# Genetic Differentiation and Evolutionary Adaptation in Cryptomeria japonica

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ABSTRACT Local adaptation of plant species is a central issue for survival during global climate change, especially for long-lived forest trees, with their lengthy regeneration time and spatially limited gene flow. Identification of loci and/or genomic regions associated with local adaptation is necessary for knowledge of both evolution and molecular breeding for climate change. Cryptomeria japonica is an important species for forestry in Japan; it has a broad natural distribution and can survive in a range of different environments. The genetic structure of 14 natural populations of this species was investigated using 3930 SNP markers. Populations on the Pacific Ocean side of Japan are clearly different from those on the Japan Sea side, as discussed in previous studies. Structure analysis and population network trees show that peripheral populations, including the most northerly and southerly ones, have unique features. We found that the genetic differentiation coefficient is low,  $F_{ST}$  = 0.05, although it must account for the presence of important genes associated with adaptation to specific environments. In total, 208 outlier loci were detected, of which 43 were associated with environmental variables. Four clumped regions of outlier loci were detected in the genome by linkage analysis. Linkage disequilibrium (LD) was quite high in these clumps of outlier loci, which were found in linkage groups (LGs) 2, 7, 10, and 11, especially between populations of two varieties, and when interchromosomal LD was also detected. The LG7 region is characteristic of the Yakushima population, which is a large, isolated, peripheral population occupying a specific environment resulting from isolation combined with volcanic activity in the region. The detected LD may provide strong evidence for selection between varieties.

**KEYWORDS** 

LD conifer local adaptation outlier locus selection

Natural selection is the mechanism that drives the evolution of organisms, leading to individuals that are better adapted to a new environment than the original population (Endler 1986; Linhart and Grant

<sup>2</sup>Corresponding author: Department of Forest Genetics, Forestry and Forest Products Research Institute, Tsukuba, Ibaraki 305-8687, Japan. E-mail: ytsumu@ffpri.affrc.go.jp 1996). The fitness of such adapted individuals is higher than the original organisms that spread into the new habitat and, from these, a new species may evolve (Holt and Gomulkiewicz 1997). The genetic mechanism associated with local adaptation has been studied in model plant species such as *Arabidopsis thaliana* (Fournier-Level *et al.* 2011). In long-lived plant species, however, there are few studies of adaptive mechanisms, probably because of the difficulty in evaluating phenotypes over long time periods in the field and the limited genomic information available.

Forest trees have become adapted to specific environments over many generations as the natural distribution has shifted and the population size has changed in response to past global climate change (González-Martínez *et al.* 2006; Savolainen *et al.* 2007; Neale and Ingvarsson 2008; Neale and Kremer 2011). Such climate change may result in the evolution of new species because selection pressure is different depending on climatic conditions, for example, during glacial and interglacial periods. During the past one million years,

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glaciation cycles of 100,000 years duration have prevailed (Howard 1997); this means that the average temperature and precipitation have fluctuated drastically between glacial and interglacial periods every 100,000 years. During glacial periods, small and isolated populations may be left behind within environmental refugia, especially in northern parts of a species' range. Population size would be reduced under severe climatic conditions, and only resistant individuals would survive. Subsequently, offspring of the survivors would colonize out from any refugia during the interglacial period, and genetic differentiation between the isolated population and other populations is likely to have increased during their separation. Repeated glaciations would increase such genetic differentiation and drive the evolution of adaptations to survive severe conditions just like allopatric speciation.

Linkage disequilibrium (LD) throughout the genome reflects the population history, the breeding system, and the pattern of geographic subdivision, whereas LD in each genomic region reflects the history of natural selection, gene conversion, mutation, and other forces that cause gene-frequency evolution (Slatkin 2008). If we observe LD carefully in long-lived plant species, we can detect the genetic signature associated with local adaptation, even in forest tree species, because the history of the organism is recorded in the genome.

Cryptomeria japonica is an allogamous coniferous species that relies on wind-mediated pollen and seed dispersal. Modern natural forests of the species are distributed across various environments in the Japanese Archipelago, from Aomori Prefecture (40° 44' N) to Yakushima Island (30° 15' N) (Hayashi 1951). However, its distribution is discontinuous and scattered; it occupies small, restricted areas as a result of having been extensively exploited by humans over the past 1000 years (Ohba 1993). The geographical variation between natural forests of C. japonica has been investigated, focusing on morphological traits (needle length, needle curvature, and other features) (Murai 1947), diterpene components (Yasue et al. 1987), and reproductive system (Kimura et al. 2013). The results of these studies suggest that there are two main lines: ura-sugi (C. japonica var. radicans, found near the Sea of Japan) and omote-sugi (C. japonica, found near the Pacific Ocean). The ura-sugi variety has slender branchlets with soft leaves, whereas the omote-sugi variety has rough branchlets with hard leaves (Yamazaki 1995). Previous studies using 148 cleaved amplified polymorphic sequence (CAPS) loci and 1026 single nucleotide polymorphisms (SNPs) have revealed genetic differentiation between the two varieties, and four and 14 outlier loci have been identified as potential local adaptation genes, respectively (Tsumura et al. 2007, 2012); these may be associated with genetic differentiation of the varieties.

A high-throughput SNP genotyping system has been developed; using this, tens of thousands of genotypes can be obtained in only a few days. Genome scanning based on a large number of SNPs can result in an accurate evaluation of the genetic diversity and structure of natural populations and facilitates the detection of candidate loci associated with economically important traits and adaptive genes for specific environments (Vasemägi and Primmer 2005; Namroud *et al.* 2008; Holliday *et al.* 2010). This method may allow us to detect loci associated with adaptations that would be valuable for surviving climate change.

In this study, we focus on adaptive genes linked to past climate changes. We investigate the current genetic structure of natural populations of *C. japonica* using several thousand SNPs and characterize the outlier loci using linkage mapping and linkage disequilibrium methods. Then, we discuss their relationship to current genetic structure and how this species has adapted to the different climates experienced on the Japan Sea side and the Pacific Ocean side of the country.

# MATERIALS AND METHODS

## Investigated populations

We examined 14 populations comprising 186 individuals that we used in a previous study (Tsumura *et al.* 2012): seven from the Japan Sea side of Japan and seven from the Pacific Ocean side (Table 1, Figure 1). All trees sampled were growing in national forests that are *in situ* gene-conservation forests in Japan. The locations of the sampled populations covered most of the natural distribution of *C. japonica*. In addition, because *C. japonica* has been widely planted since 1945 in Japan, we only sampled relatively large, old trees that were presumed to predate the widespread planting programs.

# **SNP** genotyping

The identification of SNPs was performed previously by resequencing 5170 unique EST contigs in a discovery panel of four C. japonica individuals collected from a range-wide sample of trees (Uchiyama et al. 2012). The SNPs were also identified from transcriptome assembly of seedlings of 10 trees, range-wide, by a next-generation sequencer (Ueno et al. 2012; Uchiyama et al. 2014). All detected SNPs were EST-based coding regions. Multiplexed genotyping of SNP markers for natural populations was performed using Illumina's four set of 1536-plex (6144 SNPs) GoldenGate array according to the protocol recommended by the manufacturer (Uchiyama et al. 2014). Only SNPs with Illumina design scores more than 0.6 were used. When multiple SNPs were available within the same sequence, a single highly polymorphic SNP was selected as the target for that sequence. The quality of the GoldenGate genotype scores for individual SNPs was assessed based on their GenTrain cluster and GenCall genotype scores in GenomeStudio (Illumina Inc., San Diego, California). A minimum GenCall50 (GC50) score of 0.25 was chosen as the threshold for the inclusion of SNP loci in the final data set, and genotypic clusters were edited manually when necessary. In the present study, this threshold corresponded to SNPs with accurate scoring for at least 95% of individuals, with most successful SNPs scored for more than 99% of individuals analyzed.

# **Genetic structure**

To evaluate the within-population variation, we used the proportion of polymorphic loci (Pl) at the 95% probability level, the unbiased heterozygosity ( $H_e$ ) (Nei 1978), and the allelic richness ( $R_s$ ) calculated from the allele frequencies of all loci analyzed. Allelic richness was determined using the method of El Mousadik and Petit (1996) with a sample size of 6. The fixation indices,  $F_{\rm IS} = 1 - H_{\rm o} / H_{\rm e}$ , for polymorphic loci and their averages over all loci were determined to compare the observed genotype frequencies with expectations based on Hardy-Weinberg equilibrium (Wright 1922; Nei 1977; Nei and Chesser 1983). Deviations from such expectations were analyzed using Fisher's exact test. Coefficients of gene differentiation,  $F_{ST}$ , between populations were calculated to determine how gene diversity was partitioned at each level (Wright 1978). These analyses were conducted using the FSTAT software package version 2.9.4 (Goudet 2003) and GenAlEx version 6.5 (Peakall and Smouse 2012). To determine any population structure and infer the most appropriate number of subpopulations (K) for interpreting the data without prior information about the number of locations at which the populations were sampled, we used the F-model of the Bayesian clustering approach, STRUCTURE, proposed by Pritchard et al. (2000). Ten independent runs with K values ranging from 1 to 10 were performed using 2×10<sup>6</sup> MCMC (Markov Chain Monte Carlo) sampling after a burn-in period of 50,000 iterations. The posterior probability was

