



Cardiac Fibroblasts and Cardiac Fibrosis: Precise Role of Exosomes

Prabhat Ranjan¹, Rajesh Kumari¹ and Suresh Kumar Verma^{1,2*}

¹ Division of Cardiovascular Disease, The University of Alabama at Birmingham, Birmingham, AL, United States, ² Department of Biomedical Engineering, The University of Alabama at Birmingham, Birmingham, AL, United States

OPEN ACCESS

Edited by:

Yaoliang Tang, Augusta University, United States

Reviewed by:

Kishore K. Wary, University of Illinois at Chicago, United States Mohsin Khan, Temple University, United States

> *Correspondence: Suresh Kumar Verma sverma@uabmc.edu

Specialty section:

This article was submitted to Stem Cell Research, a section of the journal Frontiers in Cell and Developmental Biology

Received: 25 September 2019 Accepted: 20 November 2019 Published: 04 December 2019

Citation:

Ranjan P, Kumari R and Verma SK (2019) Cardiac Fibroblasts and Cardiac Fibrosis: Precise Role of Exosomes. Front. Cell Dev. Biol. 7:318. doi: 10.3389/fcell.2019.00318 Exosomes are a group of extracellular microvesicles that deliver biologically active RNAs, proteins, lipids and other signaling molecules to recipient cells. Classically, exosomes act as a vehicle by which cells or organs communicate with each other to maintain cellular/tissue homeostasis and to respond to pathological stress. Most multicellular systems, including the cardiovascular system, use exosomes for intercellular communication. In heart, endogenous exosomes from cardiac cells or stem cells aid in regulation of cell survival, cell proliferation and cell death; and thus tightly regulate cardiac biology and repair processes. Pathological stimulus in heart alters secretion and molecular composition of exosomes, thus influencing the above processes. The past decade has yielded increasing interest in the role of exosomes in the cardiovascular system and significant contribution of cardiac fibroblast (CF) and mediated cardiac fibrosis in heart failure, in this review we had overviewed the relevant literatures about fibroblast exosomes, its effect in the cardiovascular biology and its impact on cardiovascular disease (CVD). This review briefly describes the communication between fibroblasts and other cardiac cells via exosomes, the influence of such on myocardial fibrosis and remodeling, and the possibilities to use exosomes as biomarkers for acute and chronic heart diseases.

Keywords: heart failure, paracrine signaling, exosome, miR, fibrosis

Abbreviations: α-SMA, α-smooth muscle actin; AP-1, activating protein-1; Azin1, antizyme inhibitor 1; CD, cluster of differentiation; CFs, cardiac fibroblasts; CM, cardiomyopathy; CVD, cardiovascular disease; DDR2, discoidin domain receptor 2; DNA, deoxyribonucleic acid; ECM, extracellular matrix; EPC, endothelial progenitor cell; EV, extracellular vesicle; EXO, exosome; Gal-3, Galectin-3; Hsp, heat-shock proteins; IGF1, insulin-like growth factor-1; IL-10, interleukin-10; IR, ischemia-reperfusion; Jnk1, c-Jun N-terminal kinases 1; KO, knock out; lncRNAs, long non-coding RNAs; MAPKs, mitogen-activated protein kinases; MI, myocardial infarction; miR, Micro RNA; MMPs, matrix metalloproteinases; NcRNAs, non-coding RNAs; NF-κB, nuclear factor κB; NFAT, nuclear factor of activated T-cell; Nm, nano meter; PDGFs, platelet derived growth factors; PDLIM5, PDZ and LIM domain 5; PPAR-α, peroxisome proliferators-activated receptor alfa; RAS, renin-angiotensin system; RNAs, ribonucleic acids; ROS, reactive oxygen species; SDF-1, stromal-derived factor-1; SORBS2, sorbin and SH3 domain-containing protein 2; TGF-β, transforming growth factor-β; TIMPs, tissue inhibitors of metalloproteinases; TNF-α, tumor necrosis factor-α.

INTRODUCTION

Intracellular communication is important in proficient and appropriate organization and function of various cells in multicellular organs. Multiple fundamental mechanisms are involved in the interactions between cells or even between different organs. For example growth factors, chemokines, adiponectin, small peptides, ECM proteins or sometimes direct cell-cell interaction are important for cellular communications (Corrado et al., 2013). However, in the last decade a considerable amount of experimental evidence has suggested that cells use a sophisticated method of communication using microvesicles called exosomes (Corrado et al., 2013; Maia et al., 2018). Exosomes are 30-120 nm size nanovesicles and have been identified in multiple cell types including stem cells for efficient intracellular communications (Mathivanan et al., 2010). Promising literature has shown that exosomes play a critical role in the shuttling of extraordinary sets of bioactive and signaling molecules which include membrane receptors, genetic materials, enzymes, cytokines and different bioactive materials in cells (Corrado et al., 2013; Cerezo-Magana et al., 2019). Thorough knowledge of a critical role for exosomes in the cardiovascular system is still developing, but establishment of novel tools and techniques in the past decade have boosted this research area significantly. Seminal work from others and our group has suggested that exosome-mediated intracellular signaling plays an important role in stem cell-mediated cardiac protection both in ischemic and hypertrophic heart failure (Sahoo et al., 2011; Mackie et al., 2012; Khan et al., 2015; Tseliou et al., 2015; Garikipati et al., 2018). Exosomes derived from stem cells provide an excellent cell-free system to improve cardiac function without significant immune response. Furthermore, cardioprotective factors such as miRs (Figure 1) and proteins packaged in stem cell exosomes may enhance the regenerative potential of stem cells to improve the endogenous repair process. Recently, it was shown that exosomes derived from IL-10-depleted EPCs exhibit altered exosomal content, which ultimately impairs the EPC's cardiac repair property (Garikipati et al., 2017). Interestingly, modulation of miR-375 using a miRNA antagomir in IL-10KO exosomes partially rescued endothelial cell function (Yue et al., 2017). These studies clearly indicate that the direct role of exosomes in CVDs and repair processes and alterations in exosomal contents could be beneficial in the treatment of heart disease.

Numerically, heart consists mainly of CFs (Zhou and Pu, 2016) and during ischemic/hypertrophic insults these fibroblasts become activated and involved in cardiac fibrosis and remodeling (Travers et al., 2016). Bang et al. (2014) have shown that fibroblast-derived exosomes have the ability to enhance cardiac myocytes hypertrophy in pressure-overloaded myocardium. The constituent analysis of these exosomes indicates that they are rich in passenger strands of miR such as miR-21*, a major signaling molecule which leads the hypertrophic signaling in heart. Interestingly, inhibition of miR-21 significantly reduced the cardiac hypertrophy and remodeling in this study (Bang et al., 2014). Furthermore, elevated level of miR-155 was found in macrophage-derived exosomes during heart injury (Wang et al., 2017). Intriguingly, Wang et al. (2017) has

suggested that miR-155 in macrophage exosomes has potential to enhance proliferation and differentiation of resident fibroblasts and further exacerbate inflammation. These findings suggest that targeting selective molecules in cardiac fibroblast-derived (CF)-exosomes or inhibition of exosome secretion could be potential therapeutic approaches in heart failure treatment. It is also possible that exosomes from other cells such as immune cells can stimulate transition of naïve fibroblasts to activated myofibroblasts. Very limited literature is available regarding the activated fibroblasts exosomes and exosome-mediated paracrine signaling in cardiac fibrosis and remodeling. We hope that future rigorous studies on CF exosomes and mediated intercellular communications in the heart (between CFs and other cells or vice versa) will provide better understanding to develop novel therapies for CVDs. In this review article, we explore the current understanding of CFs; cardiac fibrosis; exosomes; exosomal biogenesis, structure, composition and involvement in cardiac fibrosis during heart failure. Additionally, we will discuss possibilities of exosomes as biomarkers for cardiac fibrosis and remodeling.

CARDIAC FIBROBLASTS AND CARDIAC FIBROSIS

Excessive cardiac fibrosis is a major problem in nearly all types of heart disease and significantly attributed by activation and excessive proliferation of CFs (Ali et al., 2014; Travers et al., 2016). During development, the CF population changes dramatically and regulates cardiomyocyte proliferation through multiple signaling pathways (Banerjee et al., 2007; Ieda et al., 2009). However, during disease states, excessive ECM proteins such as collagen accumulate and expand in the cardiac interstitium, which disrupts heart contractile capacity and impairs its systolic and diastolic function (Janicki and Brower, 2002; Berk et al., 2007; Kong et al., 2013). Upon cardiac insult, for example acute MI, cardiomyocytes die and a massive inflammatory and fibrogenic responses are triggered to develop a fibrotic scar as a reparative response (Frangogiannis, 2012; Humeres and Frangogiannis, 2019). Activated CFs, termed myofibroblasts, are phenotypically modified cells with differential expression of excessive ECM proteins including collagens, MMPs, and their inhibitors (Cleutjens et al., 1995; Kawaguchi et al., 2011). As compared to CFs, myofibroblasts are more contractile and express significantly more α -SMA and periostin. At an early stage of cardiac stress, these changes contribute to an adaptive repair process but eventually lead to adverse cardiac remodeling and progression toward heart failure (Tomasek et al., 2002; Hinz, 2007, 2010; Shinde et al., 2017). With or without cardiac injury, aging, chronic kidney disease, diabetes and obesity may also trigger cardiac fibrosis and remodeling (Biernacka and Frangogiannis, 2011; Cavalera et al., 2014; Hayer et al., 2018). The homeostasis of collagen turnover is tightly regulated by CFs and any imbalance in collagen metabolism leads to cardiac fibrosis (Janicki and Brower, 2002; Rathod et al., 2016). Cardiac fibrosis is mainly categorized into four types, based on the location and cause. The most prevalent two forms are the reactive

