Taylor & Francis

OPEN ACCESS Check for updates

Drug release and magneto-calorific analysis of magnetic lipid microcapsules for potential cancer therapeutics

Hongmei Bi^{a,b}, Zeqin Chen^a and Jiaqin Qiu^a

^aCollege of Biological and Food Engineering, Guangdong University of Petrochemical Technology, Maoming, China; ^bCollege of Science, Heilongjiang Bayi Agricultural University, Daqing, China

ABSTRACT

Magnetic nanoparticles (MNPs) with safety, stability and excellent magneto-calorific effect are the precondition for the smart magnetic drug carriers' fabrication and controllable drug release at a specific target in clinical treatment. In this study, the drug release and magneto-calorific effect of two types of magnetic lipid microcapsules (MLMs) loading lipid-coated MNPs and uncoated MNPs respectively were compared deeply in experimental analysis and theoretical simulation. The simulation results revealed that almost same magnetic heat effect and temperature increasing exist between lipid-coated and uncoated MNPs, which was consistent with the experimental drug release results. Coating lipid on MNPs didn't affect the magnetic heat and heat transfer of the MNPs. Because of the heat transfer between MNPs and water, MLMs and water around, the temperature increasing of whole sample solution is lower than that of the MNPs themselves. Our results provide a reliable theoretical basis for the development of healthy, safe, and biocompatible drug delivery systems.

ARTICLE HISTORY

Received 5 April 2021 Accepted 10 May 2021

KEYWORDS

Lipid-coated MNPs; drug release; magneto-calorific effect; heat transfer

1 Introduction

Drug delivery carriers based on MNPs are explored intensively in biomedical field to amplify drug efficacy in clinical treatment such as remote-controlled drug release at the desired targeted regions or active pathology by magnetic hyperthermia under an alternating magnetic field (AMF) [1]. Because of their disadvantages of aggregation and precipitation[2], safe and stable MNPs are the precondition to expand their actual clinic application [3,4]. Organic materials [5,6], inorganic materials [7,8], and biomaterials [9,10] have been designed to composite with MNPs and further fabricate magnetic drug delivery carriers. Among them, lipid is a promising material in medicinal carrier application because it is the main component of the cytomembrane and show excellent biocompatibility and biosafety. Magnetoliposomes (MLs) or lipid/ drug/materials hybrid structures were also exploited for the drug delivery and release application [11]. Although different release mechanisms of MLs have been presented such as changed permeability or disruption of the lipid membrane in an AMF, the magnetic heat of loading MNPs in their lumen or embedded/absorbed MNPs in lipid membrane played an important role in payload release [12]. Our group introduced a novel kind of lipid-coated MNPs into

drug release system [9]. The dispersion, stability, and biocompatibility of the lipid-coated MNPs were enhanced, but no significant temperature increasing of the sample solution was observed in the drug release process. Other groups have also reported the payload release of MLs without a significant increase in local temperatures, regardless of the release mechanism [13,14]. It is necessary to reveal the essence of magnetic heat of MNPs and whole system in theoretically. So, in order to explore them in AMF deeply, the drug release behaviors of MLMs loaded with lipid-coated and uncoated MNPs in their lumen respectively were compared in experimentally. The magnetic heat of the two types of MNPs was further investigated by theoretical simulation using COMSOL software. The simulation results revealed almost the same magnetic heat effect and temperature increase for the lipid-coated and uncoated MNPs. Coating lipid on the MNPs did not affect their magnetic heat. While because of the existent heat transfer between MNPs and water, MLMs and water around, the temperature increasing of whole sample solution is lower than that of the MNPs themselves. Our results provide a reliable theoretical basis for the development of healthy, safe, and biocompatible drug delivery systems.