Table 1 Lc	ocations fo	or 14 inve	stigated po	pulations	s of C. J	japonica and t	heir current a	ind last glacie	al period envir	onment vari	ables			
Population	Abbrev	Lat	Long	Alt (m)	Z	Annual Mean Temp (°C)	Max Temp of Warmest Month (°C)	Min Temp of Coldest Month (°C)	Annual Precip (mm)	Deepest Snow (cm)	Annual Mean Temp in LGM (°C)	Max Temp of Warmest Month in LGM (°C)	Min Temp of Coldest Month in LGM (°C)	Annual Precip in LGM (mm)
Ajigasawa	AJG	40.6756	140.2053	319	13	9.4	26.5	-5.4	1447	70	8.5	27.3	-8.3	1304
Nibetsu	NBT	39.8061	140.2600	315	15	9.8	27.8	-5.6	1683	37	8.9	28.6	-8.5	1554
lshinomaki	ISN	38.3286	141.4919	195	13	11.3	26.7	-3.2	1225	7	10.4	27.4	-5.6	1099
Donden	DND	38.1397	138.3833	725	14	8.4	25.3	-6.1	1924	66	7.6	26.1	-8.5	1757
Bijodaira	BJD	36.5761	137.4589	628	18	8.7	26.6	-7.7	1669	171	8.0	28.6	-9.7	1520
Ashitaka	AST	35.2322	138.8361	690	6	11.7	27.3	-4.8	2044	37	11.0	29.1	-6.8	1923
Kawazu	KWZ	34.8314	139.0000	664	16	12.6	26.4	-1.4	2327	<del>.                                    </del>	11.8	27.8	-3.4	2213
Ashu	ASH	35.3078	135.7739	802	13	11.2	27.6	-3.5	1971	55	10.5	29.4	-5.6	1839
<u>Oki</u>	OKI	36.2683	133.3292	363	17	12.6	28.2	-1.2	1908	29	11.5	28.8	-4.1	1731
Azouji	ΓZΥ	34.4820	131.9634	1112	ω	8.6	23.9	-6.0	2245	43	7.6	25.1	-8.6	2085
Shingu	SNG	33.8900	135.7100	592	14	13.9	27.9	-0.1	2600	2	13.1	29.1	-2.2	2472
Kochi	KCH	33.5928	134.0958	615	9	12.8	26.4	-0.9	2272	2	11.9	27.2	-3.2	2134
Oninome	NNO	32.6984	131.5182	1170	4	9.4	22.9	-5.3	3227	2	8.4	23.6	-7.7	3076
Yakushima	YKU	30.3035	130.5731	1071	15	13.8	24.5	2.3	3411	0	12.7	25.0	0.3	3321
Abbrev, abbrev	iation; Lat, I	latitude; Lon	ig, longitude;	Alt, altituc	de; Tem	o, temperature; N	Vax, maximum;	Min, minimum;	Precip, precipita	tion.				

then calculated for each value of *K* using the estimated log-likelihood of *K* to identify the optimal value. The optimal value of *K* was the one at which the log likelihood of the data,  $\ln P(X|K)$  (Pritchard *et al.* 2000), or delta *K*, the rate of change of  $\ln P(X|K)$  between successive *K* values (Evanno *et al.* 2005), was maximal. To examine the genetic differentiation between two groups representing the two varieties, *C. japonica* (omote-sugi) and *C. japonica* var. *radicans* (ura-sugi), we performed a hierarchical analysis of molecular variance (AMOVA) (Excoffier *et al.* 1992) in which the significance levels for variance components were tested using permutations.

We performed a phylogeographic analysis using the Neighbor-net method (Bryant and Moulton 2004) implemented in SplitsTree4 (Huson and Bryant 2006) based on the pairwise  $F_{ST}$  between populations to reveal more details of the evolutionary relationship between populations, for example, hybridization between populations.

Methods for detecting candidate loci for divergence: We used a number of approaches to allow us to detect signs of natural selection and to identify, tentatively, the environmental parameters associated with the corresponding selective pressures acting on C. japonica populations. We compared the distributions of the FST values over all loci to their expected distributions under the assumption of neutrality. Beaumont and Nichols (1996) have shown that the distribution of  $F_{ST}$  as a function of heterozygosity in the context of an island model is quite robust, i.e., it is insensitive to variations in factors such as population structure, demographic structure, and mutation level. We therefore used this method to identify markers that deviated from the null hypothesis of neutral evolution. All calculations for identifying potential nonneutral genes in the populations studied were performed using the FDIST2 approach of Beaumont and Nichols (1996) implemented in LOSITAN (Antao et al. 2008) and Arlequin version 3.11 (Excoffier and Lischer 2010). The Arlequin program allowed us to conduct hierarchical analysis using two genetic lines, here the ura-sugi and omote-sugi varieties. In the analysis using FDIST2, we identified the outlier loci using not only all 14 populations but also each type of population, *i.e.*, the urasugi populations and the omote-sugi populations.

An alternative method for identifying candidate loci is BayeScan version 2.1, which uses a Bayesian method to estimate a posterior probability for each locus directly and a reversible-jump MCMC approach to selection (Foll and Gaggiotti 2008). The main advantage of BayeScan is that it estimates population-specific FST coefficients and therefore accommodates differences in demographic history and the extent of genetic drift between populations. This method is an extension of that proposed by Beaumont and Balding (2004), and it is based on a logistic regression model that decomposes genetic variation into population and locus-specific effects. Preliminary tests were conducted using a burn-in of 10,000 iterations, a thinning interval of 50, and a sample size of 10,000 (Foll and Gaggiotti 2008). Four independent runs were performed for each of the two datasets to account for the consistency of the detected outliers. The loci were ranked according to their estimated posterior probability and all loci with a value more than 0.990 were retained as outliers. This corresponds to a log<sub>10</sub> Bayes factor of more than 2, making it possible to be confident in the validity of the model.

*Environmental data and the association with genotypes:* Environmental data regarding the area of origin of each population, including longitude, latitude, altitude, and 19 bioclimatic variables over 50 years (1950–2000), were obtained from WorldClim (Hijmans *et al.* 2005). Past climate data for the interglacial period (LIG) and last glacial maximum based on the MIROC model were obtained from the Paleoclimate Modeling Intercomparison Project Phase II (http://pmip2.lsce.ipsl.fr/).



**Figure 1** Natural distribution of *Cryptomeria japonica* in Japan (shaded areas) (Hayashi 1951) and the locations of the 14 natural populations surveyed in this study. The dotted line indicates the coastline approximately 18,000 years ago. Areas shaded in bold or within thin diagonal lines indicate established refugia (Izu Peninsula, Wakasa Bay, Oki Island, and Yakushima Island) and probable refugia, respectively, at that time (Tsukada 1986).

Some of the climate variables and geography such as longitude, latitude, and altitude are correlated with each other, as shown by Tsumura *et al.* (2012); therefore, principal component analysis (PCA) was used to reduce dimensionality for the climate variables and geography. Principal components (PCs) with eigenvalues greater than 1.0 were used for further analysis and PC scores were used for association analysis. All analyses were performed in R 2.12.1 (R Core Development Team 2010) using the prcomp function in the statistics package (http://cran.r-project.org/).

To calculate correlations between SNP allele frequencies and current and past climate variables, we used the Bayesian linear model method of Coop et al. (2010), which controls for population history by incorporating a covariance matrix of populations and accounts for differences in sample size between populations. Using the full set of SNPs, we estimated a covariance matrix of allele frequencies across populations. This matrix was used as the basis of the null model for the transformed allele frequencies at each SNP to be tested. For each tested SNP, the method generates a Bayes factor as a measure of the support for the alternative model relative to the null model in which the transformed population allele frequency distribution is dependent on population structure alone. The significance threshold for ranked Bayes factors was set to 2.0 and the environmental variables considered were latitude, longitude, and 19 climate variables-current and past based on a model with an interglacial and two LGMs-which were transformed to PC scores as described above. Putative functions for the detected outlier loci were identified by performing BLASTx searches against the NR data in the NCBI database using Blast2GO (Conesa et al. 2005) and the Arabidopsis genome annotation (TAIR10) (http://www.arabidopsis.org/).

Mapping the outlier loci on the linkage map and linkage disequilibrium: The YI pedigree used to determine the positions of the outlier loci of *C. japonica* has previously been used to construct a linkage map in which a total of 1261 markers were assigned to 11

large linkage groups, giving a total map length of 1405.2 cM (Tani *et al.* 2003, Moriguchi *et al.* 2012). The mapped population was genotyped using Illumina's 1536-plex GoldenGate array, as discussed above. All linkage analyses and map estimations were performed using the JoinMap v4.1 software package (Van Ooijen and Voorrips 2001). During map construction, markers were assigned to tentative linkage groups by comparing the linkages formed at logarithm of odds (LOD) thresholds ranging from 3.0 to 9.0, increasing in increments of 1.0. Finally, the markers were ordered at the LOD threshold of 8.0. Map distances were calculated using the Kosambi mapping function (Kosambi 1944).