Mesenchymal stem cells	miR-19, miR-223	Cardioprotective Cardioprotective Cardioprotective	
Induced pluripotent stem cells	miR-21, miR-210		
Embryonic stem cells	miR-290 family		
Progenitor cells	Exosomal miRNA Cardiovascular ef		
Endothelial progenitor cells	miR-375, miR-126-3p	Cardioprotective	
Cardiac progenitor cells	miR-451, miR-146a, miR181b, miR-210, miR-132		
Cardiac cells	Exosomal miRNA	Cardiovascular effects	
Fibroblasts	miR-21-3p, miR-29b, miR-30c	Cardiomyocyte hypertrophy	
Cardiomyocytes	miR-320, miR-133, miR-30	Cardiomyopathy, Cardiomyocyte apoptosis	
Macrophages	miR-155, miR-21-5p	Fibrogenesis	
Endothelial cells	miR-214, miR-146a	Regulate senescence & angiogenesis	

interstitial and the replacement fibrosis. Interstitial fibrosis mainly describes the expansion of endomysium and perimysium, is caused by progressive deposition of extracellular proteins in the interstitial space, and leads to cardiomyocyte death. Whereas, replacement fibrosis occurs by necrosis of cardiomyocytes and is associated with systolic ventricular dysfunction, hypertrophic CM and myocarditis (Hashimura et al., 2017; Liu et al., 2017; Frangogiannis, 2019). A third category is infiltrative interstitial fibrosis, which occurs due to infiltration of inflammatory cells in right ventricles of systemic sclerosis-associated pulmonary arterial hypertension (Overbeek et al., 2010). The fourth type, termed endomyocardial fibrosis, is a primary cause of congestive heart failure in children under 2 years of age and underlying causes for this type are not well established but include infections, autoimmunity, genetic factors, and nutritional deficiencies etc. (Rohit et al., 2013; Duraes et al., 2019). The pathophysiology of cardiac fibrosis is mostly attributed to excessive synthesis and accumulation of ECM proteins by activated myofibroblasts. Even though cardiac fibrosis is involved in most forms of CVD, clinical interventions targeting cardiac fibrosis are not yet in hand. Cell population heterogeneity and lack of identification of cell-specific markers add to the complexity of designing and improving therapeutic intervention to reduce cardiac fibrosis.

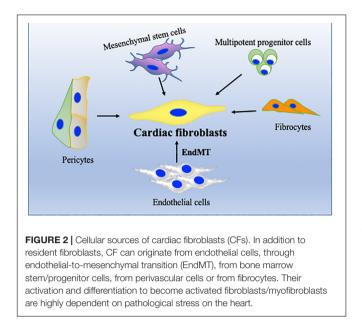
Origin and Activation of Fibroblasts

Regardless of etiology, the origin of myofibroblasts remains controversial (Kong et al., 2014). Recent studies with lineage

tracing strategies have suggested that cardiac fibrosis is primarily mediated by resident fibroblasts; however, other cell types (Figure 2) including monocytes/macrophages, endothelial cells, and hematopoietic fibroblast progenitors may also contribute to pathological fibrosis in heart (Ali et al., 2014; Kong et al., 2014; Moore-Morris et al., 2014, 2018). Endothelial cells and *a*-SMA-expressing mesenchymal cells have been shown to significantly contribute to fibrosis via their canonical Wnt-mediated endothelial-to-mesenchymal transition (EndMT) (Aisagbonhi et al., 2011). We have recently shown that bone marrow cells migrate to heart and transdifferentiate into myofibroblasts after myocardial damage, and thus contribute to tissue remodeling. In our study, we found that inflammatory stimulus acts as a catalyst for enhanced mobilization and homing of these bone marrow progenitor cells (Verma et al., 2017). In heart, cardiomyocytes/fibroblasts/resident macrophages secrete several chemokines such as SDF-1 which play an important role in migration of these cells to the heart (Mollmann et al., 2006; Chu et al., 2010). In addition to bone marrow cells, cardiac endothelial cells may also become myofibroblast-like cells by a process called EndMT and may be involved in pathological fibrosis during hypertrophic heart failure (Zeisberg et al., 2007).

Stimulator of Cardiac Fibrosis

Inflammatory stimulus and cardiomyocyte death are often the initial factors which stimulate a profibrotic signaling cascade in resident fibroblasts which secrete excess ECM proteins



(Tomasek et al., 2002). The potential mechanism and type of profibrotic stimulation which leads this process mostly depends on the type of cardiac injury. Many different growth factors (TNFa, PDGFs, TGFB), cytokines (IL-1, IL-10, IL-11), renin angiotensin system (RAS), and microRs are key modulators in initiation and progression of cardiac fibrosis (Saxena et al., 2013; Thum, 2014; Frangogiannis, 2015, 2019; Tao H. et al., 2016; Verma et al., 2017). RAS and TGF^β signaling are perhaps the most studied among all fibrotic pathways. Regardless of the type of cardiac injury, components of RAS are mostly produced by macrophages and resident fibroblasts and ultimately stimulate cardiac fibrosis by TGF^β signaling pathway. Upon activation, TGFβ modulates cellular functions in various cells including cardiomyocytes and fibroblasts and promotes the myofibroblast phenotype via canonical and non-canonical pathways (Kong et al., 2014). TGF_β-independent activation of human fibroblasts to myofibroblasts has also been studied and is involved in cardiac remodeling (Baranyi et al., 2019). Other than these stimulators, the roles of metalloproteinases (MMPs), TIMPs, and NF-kB are also well established in mediating cardiac fibrosis (Creemers et al., 2001; Kumar et al., 2011; George et al., 2016). As myofibroblasts are a major source of excessive ECM production, in addition to fibroblasts, macrophages also play an important role in ECM production and remodeling. Upon cardiac injury, macrophages adopt a more fibrotic M2 phenotype which has reduced expression of inflammatory cytokines like TNFa and interleukin-6 (IL-6) and increased secretion of IL-10, IGF1, TGFB, and Gal-3 that are really key for ECM remodeling (MacKinnon et al., 2008; Suthahar et al., 2017). Non-immune cells like cardiomyocytes and CFs secrete proinflammatory cytokines to which myofibroblasts mostly respond and leads to excessive cardiac fibrosis (Yamauchi-Takihara et al., 1995; Porter and Turner, 2009; Aoyagi and Matsui, 2011). In addition to these inflammatory and profibrotic factors, GSK3β, β-catenin, TGFβ/SMAD-4, Wnt/β-catenin, MAPKs, and

AKT signaling molecules and pathways play pivotal roles in cardiac fibrosis by regulating ECM metabolism, cardiomyocyte survival and proliferation, and maintaining wound healing after cardiac injury (Deb, 2014; Ma et al., 2018; Singh et al., 2019; Umbarkar et al., 2019; Figure 3). Among intracellular pathways, oxidative stress is a key factor that enhances cardiac fibrosis by triggering fibroblast proliferation and fibrotic signaling in heart. ROS generation has a dual role in cardiac fibrosis as studies reported both matrix-degradation and matrix-synthesis effects. High ROS levels increase TGF^β production and enhance CFs proliferation; paradoxically, high ROS also stimulates MMPs which facilitate ECM degradation (Siwik et al., 2001; Purnomo et al., 2013). Intriguingly, in the last decade, the role of ncRNAs, such as microRNA (miRNA), circular RNA and lncRNA, have been explored extensively in cardiovascular research and are suggested as an important trigger for various cardiovascular events including cardiac fibrosis and remodeling. Among these, microRNs are the most extensively studied NcRNA and play a critical role in regulation of fibroblast proliferation and fibrosis (Thum and Condorelli, 2015; Piccoli et al., 2016). For example, miR-21 is expressed in all heart cells and well characterized as profibrotic miRNA. It targets the expression of sprouty homology 1 (SPRY1), PTEN, and TGF^β receptor III (Huang et al., 2015). In contrast, miR-29 has been shown to reduce fibrosis by down-regulating expression of ECM genes, though in other studies miR-29 expression decreased during heart failure (van Rooij et al., 2008). Other miRNAs like miR-126, miR-15, miR-499, miR-24, miR-378, miR-1, miR-133, miR-26, miR-22, miR-199, miR-23, and miR-208 have also been studied considerably in different heart failure models (Montgomery et al., 2011; Melman et al., 2014; Tijsen et al., 2014; Suresh Babu et al., 2016; Figure 3).

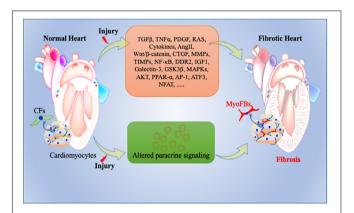


FIGURE 3 Pathophysiology of cardiac fibrosis. During normal remodeling, adequate ECM proteins secreted by cardiac fibroblasts (CFs) are important to maintain cellular integrity in heart. However, upon cardiac injury, CFs transdifferentiate into activated myofibroblasts, secrete excessive amounts of ECM proteins, and expand the cardiac interstitium as a wound healing process. Many key molecular determents are involved in this repair process including exosomes. Altered molecular and paracrine signaling pathways can contribute to exacerbated activation and *trans*-differentiation of fibroblasts to myofibroblasts and to adverse/pathological cardiac remodeling and heart failure. CFs = Cardiac fibroblasts; myoFBs: myofibroblasts.