CONTACT Hongmei Bi Angmei_bi@126.com College of Biological and Food Engineering, Guangdong University of Petrochemical Technology, Maoming, 525000, China, College of Science, Heilongjiang Bayi Agricultural University, Daqing, 163319

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

2 Materials and methods

2.1 Materials

1,2-dioleoyl-sn-glycero-3-phosphocholine (DOPC) was purchased from Avanti Polar Lipids (USA). Fluorescencelabeled 1,2-dioleoyl-sn-glycero-3-phosphoethanolamine-N-(7-nitro-2-1,3-benzoxadiazol-4-yl) (NBD PE), Texas red-labeled 1.2-dihexadecanovl-sn-glycero -3-phosphoethanolamine triethylammonium salt (TR-DHPE), and 5-(6)-carboxylfluorescein (CF) were purchased from Invitrogen (Eugene, OR). Indium tin oxide (ITO, sheet resistance \approx 8–12 Ω , thickness \approx 160 nm) electrodes were purchased from Hangzhou Yuhong Technology Co. Ltd. (China). Trisodium citrate, Triton X-100, ferric chloride, ferrous chloride, and chloroform were purchased from Sigma (China). Ethanol and ammonium hydroxide (all analytical grade, purity > 99.5%) were purchased from FuYu Chemicals (China). Millipore Milli-Q water with a resistivity of 18.0 M Ω ·cm was used for solution preparation in all experiments.

2.2 Preparation and modification of MNPs

Hydrophilic MNPs were synthesized by co-precipitation of FeCl₃ and FeCl₂ in alkaline condition[15]. Firstly, a mixed solution containing 1.0 g of $FeCl_3 \cdot 6H_2O$ and 0.5 g of FeCl₂ · 4H₂O was stirred for 5 min at 80 °C. Then 5.0 mL of concentrated NH₃·H₂O was added into the solution, which was further stirred for 30 min. Another 5.0 mL of 17 % trisodium citrate solution was added, then the temperature was increased to 95 °C for 90 min under vigorous stirring. The citrate-stabilized uncoated MNPs were obtained by collecting the precipitate and washing with water. Lipid-coated MNPs were prepared by the disruption of DOPC vesicles on uncoated MNPs. The DOPC vesicles were first prepared by an extrusion approach[16]. Dried lipid film in a glass flask (1 mg DOPC) was hydrated in deionized water by vortex, which was followed by extrusion through a 100-nmpore-size polycarbonate membrane 21 times using a mini-extruder (Avanti Polar Lipids, Inc.). Fresh citratestabilized MNPs solution was mixed with freshly prepared DOPC vesicle solution and PBS (0.2 mol/L, pH 7.4) buffer solution, which was followed by for 2 h incubation. Lipid-coated MNPs were obtained after thoroughly washing with distilled water.

2.3 *Preparation of fluorescent MLMs and CF release in AMF*

MLMs were prepared by the electro-formation method using Indium Tin Oxide (ITO)- electrode. Dried lipid film was prepared by flatting a certain volume of 5.0 mg/mL DOPC (0.5% TR-DHPE, mass ratio) chloroform solution onto an ITO electrode surface. After drying under vacuum for 2 h, the chamber was filled with mixed solution of CF (0.05 mg/mL) and MNPs. Finally, an AC electric field of 5 V and 10 Hz was applied for 3 h to induce MLMs formation. Then the generated MLMs were filtered with a polycarbonate membrane (400 nm) to remove the unpacked MNPs and CF. The fluorescence intensity of released CF (excitation 492 nm, emission 517 nm) of the sample solutions $(1.2 \sim 1.4 \times 10^6 \text{ MLMs/})$ mL) was measured by a fluorescence spectrometer (PerkinElmer, LS55) upon applying an AMF (WT20C, China). The emission fluorescence spectra of CF were recorded between 500 and 650 nm with excitation wavelength at 492 nm and 10.0 nm slit (PerkinElmer, LS55). Triton X-100 was used to completely disrupt the MLMs and measure the total fluorescence intensity. The release percentage of CF from MLMs was calculated using below equation:

$$ext{%release} = rac{I_{ML}(ext{t}) - I_{ML}(ext{0})}{I_{MAX}(ext{t}) - I_{ML}(ext{0})} imes 100\%$$

where I_{ML} (t) is fluorescence intensity of released CF from the treated sample; $I_{ML}(0)$ is the fluorescence intensity of CF from the control sample; and I_{MAX} is the total fluorescence intensity of the sample.

