Review Articular cartilage and changes in arthritis Noncollagenous proteins and proteoglycans in the extracellular matrix of cartilage

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Abstract

Cartilage contains numerous noncollagenous proteins in its extracellular matrix, including proteoglycans. At least 40 such molecules have been identified, differing greatly in structure, distribution, and function. Some are present in only selected cartilages or cartilage zones, some vary in their presence with a person's development and age, and others are more universal in their expression. Some may not even be made by the chondrocytes, but may arise by absorption from the synovial fluid. In many cases, the molecules' function is unclear, but the importance of others is illustrated by their involvement in genetic disorders. This review provides a selective survey of these molecules and discusses their structure, function, and involvement in inherited and arthritic disorders.

Keywords: cartilage, extracellular matrix, protein, proteoglycan

Introduction

The extracellular matrix of articular cartilage contains a large variety of noncollagenous proteins. Many of these are listed in Table 1, and while this list is by no means exhaustive, it does include those that have been studied in the most detail. It is impossible to give any common feature that unites this group of molecules, as they vary greatly in structure and function, and in some cases it is not clear that they are even made by the chondrocytes. Many of the molecules are proteoglycans, bearing glycosaminoglycan chains, whereas others are glycoproteins or even nonglycosylated proteins. Some of the molecules represent degradation products of larger precursors that accumulate because of their interaction with other matrix components. Many of the molecules play a structural role, whereas others may be involved in regulating cell function. In addition, many of the molecules vary in their abundance

and structure with anatomical site or the person's age, and many are not unique to cartilage. The importance of many of the molecules to cartilage function is illustrated in Table 2, which shows their association with pathology when they are produced in a mutant form.

Proteoglycans of the cartilage extracellular matrix

Aggregating proteoglycans

Among the noncollagenous proteins of cartilage, aggrecan has undoubtedly received the greatest attention, because of its high abundance in cartilage, its close association with the ability of the tissue to resist compression, and its modification in many cartilage disorders. Aggrecan belongs to the family of aggregating proteoglycans that form large, multimolecular complexes with hyaluronan [1]. The family also includes versican, neurocan, and brevican,

CILP = cartilage intermediate-layer protein; CMP = cartilage matrix protein; COMP = cartilage oligomeric matrix protein; CS = chondroitin sulfate; CS1/CS2 = chondroitin-sulfate-attachment regions of aggrecan; G1/G2/G3 = globular regions [of aggrecan]; IL-1 = interleukin-1; PRELP = proline- and arginine-rich end leucine-rich repeat protein; SLRP = small leucine-rich repeat proteoglycan.

Table 1

Proteoglycans (PGs) and proteins of the cartilage extracellular matrix

Proteoglycans	Proteins
Aggregating Aggrecan Versican Link protein Leucine-rich repeat Biglycan (DS-PGI) Decorin (DS-PGII) Epiphycan (DS-PGIII) Fibromodulin Lumican Other Perlecan SZP/Lubricin	Structural COMP (Thrombospondin-5) Thrombospondin-1 and -3 CMP (Matrilin-1) Matrilin-3 CILP C-type lectin Fibronectin PRELP Chondroadherin Tenascin-C Fibrillin Elastin Regulatory gp-39/YKL-40 Matrix gla protein/ MGP Pleiotrophin Chondromodulin-I/SCGP Chondromodulin-II CD-RAP Growth factors Other Chondrocalcin PARP Lysozyme Phospholipase A2 Proteinases and inhibitors

CD-RAP = cartilage-derived retinoic acid responsive protein; CILP = cartilage intermediate layer protein; CMP = cartilage matrix protein; COMP = cartilage oligomeric matrix protein; DS-PG(I, II, III) = dermatan sulfate proteoglycan (I, II, III); gla = gamma-carboxyglutamic acid; gp = glycoprotein; PARP = proline- and arginine-rich protein; PRELP = proline- and arginine-rich end leucine-rich repeat protein; SZP = superficial zone protein.

though of these only versican has been shown to be expressed in cartilage, and at much lower levels than aggrecan. All the members of the family have an amino-terminal globular domain, which is responsible for interaction with hyaluronan, and a carboxy-terminal globular domain, which has lectin-like homology. These features have resulted in the family being termed hyalectans or lecticans.