PARACRINE SIGNALING AND ITS ROLE IN CARDIAC FIBROSIS

Recently, enormous efforts have been made to explore the involvement of paracrine signaling in heart pathophysiology (Anthony and Shiels, 2013; Gartz and Strande, 2018). EVs, as a means of paracrine signaling, are bilayer membrane-bound cargoes and include exosomes, microvesicles and apoptotic bodies. Exosomes are extensively studied paracrine factors and can deliver various functional modulators such as proteins, lipids, DNA, mRNA, and miRNA. These transporter vesicles regulate a plethora of functions like cell differentiation, proliferation, senescence, cell death, cell-cell communication, angiogenesis, recycling of membrane lipids and proteins, and immunomodulation (Janowska-Wieczorek et al., 2005; Valadi et al., 2007; El Andaloussi et al., 2013; Xu and Tahara, 2013; De Jong et al., 2014). In the following section, we will focus on recent advancements in exosome research, especially on fibroblast biology and its role in myocardial fibrosis.

Exosomes: Structure and Function

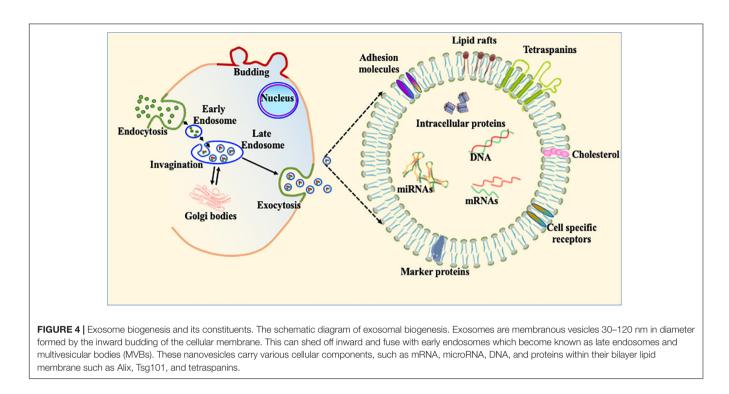
Most living and actively functional cells secret 100-1,000 nm size micro-particles. Based on size, these microparticles are classified into three main subpopulations. The smallest particles (ranging from 30-120 nm) are termed exosomes (Figure 4). Exosomes were first discovered in 1946 as cellular waste and described as nanosized vesicles in 1981 (Chargaff and West, 1946; Valadi et al., 2007). Recent advances in biological science suggest that exosomes play important roles in multiple biological and pathological processes. Due to much complex biology and physiology in heart, exosome-mediated intracellular communication is an under-developed area in the cardiovascular field. Once exosomes are secreted into the extracellular system, they are stable for a relatively long period of time and can transfer cell specific signature signaling molecules to the target or recipient cells (Aryani and Denecke, 2016). Exosomes are mostly involved in cellular communication between different cell populations in multicellular organs. The main purpose of exosome-mediated intracellular signaling is to maintain cellular homeostasis and appropriate response to physiological stress. These nanovesicles contain various cellular components, such as mRNA, microRNA, DNA and membrane-bound or embedded proteins such as Alix, Tsg101 and tetraspanins (Frydrychowicz et al., 2015; Gurunathan et al., 2019). Tetraspanins are a group of transmembrane signaling proteins present in exosomes and in viable cells. The most common tetraspanins on exosomes are CD9, CD63, CD81, and CD82 and are commonly used as markers for characterization of exosomes. Exosomes may also contain Hsp (Hsp70 and Hsp90) and several intercellular adhesion molecules such as CD11a, CD11b, CD11c, CD18, CD146, CD166, and LFA-3/CD58 from their parent cells (Bobrie et al., 2011). In addition to these cellular proteins, exosomes contain a variety of genetic materials (mRNA and miRs) and are involved in angiogenesis, epigenetics, and gene regulation (Bobrie et al., 2011; Frydrychowicz et al., 2015). Although it was initially considered that exosomes are the entities whose

primary function is to clear cellular waste, with increasing understanding of structural details and physiological function, study of cell-specific exosomes is currently a very hot area to explore disease pathobiology (**Figure 4**; Johnstone et al., 1987; Chang and Wang, 2019; Kelemen et al., 2019; Sole et al., 2019; Wang et al., 2019). Based on the broad array of content packaged in exosomes, we think that regulation of exosomal contents is a potential therapeutic strategy in heart disease treatment.

Role of Exosomes in Cardiac Fibrosis

Emerging evidence suggests that exosomes are secreted by most cardiac, vascular and stem cells in heart (Sahoo et al., 2011; Mackie et al., 2012; Khan et al., 2015; Tseliou et al., 2015; Garikipati et al., 2018). Thus, we surmise that all the various cells in heart use exosomes to communicate with each other viz. cardiac myocytes to endothelial cells, endothelial cells to smooth muscle cells, fibroblasts to cardiac myocytes and vice versa to regulate physiological or pathophysiological processes (Hergenreider et al., 2012; Bang et al., 2014; Wang et al., 2014). Thus, any alteration either in signaling molecules packaged in exosomes or in exosomal machinery can affect physiological homeostasis which ultimately results in heart disease. Exosomes play a central role in many cardiac diseases including MI, hypertrophy and ischemia (Xu et al., 2017). Recent evidence has shown that cardiac cell communication via exosomes is altered during fibrosis, a key mediator for heart diseases (Cosme et al., 2017). These novel findings are driving active research to study the role of fibroblast-derived exosomes and the effects of exosomes from other cells on fibroblasts to understand the pathophysiological consequences in fibrosis and heart disease.

Normally CFs contribute \sim 70% of the cardiac cells and support cardiomyocytes by producing ECM and by regulating proliferation and migration of other cardiac cells (Furtado et al., 2016). Thus, fibroblasts play an important role in cardiac repair. However, in certain circumstances excessive proliferation and differentiation of fibroblasts lead to fibrosis and heart failure. Recent literature has shown that exosomes may also modify cardiac repair and fate of fibrosis via modulation of fibroblast function (Barile et al., 2017; Wang et al., 2017). Exosomes derived from cardiac progenitor cells (CPC) have potential to activate naive fibroblasts to initiate the wound healing process for myocardial repair (Barile et al., 2017). During cardiac injury, activated macrophages use exosomes enriched in miR-155 to regulate fibroblast differentiation to myofibroblasts resulting in more fibrosis. Thus, macrophage-specific inhibition of miR-155 or direct inhibition of this miRNA could be potential therapeutic approaches for regulation of cardiac injury (Barile et al., 2017). Recently, Yang et al. (2018) have found that cardiomyocytederived exosomes can promote cardiac fibrosis via myocytefibroblast cross-talk. It has been shown that injured epithelial cells secrete exosomes enriched with profibrotic factors, which can lead to fibrosis (Tomasek et al., 2002; Lee and Kalluri, 2010; Borges et al., 2013). Recently, we have shown that EPCs from IL-10KO mice secrete exosomes which are enriched with profibrotic and antiangiogenic factors and miRs. Alteration in exosomal contents significantly reduced fibrotic signaling after exosome



transplantation in heart post-MI (Garikipati et al., 2017; Yue et al., 2017). Thus, these publications indicate that exosomes from different cellular sources are acting as a significant determinant in regulating cardiac fibrosis. Though exosomes contain several important molecular signatures, the role of exosomal miRs and its significance in regulation of CVDs are recently highlighted in many articles (Valadi et al., 2007). We believe that modification of miRs using selective antigomir/mimic in exosomes can regulate pathological fibrosis and remodeling. To date, several miRs are identified which regulate fibroblast proliferation, differentiation and thus fibrosis (Garikipati et al., 2018; **Table 1**). Therefore, targeting those miRs in exosomes could be beneficial in reducing fibrosis and restoring heart function.

miR-Enriched Exosomes and Their Role in Cardiac Fibrosis

MicroRNAs (miRNAs, miRs) are highly conserved 21–25nucleotide small NcRNA. They regulate target gene expression by binding to mRNAs and regulating the translation process (Thum and Condorelli, 2015; Piccoli et al., 2016). miRNA expression is altered in many CVDs including cardiac fibrosis and remodeling (Li et al., 2013; Tao L. et al., 2016). Recent advancements in technology helped to us to understand the direct role of miRs in cardiac biology and functions. For instance, during ischemic heart diseases, miRs play important roles in cardiac myocyte survival and thus improve heart function. Few miRs are exclusively expressed in muscle tissue (such as cardiac myocytes and skeletal muscles) and involved in cardiomyocyte contractility, survival and function. These miRs are termed myomiR (such as miR-1, miR-133, miR-206, miR-208, miR-486, and miR-499) (McCarthy, 2011; Chistiakov et al., 2016). As myomiRs are mainly expressed in muscle cells and play important roles in cell function, during myocardial damage, myomiR expression is altered tremendously (Gidlof et al., 2011). Interestingly, altered expression of many miRs such as miR-15, miR-21, miR-208a, miR-195, miR-29a, and miR-497 has potential to impair cardiac function post-injury (Hydbring and Badalian-Very, 2013; Porrello et al., 2013; Lin et al., 2015). Previous studies have suggested that CF activation and cardiac fibrosis are tightly regulated by sets of miRs termed fibrosis-associated miRNAs (Please see Table 1). Any alteration in these specific miRs can lead to exaggerated fibrosis. Interestingly, miR-433 is highly increased during myocardial ischemia and leads to cardiac fibrosis. This specific miR regulates MAPK 1 and TGFβ signaling pathways and thus enhances profibrotic signaling (Tao L. et al., 2016). Recently, Moghaddam et al. (2019) have mentioned that many miRs (such as miR-21, miR-22, and miR-24) are highly upregulated during acute ischemic injury. In addition, this group has also mentioned that miR-15, miR-34, miR-130, and miR-378 expression are noticeably reduced and are mainly responsible for the cardiac fibrosis after acute MI and IR injury models (Moghaddam et al., 2019). In summary, miRs from different cellular sources (Please see Table 1) can have ability to alter multiple molecular and cellular processes including cardiac fibrosis. Some sets of miRs can induce fibrosis and others can reduce it. Therefore, a balanced expression of these miRNAs is critically important during appropriate cardiac healing processes after any type of cardiac injury.