2.4 Finite element analysis of Magneto-calorific effect

The heat transfer/conjugate heat transfer module and the magnetic induction heating module in COMSOL Multiphysics 4.3 were used to simulate the magnetocalorific. Fluid convection was ignored in the temperature range of the magneto-calorific simulation. [17] All simulations were performed with a triangular grid mesh. 2.7×10^5 triangular elements were involved in the simulation. The mesh was adjusted to obtain the required convergence within a reasonable computing time scale with the required accuracy. The input parameters in simulation are shown in Table S1.

3 Results and discussion

3.1 Preparation of MNPs and MLMs

MNPs prepared by the co-precipitation method have desirable water solubility and size controllability[10]. TEM images, FTIR spectra, X-ray powder diffraction (XRD), and magnetic hysteresis loop analysis confirmed that these uncoated MNPs are superparamagnetic Fe_3O_4 with surface functional carboxyl (COOH) groups. (shown in Figures 1-Figure S5) The mean diameter of uncoated



Figure 1. Histogram of MLMs diameter distributions. The statistical samples were selected randomly over 200 for each distribution.

MNPs is about 13.2 ± 2.1 nm, the thickness of a lipid film on lipid-coated MNPs is about 5.3 ± 0.6 nm. Excellent dispersion and stability of these lipid-coated MNPs were also shown after one month storage. The response to external magnetic field of lipid-coated MNPs showed their excellent superparamagnetic character, which is very important for heat generation in AMF irradiation. The same Zeta potential of lipid-coated MNPs and pure DOPC vesicles also indicate the MNPs were wrapped with lipid successfully. In order to compare the drug release and magnetic heat effectively, MLMs loading uncoated and lipid-coated were prepared again in same experimental conditions using the same method reported before^[9]. The size distributions of these two MLMs according to the results are plotted in Figure 1. The average diameter was similar for both types of MLMs with 17 \pm 2.7 μ m (loading uncoated MNPs) and $18 \pm 3.2 \ \mu m$ (loading lipid-coated MNPs) respectively. Dry lipid films swell and hump to grow a sphere in an alternating electric field and MNPs enter into the lumen of MLMs during this swelling process. Maybe the weak attraction between uncoated MNPs and lipid membrane restricts the growth of MLMs, while there was no attraction between lipid-coated MNPs and lipid membrane, which lead to the greater size of formed MLMs loading lipid-coated MNPs as shown in Figure 1.

3.2 CF release of MLMs under AMF

Polycarbonate membrane was used to filter away nonencapsulated nanoparticles and CF to obtain an ideal sample solution. AMF with frequency of 100 Hz, 500 Hz, 1 kHz, and 5 kHz respectively was applied to induce the release of the included CF. The release behavior of CF



Figure 2. CF release of MLMs loading lipid-coated MNPs and uncoated MNPs at different frequencies. The error bars show the standard deviation (SD) of the results for three samples.

was plotted in Figure 2, where the solid and dotted lines represent the CF released from MLMs loaded with uncoated and lipid-coated MNPs, respectively. Dotted lines serves as the contrast curves in this study, which are almost the same results as before^[9].