Coefficients of linkage disequilibrium were calculated as described by Weir (1996) using squared allele-frequency correlations ( $r^2$ ) for pairs of loci on the basis of genotype data from the investigated populations. The differences from equilibrium were verified by chisquared tests with a false discovery rate of 0.01 (Weir 1996). These analyses were performed using the R-package "genetics" (http://cran. r-project.org/web/packages/genetics/).

Assuming a random distribution of markers, if the genome was divided into N intervals, then the number of markers per interval would follow a Poisson distribution. To determine whether outlier loci were randomly distributed, every linkage group was divided into 15-, 20-, 25-, and 50-cM intervals. We used the chi-square test to compare the actual distribution of outlier loci to that expected for a Poisson distribution, as described by Kang *et al.* (2010).

## RESULTS

# **SNP** genotyping

Of the 6144 SNPs, 3934 (64.0%) yielded data that met our quality thresholds according to the GoldenGate genotyping system (Uchiyama *et al.* 2012). The median  $GC_{50}$  score across all usable SNPs was 0.80, with an average call rate of 99.4%. Of those 3934 SNPs, four were monomorphic and were therefore discarded. The remaining

3930 SNPs were used in the subsequent analyses. Only 216 SNPs (5.50%, P < 0.05) differed significantly from Hardy-Weinberg equilibrium after Bonferroni correction.

# **Genetic structure**

The Pl values for the SNPs ranged from 0.705 to 0.945, with an average of 0.898. The  $R_s$  values also varied, from 1.542 to 1.610, with an average of 1.591, and the  $H_e$  values ranged from 0.301 to 0.339, with an average of 0.328 (Table 2). In contrast to findings from previous studies using CAPS and SSR, these parameters do not reveal any clear geographical trends (Tsumura et al. 2007; Takahashi et al. 2005), as reported by Tsumura *et al.* (2012) using 1026 SNPs. The average  $F_{IS}$ value for all but 47 loci was not significantly different from expectations under Hardy-Weinberg equilibrium (P < 0.01). The overall genetic differentiation among populations at the 3930 loci was low  $(F_{\rm ST} = 0.0506)$ . We also conducted an AMOVA to determine the variation within and among groups (the two varieties) and populations, and to test the significance of the among-population variation. In this analysis, the ONN population was included in the ura-sugi variety because the result of STRUCURE analysis and the phylogenetic network clearly indicated that this was where the population belonged. Therefore, there were eight ura-sugi and six omote-sugi populations. The variation among populations was 5.06% and was highly significant (P < 0.001), and the proportions of variance among varieties and among populations within varieties were also significant, i.e., 1.98% and 3.05%, respectively (Table 3). Bayesian clustering of the information from the 3930 loci demonstrated that the models with K = 2 and K = 4 both provided satisfactory explanations of the observed data, based on the delta K and the highest log-likelihood values, respectively (Figure 2). Both of the bar plots for K = 2 and K = 4revealed clear differences between populations of the two varieties with the exception of the ONN population (Figure 2), as found in a previous study (Tsumura et al. 2012). In addition, the bar plot for K = 4 separated the populations within each variety. The ura-sugi populations were divided into two groups, with the first consisting of the most northern population and the second containing the others. The omote-sugi populations were divided into two major groups, namely YKU and the others.

The network tree constructed using Neighbor-net indicated that the two varieties were clearly divided, and that the ONN population on Kyushu Island belonged to the ura-sugi variety, which was closed and introgressive into the neighboring AZJ population (Figure 3). The SNG population of omote-sugi that was geographically closest to the ura-sugi variety populations seemed to subject to introgression from the ura-sugi variety. The branch length of the ONN population and the YKU population suggested long-term isolation or a bottleneck.

# **Outlier** loci

We identified 407 loci lying outside the 99% confidence interval (C.I.), of which 199 were above the C.I., suggesting positive selection using the  $F_{ST}$ -based method (FDIST2) (Supporting Information, Table S1). BLASTx searches identified putative functions or annotations for 178 of these loci (85.6%) (Table S1). The method of considering of the hierarchical structure, *i.e.*, the ura-sugi and omote-sugi varieties, using Arlequin identified 229 loci, of which 131 were above the C.I. (Table S2).

Using the Bayesian method implemented in BayeScan, we identified 79 loci as outliers with a  $log_{10}$  Bayes factor above 1 after correction using a false discovery rate of 0.05. All but eight of these loci were also detected by the  $F_{ST}$ -based method (Table S1, Figure S1). Of these outliers, 42 had  $log_{10}$  Bayes factor values above 2 (Figure 4); 33 of these 42 were also identified as outliers by the  $F_{ST}$ -based method (Table S1). Thirty-five of the 42 loci had  $log_{10}$  Bayes factors above 3, corresponding to a posterior probability of locus effects above 0.999; 27 had a  $log_{10}$  Bayes factor of more than 5, corresponding to a posterior probability of 1.000. BLASTx searches identified putative functions for 36 of the 42 loci with  $log_{10}$  Bayes factors above 2 (85.7%) (Table S1).

Within-variety  $F_{\rm ST}$  analysis identified 221 outlier loci. Only four of the identified outlier loci were detected in both the omote-sugi and ura-sugi populations; these were estSNPg00691, estSNPg02482, SNPg01839, and SNPg03850 (Table S2). Using the Bayesian method, 38 loci were identified in either the omote-sugi or ura-sugi populations, but no common outlier loci were present in both populations. All but eight of the loci identified by Bayscan were also detected in the  $F_{\rm ST}$  analysis.

#### Environment variables and their association with loci

Annual precipitation and mean temperature exhibit a clear trend from north to south in Japan, and the snow depth differs greatly between the Japan Sea side and the Pacific Ocean side (Table 1). Cumulatively, the first three principal components explained 86.16% of the total variance in current environmental conditions (Table S3). Principal component 1 (PC1) was strongly related to the location of populations

Population	Ν	Но	He	F <sub>IS</sub>	PI	Rs	No. of Private Alleles
AJG	13	0.326	0.324	-0.008	0.901	1.583	3
NBT	15	0.324	0.330	0.019	0.922	1.593	2
ISN	13	0.329	0.333	0.014	0.915	1.599	0
DND	14	0.328	0.332	0.012	0.924	1.598	2
BJD	18	0.333	0.335	0.006	0.945	1.603	2
AST	9	0.334	0.332	-0.007	0.884	1.597	0
KWZ	16	0.330	0.333	0.009	0.934	1.599	4
ASH	13	0.331	0.338	0.019	0.940	1.608	2
OKI	17	0.325	0.333	0.024	0.940	1.599	1
AZJ	8	0.339	0.332	-0.023	0.879	1.598	2
SNG	14	0.327	0.335	0.023	0.932	1.603	2
КСН	6	0.325	0.339	0.045	0.858	1.610	2
ONN	4	0.303	0.301	-0.010	0.705	1.542	0
YKU	15	0.299	0.303	0.013	0.898	1.547	7
Mean	12.5	0.325	0.328	0.010	0.898	1.591	2.1

Table 2 Genetic diversity of 14 investigated populations of C. japonica based on 3930 SNPs

n, number of investigated individuals; Ho, the observed number of heterozygosity; He, the expected number of heterozygosity; F<sub>IS</sub>, fixation idenx; PI, the number of polymorphic loci; Rs, allelic richness.

#### Table 3 Results of analysis of molecular variance

Source of Variation	d.f.	Sum of Squares	Variance Components	Percentage of Variation
Among groups	1	3578.128	13.40222	1.98
Among populations within groups	12	14042.649	20.61734	3.08
Within populations	346	222325.545	642.55938	94.94

Two varieties, *C. japonica* and *C. japonica* var. *radicans*, are used as different groups. ISN, AST, KWZ, SNG, KCH, and YKU are included into *C. japonica* (omote-sugi). The others, AJG, NBT, DND, BJD, ASH, OKI, AZJ, and ONN, are included in *C. japonica* var. *radicans* (ura-sugi).

such as latitude, and PC2 and PC3 were explained mainly by temperature and precipitation in the dry season, respectively. The result of the PCA for the interglacial period (LIG) was very similar to that for current environmental conditions (Table S3). However, the results of the PCA for the last glacial maximum (LGM) were related to the temperature in the warmest season (PC2), precipitation in the dry season (PC2), average temperature in the dry season (PC3), and seasonal precipitation (PC3), although PC1 was highly related to latitude and seasonal temperature in the MIROC model.

In total, 38 loci were associated with environmental variables in at least one of the three climate condition scenarios: current, LIG, or LGM (Table 4). Of theses, 18 loci were detected in association with all three climatic conditions. With the exception of five loci, the detected loci in the current scenario were the same as those associated with the LIG, because the climatic conditions were similar. Eleven loci were only associated with the LGM.

All loci found to be related to an environmental variable were also identified as outliers in the Lositan, Arlequin, or BayeScan analyses. BLASTx searches identified putative functions for 37 of the 43 environment-associated loci.

## Linkage mapping of outlier loci

In total, 208 loci were identified as outliers or loci associated with environmental variables for all populations; 101 of these loci were successfully mapped on the *C. japonica* linkage map, and their distribution was investigated (Figure 5) (Moriguchi *et al.* 2012). The numbers of mapped outlier loci in each linkage group varied substantially. For example, linkage groups (LGs) 4 and 5 contained only four loci, whereas LG2 contained 21. The observed distribution of loci deviated significantly from the Poisson distribution for all marker intervals considered (15, 20, 25, and 50 cM) and, therefore, was not random (P < 0.001). Thus, the outlier loci clearly formed clumps associated with specific linkage groups, particularly LG2, LG7, LG10, and LG11 (Figure 5). The outlier distribution was related to the  $F_{ST}$  distribution; regions with high  $F_{ST}$  values had high numbers of outlier loci.