As we discussed, exosomes contain an extensive repertoire of genetic material including miRs. Recent reports have suggested that cells can also exchange miRs via exosomes, which can significantly alter the recipient cell's biology and function (Guay et al., 2015; Khalyfa et al., 2016; Qiao et al., 2019).

We have previously shown that miR-125b is an important miR in heart and plays an important role in activation of fibroblasts (Nagpal et al., 2016). Recently, Yang et al. (2018) found that exosomes derived from cardiomyocytes are enriched with miR-208a. At the molecular level, this study suggests that miR-208a enhances NFAT phosphorylation by targeting Dyrk2, preventing its entry into the nucleus in CFs, and therefore triggering fibrosis. Furthermore, Chaturvedi et al. (2015) have demonstrated that during exercise miRNA-29b and miRNA-455enriched exosomes from cardiomyocytes can prevent fibrosis by downregulating MMP9 levels in diabetic mice. Furthermore, Bang et al. (2014) reported that in CFs, exosomal miRs are enriched with many miR passenger strands. In this study they found that fibroblast exosomes are enriched with miR-21* which has potential to induce cardiomyocyte hypertrophy by silencing expression of SORBS2 or PDLIM5 (Bang et al., 2014). Furthermore, Ang II-induced cardiac hypertrophy was effectively controlled by miR-21* inhibition in mice (Bang et al., 2014). In a very similar study, Lyu et al. (2015), showed that activated CF exosomes enhanced RAS signaling in cardiomyocytes, whereas inhibition of CF-exosome secretion by GW4869 (a potent EV inhibitor) significantly reversed Ang II-induced cardiac hypertrophy and remodeling. In addition, Wang et al. (2017) demonstrated that exosomal miR-155 inhibits both SOS and Suppressor of Cytokine Signaling 1 (SCS) expression, respectively, in fibroblasts and macrophages, thus regulating their proliferation. These studies suggest exosomes play important roles in fibroblast-mediated paracrine signaling. Furthermore, regulation of exosome biogenesis or content using pharmacological or molecular approaches could provide valuable therapeutic tools in regulation of heart failure.

Exosomes May Act as a Potential Biomarker in Cardiac Fibrosis

Recently, attempts have been made to use miRNAs or other signaling molecules in serum or plasma as diagnostic biomarkers for cancer. Researchers have found that the molecular constituents of exosomes are highly associated with parent cell phenotype and concurrent physiological/pathological condition. Thus, we can believe that exosomes are replicas of parent cells in regard to their molecular constituents. During biogenesis, exosomes receive multiple proteins via processing through endosomal pathways. These proteins are displayed on the exosome surface and include, but are not limited to, tetraspanins, heat shock proteins (HSP70) and proteins from the Rab family, Tsg101 and Alix (Bobrie et al., 2011; van der Pol et al., 2012). Therefore, proteins on the exosome surface may be utilized as diagnostic tools, as they have been proven very specific and clinically relevant (Lin et al., 2015). It has been shown that body fluids are rich in exosomes, and the specific biomolecules inside of, or on the surface of, exosomes can act as a biomedical tool to determine the disease stage or progression (Takata et al., 2008; Street et al., 2011). Careful investigation of exosomes in body fluids of patients with high risk factors for CVDs may provide us clinically useful information to diagnose these diseases at much earlier stages than previously possible. For example, cardiomyocytes secrete various muscle-specific miRNAs through

S. no.	miRNA	miRNA level	Cardiovascular disease	Target gene/pathway	Cardiovascular effects	References
1	miR-433	Overexpression	Cardiac fibrosis	AZIN1 and JNK1	Induce CF	Tao L. et al., 2016
2	miR-21-5p	Overexpression	Left ventricular hypertrophy	PPARα	Induce Hypertrophy	Chuppa et al., 2018; Moghaddam et al., 2019
3	miR-21-5p, miR-135b	Overexpression	Left ventricular hypertrophy, Cardiomyopathy	Wnt and Hippo pathway	Induce Fibrosis	Zhang H. et al., 2016; Moghaddam et al., 2019
4	miR-22	Downregulation	Cardiac fibrosis	TGFβRI	Induce CF	Jazbutyte et al., 2013; Hong et al., 2016
ō	miR-29	Downregulation	Cardiac fibrosis	TGFβ/BNP	Induce CF	van Rooij et al., 2008; Chaturvedi et al., 2015
6	miR-34a	Overexpression	Cardiac fibrosis after MI and IR injury	SMAD4	Induce CF	Huang et al., 2014
7	miR-208a	Overexpression	Cardiac fibrosis	Dyrk2	Induce CF	Shyu et al., 2015; Yang et al., 2018
3	miR-132	Overexpression	Cardiac fibrosis	PTEN gene, PI3K/Akt	Inhibit CF	Zhang et al., 2018
Э	miR-29a-c	Downregulation	Cardiac fibrosis	TGF-β/Smad3	Induce CF	Roncarati et al., 2014; Zhang et al., 2014; Harmanci et al., 2017
10	miR-669a	Downregulation	Cardiac fibrosis	MyoD	Induce CF	Quattrocelli et al., 2013
11	miR-455	Overexpression	Cardiac fibrosis	CTGF, LncRNA H19	Inhibit CF	Chaturvedi et al., 2015; Huang et al., 2017
12	miR-155	Overexpression	Cardiac fibrosis	TGF-β1–Smad 2	Induce CF	Zhang D. et al., 2016; Wang et al., 2017
13	miR-425, miR-744	Downregulation	Cardiac fibrosis	TGFB1 3'UTR	Induce CF	Wang et al., 2018

exosomes. During cardiac injury, elevated levels of these miRNAs can be detected in blood exosomes much earlier than detection of cardiac troponins or other markers (Xu et al., 2019). Enhanced levels of fibrosis-associated miRNAs (Table 1) such as miR-21, miR-425, miR-744, miR-208a, and others in plasma exosomes can also act as biomarkers in early diagnosis of hypertrophic heart diseases (Wang et al., 2018). In coronary bypass, patients' plasma exosomes are enriched with miR-1 and miR-133 and thus these miRs can be used to indicate disease progress (Emanueli et al., 2016). Matsumoto et al. (2013) has suggested that microRNAs miR-34a, miR-192, and miR-194 can be used as biomarkers to determine heart failure as well. Most studies mentioned here, and many more, have clearly indicated that miRNAs or other molecules packaged in exosomes may act as prognostic markers for heart diseases. However, rigorous investigations must be carried out in large cohorts of human patients before reaching at any final conclusion. We are optimistic that, in the near future, exosomes will be a powerful diagnostic marker to determine the progress of heart disease at early stages and will help our fellow clinicians manage this deadly disease in a more efficient manner.

CONCLUDING AND PROSPECTIVE REMARKS

Influence of exosome-mediated cardiovascular signaling and its role in CVDs have been rigorously studied and many insightful studies have been conducted and published in the recent past; however, we are still far from developing the exosomebased therapeutic for the treatment of CVDs. More in-depth research is warranted for fully understanding the biological aspects of loading, targeting, and delivery of exosomes, and for identifying the endogenous content of exosomes. Several

REFERENCES

- Aisagbonhi, O., Rai, M., Ryzhov, S., Atria, N., Feoktistov, I., and Hatzopoulos, A. K. (2011). Experimental myocardial infarction triggers canonical Wnt signaling and endothelial-to-mesenchymal transition. *Dis. Model. Mech.* 4, 469–483. doi: 10.1242/dmm.006510
- Ali, S. R., Ranjbarvaziri, S., Talkhabi, M., Zhao, P., Subat, A., Hojjat, A., et al. (2014). Developmental heterogeneity of cardiac fibroblasts does not predict pathological proliferation and activation. *Circ. Res.* 115, 625–635. doi: 10.1161/ CIRCRESAHA.115.303794
- Anthony, D. F., and Shiels, P. G. (2013). Exploiting paracrine mechanisms of tissue regeneration to repair damaged organs. *Transplant Res.* 2:10. doi: 10.1186/2047-1440-2-10
- Aoyagi, T., and Matsui, T. (2011). The cardiomyocyte as a source of cytokines in cardiac injury. J. Cell Sci. Ther. 2012:003.
- Aryani, A., and Denecke, B. (2016). Exosomes as a nanodelivery system: a key to the future of neuromedicine? *Mol. Neurobiol.* 53, 818–834. doi: 10.1007/s12035-014-9054-5
- Banerjee, I., Fuseler, J. W., Price, R. L., Borg, T. K., and Baudino, T. A. (2007). Determination of cell types and numbers during cardiac development in the neonatal and adult rat and mouse. *Am. J. Physiol. Heart Circ. Physiol.* 293, H1883–H1891. doi: 10.1152/ajpheart.00514.2007
- Bang, C., Batkai, S., Dangwal, S., Gupta, S. K., Foinquinos, A., Holzmann, A., et al. (2014). Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. J. Clin. Invest. 124, 2136–2146. doi: 10.1172/JCI70577

unanswered questions remain to be addressed, such as (1) What regulates exosome biogenesis during heart failure? (2) How do cell type-specific exosomes exert their effect at the time of heart injury? (3) Does intense fibrotic response alter exosomemediated signaling during CVD? (4) Are there qualitative and/or quantitative differences among fibroblast exosomes from various regions of the myocardium? and (5) Is it possible to alter exosomes to attenuate their detrimental effects and to enhance the benefits? It would be greatly beneficial to develop alternative strategies to engineer fibroblast (or any cell-specific) exosomes to enrich them with factors that target exosomes to the heart and appropriately repair the injury. Recent advancements in cardiovascular research indicate that exosomes may be used as a biomarker to determine heart disease at a much earlier stage than previously used biomarkers. As heart failure is a leading cause of morbidity and mortality both in developed and developing countries, developing novel biomarkers in the form of exosomes will meet a tremendous need to manage this number one lethal disease in a better manner.