Aggrecan has an additional globular domain (G2) that is separated from the amino-terminal globular domain (G1) by a short, interglobular domain [2]. The G2 domain is separated from the carboxy-terminal globular domain (G3) by a keratan sulfate attachment domain and two chondroitin sulfate (CS) attachment domains (CS1 and CS2). Over 100 CS and keratan sulfate chains may be present in the three glycosamino-glycan attachment domains, though it is not clear at present whether all potential attachment sites are always occupied or whether variation may occur among individuals. The high CS and keratan sulfate content of aggrecan and its ability to interact with hyaluronan are essential features for normal articular

Table 2

Genetic disorders and the mutant cartilage matrix proteoglycans and proteins with which they are associated

Mutant matrix molecule	Associated disorder ¹
Aggrecan (impaired core protein synthesis)	Cartilage matrix deficiency (in mice) Nanomelia (in chickens) Brachymorphism (in mice)
Aggrecan (impaired glycosaminoglycan sulfation)	Diastrophic dysplasia Achondrogenesis 1B Atelosteogenesis type II
Perlecan	Chondrodystrophic myotonia Dyssegmental dysplasia
SZP	Camptodactyly-arthropathy- coxa vara-pericarditis syndrome
COMP	Pseudoachondroplasia Multiple epiphyseal dysplasia

¹Human disorder unless indicated otherwise. COMP = cartilage oligomeric protein; SZP = superficial zone protein.

cartilage function, as they provide the rheological properties necessary for resisting compression. The function of the G3 domain of aggrecan is unclear. Its lectin-like properties suggest the possibility of interaction with other components of the extracellular matrix [3], though it has also been suggested that it is involved in intracellular trafficking during aggrecan synthesis. Mutations in the aggrecan gene that prevent core protein synthesis form the basis of chondrodysplasias in mice (cartilage matrix deficiency) and chicks (nanomelia) [3]. In addition, impaired glycosaminoglycan sulfation on aggrecan causes the chondrodysplastic phenotypes associated with the brachymorphic mouse and diastrophic dysplasia in humans.

An interesting feature of the human aggrecan gene is the existence of polymorphism in the region encoding the CS1 domain. This region is composed of repeat sequences, which may range in number from 13 to 33 [4]. Individuals with the shortest alleles will have the lowest proportion of CS on their aggrecan molecules, and may be at risk for cartilage degeneration due to impaired aggrecan function. Irrespective of such polymorphism, the glycosaminoglycan composition of aggrecan varies considerably during juvenile development, as both the size and sulfation pattern of the CS and keratan sulfate change, though the functional consequence of this change is unclear. In addition, size heterogeneity is generated in the aggrecan core protein by the action of proteinases, with those fragments bearing a G1 domain being selectively retained in the tissue matrix. Proteolysis ultimately results in the accumulation of free G1 domains that have a long half-life in the tissue [5]. Many proteinases are able to degrade aggrecan if they gain access to the cartilage matrix, but most physiological and pathological degradation of articular cartilage is associated with the action of matrix metalloproteinases and aggrecanases [6]. Degradation products resulting from the action of both classes of proteinase accumulate in the synovial fluid of patients with arthritis [7,8] and have been used as markers of tissue destruction. Aggrecan synthesized in the arthritic joint has a CS sulfation pattern more akin to that in the normal juvenile than the normal adult. The appearance of this immature CS structure has also been used as a marker of the arthritic joint, and in particular of the reparative process that is being mounted. The G1-containing aggrecan fragments that accumulate with age or tissue degeneration may play a role in the induction of an autoimmune polyarthritis in susceptible individuals [9].

The interaction of aggrecan with hyaluronan is stabilized by the presence of link proteins. As with aggrecan, these proteins undergo proteolytic modification throughout life and can be used as an indicator of proteinase action. They provide evidence of the action of matrix metalloproteinase throughout juvenile development, and the participation of additional agents in the adult [10,11]. The link proteins are not susceptible to cleavage by the aggrecanase produced under cytokine stimulation of cartilage [12], and there is no evidence that any of the proteolytically modified link proteins have impaired function. Link protein can be lost from the cartilage matrix during periods of tissue degeneration, but such loss is most likely due to depolymerization of hyaluronan and involves concomitant loss of aggrecan. The importance of link protein in proteoglycan aggregate function is demonstrated by the impaired cartilage development observed in the link-protein-null mouse [13].