## Linkage disequilibrium and the associated loci

Linkage disequilibrium (LD) was detected for 491,563 (6.37%, q < 0.1) of the 7,720,485 possible combinations of all 3930 loci with the false discovery rate of 0.1. However, the proportion of significant LD among the 208 outlier loci in all 14 populations was very high, 38.852% (q < 0.1) (Table 5). In particular, a significant LD was detected among the loci of the clumped regions, but inter-chromosomal LD was also detected, including for LG2 and LG10, LG10 and LG11 (Figure 6). The relatively higher  $r^2$  values (>0.1) between linkage groups were not frequent, which were gSNP09342 in LG10 and two loci in LG2, gSNP09342 in LG10 and three loci in LG11, gSNP09193 in LG11 and four loci in LG7, and estSNP02482 in LG2 and gSNP06850 in LG10. However, the proportion of significant LD was reduced to less than the half when we calculated the LD in each of the variety populations (ura-sugi and omote-sugi) and also the 13 populations excluding the YKU population (Table 5).

We carefully recorded the allele frequency of each outlier loci in the four clumped regions and found that the genotypic frequency of most of loci is very different between populations of the two varieties (Table 6). The heterozygosity of populations of ura-sugi is lower in the three clumped regions than that of the omote-sugi populations for approximately 60% of the outlier loci. The seven outlier loci associated with LG7 formed a unique group that was specific to the YKU population, three loci of which (SNPg04215, SNPg07948, and SNPg06867) showed lower heterozygosity in all populations compared with the Yakushima population. Two loci (SNPg11760 and SNPg01044) exhibited very low heterozygosity only in the YKU population, and for the other two loci the allele frequency of the YKU population was different from that of the others.

#### DISCUSSION

### Factors determining the genetic structure of C. japonica

The formation of the Japanese archipelago began approximately 17 million years ago, with the earliest incarnation of the Japan Sea

**Figure 2** Genetic relationships among the 14 populations surveyed using STRUCTURE (Pritchard *et al.* 2000) based on 3930 SNPs. The models with K = 2 and K = 4 were optimal based on the delta K value and the highest log-likelihood value, respectively.



**Figure 3** A population network based on pair-wise  $F_{ST}$  values between populations constructed by the Neighbor-net.

forming approximately 7 million years ago (Taira 2001). A *Cryptomeria anglica* fossil dated to the later phase of the Miocene has been found in Japan and is a probable ancestor of *C. japonica*. Moreover, *C. japonica* fossils have been found to date back to the Pleiocene, thus postdating the formation of the Japanese archipelago (Uemura 1981). *C. japonica* has therefore adapted to climatic differences over extended periods of time while shifting its distribution in the face of global climate changes, including several glacial and interglacial periods.

The estimated current potential natural distributions are much wider than the observed natural distribution (Kimura *et al.* in press) (Figure S2), which means that interspecific competition, for example, competition with evergreen tree species in western Japan (Takahara 1998), and historical human disturbance associated with timber extraction strongly influenced the distribution we see today. The estimated natural distribution using the MIROC model suggests small northern refugia during the LGM (Figure S2).

The network tree and the structure results clearly divided the 14 study populations into two variety-based groups, as have previous studies (Tsumura et al. 2007, 2012). The ONN population belongs to the ura-sugi and is close to the AZJ population; however, the long branch-length in the network tree suggests a bottleneck and isolation. In fact, a limited number of individuals survive in the ONN population at relatively higher elevations, the sole natural population on Kyushu Island. We selected the 14 populations investigated to cover the whole natural distribution area, based on geographical location. The ONN population is closer to the Pacific Ocean side of the country, and thus this population was included as an omote-sugi population. Its phenotype, especially for the needles, is intermediate between the ura-sugi and omote-sugi varieties (Nakao et al. 1986). The AST, KWZ, and YKU populations were mostly fixed within the ura-sugi cluster, representing the main refugia during the LGM; however, the ISN, SNG, and KCH populations have some associations with the



**Figure 4** Plot of  $F_{ST}$  values and Bayes factors ( $log_{10}$ ) for 3930 loci identified using the BayeScan outlier test. Dashed lines indicate the  $log_{10}$  of the Bayes factor threshold that provides "decisive" evidence for selection corresponding to a posterior probability of 0.99.

Table 4 Strongly associated loci between the results of principal component analysis of environmental variables

utative Function	anonymous locus	_02 genomic sequence	ess-ripening protein			e pvrophosphorvlase	nal protein 19	natoria lo		main-containing	tion factor			n-induced protein	id responsive elements-	factor 2	n-induced protein	omplex component csl4	atase hydrolase family	1	uced protein-like	_	transcription factor	na storekeeper	tional requilator	notal regulator atoin lianco mmc21 liko	orent ngase minisz i-inke	an-protein synuase	al protein			o biotic stimulus	ndent phosphohydrolase	in-containing protein	onstituent of	ienal double-bond	0	protein 113a	dient regulation 5	transcription factor	ted r2r3 mvb	tion partial	stein	d protein kinase	y subunit beta-2-like		in homolog	(continued)
Βr	Pinus taeda	0_14423_	Abscisic str	3-like	No hit	udp-alucos	50s riboson	50e riboeon		apz en dor	transcript	No hit	No hit	pi starvatio	Abscisic aci	binding f	pi starvatio	Exosome co	Isochorismé	protein	Harpin-indu	No hit	R2R3-MYB	DNA-bindir	transcript		Alaba alito	Alpha-gluc	Hypothetic	No hit	No hit	Response t	Metal-depei	hd domai	structural c	Phenylprop	reductase	Ribosomal	Proton grac	myb family	Tannin-rela:	transcript	pdi-like prc	snf1-relatec	regulator	Proteolysis	DnaJ prote	
Linkage Group			7		10	Ø			1	o		7	2	6	I			ъ			10		11	5				(	71	ŋ	10	2	2		с				I	6			6	10		2	2	
F <sub>ST</sub>	0.1459		0.1482		0.2450	0.1922	0.2338	0.0104	70700	U.2400		0.1574	0.1373	0.1627	0.2397		0.1627	0.1242	0.2224		0.1415	0.2228	0.2556	0.1539		0 2113	0 2 1 0 4	1001000	0.1405	0.2/96	0.2689	0.2365	0.2419		0.1612	0.1657		0.1876	0.1342	0.1561	0.2334		0.1503	0.2250		0.2508	0.1772	
LGM_MIR_PC2					*									*			*	*	*				××	**		*	*	÷			¥	**	¥		*			**	*	*			*			*	*	
LGM_MIR_PC1	*		×		*	××	*	*	*	÷		*	*														*	-								*								*				
LIG_PC3																							*	*		*	*	•	6		* *	¥	* *								*						*	
LIG_PC2					*									*	*		*		*		*	*	**																									
LIG_PC1	×		¥			×	*	*	×			*																																				
Current_PC3																							*	*		*	*		÷ ·	*	*	**	×															
Current_PC2					*									*	*		*	*	*		*	*	**																									
Current_PC1	**		××		*	*	*	*	*	ł		*	*																																			
Locus	gSNP06309	)	gSNP11760		gSNP06850	estSNP00328	estSNP00402	octon 001043		estanruzzug		gSNP03082	gSNP06599	estSNP00454	estSNP01816		estSNP02019	estSNP02622	estSNP03061		gSNP01042	gSNP01385	aSNP04024	estSNP00558		0c+CNIDO11111		estonru 239	estSNPUZ553	gsNPU////	gSNP09342	estSNP02046	estSNP02882		estSNP00015	estSNP00110		estSNP00178	estSNP00338	estSNP00625	estSNP00691		estSNP01232	estSNP01913		estSNP02482	estSNP03062	

Locus	Current_PC1	Current_PC2	Current_PC3	LIG_PC1	LIG_PC2	LIG_PC3	LGM_MIR_PC1	LGM_MIR_PC2	$F_{\rm ST}$	Linkage Group	Putative Function
gSNP01330							*		0.2382	11	Adenosine 3-phospho
gSNP03195							*		0.1558	Ι	btb poz domain-containing
gSNP11860							*		0.1571	4	protein augupo/p-like 3-ketoacyl-synthase
Loci detected b detected by bla:	y Principal compo stx search (E-value	nent analysis (PCA cut off $\leq 1 - 10^{-3}$	) of environmenta 3). The "current" is	I variables we	ere also dete climate data	cted by both between 195	BayeScan (Bayes fa 0 and 2000, "LIG" is	ctor >2) and Lositan the average climate	(P < 0.001) data of last	), and their interglacial	inkage group and putative function were between ${\sim}120,000$ and 140,000 years BP,

and LGM is the average climate data of last glacial maximum at  ${\sim}21,000$  years BP. \*:  ${
m p<}0.01$ , \*\*: ${
m p<}0.001$ 

ura-sugi cluster, probably because they experienced introgression from ura-sugi populations during population expansion after LGM. From fossil pollen data, Takahara (1998) suggested that, in western Japan, especially on Sikoku Island and probably Kyushu Island, the expansion of natural forests of C. japonica along the Pacific Ocean side may have been much slower than along the Japan Sea side because of the rapid expansion of evergreen broadleaf forest during the postglacial period. This may be one reason why the origin of the sole natural population on Kyushu Island, the ONN population, is on the Japan Sea side. The YKU population also has a long branch-length in the network tree, suggesting long-term isolation. The YKU population is the most southerly population and an extensive natural population has been maintained for a long time because it occurs on a small isolated island with steep mountains, offering a preferred habitat for this species (Tsumura and Ohba 1993). It is thought that this island became isolated from Kyushu Island during the LGM. However, the vegetation was severely damaged by the eruption of Kikai caldera approximately 40 km north from Yakushima Island, 7300 years ago, and it is likely that only the vegetation of the southern coastal area survived this event (Geshi 2009). This reduction in population size with the isolation from Kyushu Island may explain the characteristics of the YKU population. The Neighbor-net output suggests some degree of reticulation between the two varieties; this suggestion is also supported by the admixture revealed by the STRUCTURE analysis.