AUTHOR CONTRIBUTIONS

PR generated the illustration. PR, RK, and SV wrote the manuscript. SV edited final draft of the manuscript. All authors drafted the manuscript.

FUNDING

This work was supported by the National Institute of Health grant HL135060 and American Heart Association scientist development grant 14SDG20480104 to SV.

- Baranyi, U., Winter, B., Gugerell, A., Hegedus, B., Brostjan, C., Laufer, G., et al. (2019). Primary human fibroblasts in culture switch to a myofibroblastlike phenotype independently of TGF beta. *Cells* 8:E721. doi: 10.3390/cells80 70721
- Barile, L., Milano, G., and Vassalli, G. (2017). Beneficial effects of exosomes secreted by cardiac-derived progenitor cells and other cell types in myocardial ischemia. *Stem Cell Investig.* 4:93. doi: 10.21037/sci.2017.11.06
- Berk, B. C., Fujiwara, K., and Lehoux, S. (2007). ECM remodeling in hypertensive heart disease. J. Clin. Invest. 117, 568–575. doi: 10.1172/JCI31044
- Biernacka, A., and Frangogiannis, N. G. (2011). Aging and cardiac fibrosis. Aging Dis. 2, 158–173.
- Bobrie, A., Colombo, M., Raposo, G., and Thery, C. (2011). Exosome secretion: molecular mechanisms and roles in immune responses. *Traffic* 12, 1659–1668. doi: 10.1111/j.1600-0854.2011.01225.x
- Borges, F. T., Melo, S. A., Ozdemir, B. C., Kato, N., Revuelta, I., Miller, C. A., et al. (2013). TGF-beta1-containing exosomes from injured epithelial cells activate fibroblasts to initiate tissue regenerative responses and fibrosis. J. Am. Soc. Nephrol. 24, 385–392. doi: 10.1681/ASN.2012101031
- Cavalera, M., Wang, J., and Frangogiannis, N. G. (2014). Obesity, metabolic dysfunction, and cardiac fibrosis: pathophysiological pathways, molecular mechanisms, and therapeutic opportunities. *Transl. Res.* 164, 323–335. doi: 10.1016/j.trsl.2014.05.001
- Cerezo-Magana, M., Bang-Rudenstam, A., and Belting, M. (2019). The pleiotropic role of proteoglycans in extracellular vesicle mediated communication in the tumor microenvironment. *Semin. Cancer Biol.* doi: 10.1016/j.semcancer.2019. 07.001 [Epub ahead of print].

- Chang, W., and Wang, J. (2019). Exosomes and their noncoding RNA cargo are emerging as new modulators for diabetes mellitus. *Cells* 8:E853. doi: 10.3390/ cells8080853
- Chargaff, E., and West, R. (1946). The biological significance of the thromboplastic protein of blood. *J. Biol. Chem.* 166, 189–197.
- Chaturvedi, P., Kalani, A., Medina, I., Familtseva, A., and Tyagi, S. C. (2015). Cardiosome mediated regulation of MMP9 in diabetic heart: role of mir29b and mir455 in exercise. *J. Cell Mol. Med.* 19, 2153–2161. doi: 10.1111/jcmm. 12589
- Chistiakov, D. A., Orekhov, A. N., and Bobryshev, Y. V. (2016). Cardiac-specific miRNA in cardiogenesis, heart function, and cardiac pathology (with focus on myocardial infarction). J. Mol. Cell Cardiol. 94, 107–121. doi: 10.1016/j.yjmcc. 2016.03.015
- Chu, P. Y., Mariani, J., Finch, S., McMullen, J. R., Sadoshima, J., Marshall, T., et al. (2010). Bone marrow-derived cells contribute to fibrosis in the chronically failing heart. Am. J. Pathol. 176, 1735–1742. doi: 10.2353/ajpath.2010. 090574
- Chuppa, S., Liang, M., Liu, P., Liu, Y., Casati, M. C., Cowley, A. W., et al. (2018). MicroRNA-21 regulates peroxisome proliferator-activated receptor alpha, a molecular mechanism of cardiac pathology in cardiorenal syndrome Type 4. *Kidney Int.* 93, 375–389. doi: 10.1016/j.kint.2017. 05.014
- Cleutjens, J. P., Verluyten, M. J., Smiths, J. F., and Daemen, M. J. (1995). Collagen remodeling after myocardial infarction in the rat heart. Am. J. Pathol. 147, 325–338.
- Corrado, C., Raimondo, S., Chiesi, A., Ciccia, F., De Leo, G., and Alessandro, R. (2013). Exosomes as intercellular signaling organelles involved in health and disease: basic science and clinical applications. *Int. J. Mol. Sci.* 14, 5338–5366. doi: 10.3390/ijms14035338
- Cosme, J., Guo, H., Hadipour-Lakmehsari, S., Emili, A., and Gramolini, A. O. (2017). Hypoxia-induced changes in the fibroblast secretome, exosome, and whole-cell proteome using cultured, cardiac-derived cells isolated from neonatal mice. J. Proteome Res. 16, 2836–2847. doi: 10.1021/acs.jproteome. 7b00144
- Creemers, E. E., Cleutjens, J. P., Smits, J. F., and Daemen, M. J. (2001). Matrix metalloproteinase inhibition after myocardial infarction: a new approach to prevent heart failure? *Circ. Res.* 89, 201–210. doi: 10.1161/hh1501. 094396
- De Jong, O. G., Van Balkom, B. W., Schiffelers, R. M., Bouten, C. V., and Verhaar, M. C. (2014). Extracellular vesicles: potential roles in regenerative medicine. *Front. Immunol.* 5:608. doi: 10.3389/fimmu.2014.00608
- Deb, A. (2014). Cell-cell interaction in the heart via Wnt/beta-catenin pathway after cardiac injury. *Cardiovasc. Res.* 102, 214–223. doi: 10.1093/cvr/ cvu054
- Duraes, A. R., de Souza Lima Bitar, Y., Roever, L., and Neto, M. G. (2019). Endomyocardial fibrosis: past, present, and future. *Heart Fail. Rev.* doi: 10.1007/ s10741-019-09848-4 [Epub ahead of print].
- El Andaloussi, S., Lakhal, S., Mager, I., and Wood, M. J. (2013). Exosomes for targeted siRNA delivery across biological barriers. *Adv. Drug Deliv. Rev.* 65, 391–397. doi: 10.1016/j.addr.2012.08.008
- Emanueli, C., Shearn, A. I., Laftah, A., Fiorentino, F., Reeves, B. C., Beltrami, C., et al. (2016). Coronary artery-bypass-graft surgery increases the plasma concentration of exosomes carrying a cargo of cardiac MicroRNAs: an example of exosome trafficking out of the human heart with potential for cardiac biomarker discovery. *PLoS One* 11:e0154274. doi: 10.1371/journal.pone. 0154274
- Frangogiannis, N. G. (2012). Regulation of the inflammatory response in cardiac repair. Circ. Res. 110, 159–173. doi: 10.1161/CIRCRESAHA.111. 243162
- Frangogiannis, N. G. (2015). Pathophysiology of myocardial infarction. Compr. Physiol. 5, 1841–1875. doi: 10.1002/cphy.c150006
- Frangogiannis, N. G. (2019). Cardiac fibrosis: cell biological mechanisms, molecular pathways and therapeutic opportunities. *Mol. Aspects Med.* 65, 70–99. doi: 10.1016/j.mam.2018.07.001
- Frydrychowicz, M., Kolecka-Bednarczyk, A., Madejczyk, M., Yasar, S., and Dworacki, G. (2015). Exosomes - structure, biogenesis and biological role in non-small-cell lung cancer. *Scand. J. Immunol.* 81, 2–10. doi: 10.1111/sji. 12247