Small leucine-rich repeat proteoglycans

The small leucine-rich repeat proteoglycans (SLRPs) are characterized by a central domain composed of a series of adjacent leucine-rich repeats bordered at each end by disulfide-bonded domains [1]. The family may be divided into two subfamilies, depending on the presence of dermatan sulfate chains or keratan sulfate chains. Human cartilage has been shown to contain three dermatan sulfate proteoglycans (also called DS-PGs) - biglycan (DS-PGI), decorin (DS-PGII), and epiphycan (DS-PGIII) and in all of these, the dermatan sulfate chains are in the amino-terminal region of the core proteins. Only decorin and biglycan have been found in articular cartilage, and they are present throughout life. Whereas decorin remains in its intact form at all ages, biglycan exhibits age-related proteolytic processing that results in removal of the aminoterminal region bearing the dermatan sulfate chains. Such nonglycanated biglycan accumulates in the cartilage matrix with age, but it is not clear whether this is of any functional consequence [14]. Decorin and biglycan also have short, amino-terminal propeptides that are removed in the extracellular matrix by procollagen-C proteinase, the same enzyme responsible for removing the carboxy propeptide from type II collagen. Propeptide removal is

incomplete in adult cartilage [15], but again, the functional consequence, if any, is unclear.

Human articular cartilage contains two potential keratan sulfate proteoglycans, fibromodulin and lumican. Like decorin and biglycan, fibromodulin is present in articular cartilage throughout life, though it contains keratan sulfate chains only in the fetus and juvenile [16]. In the adult, it exists as a glycoprotein devoid of keratan sulfate. In contrast, lumican is not present in articular cartilage of the fetus or young juvenile [17]; in the adult, it is present in predominantly a glycoprotein form. It is unclear whether the presence or absence of keratan sulfate influences the function of these proteoglycans in cartilage. All SLRPs have all been shown to interact with the fibrillar collagens of the extracellular matrix, though their site and strength of interaction may vary. The importance of these molecules in matrix organization is illustrated by the abnormalities associated with SLRP-null mice [18-21], though these abnormalities are perhaps less severe than might have been expected and it is possible that there is a functional redundancy between some family members. Unlike aggrecan, the SLRPs of the cartilage matrix appear relatively resistant to extensive proteolytic modification and do not show a ready sensitivity towards cytokine-induced damage [12]. Fragments have, however, been observed in the matrix of arthritic cartilage.

Other proteoglycans

The cartilage matrix also contains the proteoglycan perlecan. This is somewhat surprising, because perlecan is commonly thought of as a basement membrane proteoglycan [1], yet articular cartilage is devoid of basement membranes. Basement membrane perlecan is characterized by the presence of heparan sulfate chains in its amino-terminal region, though it has been reported that cartilage perlecan may exist in a nonglycanated form [22]. The perlecan core protein is extremely large and might be expected to be a good candidate for proteolytic processing, but at present there is no information available on structural changes with either age or arthritis. The importance of perlecan to cartilage function is demonstrated by the perlecan-null mouse [23], in which severe chondrodysplasia is a major part of the phenotype in addition to basement membrane defects affecting heart and brain development. In the human, mutations in the perlecan gene have been associated with Schwartz-Jampel syndrome (chondrodystrophic myotonia) [24], and have recently been reported in dyssegmental dysplasia. At present, the function of perlecan in cartilage, and in particular in the growth plates, is unknown.

A final proteoglycan associated with cartilage has been termed superficial zone protein [25]. It is synthesized by the superficial chondrocytes of articular cartilage and by synoviocytes, and has an attachment site for a CS chain. It is identical to the precursor protein of megakaryocytestimulating factor, and probably is the same as a protein originally described as lubricin, which is responsible for the lubrication and frictionless motion of the cartilage surface. While some superficial zone protein may be retained in the extracellular matrix, most is destined for secretion into the synovial cavity. The synthesis of this protein is impaired in the arthritic joint, where alternative splicing has been reported, and production is downregulated by the presence of inflammatory cytokines such as IL-1. Gene defects in this protein have been associated with camptodactyly-arthropathy-coxa vara-pericarditis syndrome [26]. In addition to its role as a lubricant, the protein may play a role in regulating synovial cell proliferation, as this syndrome and various forms of arthritis are associated with synovial hyperplasia. In the case of camptodactyly-arthropathy-coxa vara-pericarditis syndrome, hyperplasia occurs in the absence of inflammation.