According to fossil pollen data (Tsukada 1983, 1988), the refugial areas of this species during the last glacial period (approximately 18,000 years ago) were along Wakasa Bay to Oki Island (ura-sugi variety), on the Izu Peninsula, Yakushima Island, and (probably) the southern Kii Peninsula and Shikoku Island (omote-sugi variety) (Figure 1). Before the LGM, the two varieties had already diverged (Kimura et al. in press) and the genetic structure developed further as a result of global climate change, for example, during the glacial and interglacial periods. These northern populations are thought to have established approximately 6000 years ago by migration from refugia, based on inferences from fossil pollen data (Tsukada 1986). If so, then the genetic diversity of the most northerly marginal forests was reduced by bottleneck effects. However, a few populations in northern areas have relatively high genetic diversity compared with those of the other northern populations, suggesting that some small populations may have survived in the northern area even during the last glacial period (Takahara 1998). C. japonica trees can sometimes survive severe climate conditions, such as low temperatures and heavy snowfalls, which are common at high elevations. For example, at 2050 m (Taira et al. 1993), most seedlings cannot survive but a few trees are able to regenerate by layering, and this can allow them to persist for a long time (Taira et al. 1997; Moriguchi et al. 2001). Such small populations may have survived in refugia in northern Japan during glacial periods; this suggestion is supported by the structure result for K = 4.

Genetic differentiation between populations was small (average  $F_{ST} = 0.0506$ ), as found in previous studies using SSR and CAPS (Takahashi *et al.* 2005; Tsumura *et al.* 2007). The distribution has shifted because of climate change, and small refugia in marginal areas may have supported founder populations; thereafter, the genetic differentiation could be expected to have been large because of the founder effect. However, the long life-cycle of tree species may have provided sufficient pollen flow between populations, so that the founder effect was limited and the genetic differentiation was suppressed (Austerlitz *et al.* 2000). However, even the limited genetic differentiation is associated with some important loci linked to adaptation to the different

Table 4, continued



**Figure 5** The distribution of markers identified as being potentially involved in local adaptation across the genome of *Cryptomeria japonica*. The figure shows plots of marker position (based on 1255 mapped SNPs or the other DNA markers; Moriguchi *et al.* 2012; Y. Moriguchi, T. Ujino-Ihara, K. Uchiyama, N. Futamura, S. Ueno, A. Matsumoto, and Y. Tsumura unpublished data) in cumulative centimorgans (cM) vs. (A) Bayes factor ( $log_{10}$ ), as determined by BayeScan analysis, and (B) *F*-statistic value, *F*<sub>ST</sub>. Vertical dashed lines demarcate the 11 linkage groups. The four gray zones indicate candidate regions associated with local adaptation.

environments experienced during the winter on the Japan Sea side and Pacific Ocean side of the country.

# How did natural selection influence to the genetic structure?

The natural C. japonica forest has a wide distribution, from 30°N, 130°E to 40°N, 140°E (Figure 1). There is heavy snowfall on the Japan Sea side of the country in the winter, but the Pacific Ocean side is quite dry in the winter, and the annual precipitation is much higher in the southern part of Japan than elsewhere in the country. Thus, there are significant differences in the environmental factors affecting the different regions examined in this work, especially in terms of annual precipitation and the maximum depth of fallen snow (Table 1). There is a cline of environmental variables including precipitation, temperature, and snowfall across the Japanese archipelago from the northeast to southwest. The key environmental difference between the Japan Sea and Pacific Ocean sides of Japan is snowfall. This difference in environments exerts an important selection pressure. The results of our PCA indicate that the current natural distribution is positively linked to winter precipitation; the natural distribution during the LGM was also positively linked to the maximum and mean temperatures during the warmest period of the year. C. japonica has adapted well to temperate climates and moist regions. Normally, it can grow in areas

with minimum temperatures during the coldest month of approximately  $-7^{\circ}$  to 2°, maximum temperatures during the warmest month of 22° to 28°, and effective annual precipitation exceeding 1500 mm. The current distribution seems to be strongly restricted by precipitation, especially in winter, which is the coldest and driest season. The LGM distribution was probably limited by maximum temperature and also precipitation. The results of the species distribution model support this conclusion (Figure S2).

Two-hundred eight outlier loci were detected using two different methods—Lositan and Bayscan (Table S1). Of these, 43 loci were also associated with environmental variables (Table 4). The detected outlier loci that were associated with environmental variables represent only 1.1% of all analyzed loci; this is a similar proportion to that reported in previous studies (2.7% and 1.4%) (Tsumura *et al.* 2007, 2012). Eckert *et al.* (2010) identified 22 candidate loci (BF  $log_{10} > 2.0$ ) associated with climatic variables from a total of 1730 loci in 682 loblolly pine tree samples covering the full range of the species. Prunier *et al.* (2011) also identified 10 candidate loci above the 99% C.I. in black spruce. The proportion of detected loci (1.3% and 1.7%, respectively) associated with the climatic variables was very similar to that found in the current study. The detected outlier loci among the current, last interglacial (LIG), and LGM climate variables were very similar to each other, especially the first two because of their similar

Table 5 The proportion	n of significant LD	among the detected	outlier loci and all loci
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							Proportion of	Significant LD
	Investigated Population	n	loci	Average of $r^2$	SD	r² median	P < 0.01	q < 0.1*
Outlier loci	All 14 populations	172	208	0.01729	0.04024	0.00786	0.27397	0.38852
	13 populations excluding Yakushima	158	205	0.01498	0.03920	0.00614	0.21019	0.29139
	Ura-sugi, 8 populations	101	202	0.01255	0.03895	0.00482	0.07950	0.06606
	Omote-sugi, 6 populations	71	208	0.02125	0.04449	0.00891	0.11994	0.13643
	Omote-sugi, 5 populations excluding Yakushima	54	205	0.01936	0.04164	0.00848	0.06432	0.04103
All loci	All 14 populations for all loci	172	3926	0.00646	0.01206	0.00290	0.07964	0.06367
	13 populations excluding Yakushima for all loci	158	3915	0.00684	0.01254	0.00308	0.07598	0.05733
	Ura-sugi, 8 populations for all loci	101	3875	0.01036	0.01668	0.00475	0.07195	0.05004
	Omote-sugi, 6 populations for all loci	71	3893	0.01493	0.02221	0.00702	0.07337	0.05136
	Omote-sugi, 5 populations excluding Yakushima for all loci	54	3868	0.01809	0.02628	0.00856	0.06933	0.04418

\*False discovery rate (FDR 0.1).

Table 6 The outlier loci in	four clumped regions o	of linkage map, their F <sub>s</sub>	T, the heterozygosities,	and the allele frequencies