- Furtado, M. B., Nim, H. T., Boyd, S. E., and Rosenthal, N. A. (2016). View from the heart: cardiac fibroblasts in development, scarring and regeneration. *Development* 143, 387–397. doi: 10.1242/dev.120576
- Garikipati, V. N. S., Shoja-Taheri, F., Davis, M. E., and Kishore, R. (2018). Extracellular vesicles and the application of system biology and computational modeling in cardiac repair. *Circ. Res.* 123, 188–204. doi: 10.1161/CIRCRESAHA.117.311215
- Garikipati, V. N. S., Verma, S. K., Jolardarashi, D., Cheng, Z., Ibetti, J., Cimini, M., et al. (2017). Therapeutic inhibition of miR-375 attenuates post-myocardial infarction inflammatory response and left ventricular dysfunction via PDK-1-AKT signalling axis. *Cardiovasc. Res.* 113, 938–949. doi: 10.1093/cvr/cvx052
- Gartz, M., and Strande, J. L. (2018). Examining the paracrine effects of exosomes in cardiovascular disease and repair. J. Am. Heart Assoc. 7:e007954. doi: 10.1161/ JAHA.117.007954
- George, M., Vijayakumar, A., Dhanesh, S. B., James, J., and Shivakumar, K. (2016). Molecular basis and functional significance of Angiotensin II-induced increase in discoidin domain receptor 2 gene expression in cardiac fibroblasts. J. Mol. Cell Cardiol. 90, 59–69. doi: 10.1016/j.yjmcc.2015.12.004
- Gidlof, O., Andersson, P., van der Pals, J., Gotberg, M., and Erlinge, D. (2011). Cardiospecific microRNA plasma levels correlate with troponin and cardiac function in patients with ST elevation myocardial infarction, are selectively dependent on renal elimination, and can be detected in urine samples. *Cardiology* 118, 217–226. doi: 10.1159/000328869
- Guay, C., Menoud, V., Rome, S., and Regazzi, R. (2015). Horizontal transfer of exosomal microRNAs transduce apoptotic signals between pancreatic betacells. *Cell Commun. Signal.* 13:17. doi: 10.1186/s12964-015-0097-7
- Gurunathan, S., Kang, M. H., Jeyaraj, M., Qasim, M., and Kim, J. H. (2019). Review of the isolation, characterization, biological function, and multifarious therapeutic approaches of exosomes. *Cells* 8:307. doi: 10.3390/cells8040307
- Harmanci, D., Erkan, E. P., Kocak, A., and Akdogan, G. G. (2017). Role of the microRNA-29 family in fibrotic skin diseases. *Biomed. Rep.* 6, 599–604. doi: 10.3892/br.2017.900
- Hashimura, H., Kimura, F., Ishibashi-Ueda, H., Morita, Y., Higashi, M., Nakano, S., et al. (2017). Radiologic-pathologic correlation of primary and secondary cardiomyopathies: MR Imaging and histopathologic findings in hearts from autopsy and transplantation. *Radiographics* 37, 719–736. doi: 10.1148/rg. 2017160082
- Hayer, M. K., Price, A. M., Liu, B., Baig, S., Ferro, C. J., Townend, J. N., et al. (2018). Diffuse myocardial interstitial fibrosis and dysfunction in early chronic kidney disease. Am. J. Cardiol. 121, 656–660. doi: 10.1016/j.amjcard.2017.11.041
- Hergenreider, E., Heydt, S., Treguer, K., Boettger, T., Horrevoets, A. J., Zeiher, A. M., et al. (2012). Atheroprotective communication between endothelial cells and smooth muscle cells through miRNAs. *Nat. Cell Biol.* 14, 249–256. doi: 10.1038/ncb2441
- Hinz, B. (2007). Formation and function of the myofibroblast during tissue repair. J. Invest. Dermatol. 127, 526–537. doi: 10.1038/sj.jid.5700613
- Hinz, B. (2010). The myofibroblast: paradigm for a mechanically active cell. *J. Biomech.* 43, 146–155. doi: 10.1016/j.jbiomech.2009.09.020
- Hong, Y., Cao, H., Wang, Q., Ye, J., Sui, L., Feng, J., et al. (2016). MiR-22 may suppress fibrogenesis by targeting TGFbetaR I in cardiac fibroblasts. *Cell Physiol. Biochem.* 40, 1345–1353. doi: 10.1159/000453187
- Huang, Y., He, Y., and Li, J. (2015). MicroRNA-21: a central regulator of fibrotic diseases via various targets. *Curr. Pharm. Des.* 21, 2236–2242. doi: 10.2174/ 1381612820666141226095701
- Huang, Y., Qi, Y., Du, J. Q., and Zhang, D. F. (2014). MicroRNA-34a regulates cardiac fibrosis after myocardial infarction by targeting Smad4. *Expert Opin. Ther. Targets* 18, 1355–1365. doi: 10.1517/14728222.2014.961424
- Huang, Z. W., Tian, L. H., Yang, B., and Guo, R. M. (2017). Long noncoding RNA H19 acts as a competing endogenous RNA to mediate CTGF expression by sponging miR-455 in cardiac fibrosis. DNA Cell Biol. 36, 759–766. doi: 10.1089/dna.2017.3799
- Humeres, C., and Frangogiannis, N. G. (2019). Fibroblasts in the infarcted, remodeling, and failing heart. JACC Basic Transl. Sci. 4, 449–467. doi: 10.1016/ j.jacbts.2019.02.006
- Hydbring, P., and Badalian-Very, G. (2013). Clinical applications of microRNAs. F1000Res 2:136. doi: 10.12688/f1000research.2-136.v3
- Ieda, M., Tsuchihashi, T., Ivey, K. N., Ross, R. S., Hong, T. T., Shaw, R. M., et al. (2009). Cardiac fibroblasts regulate myocardial proliferation through

beta1 integrin signaling. Dev. Cell 16, 233-244. doi: 10.1016/j.devcel.2008. 12.007

- Janicki, J. S., and Brower, G. L. (2002). The role of myocardial fibrillar collagen in ventricular remodeling and function. J. Card. Fail. 8(6 Suppl.), S319–S325. doi: 10.1054/jcaf.2002.129260
- Janowska-Wieczorek, A., Wysoczynski, M., Kijowski, J., Marquez-Curtis, L., Machalinski, B., Ratajczak, J., et al. (2005). Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *Int. J. Cancer* 113, 752–760. doi: 10.1002/ijc.20657
- Jazbutyte, V., Fiedler, J., Kneitz, S., Galuppo, P., Just, A., Holzmann, A., et al. (2013). MicroRNA-22 increases senescence and activates cardiac fibroblasts in the aging heart. Age 35, 747–762. doi: 10.1007/s11357-012-9407-9
- Johnstone, R. M., Adam, M., Hammond, J. R., Orr, L., and Turbide, C. (1987). Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). J. Biol. Chem. 262, 9412–9420.
- Kawaguchi, M., Takahashi, M., Hata, T., Kashima, Y., Usui, F., Morimoto, H., et al. (2011). Inflammasome activation of cardiac fibroblasts is essential for myocardial ischemia/reperfusion injury. *Circulation* 123, 594–604. doi: 10. 1161/CIRCULATIONAHA.110.982777
- Kelemen, E., Danis, J., Goblos, A., Bata-Csorgo, Z., and Szell, M. (2019). Exosomal long non-coding RNAs as biomarkers in human diseases. *EJIFCC* 30, 224–236.
- Khalyfa, A., Kheirandish-Gozal, L., Khalyfa, A. A., Philby, M. F., Alonso-Alvarez, M. L., Mohammadi, M., et al. (2016). Circulating plasma extracellular microvesicle MicroRNA cargo and endothelial dysfunction in children with obstructive sleep apnea. Am. J. Respir. Crit. Care Med. 194, 1116–1126. doi: 10.1164/rccm.201602-0323OC
- Khan, M., Nickoloff, E., Abramova, T., Johnson, J., Verma, S. K., Krishnamurthy, P., et al. (2015). Embryonic stem cell-derived exosomes promote endogenous repair mechanisms and enhance cardiac function following myocardial infarction. *Circ. Res.* 117, 52–64. doi: 10.1161/CIRCRESAHA.117.305990
- Kong, P., Christia, P., and Frangogiannis, N. G. (2014). The pathogenesis of cardiac fibrosis. *Cell Mol. Life Sci.* 71, 549–574. doi: 10.1007/s00018-013-1349-6
- Kong, P., Christia, P., Saxena, A., Su, Y., and Frangogiannis, N. G. (2013). Lack of specificity of fibroblast-specific protein 1 in cardiac remodeling and fibrosis. *Am. J. Physiol. Heart Circ. Physiol.* 305, H1363–H1372. doi: 10.1152/ajpheart. 00395.2013
- Kumar, S., Seqqat, R., Chigurupati, S., Kumar, R., Baker, K. M., Young, D., et al. (2011). Inhibition of nuclear factor kappaB regresses cardiac hypertrophy by modulating the expression of extracellular matrix and adhesion molecules. *Free Radic. Biol. Med.* 50, 206–215. doi: 10.1016/j.freeradbiomed.2010. 10.711
- Lee, S. B., and Kalluri, R. (2010). Mechanistic connection between inflammation and fibrosis. *Kidney Int. Suppl.* 119, S22–S26. doi: 10.1038/ki.2010.418
- Li, J., Xu, J., Cheng, Y., Wang, F., Song, Y., and Xiao, J. (2013). Circulating microRNAs as mirrors of acute coronary syndromes: MiRacle or quagMire? J. Cell. Mol. Med. 17, 1363–1370. doi: 10.1111/jcmm.12148
- Lin, J., Li, J., Huang, B., Liu, J., Chen, X., Chen, X. M., et al. (2015). Exosomes: novel biomarkers for clinical diagnosis. *Sci. World J.* 2015, 657086. doi: 10.1155/2015/ 657086
- Liu, T., Song, D., Dong, J., Zhu, P., Liu, J., Liu, W., et al. (2017). Current understanding of the pathophysiology of myocardial fibrosis and its quantitative assessment in heart failure. *Front. Physiol.* 8:238. doi: 10.3389/ fphys.2017.00238
- Lyu, L., Wang, H., Li, B., Qin, Q., Qi, L., Nagarkatti, M., et al. (2015). A critical role of cardiac fibroblast-derived exosomes in activating renin angiotensin system in cardiomyocytes. *J. Mol. Cell Cardiol.* 89(Pt B), 268–279. doi: 10.1016/j.yjmcc. 2015.10.022
- Ma, Z. G., Yuan, Y. P., Wu, H. M., Zhang, X., and Tang, Q. Z. (2018). Cardiac fibrosis: new insights into the pathogenesis. *Int. J. Biol. Sci.* 14, 1645–1657. doi: 10.7150/ijbs.28103
- Mackie, A. R., Klyachko, E., Thorne, T., Schultz, K. M., Millay, M., Ito, A., et al. (2012). Sonic hedgehog-modified human CD34+ cells preserve cardiac function after acute myocardial infarction. *Circ. Res.* 111, 312–321. doi: 10.1161/ CIRCRESAHA.112.266015