Proteins of the cartilage extracellular matrix Structural proteins

The extracellular matrix of cartilage contains numerous proteins that are neither collagens nor proteoglycans [27], and several of these are thought to play a structural role in the matrix. Cartilage oligomeric matrix protein (COMP) is perhaps the best studied of these proteins. It belongs to the thrombospondin family and has been termed thrombospondin-5, and is structurally more closely related to thrombospondins 3 and 4 than to thrombospondins 1 and 2 [28]. Other members of the thrombospondin family have been detected in cartilage, though not at the same level or widespread distribution as COMP. This protein is present in all cartilages, being most abundant in growth plate during development, but also in mature articular cartilage. It exists as a disulfide-bonded pentamer linked near its amino-terminal region, and the projecting carboxy-terminal regions are suggested to interact with collagen. The need for COMP in cartilage is best illustrated by the presence of pseudoachondroplasia or multiple epiphyseal dysplasia in individuals bearing a mutation in the COMP gene [29]. A phenotype of multiple epiphyseal dysplasia can also arise by mutations in a type IX collagen gene, and this may indicate an association between COMP and type IX collagen. During cartilage turnover, COMP undergoes degradation, and fragments are released into the synovial fluid. An increase in such fragments has been observed in the synovial fluid of patients suffering from joint trauma and those in the early stages of primary osteoarthritis [30], and it has been suggested that elevated levels of COMP in synovial fluid may serve as a marker of such disorders.

Cartilage matrix protein (CMP) is also thought to serve a structural role in the extracellular matrix [31]. It belongs to the matrilin family and has also been termed matrilin-1. Matrilin-3 has also been detected in some cartilages. CMP exists in the cartilage matrix as a disulfide-bonded trimer,

joined near the carboxy terminus of its subunits. While CMP is present in skeletal cartilages during development, it is most abundant in extraskeletal cartilages in the adult and is deficient in articular cartilage. This protein is known to interact with both type II collagen and aggrecan, though its precise function remains unclear. Indeed, CMP-null mice do not exhibit any obvious skeletal phenotype and appear to develop normally [32], which may imply a functional redundancy between CMP and matrilin-3. Although CMP is not detected in normal articular cartilage, it is produced by the chondrocytes of arthritic cartilage [33].

Articular cartilages have a matrix protein that is most abundant in the mid-zone of the tissue but deficient in the deepest and superficial zones [34]. On the basis of this localization, the protein has been termed cartilage intermediate-layer protein (CILP). CILP is more abundant in adult than in juvenile articular cartilage, but the relevance of the site- and age-related distribution to function is unknown. CILP production has also been reported to be increased in osteoarthritic cartilage. Interestingly, the transcript from the CILP gene encodes two proteins. The amino-terminal portion of the message encodes CILP, while the carboxyterminal portion encodes nucleotide pyrophosphohydrolase (NTPPHase) [35]. The initial translation product contains both proteins, which are separated by proteolytic cleavage within the chondrocytes. The relevance of this phenomenon and the function of CILP are at present unknown, and CILP does not appear to have a close structural relationship to any other protein yet described.

Other structural proteins are thought to be involved in cell-matrix interactions rather than matrix-matrix interactions. Among these, fibronectin deserves particular mention. Fibronectin is present in many tissues and exists as a disulfide-bonded dimer joined at the carboxy terminus of its subunits [36]. Fibronectin can exist in multiple isoforms, because of alternative splicing of its gene, and chondrocytes appear to produce a characteristic splice variant [37]. The abundance of fibronectin increases about 10-fold in osteoarthritic cartilage [38], though the functional significance of this is unclear. However, it is interesting that fibronectin fragments, resulting from proteolytic degradation, are able to propagate degradation of aggrecan at the same sites as expected for the action of aggrecanase [39]. It has been suggested that the fibronectin fragments that may accumulate in the arthritic joint may stimulate the local production of inflammatory cytokines, such as IL-1, that upregulate aggrecanase expression.

Another molecule of interest is proline- and arginine-rich end leucine-rich repeat protein (PRELP), which is closely related in protein structure and gene organization to fibromodulin and lumican but is devoid of keratan sulfate chains. PRELP shows selective distribution among cartilagenous tissues and is not present in fetal and young juvenile human cartilage [40]. The unique amino-terminal region of PRELP may facilitate interaction with heparan sulfate proteoglycans on cell membranes [41]. A final protein worthy of note is chondroadherin, which bears neither glycosaminoglycan chains nor N-linked oligosaccharides and, in common with elastin, may be devoid of carbohydrate. Chondroadherin also belongs to the family of leucine-rich repeat proteins [42] and, in common with PRELP, is thought to play a role in mediating cell-matrix interactions.

Regulatory proteins

Several proteins in the extracellular matrix are thought to influence cell proliferation or metabolism rather than play a structural role in the matrix (see Table 1), but a discussion of their properties is beyond the scope of this review. However, one of these proteins, termed gp-39, deserves special recognition. It is related to the chitinase family but does not have enzymic activity. It is not detected in normal articular cartilage, but is produced by chondrocytes in culture and is present in arthritic cartilage [43]. As such, it may reflect situations in which rapid tissue remodelling is occurring and may be indicative of the capacity of chondrocytes to recognize an abnormal environment and initiate a repair response.

Other proteins

This category includes proteinases and their inhibitors, degradation products of collagen, and basic proteins that associate with the extracellular matrix. Two products of collagen degradation have been reported to accumulate in cartilage [27]. One is chondrocalcin, which represents the carboxy-propeptide of type II collagen, and the second is proline-arginine-rich protein, which represents the aminopropeptide domain of the $\alpha 2(XI)$ chain of type XI collagen. It is possible that these molecules are not merely innocent bystanders but are involved in feedback regulation of collagen synthesis. The abundance of chondrocalcin in cartilage is often used as an indication of new collagen synthesis. Finally, lysozyme [44] and phospholipase A2 [45] are worthy of mention. Both are cationic proteins that may owe their presence in the cartilage matrix to the high content of anionic aggrecan. In the case of lysozyme, it is likely that much of it is not produced by the chondrocytes but rather is absorbed from the synovial fluid.

Conclusion

It is apparent from this brief review that the extracellular matrix of cartilage contains many noncollagenous proteins and proteoglycans whose precise functions are only just beginning to be understood. These molecules may serve a structural or regulatory role, and in some cases may do both, as degradation products of some of the structural molecules are known to influence the chondrocyte. The recognition of genetic disorders in which synthesis of the matrix molecules is perturbed has aided greatly in our understanding of their functional role, but the reason for many site- and age-related restrictions in expression remains unclear. The role of many of the molecules in the arthritic joint is also unclear, as in many cases they may be pawns of the disease, undergoing destruction, yet in others they may be actively involved in propagating destruction or initiating repair. This is an area where there is still a wealth of information to be mined.

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References

- 1. lozzo RV: Matrix proteoglycans: from molecular design to cellular function. Annu Rev Biochem 1998, 67:609-652.
- Watanabe H, Yamada Y, Kimata K: Roles of aggrecan, a large chondroitin sulfate proteoglycan, in cartilage structure and function. J Biochem 1998, 124:687-693.
- 3. Vertel B: The ins and outs of aggrecan. *Trends Cell Biol* 1995, 5:458-464.
- Doege KJ, Coulter SN, Meek LM, Maslen K, Wood JG: A humanspecific polymorphism in the coding region of the aggrecan gene. Variable number of tandem repeats produce a range of core protein sizes in the general population. *J Biol Chem* 1997, 21:13974-13979.
- Maroudas A, Bayliss MT, Uchitel-Kaushansky N, Schneiderman R, Gilav E: Aggrecan turnover in human articular cartilage: use of aspartic acid racemization as a marker of molecular age. Arch Biochem Biophys 1998, 350:61-71.
- Caterson B, Flannery CR, Hughes CE, Little CB: Mechanisms involved in cartilage proteoglycan catabolism. *Matrix Biol* 2000, 19:333-344.
- Lohmander LS, Neame PJ, Sandy JD: The structure of aggrecan fragments in human synovial fluid. Evidence that aggrecanase mediates cartilage degradation in inflammatory joint disease, joint injury, and osteoarthritis. *Arthritis Rheum* 1993, 9:1214-1222.
- Fosang AJ, Last K, Maciewicz RA: Aggrecan is degraded by matrix metalloproteinases in human arthritis. Evidence that matrix metalloproteinase and aggrecanase activities can be independent. J Clin Invest 1996, 98:2292-2299.
- Glant TT, Cs-Szabó G, Nagase H, Jacobs JJ, Mikecz K: Progressive polyarthritis induced in BALB/c mice by aggrecan from normal and osteoarthritic human cartilage. *Arthritis Rheum* 1998, 41:1007-1018.
- Nguyen Q, Liu J, Roughley PJ, Mort JS: Link protein as a monitor in situ of endogenous proteolysis in adult human articular cartilage. *Biochem J* 1991, 278:143-147.
- Hughes CE, Caterson B, White RJ, Roughley PJ, Mort JS: Monoclonal antibodies recognizing protease-generated neoepitopes from cartilage proteoglycan degradation. Application to studies of human link protein cleavage by stromelysin. J Biol Chem 1992, 267:16011-16014.
- Sztrolovics R, White RJ, Poole AR, Mort JS, Roughley PJ: Resistance of small leucine-rich repeat proteoglycans to proteolytic degradation during interleukin-1-stimulated cartilage catabolism. *Biochem J* 1999, 339:571-577.
- Watanabe H, Yamada Y: Mice lacking link protein develop dwarfism and craniofacial abnormalities. Nat Genet 1999, 21: 225-229.
- Roughley PJ, White RJ, Magny MC, Liu J, Pearce RH, Mort JS: Non-proteoglycan forms of biglycan increase with age in human articular cartilage. *Biochem J* 1993, 295:421-426.
- Roughley PJ, White RJ, Mort JS: Presence of pro-forms of decorin and biglycan in human articular cartilage. *Biochem J* 1996, **318**:779-784.
- Roughley PJ, White RJ, Cs-Szabó G, Mort JS: Changes with age in the structure of fibromodulin in human articular cartilage. Osteoarthritis Cart 1996, 4:153-161.
- 17. Grover J, Chen XN, Korenberg JR, Roughley PJ: The human lumican gene. Organization, chromosomal location, and expression in articular cartilage. *J Biol Chem* 1995, 270: 21942-21949.

- Xu T, Bianco P, Fisher LW, Longenecker G, Smith E, Goldstein S, Bonadio J, Boskey A, Heegaard AM, Sommer B, Satomura K, Dominguez P, Zhao C, Kulkarni AB, Robey PG, Young MF: Targeted disruption of the biglycan gene leads to an osteoporosis-like phenotype in mice. *Nat Genet* 1998, 20:78-82.
- Svensson L, Aszódi A, Reinholt FP, Fassler R, Heinegård D, Oldberg Å: Fibromodulin-null mice have abnormal collagen fibrils, tissue organization, and altered lumican deposition in tendon. J Biol Chem 1999, 14:9636-9647.
- Chakravarti S, Magnuson T, Lass JH, Jepsen KJ, LaMantia C, Carroll H: Lumican regulates collagen fibril assembly: skin fragility and corneal opacity in the absence of lumican. J Cell Biol 1998, 141:1277-1286.
- Iozzo RV, Cohen IR, Grassel S, Murdoch AD: The biology of perlecan: the multifaceted heparan sulphate proteoglycan of basement membranes and pericellular matrices. *Biochem J* 1994, 302:625-639.
- Costell M, Gustafsson E, Aszódi A, Morgelin M, Bloch W, Hunziker E, Addicks K, Timpl R, Fassler R: Perlecan maintains the integrity of cartilage and some basement membranes. *J Cell Biol* 1999, 147:1109-1122.
- Nicole S, Davoine CS, Topaloglu H, Cattolico L, Barral D, Beighton P, Hamida C, Hammouda H, Cruaud C, White PS, Samson D, Urtizberea JA, Lehmann-Horn F, Weissenbach J, Hentati F, Fontaine B: Perlecan, the major proteoglycan of basement membranes, is altered in patients with Schwartz-Jampel syndrome (chondrodystrophic myotonia). Nat Genet 2000, 26:480-483.
- Flannery CR, Hughes CE, Schumacher BL, Tudor D, Aydelotte MB, Kuettner KE, Caterson B: Articular cartilage superficial zone protein (SZP) is homologous to megakaryocyte stimulating factor precursor and is a multifunctional proteoglycan with potential growth-promoting, cytoprotective, and lubricating properties in cartilage metabolism. *Biochem Biophys Res Commun* 1999, 254:535-541.
- Marcelino J, Carpten JD, Suwairi WM, Gutierrez OM, Schwartz S, Robbins C, Sood R, Makalowska I, Baxevanis A, Johnstone B, Laxer RM, Zemel L, Kim CA, Herd JK, Ihle J, Williams C, Johnson M, Raman V, Alonso LG, Brunoni D, Gerstein A, Papadopoulos N, Bahabri SA, Trent JM, Warman ML: CACP, encoding a secreted proteoglycan, is mutated in camptodactyly-arthropathy-coxa vara-pericarditis syndrome. Nat Genet 1999, 23:319-322.
- Neame PJ, Tapp H, Azizan A: Noncollagenous, nonproteoglycan macromolecules of cartilage. *Cell Mol Life Sci* 1999, 55:1327-1340.
- Bornstein P, Sage EH: Thrombospondins. Meth Enzymol 1994, 245:62-85.
- