Surgery*, vol. 24, no. 6, pp. 680–683, 2006.
- [76] T. Dai, A. Gupta, Y.-Y. Huang et al., "Blue light rescues mice from potentially fatal pseudomonas aeruginosa burn infection: efficacy, safety, and mechanism of action," *Antimicrobial Agents* and Chemotherapy, vol. 57, no. 3, pp. 1238–1245, 2013.

- [77] A. Mamalis, M. Garcha, and J. Jagdeo, "Light emitting diodegenerated blue light modulates fibrosis characteristics: fibroblast proliferation, migration speed, and reactive oxygen species generation," *Lasers in Surgery and Medicine*, vol. 47, no. 2, pp. 210–215, 2015.
- [78] F. Liebel, S. Kaur, E. Ruvolo, N. Kollias, and M. D. Southall, "Irradiation of skin with visible light induces reactive oxygen species and matrix-degrading enzymes," *Journal of Investigative Dermatology*, vol. 132, no. 7, pp. 1901–1907, 2012.
- [79] S. Vandersee, M. Beyer, J. Lademann, and M. E. Darvin, "Blue-violet light irradiation dose dependently decreases carotenoids in human skin, which indicates the generation of free radicals," Oxidative Medicine and Cellular Longevity, vol. 2015, Article ID 579675, 7 pages, 2015.
- [80] N. Maddodi, A. Jayanthy, and V. Setaluri, "Shining light on skin pigmentation: the darker and the brighter side of effects of UV radiation," *Photochemistry and Photobiology*, vol. 88, no. 5, pp. 1075–1082, 2012.
- [81] L. Duteil, N. Cardot-Leccia, C. Queille-Roussel et al., "Differences in visible light-induced pigmentation according to wavelengths: a clinical and histological study in comparison with UVB exposure," *Pigment Cell & Melanoma Research*, vol. 27, no. 5, pp. 822–826, 2014.
- [82] P. Ramaswamy, J. G. Powers, J. Bhawan, I. Polyak, and B. A. Gilchrest, "Effective blue light photodynamic therapy does not affect cutaneous langerhans cell number or oxidatively damage DNA," *Dermatologic Surgery*, vol. 40, no. 9, pp. 979–987, 2014.
- [83] D. Becker, E. Langer, M. Seemann et al., "Clinical efficacy of blue light full body irradiation as treatment option for severe atopic dermatitis," PLoS ONE, vol. 6, no. 6, Article ID e20566, 2011.
- [84] M. R. Fischer, M. Abel, S. Lopez Kostka, B. Rudolph, D. Becker, and E. von Stebut, "Blue light irradiation suppresses dendritic cells activation in vitro," *Experimental Dermatology*, vol. 22, no. 8, pp. 558–560, 2013.
- [85] G. Monfrecola, S. Lembo, M. Cantelli et al., "The effect of visible blue light on the differentiation of dendritic cells in vitro," *Biochimie*, vol. 101, no. 1, pp. 252–255, 2014.
- [86] M. E. Darvin, S. F. Haag, J. Lademann, L. Zastrow, W. Sterry, and M. C. Meinke, "Formation of free radicals in human skin during irradiation with infrared light," *The Journal of Investigative Dermatology*, vol. 130, no. 2, pp. 629–631, 2010.
- [87] P. Schroeder, C. Pohl, C. Calles, C. Marks, S. Wild, and J. Krutmann, "Cellular response to infrared radiation involves retrograde mitochondrial signaling," Free Radical Biology and Medicine, vol. 43, no. 1, pp. 128–135, 2007.
- [88] P. Schroeder, J. Lademann, M. E. Darvin et al., "Infrared radiation-induced matrix metalloproteinase in human skin: implications for protection," *Journal of Investigative Dermatology*, vol. 128, no. 10, pp. 2491–2497, 2008.
- [89] C. Jantschitsch, S. Majewski, A. Maeda, T. Schwarz, and A. Schwarz, "Infrared radiation confers resistance to UV-induced apoptosis via reduction of DNA damage and upregulation of antiapoptotic proteins," *The Journal of Investigative Dermatology*, vol. 129, no. 5, pp. 1271–1279, 2009.
- [90] C. Jantschitsch, M. Weichenthal, A. Maeda, E. Proksch, T. Schwarz, and A. Schwarz, "Infrared radiation does not enhance the frequency of ultraviolet radiation-induced skin tumors, but their growth behaviour in mice," *Experimental Dermatology*, vol. 20, no. 4, pp. 346–350, 2011.
- [91] C.-H. Lee, C.-H. Hong, W.-T. Liao, and H.-S. Yu, "Differential immunological effects of infrared irradiation and its associated

- heat in vivo," *Journal of Photochemistry and Photobiology B: Biology*, vol. 155, pp. 98–103, 2016.
- [92] S. Y. Celine Lee, I.-W. Seong, J.-S. Kim et al., "Enhancement of cutaneous immune response to bacterial infection after low-level light therapy with 1072 nm infrared light: a preliminary study," *Journal of Photochemistry and Photobiology B: Biology*, vol. 105, no. 3, pp. 175–182, 2011.
- [93] N. R. S. Santos, J. B. M. Sobrinho, P. F. Almeida et al., "Influence of the combination of infrared and red laser light on the healing of cutaneous wounds infected by *Staphylococcus aureus*," *Photomedicine and Laser Surgery*, vol. 29, no. 3, pp. 177– 182, 2011.
- [94] K. A. Muili, S. Gopalakrishnan, S. L. Meyer, J. T. Eells, and J.-A. Lyons, "Amelioration of experimental autoimmune encephalomyelitis in C57BL/6 mice by photobiomodulation induced by 670 nm light," *PLoS ONE*, vol. 7, no. 1, Article ID e30655, 2012.
- [95] M. R. Hamblin, "Shining light on the head: photobiomodulation for brain disorders," BBA Clinical, vol. 6, pp. 113–124, 2016.
- [96] E. M. Cela, A. Friedrich, M. L. Paz, S. I. Vanzulli, J. Leoni, and D. H. González Maglio, "Time-course study of different innate immune mediators produced by UV-irradiated skin: comparative effects of short and daily versus a single harmful UV exposure," *Immunology*, vol. 145, no. 1, pp. 82–93, 2015.
- [97] S. P. Hong, M. J. Kim, M.-Y. Jung et al., "Biopositive effects of low-dose UVB on epidermis: coordinate upregulation of antimicrobial peptides and permeability barrier reinforcement," *Journal of Investigative Dermatology*, vol. 128, no. 12, pp. 2880– 2887, 2008.
- [98] N. M. Khaskhely, M. Maruno, H. Uezato et al., "Low-dose UVB contributes to host resistance against *Leishmania amazonensis* infection in mice through induction of gamma interferon and tumor necrosis factor alpha cytokines," *Clinical and Diagnostic Laboratory Immunology*, vol. 9, no. 3, pp. 677–686, 2002.