- MacKinnon, A. C., Farnworth, S. L., Hodkinson, P. S., Henderson, N. C., Atkinson, K. M., Leffler, H., et al. (2008). Regulation of alternative macrophage activation by galectin-3. J. Immunol. 180, 2650–2658. doi: 10.4049/jimmunol.180.4.2650
- Maia, J., Caja, S., Strano Moraes, M. C., Couto, N., and Costa-Silva, B. (2018). Exosome-based cell-cell communication in the tumor microenvironment. *Front. Cell Dev. Biol.* 6:18. doi: 10.3389/fcell.2018.00018
- Mathivanan, S., Ji, H., and Simpson, R. J. (2010). Exosomes: extracellular organelles important in intercellular communication. J. Proteomics 73, 1907–1920. doi: 10.1016/j.jprot.2010.06.006
- Matsumoto, S., Sakata, Y., Suna, S., Nakatani, D., Usami, M., Hara, M., et al. (2013). Circulating p53-responsive microRNAs are predictive indicators of heart failure after acute myocardial infarction. *Circ. Res.* 113, 322–326. doi: 10.1161/CIRCRESAHA.113.301209
- McCarthy, J. J. (2011). The MyomiR network in skeletal muscle plasticity. *Exerc.* Sport Sci. Rev. 39, 150–154. doi: 10.1097/JES.0b013e31821c01e1
- Melman, Y. F., Shah, R., and Das, S. (2014). MicroRNAs in heart failure: is the picture becoming less miRky? *Circ. Heart Fail.* 7, 203–214. doi: 10.1161/ CIRCHEARTFAILURE.113.000266
- Moghaddam, A. S., Afshari, J. T., Esmaeili, S. A., Saburi, E., Joneidi, Z., and Momtazi-Borojeni, A. A. (2019). Cardioprotective microRNAs: lessons from stem cell-derived exosomal microRNAs to treat cardiovascular disease. *Atherosclerosis* 285, 1–9. doi: 10.1016/j.atherosclerosis.2019.03.016
- Mollmann, H., Nef, H. M., Kostin, S., von Kalle, C., Pilz, I., Weber, M., et al. (2006). Bone marrow-derived cells contribute to infarct remodelling. *Cardiovasc. Res.* 71, 661–671. doi: 10.1016/j.cardiores.2006.06.013
- Montgomery, R. L., Hullinger, T. G., Semus, H. M., Dickinson, B. A., Seto, A. G., Lynch, J. M., et al. (2011). Therapeutic inhibition of miR-208a improves cardiac function and survival during heart failure. *Circulation* 124, 1537–1547. doi: 10.1161/CIRCULATIONAHA.111.030932
- Moore-Morris, T., Cattaneo, P., Guimaraes-Camboa, N., Bogomolovas, J., Cedenilla, M., Banerjee, I., et al. (2018). Infarct fibroblasts do not derive from bone marrow lineages. *Circ. Res.* 122, 583–590. doi: 10.1161/CIRCRESAHA. 117.311490
- Moore-Morris, T., Guimaraes-Camboa, N., Banerjee, I., Zambon, A. C., Kisseleva, T., Velayoudon, A., et al. (2014). Resident fibroblast lineages mediate pressure overload-induced cardiac fibrosis. J. Clin. Invest. 124, 2921–2934. doi: 10.1172/ JCI74783
- Nagpal, V., Rai, R., Place, A. T., Murphy, S. B., Verma, S. K., Ghosh, A. K., et al. (2016). MiR-125b is critical for fibroblast-to-myofibroblast transition and cardiac fibrosis. *Circulation* 133, 291–301. doi: 10.1161/CIRCULATIONAHA. 115.018174
- Overbeek, M. J., Mouchaers, K. T., Niessen, H. M., Hadi, A. M., Kupreishvili, K., Boonstra, A., et al. (2010). Characteristics of interstitial fibrosis and inflammatory cell infiltration in right ventricles of systemic sclerosis-associated pulmonary arterial hypertension. *Int. J. Rheumatol.* 2010:604615. doi: 10.1155/ 2010/604615
- Piccoli, M. T., Bar, C., and Thum, T. (2016). Non-coding RNAs as modulators of the cardiac fibroblast phenotype. J. Mol. Cell Cardiol. 92, 75–81. doi: 10.1016/j. yjmcc.2015.12.023
- Porrello, E. R., Mahmoud, A. I., Simpson, E., Johnson, B. A., Grinsfelder, D., Canseco, D., et al. (2013). Regulation of neonatal and adult mammalian heart regeneration by the miR-15 family. *Proc. Natl. Acad. Sci. U.S.A.* 110, 187–192. doi: 10.1073/pnas.1208863110
- Porter, K. E., and Turner, N. A. (2009). Cardiac fibroblasts: at the heart of myocardial remodeling. *Pharmacol. Ther.* 123, 255–278. doi: 10.1016/j. pharmthera.2009.05.002
- Purnomo, Y., Piccart, Y., Coenen, T., Prihadi, J. S., and Lijnen, P. J. (2013). Oxidative stress and transforming growth factor-beta1-induced cardiac fibrosis. *Cardiovasc. Hematol. Disord. Drug Targets* 13, 165–172. doi: 10.2174/ 1871529x11313020010
- Qiao, L., Hu, S., Liu, S., Zhang, H., Ma, H., Huang, K., et al. (2019). microRNA-21-5p dysregulation in exosomes derived from heart failure patients impairs regenerative potential. *J. Clin. Invest.* 129, 2237–2250. doi: 10.1172/JCI123135
- Quattrocelli, M., Crippa, S., Montecchiani, C., Camps, J., Cornaglia, A. I., Boldrin, L., et al. (2013). Long-term miR-669a therapy alleviates chronic dilated cardiomyopathy in dystrophic mice. *J. Am. Heart Assoc.* 2:e000284. doi: 10. 1161/JAHA.113.000284

Rathod, R. H., Powell, A. J., and Geva, T. (2016). Myocardial fibrosis in congenital heart disease. *Circ. J.* 80, 1300–1307. doi: 10.1253/circj.CJ-16-0353

- Rohit, M., Gupta, A., and Talwar, K. K. (2013). Heart failure in children in tropical regions. Curr. Heart Fail. Rep. 10, 277–284. doi: 10.1007/s11897-013-0158-4
- Roncarati, R., Viviani Anselmi, C., Losi, M. A., Papa, L., Cavarretta, E., Da Costa Martins, P., et al. (2014). Circulating miR-29a, among other up-regulated microRNAs, is the only biomarker for both hypertrophy and fibrosis in patients with hypertrophic cardiomyopathy. J. Am. Coll. Cardiol. 63, 920–927. doi: 10. 1016/j.jacc.2013.09.041
- Sahoo, S., Klychko, E., Thorne, T., Misener, S., Schultz, K. M., Millay, M., et al. (2011). Exosomes from human CD34(+) stem cells mediate their proangiogenic paracrine activity. *Circ. Res.* 109, 724–728. doi: 10.1161/CIRCRESAHA.111. 253286
- Saxena, A., Chen, W., Su, Y., Rai, V., Uche, O. U., Li, N., et al. (2013). IL-1 induces proinflammatory leukocyte infiltration and regulates fibroblast phenotype in the infarcted myocardium. *J. Immunol.* 191, 4838–4848. doi: 10.4049/jimmunol. 1300725
- Shinde, A. V., Humeres, C., and Frangogiannis, N. G. (2017). The role of alpha-smooth muscle actin in fibroblast-mediated matrix contraction and remodeling. *Biochim. Biophys. Acta Mol. Basis Dis.* 1863, 298–309. doi: 10.1016/ j.bbadis.2016.11.006
- Shyu, K. G., Wang, B. W., Cheng, W. P., and Lo, H. M. (2015). MicroRNA-208a increases myocardial endoglin expression and myocardial fibrosis in acute myocardial infarction. *Can. J. Cardiol.* 31, 679–690. doi: 10.1016/j.cjca.2014.12. 026
- Singh, A. P., Umbarkar, P., Guo, Y., Force, T., Gupte, M., and Lal, H. (2019). Inhibition of GSK-3 to induce cardiomyocyte proliferation: a recipe for in situ cardiac regeneration. *Cardiovasc. Res.* 115, 20–30. doi: 10.1093/cvr/cvy255
- Siwik, D. A., Pagano, P. J., and Colucci, W. S. (2001). Oxidative stress regulates collagen synthesis and matrix metalloproteinase activity in cardiac fibroblasts. *Am. J. Physiol. Cell Physiol.* 280, C53–C60. doi: 10.1152/ajpcell.2001.280.1.C53
- Sole, C., Moline, T., Vidal, M., Ordi-Ros, J., and Cortes-Hernandez, J. (2019). An exosomal urinary miRNA signature for early diagnosis of renal fibrosis in lupus nephritis. *Cells* 8:773. doi: 10.3390/cells8080773
- Street, J. M., Birkhoff, W., Menzies, R. I., Webb, D. J., Bailey, M. A., and Dear, J. W. (2011). Exosomal transmission of functional aquaporin 2 in kidney cortical collecting duct cells. *J. Physiol.* 589(Pt 24), 6119–6127. doi: 10.1113/jphysiol. 2011.220277
- Suresh Babu, S., Thandavarayan, R. A., Joladarashi, D., Jeyabal, P., Krishnamurthy, S., Bhimaraj, A., et al. (2016). MicroRNA-126 overexpression rescues diabetesinduced impairment in efferocytosis of apoptotic cardiomyocytes. *Sci. Rep.* 6:36207. doi: 10.1038/srep36207
- Suthahar, N., Meijers, W. C., Sillje, H. H. W., and de Boer, R. A. (2017). From inflammation to fibrosis-molecular and cellular mechanisms of myocardial tissue remodelling and perspectives on differential treatment opportunities. *Curr. Heart Fail. Rep.* 14, 235–250. doi: 10.1007/s11897-017-0343-y
- Takata, K., Matsuzaki, T., Tajika, Y., Ablimit, A., and Hasegawa, T. (2008). Localization and trafficking of aquaporin 2 in the kidney. *Histochem. Cell Biol.* 130, 197–209. doi: 10.1007/s00418-008-0457-0
- Tao, H., Yang, J. J., Shi, K. H., and Li, J. (2016). Wnt signaling pathway in cardiac fibrosis: new insights and directions. *Metabolism* 65, 30–40. doi: 10.1016/j. metabol.2015.10.013
- Tao, L., Bei, Y., Chen, P., Lei, Z., Fu, S., Zhang, H., et al. (2016). Crucial role of miR-433 in regulating cardiac fibrosis. *Theranostics* 6, 2068–2083. doi: 10.7150/ thno.15007
- Thum, T. (2014). Noncoding RNAs and myocardial fibrosis. *Nat. Rev. Cardiol.* 11, 655–663. doi: 10.1038/nrcardio.2014.125
- Thum, T., and Condorelli, G. (2015). Long noncoding RNAs and microRNAs in cardiovascular pathophysiology. *Circ. Res.* 116, 751–762. doi: 10.1161/ CIRCRESAHA.116.303549
- Tijsen, A. J., van der Made, I., van den Hoogenhof, M. M., Wijnen, W. J., van Deel, E. D., de Groot, N. E., et al. (2014). The microRNA-15 family inhibits the TGFbeta-pathway in the heart. *Cardiovasc. Res.* 104, 61–71. doi: 10.1093/ cvr/cvu184
- Tomasek, J. J., Gabbiani, G., Hinz, B., Chaponnier, C., and Brown, R. A. (2002). Myofibroblasts and mechano-regulation of connective tissue remodelling. *Nat. Rev. Mol. Cell Biol.* 3, 349–363. doi: 10.1038/nrm809

- Travers, J. G., Kamal, F. A., Robbins, J., Yutzey, K. E., and Blaxall, B. C. (2016). Cardiac fibrosis: the fibroblast awakens. *Circ. Res.* 118, 1021–1040. doi: 10.1161/ CIRCRESAHA.115.306565
- Tseliou, E., Fouad, J., Reich, H., Slipczuk, L., de Couto, G., Aminzadeh, M., et al. (2015). Fibroblasts rendered antifibrotic, antiapoptotic, and angiogenic by priming with cardiosphere-derived extracellular membrane vesicles. J. Am. Coll. Cardiol. 66, 599–611. doi: 10.1016/j.jacc.2015.05.068
- Umbarkar, P., Singh, A. P., Gupte, M., Verma, V. K., Galindo, C. L., Guo, Y., et al. (2019). Cardiomyocyte SMAD4-Dependent TGF-beta signaling is essential to maintain adult heart homeostasis. *JACC Basic Transl. Sci.* 4, 41–53. doi: 10.1016/ j.jacbts.2018.10.003
- Valadi, H., Ekstrom, K., Bossios, A., Sjostrand, M., Lee, J. J., and Lotvall, J. O. (2007). Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 9, 654–659. doi: 10.1038/ncb1596
- van der Pol, E., Boing, A. N., Harrison, P., Sturk, A., and Nieuwland, R. (2012). Classification, functions, and clinical relevance of extracellular vesicles. *Pharmacol. Rev.* 64, 676–705. doi: 10.1124/pr.112.005983
- van Rooij, E., Sutherland, L. B., Thatcher, J. E., DiMaio, J. M., Naseem, R. H., Marshall, W. S., et al. (2008). Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. *Proc. Natl. Acad. Sci.* U.S.A. 105, 13027–13032. doi: 10.1073/pnas.0805038105
- Verma, S. K., Garikipati, V. N. S., Krishnamurthy, P., Schumacher, S. M., Grisanti, L. A., Cimini, M., et al. (2017). Interleukin-10 inhibits bone marrow fibroblast progenitor cell-mediated cardiac fibrosis in pressure-overloaded myocardium. *Circulation* 136, 940–953. doi: 10.1161/CIRCULATIONAHA.117.027889
- Wang, C., Zhang, C., Liu, L., A, X., Chen, B., Li, Y., et al. (2017). Macrophagederived mir-155-containing exosomes suppress fibroblast proliferation and promote fibroblast inflammation during cardiac injury. *Mol. Ther.* 25, 192–204. doi: 10.1016/j.ymthe.2016.09.001
- Wang, L., Liu, J., Xu, B., Liu, Y. L., and Liu, Z. (2018). Reduced exosome miR-425 and miR-744 in the plasma represents the progression of fibrosis and heart failure. *Kaohsiung J. Med. Sci.* 34, 626–633. doi: 10.1016/j.kjms.2018.05.008
- Wang, M., Zhou, L., Yu, F., Zhang, Y., Li, P., and Wang, K. (2019). The functional roles of exosomal long non-coding RNAs in cancer. *Cell Mol. Life Sci.* 76, 2059–2076. doi: 10.1007/s00018-019-03018-3
- Wang, X., Huang, W., Liu, G., Cai, W., Millard, R. W., Wang, Y., et al. (2014). Cardiomyocytes mediate anti-angiogenesis in type 2 diabetic rats through the exosomal transfer of miR-320 into endothelial cells. J. Mol. Cell Cardiol. 74, 139–150. doi: 10.1016/j.yjmcc.2014.05.001
- Xu, D., and Tahara, H. (2013). The role of exosomes and microRNAs in senescence and aging. Adv. Drug Deliv. Rev. 65, 368–375. doi: 10.1016/j.addr.2012.07.010
- Xu, J. Y., Chen, G. H., and Yang, Y. J. (2017). Exosomes: a rising star in falling hearts. Front. Physiol. 8:494. doi: 10.3389/fphys.2017.00494
- Xu, M. Y., Ye, Z. S., Song, X. T., and Huang, R. C. (2019). Differences in the cargos and functions of exosomes derived from six cardiac cell types: a systematic review. *Stem Cell Res. Ther.* 10:194. doi: 10.1186/s13287-019-1297-7
- Yamauchi-Takihara, K., Ihara, Y., Ogata, A., Yoshizaki, K., Azuma, J., and Kishimoto, T. (1995). Hypoxic stress induces cardiac myocyte-derived interleukin-6. *Circulation* 91, 1520–1524. doi: 10.1161/01.cir.91.5.1520
- Yang, J., Yu, X., Xue, F., Li, Y., Liu, W., and Zhang, S. (2018). Exosomes derived from cardiomyocytes promote cardiac fibrosis via myocyte-fibroblast crosstalk. Am. J. Transl. Res. 10, 4350–4366.
- Yue, Y., Garikipati, V. N. S., Verma, S. K., Goukassian, D. A., and Kishore, R. (2017). Interleukin-10 deficiency impairs reparative properties of bone marrowderived endothelial progenitor cell exosomes. *Tissue Eng. Part A* 23, 1241–1250. doi: 10.1089/ten.TEA.2017.0084
- Zeisberg, E. M., Tarnavski, O., Zeisberg, M., Dorfman, A. L., McMullen, J. R., Gustafsson, E., et al. (2007). Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat. Med.* 13, 952–961. doi: 10.1038/nm1613
- Zhang, C. J., Huang, Y., Lu, J. D., Lin, J., Ge, Z. R., and Huang, H. (2018). Upregulated microRNA-132 rescues cardiac fibrosis and restores cardiocyte proliferation in dilated cardiomyopathy through the phosphatase and tensin homolog-mediated PI3K/Akt signal transduction pathway. *J. Cell Biochem.* 120, 1232–1244. doi: 10.1002/jcb.27081 [Epub ahead of print].
- Zhang, D., Cui, Y., Li, B., Luo, X., Li, B., and Tang, Y. (2016). miR-155 regulates high glucose-induced cardiac fibrosis via the TGF-beta signaling pathway. *Mol. Biosyst.* 13, 215–224. doi: 10.1039/c6mb00649c

Zhang, H., Liu, S., Dong, T., Yang, J., Xie, Y., Wu, Y., et al. (2016). Profiling of differentially expressed microRNAs in arrhythmogenic right ventricular cardiomyopathy. *Sci. Rep.* 6:28101. doi: 10.1038/srep 28101

Zhang, Y., Huang, X. R., Wei, L. H., Chung, A. C., Yu, C. M., and Lan, H. Y. (2014). miR-29b as a therapeutic agent for angiotensin II-induced cardiac fibrosis by targeting TGF-beta/Smad3 signaling. *Mol. Ther.* 22, 974–985. doi: 10.1038/mt. 2014.25

Zhou, P., and Pu, W. T. (2016). Recounting cardiac cellular composition. *Circ. Res.* 118, 368–370. doi: 10.1161/CIRCRESAHA.116.308139

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Ranjan, Kumari and Verma. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.