				He	eterozygosity(H	e)	Allele	Frequency of A	Allele
Linkage Group	*: centiMorgan	Locus	F <sub>ST</sub>	Ura-sugi Populations	Omote-sugi Populations	Yakushima Population	Ura-sugi Populations	Omote-sugi Populations	Yakushima Population
2	55.46	estSNP00091	0.1977	0.423	0.116	0.000	0.696	0.938	1.000
2	57.68	estSNP02482	0.2508	0.496	0.226	0.391	0.544	0.870	0.733
2	57.89	gSNP03082	0.1574	0.333	0.499	0.278	0.789	0.479	0.167
2	57.93	estSNP02390	0.1212	0.415	0.491	0.444	0.706	0.432	0.333
2	58.16	estSNP02046	0.2365	0.477	0.372	0.464	0.392	0.753	0.367
2	58.17	estSNP00558	0.1539	0.360	0.498	0.124	0.765	0.534	0.933
2	58.17	estSNP02882	0.2419	0.485	0.360	0.464	0.413	0.764	0.367
2	58.19	estSNP03062	0.1772	0.291	0.499	0.320	0.176	0.479	0.200
2	58.21	gSNP04589	0.1700	0.271	0.484	0.180	0.838	0.589	0.900
2	58.37	estSNP00649	0.1312	0.444	0.421	0.124	0.667	0.699	0.933
2	58.45	estSNP02061	0.1201	0.488	0.494	0.491	0.578	0.445	0.567
2	58.73	estSNP03044	0.1371	0.494	0.476	0.420	0.446	0.390	0.300
2	60.19	gSNP05191	0.1152	0.200	0.040	0.000	0.887	0.979	1.000
2	61.54	gSNP05463	0.1561	0.447	0.482	0.420	0.663	0.404	0.300
7	115.14	gSNP04215	0.5401	0.000	0.256	0.464	1.000	0.849	0.367
7	115.82	gSNP07948	0.2599	0.102	0.293	0.464	0.054	0.178	0.633
7	117.21	gSNP06867	0.2414	0.000	0.104	0.391	0.000	0.055	0.267
7	118.14	gSNP11760	0.1482	0.500	0.489	0.000	0.500	0.425	0.000
7	118.63	gSNP01044	0.1306	0.486	0.357	0.124	0.583	0.767	0.933
7	118.91	estSNP02454	0.1562	0.315	0.459	0.320	0.196	0.356	0.800
7	119.03	estSNP00146	0.1988	0.192	0.426	0.420	0.892	0.692	0.300
10	130.53	gSNP06850	0.2450	0.499	0.066	0.000	0.480	0.034	0.000
10	130.53	gSNP09342	0.2689	0.029	0.455	0.064	0.985	0.651	0.967
10	130.56	gSNP01042	0.1415	0.215	0.492	0.444	0.123	0.438	0.333
10	131.57	estSNP01913	0.2250	0.434	0.327	0.064	0.319	0.795	0.967
10	131.58	gSNP01835	0.1143	0.486	0.437	0.064	0.583	0.322	0.033
10	132.07	estSNP01295	0.1345	0.497	0.437	0.444	0.539	0.678	0.333
11	54.11	estSNP01481	0.1330	0.389	0.495	0.491	0.265	0.548	0.433
11	54.38	gSNP07270	0.1297	0.075	0.398	0.500	0.961	0.726	0.500
11	54.38	gSNP01805	0.1663	0.259	0.500	0.491	0.153	0.492	0.433
11	54.42		0.3793	0.000	0.151	0.480	0.000	0.082	0.400
11	54.77	gSNP02822	0.1725	0.044	0.324	0.000	0.977	0.797	1.000
11	54.77	gSNP04480	0.1632	0.038	0.275	0.000	0.020	0.164	0.000
11	55.09	gSNP01330	0.2382	0.048	0.378	0.358	0.025	0.253	0.233

climatic conditions. The 18 outlier loci found to be associated with conditions during the LGM may be linked to low temperature and/or limited precipitation. In fact, five of these genes are associated with stress responses. The functions of 41 of these 208 outlier loci are associated with stress responses, and nine of the 34 outlier loci in the four clumped regions of the genome are also associated with stress responses. Of these, LG2 includes five stress response genes. However, the putative functions of these outlier loci identified by the Blastx search could not be explained by the adaptive mechanisms. This indicates that these regions might be experiencing selection. However, it is important to account for hitchhiking effects properly when making suggestions of this kind, because it is possible that the genes identified may simply be closely linked to the true adaptive gene or genome region.

The mapping result clearly showed the clumped distribution of the outlier loci on the linkage map. We found four clumped regions in this study (Table 6), of which LG2 and LG11 had already been detected in a previous study (Tsumura *et al.* 2012). The four regions detected ranged from 1 cM (LG11) to 6 cM (LG2) on the genetic map, and 6 to 14 outlier loci were mapped in these regions. Their  $F_{ST}$  was more than 0.11 (average 0.19), although the average of all 3930 loci was low ( $F_{ST} = 0.0506$ ). These high  $F_{ST}$  values strongly suggest a hitch-hiking effect. These regions could be considered to represent a "genomic

island of divergence" (Nosil et al. 2009). The detected four regions might be merely controlled by four adaptive loci, and thus their surrounding loci might be detected by the hitchhiking effect of the true adaptive locus. Scotti-Saintagne et al. (2004) also identified some important regions of genomes associated with adaptation using both outlier detection and QTL mapping methods. The proportion of significant LD among the 208 outlier loci was quite high (27.40%, P <0.01) because not only the four regions but also the other outlier loci are sometimes tightly linked to each other. Such important genes sometimes comprise clusters on a genome, for example, defense-related genes (Boyko et al. 2002), which are important complex traits (Yamamoto et al. 2009; Messmer et al. 2009). The LD in conifers is thought to decay rapidly for several thousand bps (Neale and Savolainen 2004), but most data have been collected from coding genes. A recent study interestingly revealed LD in a noncoding region in C. japonica that does not decay at least for several hundred kbp (Moritsuka et al. 2012). Generally, the LD is thought to be highly structured into discrete blocks of sequences separated by hot spots of recombination (Goldstein 2001; Kim et al. 2007); therefore, the discovery of a noncoding LD may not be surprising. In this study, we investigated SNPs within coding regions; therefore, the detected LD between the outlier loci may provide evidence of selection. The proportion of significant LD among the outlier loci in all 14 of our C. japonica populations was quite high, but the significant LD in populations for each variety was low (Table 5), suggesting that selection between two varieties might have occurred. Interchromosomal LD between two varieties is also strong evidence for selection affecting them differently. The average  $r^2$  value of the outlier loci is higher than that of all loci (Figure 6, Table 5). The Yakushima (YKU) population has an effect on this high LD because of the relatively long time it has been isolated from the main population. The higher average  $r^2$  value between populations of omote-sugi is related their distribution: the populations are scattered and relatively small compared with those of ura-sugi. The average  $r^2$  value for the outlier loci of omote-sugi populations is higher than that for all 14 populations; however, the proportion of significant LD within the omote-sugi variety is low, which means some specific combinations of loci between omote-sugi populations have quite high  $r^2$  values. The average  $r^2$  value of the outlier loci is not particularly high, but the proportion of significant LD is quite high compared with that of all loci, which means that most of these outlier loci could have been detected because of the hitchhiking effect. The relatively higher  $r^2$ values (>0.1) between linkage groups only applied to 14 combinations of loci. The allele frequencies of these loci between populations of two varieties were significantly different; these were gSNP09342 in LG10 and two loci (estSNP03062 and gSNP04589) in LG2 and gSNP09342 in LG10 and three loci (gSNP01805, gSNP02822, and gSNP04480) in LG11. In addition, there were significant differences between gSNP09193 in LG11 and four loci (gSNP04215, gSNP07948, gSNP06867, and estSNP00146) in LG7; in this case, the allele frequencies were distinctive in that only the YKU population was highly heterozygote. Therefore, the LDs between these interchromosomal loci were relatively higher; however, it is still unclear whether such interchromosomal LD is created by selection or chance. Interchromosomal LD was detected in cultivated tomato, but this LD was caused by artificial selection (Robbins et al. 2011). Evolutionary mechanisms, such as co-adapted complexes of genes, might affect such LD (Dobzhansky 1970). If the detected outlier loci include many rare variants (Dickson et al. 2010), then the possibility of a falsely significant LD is increased. However, only 12.98% of outlier SNPs have MAF (minor allele frequency) < 0.1 compared with 16.84% of all SNPs with MAF < 0.1. An elevated MAF distribution in the outlier SNPs suggests these might be older SNPs rather than recently selected alleles. Identical genes have been mapped on a linkage map as different loci due to

genotyping errors, and thus we carefully checked the putative function of each mapped locus by a Blastx search. In fact, the loci, gSNP01044 and estSNP02454, were tightly linked on LG7 and are probably splicing variants of the same coding region with the same function based on the sequence data of BAC clones including the SNP; however, the other tightly linked loci did not have identical functions, which means that the mapped loci associated with the four regions are independent genes and specific adaptive haplotypes that could be maintained in each variety.

The LG7 region is characteristic of the YKU population and, therefore, the region may be the result of adaptation to a specific environment, such as the oligotrophic granite soil or the abundant precipitation (>4000 mm). The other three regions—LG2, LG10, and LG11—are related to each other. In particular, the four loci, gSNP06850, gSNP09342, gSNP01042, and estSNP01913, on LG10 have significant interchromosomal LD with loci of LG2 and LG11, for which the  $r^2$  values are relatively high. These three LD regions may be associated with differences in adaptation between the two varieties.

The selective divergence associated with the clumped regions could be older, with the selective advantage of local alleles preventing establishment of alleles entering populations by gene flow (Maynard Smith and Haigh 1974; Oleksyk *et al.* 2010). Because the heterozygosity in populations of ura-sugi is lower than that of omote-sugi populations, especially for the clumped region of LG11, most of the ura-sugi loci are fixed homozygotes (Table 6). The four regions probably contain important genes associated with adaptation to different environments and/or morphological traits that differ between the two varieties.

One adaptive mechanism of such a genome region is thought to be associated with genome inversion, because inversions can suppress recombination and thus protect adaptive alleles from gene flow (Kirkpatrick and Barton 2006; Machado *et al.* 2007; Fang *et al.* 2012). An inversion that captures two or more alleles that are adapted to the local environmental conditions has a selective advantage that can cause it to spread (Kirkpatrick 2010). The yellow monkeyflower provides a good example of local adaptation occurring in two distinct ecotypes. The inversion polymorphism has been found to affect adaptive flowering time divergence and other morphological traits in all replicated crosses between four pairs of annual and perennial populations in *Mimulus guttatus* (Lowry and Willis 2010). In our study, the





basic linkage map was constructed for the family of parent trees belonging to the omote-sugi variety (Moriguchi *et al.* 2012). We are currently unable to determine whether the inversion polymorphism is associated with adaptation in *C. japonica*; therefore, it is better to use the ura-sugi variety parents to investigate the inversion. If the inversion is detected, then we will prepare the  $F_2$  populations between the parents of the omote-sugi and ura-sugi varieties for reciprocal transplanting to examine the association between adaptive traits and the inversion polymorphism.