The control in Figure 2 denotes the CF release of MLMs without an AMF under the same experimental conditions. Because of the fluid state of the lipid bilayer and the oscillation of MNPs at room temperature, about 5% CF release occurred under the natural condition (23 ° C) without an AMF. It is noted that the CF release percentage and release rate increases gradually with the increasing frequency. CF release was sluggish at low frequency such as 100 Hz, while the rate of CF release markedly increased at higher frequencies. When the Fe concentration was fixed at 20.0 µg/mL, the maximum release percentage at 100 Hz was 8%, while it reached to 53% at a frequency of 5 kHz after 120 min. The total release rate decreased gradually with time at every freguency because the CF concentration gradient between the inside and outside of the MLMs decreased, reducing the driving force of the release [3,18]. According to magneto-caloric theory, frequency plays a key role in inducing the magnetic heating generation of MNPs in an AMF. The greater magnetic heat generated at higher frequency is conducive to the leaking of intraliposomal contents out through the destabilized lipid bilayer[19]. However, the CF release tendency and release percentage of MLMs showed little difference between the MLMs loading lipid-coated and uncoated MNPs regardless of the frequency according to our experimental results. It demonstrates that the lipid-coated and uncoated MNPs have the same magneto-calorific effect in an AFM, the lipid coating on MNPs does not affect their magneto-calorific effect and the subsequent release of content from the MLMs.

3.3 Finite element analysis of magneto-calorific effect

In order to further analyze the experimental results of CF release and compare the heating effect between lipidcoated and uncoated MNPs, the magneto-calorific effects of these two systems were simulated. The parameters in the simulation were all based on the actual magnetic field and experimental conditions[20]. Curves showing the temperature changes of the two types of MNPs generated by the magneto-calorific effect at different frequencies are plotted in Figure 3a, where the solid and dotted lines represent the temperatures of uncoated and lipid-coated MNPs, respectively. The temperature of the MNPs increased upon increasing the frequency from 100 Hz to 5 kHz. The slightly temperature increasing at 100 Hz indicate that there was no significant magnetic loss at low frequency such as 100 Hz. When the frequency was 500 Hz and above,



Figure 3. (a) Simulated temperature change curves of two types of MNPs at Fe concentration of 20.0 μ g/mL. (b) Simulation model of MLM and color maps showing temperature change based on heat transfer at 5 kHz.

a higher temperature was observed, especially at 5 kHz. Clearly, a higher frequency induced more heating of the MNPs and led to the higher temperature, which accelerated the fluidity of the lipid bilayer and promoted CF leakage as shown in Figure 2. The simulation results show good agreement with the dissipation equation based on the theory of relaxation losses [21,22]. Another notable result is that the temperature of lipidcoated MNPs was higher than uncoated MNPs at every frequency in the simulation, although the difference was small. We consider that this slight temperature difference was caused by the set value of the MNPs diameter in the simulation; because of the thin lipid layer coated on the MNPs, the lipid-coated MNPs were slightly larger than the uncoated MNPs, as revealed by TEM characterization[9], and we set the obtained mean diameter in the simulation process. Within a certain size range, larger MNPs will generate greater magnetic heat, leading to a higher temperature. In fact, the amount of released CF did not show a significant difference between the MLMs loading lipid-coated and uncoated MNPs as shown in Figure 2. This means that the lipid coating on the surface of the MNPs does not affect their heating efficiency and subsequent drug release.

Although on the basis of the strong magnetocalorific effect at high frequency, the temperature should reach to 360 K at 5 kHz according to our simulation results, we did not macroscopically monitor the temperature change of the actual sample solution as shown in Figure 3a. To consider the heat transfer between the MNPs and the surrounding water, the MLMs and the medium outside, we fabricated a spatial model of the whole system of an MLM loaded with uncoated MNPs, the color maps of temperature change at an AMF frequency of 5 kHz are shown in Figure 3b. The volume ratio of the MLM core and water shell in the spatial model was chosen in accordance with the actual MLMs sample. Initially, the MLMs and the surrounding region had the same temperature of about 275 K before AMF application, while the temperature increased to 320 K after 10 min of the 5 kHz AMF. This temperature is really lower than that of 360 K in Figure 3a. We consider the reason of different temperature increasing in these two simulations lies in the heating transfer in actual system such as MNPs and water, MLMs and water around. It clearly can be seen that the water close to MLMs was heated in the spatial model (Figure 3b). The generated magnetic heat of MNPs was fast transfer to lots of water around, so the temperature of whole system is lower than that in simulation in Figure 3a. In fact, we really did not monitor the obvious temperature increasing in