The detected outlier loci of the four regions with high LD are strong candidates for involvement in local adaptation. It is necessary to confirm whether these regions include genuine adaptive genes, comparing the sequences of these genes and surrounding regions using a BAC library to investigate the flanking regions. The identification of outlier loci with high significance levels is essential for conservation purposes and will be required for future work regarding molecular breeding.

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#### LITERATURE CITED

- Antao, T., A. Lopes, R. Lopes, A. Beja-Pereira, and G. Luikart,
   2008 LOSITAN: a workbench to detect molecular adaptation based on a Fst-outlier method. BMC Bioinformatics 9: 323.
- Austerlitz, F., S. Mariette, N. Machon, P. H. Gouyon, and B. Godelle, 2000 Effects of colonization processes on genetic diversity: differences between annual plants and tree species. Genetics 154: 1309–1321.
- Beaumont, M. A., and R. A. Nichols, 1996 Evaluating loci for use in the genetic analysis of population structure. Proc. Biol. Sci. 263: 1619–1626.
- Beaumont, M. A., and D. J. Balding, 2004 Identifying adaptive genetic divergence among populations from genome scans. Mol. Ecol. 13: 969–980.
- Bogs, J., F. W. Jaffé, A. M. Takos, A. R. Walker, and S. P. Robinson, 2007 The grapevine transcription factor VvMYBPA1 regulates proanthocyanidin synthesis during fruit development. Plant Physiol. 143: 1347–1361.
- Bryant, D., and V. Moulton, 2004 Neighbor-net: an agglomerative method for the construction of phylogenetic networks. Mol. Biol. Evol. 21: 255–265.
- Conesa, A., S. Götz, J. M. García-Gómez, J. Terol, M. Talón *et al.*,
  2005 Blast2GO: a universal tool for annotation, visualization and analysis infunctional genomics research. Bioinformatics 21: 3674.
- Coop, G., D. Witonsky, A. Di Rienzo, and J. K. Pritchard, 2010 Using environmental correlations to identify loci underlying local adaptation. Genetics 185: 1411–1423.
- Dickson, S. P., K. Wang, I. Krantz, H. Hakonarson, and D. B. Goldstein, 2010 Rare variants create synthetic genome-wide associations. PLoS Biol. 8: e1000294.
- Dobzhansky, T., 1970 Genetics of the Evolutionary Process. Columbia University Press, New York.
- Eckert, A. J., A. D. Bower, S. C. González-Martínez, J. L. Wegrzyn, G. Coop et al., 2010 Back to nature: ecological genomics of loblolly pine (*Pinus taeda*, Pinaceae). Mol. Ecol. 19: 3789–3805.
- El Mousadik, A., and R. J. Petit, 1996 High level of genetic differentiation for allelic richness among populations of the argan tree [*Argania spinosa* (L.) Skeels] endemic to Morocco. Theor. Appl. Genet. 92: 832–839.
- Endler, J. A., 1986 Natural Selection in the Wild. Princeton Univ. Press, Princeton, NJ.
- Evanno, G., S. Regnaut, and J. Goudet, 2005 Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. Mol. Ecol. 14: 2611–2620.
- Excoffier, L., and H. E. L. Lischer, 2010 Arlequin suite ver 3.5: A new series of programs to perform population genetics

analyses under Linux and Windows. Mol. Ecol. Resources 10: 564–567.

- Excoffier, L., P. E. Smouse, and J. M. Quattro, 1992 Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. Genetics 131: 479–491.
- Fang, Z., T. Pyhäjärvi, A. L. Weber, R. K. Dawe, J. C. Glaubitz *et al.*, 2012 Megabase-scale inversion polymorphism in the wild ancestor of maize. Genetics 191: 883–894.
- Foll, M., and O. Gaggiotti, 2008 A genome-scan method to identify selected loci appropriate for both dominant and codominant markers: a Bayesian perspective. Genetics 180: 977–995.
- Fournier-Level, A., A. Korte, M. D. Cooper, M. Nordborg, J. Schmitt et al., 2011 A map of local adaptation in Arabidopsis thaliana. Science 334: 86–89.
- Geshi, N., 2009 Distribution and flow mechanisms of the 7.3 ka Koya pyroclastic flow deposits covering Yakushima Island, Kagoshima Prefecture. J. Geog. 118: 1254–1260.
- Goldstein, D. B., 2001 Islands of linkage disequilibrium. Nat. Genet. 29: 109-111.
- González-Martínez, S. C., K. V. Krutovsky, and D. B. Neale, 2006 Forest tree population genomics and adaptive evolution. New Phytol. 170: 227–238.
- Goudet, J., 2003 Fstat (ver. 2.9.4), a program to estimate and test population genetics parameters. Available at: http://www2.unil.ch/popgen/softwares/ fstat.htm
- Hayashi, Y., 1951 The natural distribution of important trees, indigenous to Japan. Conifers report I, Bull. Gov. For. Exp. Station, No. 48: 1–240.
- Hijmans, R. J., S. E. Cameron, J. L. Parra, P. G. Jones, and A. Jarvis, 2005 Very high resolution interpolated climate surfaces for global land areas. Int. J. Climatol. 25: 1965–1978.
- Holliday, J. A., K. Ritland, and S. N. Aitken, 2010 Widespread, ecologically relevant genetic markers developed from association mapping of climaterelated traits in Sitka spruce (*Picea sitchensis*). New Phytol. 188: 501–514.
- Holt, R. D., and R. Gomulkiewicz, 1997 How does immigration influence local adaptation? A reexamination of a familiar paradigm. Am. Nat. 149: 563–572.
- Howard, W. R., 1997 A warm future in the past. Nature 388: 418-419.
- Huson, D. H., and D. Bryant, 2006 Application of phylogenetic networks in evolutionary studies. Mol. Biol. Evol. 23: 254–267.
- Kang, B. Y., I. K. Mann, J. E. Major, and O. P. Rajora, 2010 Near-saturated and complete genetic linkage map of black spruce (*Picea mariana*). BMC Genomics 11: 515.
- Kim, S., V. Plagnol, T. T. Hu, C. Toomajian, R. M. Clark *et al.*,
   2007 Recombination and linkage disequilibrium in *Arabidopsis thaliana*. Nat. Genet. 39: 1151–1155.
- Kimura, M., D. Kabeya, T. Saito, Y. Moriguchi, K. Uchiyama *et al.*,
   2013 Effects of genetic and environmental factors on clonal reproduction in old-growth natural populations of *Cryptomeria japonica*. For. Ecol. Manage. 304: 10–19.
- Kimura, M., K. Uchiyama, K. Nakao, Y. Moriguchi, L. S. Jose-Maldia *et al.*, in press Evidence for cryptic northern refugia in the last glacial period of *Cryptomeria japonica*. Ann. Bot. (Lond.) (in press).
- Kirkpatrick, M., 2010 How and why chromosome inversions evolve. PLoS Biol. 8: e1000501.
- Kirkpatrick, M., and N. Barton, 2006 Chromosome inversions, local adaptation and speciation. Genetics 173: 419–434.
- Kosambi, D. D., 1944 The estimation of map distances from recombination values. Ann. Eugen. 12: 172–175.
- Linhart, Y. B., and M. C. Grant, 1996 Evolutionary significance of local genetic differentiation in plants. Annu. Rev. Ecol. Syst. 27: 237–277.
- Lowry, D. B., and J. H. Willis, 2010 A widespread chromosomal inversion polymorphism contributes to a major life-history transition, local adaptation, and reproductive isolation. PLoS Biol. 8: e1000500.
- Machado, C. A., T. S. Haselkorn, and M. A. F. Noor, 2007 Evaluation of the genomic extent of effects of fixed inversion differences on intraspecific variation and interspecific gene flow in *Drosophila pseudoobscura* and *D. persimilis*. Genetics 175: 1289–1306.
- Maynard Smith, J., and J. Haigh, 1974 The hitchhiking effect of a favorable gene. Genet. Res. 23: 23–35.
- Messmer, R., Y. Fracheboud, M. Bänziger, M. Vargas, P. Stamp *et al.*, 2009 Drought stress and tropical maize: QTL-by-environment

interactions and stability of QTLs across environments for yield components and secondary traits. Theor. Appl. Genet. 119: 913–930.

Moriguchi, Y., A. Matsumoto, M. Saito, Y. Tsumura, and H. Taira, 2001 DNA analysis of clonal structure of an old growth, isolated forest of *Cryptomeria japonica* D. Don in a snowy region. Can. J. For. 31: 377–383.

Moriguchi, Y., T. Ujino-Ihara, K. Uchiyama, N. Futamura, M. Saito *et al.*, 2012 The construction of a high-density linkage map for identifying SNP markers that are tightly linked to a nuclear-recessive major gene for male sterility in *Cryptomeria japonica* D. Don. BMC Genomics 13: 95.

Moritsuka, E., Y. Hisataka, M. Tamura, K. Uchiyama, A. Watanabe *et al.*, 2012 Extended linkage disequilibrium in non-coding regions in a conifer, *Cryptomeria japonica*. Genetics 190: 1145–1148.

Murai, S., 1947 Major forestry tree species in the Tohoku region and their varietal problems. In: Kokudo Saiken Zourin Gijutsu Kouenshu, Aomoririnyukai, (eds.) pp.131–151 (in Japanese).

Nakao, T., Y. Kuroki, N. Hosoyamada, and S. Toyama, 1986 Natural cryptomeria forest in Kyushu Island—Oninome Cryptomeria forest in Mt. Ohkueyama. Jpn. J. For. Env. 28: 1–10 (in Japanese).

Namroud, M. C., J. Beaulieu, N. Juge, J. Laroche, and J. Bousquet, 2008 Scanning the genome for gene single nucleotide polymorphisms involved in adaptive population differentiation in white spruce. Mol. Ecol. 17: 3599–3616.

Neale, D. B., and O. Savolainen, 2004 Association genetics of complex traits in conifers. Trends Plant Sci. 9: 325–330.

Neale, D. B., and P. K. Ingvarsson, 2008 Population, quantitative and comparative genomics of adaptation in forest trees. Curr. Opin. Plant Biol. 11: 1–7.

Neale, D. B., and A. Kremer, 2011 Forest tree genomics: growing resources and applications. Nat. Rev. Genet. 12: 111–122.

Nei, M., 1977 F-statistics and analysis of gene diversity in subdivided populations. Ann. Hum. Genet. 41: 225–233.

Nei, M., 1978 Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics 89: 583–590.

Nei, M., and R. K. Chesser, 1983 Estimation of fixation indices and gene diversities. Ann. Hum. Genet. 47: 253–259.

Nosil, P., D. J. Funk, and D. Ortiz-Barrientos, 2009 Divergent selection and heterogeneous genomic divergence. Mol. Ecol. 18: 375–402.

Ohba, K., 1993 Clonal forestry with sugi (*Cryptomeria japonica*), pp. 66–90, in *Clonal Forestry II: Conservation and Application*, edited by M. R. Ahuja, W. J. Libby. Springer-Verlag, Berlin.

Oleksyk, T. K., M. W. Smith, and S. J. O'Brien, 2010 Genome-wide scans for footprints of natural selection. Phil. Trans. R. Soc. B 365: 185-205.

Peakall, R., and P. E. Smouse, 2012 GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. Bioinformatics 28: 2537–2539.

Pritchard, J. K., M. Stephens, and P. Donnelly, 2000 Inference of population structure using multilocus genotype data. Genetics 155: 945–959.

Prunier, J., J. Laroche, J. Beaulieu, and J. Bousquet, 2011 Scanning the genome for gene SNPs related to climate adaptation and estimating selection at the molecular level in boreal black spruce. Mol. Ecol. 20: 1702–1716.

R Core Team 2010 R: A Language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at: www.R-project.org

Robbins, M. D., S.-C. Sim, W. Yang, A. V. Deynze, E. Knaap *et al.*, 2011 Mapping and linkage disequilibrium analysis with a genome-wide collection of SNPs that detect polymorphism in cultivated tomato. J. Exp. Bot. 62: 1831–1845.

Savolainen, O., T. Pyhäjärvi, and T. Knürr, 2007 Gene flow and local adaptation in forest trees. Annu. Rev. Ecol. Evol. Syst. 38: 595–619.

Scotti-Saintagne, C., S. Mariette, I. Porth, P. G. Goicoechea, T. Barreneche et al., 2004 Genome scanning for interspecific differentiation between two closely related oak species *Quercus robur* L. & *Q. petraea* (Matt.) Liebl. Genetics 168: 1615–1626.

Slatkin, M., 2008 Linkage disequilibrium - understanding the evolutionary past and mapping the medical future. Nat. Rev. Genet. 9: 477–485.

Taira, A., 2001 Tectonic evolution of the Japanese island arc system. Annu. Rev. Earth Planet. Sci. 29: 109–134.

Taira, H., Y. Tsumura, and K. Ohba, 1993 Growing conditions and allozyme analysis of a sugi (*Cryptomeria japonica*) forest at 2,050 meters above sea level on Mount Nekomata. J. Jpn. For. Soc. 75: 541–545. Taira, H., Y. Tsumura, N. Tomaru, and K. Ohba, 1997 Regeneration system and genetic diversity of *Cryptomeria japonica* growing at different altitudes. Can. J. For. Res. 27: 447–452.

Takahara, H., 1998 Distribution history of *Cryptomeria* forest, pp. 207–223 in *Vegetation History of the Japanese Archipelago*, edited by Y. Yasuda, and N. Miyoushi. Asakura-Shoten, Tokyo (in Japanese).

Takahashi, T., N. Tani, H. Taira, and Y. Tsumura, 2005 Microsatellite markers reveal high allelic variation in natural populations of *Cryptomeria japonica* near refugial areas of the last glacial period. J. Plant Res. 118: 83–90.

Tani, N., T. Takahashi, H. Iwata, M. Yuzuru, T. Ujino-Ihara et al., 2003 A consensus linkage map for sugi (*Cryptomeria japonica*) from two pedigrees, based on microsatellites and expressed sequence taqs. Genetics 165: 1551–1568.

Tsukada, M., 1983 Vegetation and climate during the last glacial maximum in Japan. Quat. Res. 19: 212–235.

Tsukada, M., 1986 Altitudinal and latitudinal migration of *Cryptomeria japonica* for the past 20,000 years in Japan. Quat. Res. 26: 135–152.

Tsukada, M., 1988 Japan, pp. 458–518 in *Handbook of Vegetation Science*. *Vol. 7, Vegetation History*, edited by B. Huntley, T. Webb III. Kluwer, the Netherlands.

Tsumura, Y., and K. Ohba, 1993 Genetic structure of geographical marginal populations of *Cryptomeria japonica*. Can. J. For. Res. 23: 859–863.

Tsumura, Y., T. Kado, T. Takahashi, N. Tani, T. Ujino-Ihara *et al.*, 2007 Genome-scan to detect genetic structure and adaptive genes of natural populations of *Cryptomeria japonica*. Genetics 176: 2393–2403.

Tsumura, Y., K. Uchiyama, Y. Moriguchi, S. Ueno, and T. Ihara-Ujino, 2012 Genome scanning for detecting adaptive genes along environmental gradients in the Japanese conifer, *Cryptomeria japonica*. Heredity 109: 346–360.

Uchiyama, K., T. Ujino-Ihara, S. Ueno, Y. Taguchi, N. Futamura *et al.*, 2012 Single nucleotide polymorphisms in *Cryptomeria japonica*: their discovery and validation for genome mapping and diversity studies. Tree Genet. Genomes 8: 1213–1222.

Uchiyama, K., N. Miyamoto, M. Takahashi, A. Watanabe, and Y. Tsumura, 2014 Population genetic structure and the effect of historical human activity on the genetic variability of *Cryptomeria japonica* core collection in Japan. Tree Genet. Genomes 10: 1257–1270.

Uemura, K., 1981 Ancestor and change of distribution in *Cryptomeria japonica*. Iden 35: 74–79 (in Japanese).

Ueno, S., Y. Moriguchi, K. Uchiyama, T. Ujino-Ihara, N. Futamura, and T. Sakurai, K. Shinohara, and Y. Tsumura, 2012 A second generation framework for the analysis of microsatellites in expressed sequence tags and the development of EST-SSR markers for a conifer, *Cryptomeria japonica*. BMC Genomics 13: 136.

Van Ooijen, J. W., and R. E. Voorrips, 2001 JoinMap version 3.0, software for the calculation of genetic linkage maps, Plant Research International, Wageningen, The Netherlands.

Vasemägi, A., and C. R. Primmer, 2005 Challenges for identifying functionally important genetic variation: the promise of combining complementary research strategies. Mol. Ecol. 14: 3623–3642.

 Weir, B. S., 1996 Genetic data analysis II, Sinauer Associates, Sunderland, MA.
 Wright, S., 1922 Coefficients of inbreeding and relationship. Am. Nat. 56: 330–338

Wright, S., 1978 Evolution and the Genetics of Populations. Variability Within and Among Natural Populations, Vol. 4. The University of Chicago Press, Chicago.

Yamamoto, T., J. Yonemaru, and M. Yano, 2009 Towards the understanding of complex traits in rice: substantially or superficially? DNA Res. 16: 141–154.

Yamazaki, T., 1995 Cryptomeriaceae. pp. 264 in Flora of Japan. Volume I, Pteridophyta and Gymnospermae, edited by K. Iwatsuki, T. Yamazaki, D. E. Boufford, H. Ohba. Kodansha, Tokyo.

Yasue, M., K. Ogiyama, S. Suto, H. Tsukahara, F. Miyahara *et al.*, 1987 Geographical differentiation of natural *Cryptomeria* stands analyzed by diterpene hydrocarbon constituents of individual trees. J. Jpn. For. Soc. 69: 152–156.

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