experimental process, which was confirmed by the simulated results in Figure 3b. We consider that the ideal temperature increasing as theoretical simulation in Figure 3a only exist in part of the MNPs or MLMs themselves, which cannot be macroscopically measured under our experimental conditions [23,24]. The theoretical simulation and the analysis of the spatial model explained the drug release results of two MLM systems and the temperature change of the whole system.

4 Conclusions

In this work, the drug release and magneto-calorific effect of two types of MLMs loading lipid-coated MNPs and uncoated MNPs respectively were compared in depth by experimental analysis and theoretical simulation. The parameters in the simulation were all based on actual experimental conditions. The experimental results show that the release percentage and release rate of the two types of MLMs increased gradually with increasing frequency, but the difference between the two types of MLMs was small. It was proved by the magneto-calorific effect simulation of two types of MNPs, where almost equal temperature and temperature change of lipidcoated and uncoated MNPs were shown in every frequency. The magnetic heat and heat transfer of the MNPs were not affected by the lipid coating on the MNPs. The simulated results of considering the actual heat transfer showed a weak temperature increasing in the spatial model, which explained why it difficult to monitor the obvious temperature increasing of the whole system in experimental process. The results of theoretical simulation of the magneto-calorific effect and heat transfer were all consistent with the experimental results. Our results provide a reliable theoretical basis for the development of new types of MNPs and the fabrication of healthy, safe, and biocompatible drug delivery systems.

Acknowledgments

This work was supported by the Natural Science Foundation of Guangdong Province (No.2020A1515010522), Characteristic Innovation Projects of Universities in Guangdong Province (No.2019KTSCX109), Projects of Talents Recruitment of GDUPT (No.2019rc114), and Natural Science Foundation of Heilongjiang Province (No.LH2019B012).

Disclosure statement

No potential conflict of interest was reported by the author(s). The disclosure statement has been inserted. Please correct if this is inaccurate.

Funding

This work was supported by the Natural Science Foundation of Guangdong Province [2020A1515010522]; Projects of Talents Recruitment of GDUPT [2019rc114]; Characteristic Innovation Projects of Universities in Guangdong Province [2019KTSCX109]; Natural Science Foundation of Heilongjiang Province (LH2019B012).

References

- [1] Kubo T, Tachibana K, Naito T, et al. Magnetic field stimuli-sensitive drug release using a magnetic thermal seed coated with thermal-responsive molecularly imprinted polymer. ACS Biomater. Sci. Eng. 2019;5 (2):759–767.
- [2] N'Guyen TTT, Duong HTT, Basuki J, et al. Functional iron oxide magnetic nanoparticles with hyperthermia-induced drug release ability by using a combination of orthogonal click reactions. Angew. Chem. Int. Edit. 2013;52(52):14152–14156.
- [3] Nandwana V, Ryoo S-R, Zheng T, et al. Magnetic nanostructure-coated thermoresponsive hydrogel nanoconstruct as a smart multimodal theranostic platform. ACS Biomater. Sci. Eng. 2019;5(6):3049–3059.
- [4] Ahmad A, Gupta A, Ansari MM, et al. Hyperbranched polymer-functionalized magnetic nanoparticle-mediated hyperthermia and niclosamide bimodal therapy of colorectal cancer cells. ACS Biomater. Sci. Eng. 2020;6(2):1102–1111.
- [5] Soares PIP, Sousa AI, Ferreira IMM, et al. Towards the development of multifunctional chitosan-based iron oxide nanoparticles: optimization and modelling of doxorubicin release. Carbohydr Polym. 2016;153:212–221.
- [6] Hoare T, Timko BP, Santamaria J, et al. Magnetically triggered nanocomposite membranes: a versatile platform for triggered drug release. Nano Lett. 2011;11 (3):1395–1400.
- [7] Baeza A, Guisasola E, Ruiz-Hernandez E, et al. Magnetically triggered multidrug release by hybrid mesoporous silica nanoparticles. Chem. Mate. 2012;24(3):517–524.
- [8] Jiang W, Wu J, Tian RB, et al. Synthesis and characterization of magnetic mesoporous core-shell nanocomposites for targeted drug delivery applications. J. Porous. Mat. 2017;24(1):257–265.
- [9] Bi H, Han X. Magnetic field triggered drug release from lipid microcapsule containing lipid-coated magnetic nanoparticles. Chem Phys Lett. 2018;706:455–460.
- [10] Bi H, Ma S, Li Q, et al. Magnetically triggered drug release from biocompatible microcapsules for potential cancer therapeutics. J Mater Chem B. 2016;4(19):3269–3277.
- [11] Sabnis S, Sabnis NA, Raut S, et al. Superparamagnetic reconstituted high-density lipoprotein nanocarriers for magnetically guided drug delivery. Int J Nanomedicine. 2017;12:1453–1464.
- [12] Nappini S, Fogli S, Castroflorio B, et al. Magnetic field responsive drug release from magnetoliposomes in biological fluids. J. Mate. Chem. B. 2016;4(4):716–725.
- [13] Joniec A, Sek S, Krysinski P. Magnetoliposomes as potential carriers of doxorubicin to tumours. Chem- Eu. J. 2016;22(49):17715–17724.

- [14] Salvatore A, Montis C, Berti D, et al. Multifunctional magnetoliposomes for sequential controlled release. ACS Nano. 2016;10(8):7749–7760.
- [15] Massart R. Preparation of aqueous magnetic liquids in alkaline and acidic media. Magnetics, IEEE. Tran. Magn. 1981;17(2):1247–1248.
- [16] Bi H, Wang X, Han X, et al. Impact of electric fields on the nanoscale behavior of lipid monolayers at the surface of graphite in solution. Langmuir. 2018;34 (32):9561–9571.
- [17] Ondeck CL, Habib AH, Ohodnicki P, et al. Theory of magnetic fluid heating with an alternating magnetic field with temperature dependent materials properties for self-regulated heating. J Appl Phys. 2009;105.07B324.
- [18] Preiss MR, Hart A, Kitchens C, et al. Hydrophobic nanoparticles modify the thermal release behavior of liposomes. J Phys Chem B. 2017;121 (19):5040–5047.
- [19] Amstad E, Reimhult E. Nanoparticle actuated hollow drug delivery vehicles. Nanomedicine. 2012;7(1):145–164.
- [20] Habib AH, Ondeck CL, Chaudhary P, et al. Evaluation of iron-cobalt/ferrite core-shell nanoparticles for cancer thermotherapy. J Appl Phys. 2008;103.07A307.
- [21] Rosensweig RE. Towards ferrofluids with enhanced magnetization. J. Magn. Magn. Mater. 2011;323 (10):1191–1197.
- [22] Zhu L, Zhou ZY, Mao H, et al. Magnetic nanoparticles for precision oncology: theranostic magnetic iron oxide

nanoparticles for image-guided and targeted cancer therapy. Nanomedicine. 2017;12(1):73–87.

- [23] Ferjaoui Z, Dine JA, Kulmukhamedova E, et al. Superparamagnetic nanocarriers for controlled drug delivery and magnetic hyperthermia applications. ACS Appli. Mater. Inter. 2019;11(34):30610–30620.
- [24] Yin PT, Shah S, Pasquale NJ, et al. Stem cell-based gene therapy activated using magnetic hyperthermia to enhance the treatment of cancer. Biomaterials. 2016;81:46–57.

Table of Contents



Drug Release and Magneto-calorific Analysis of Magnetic Lipid Microcapsules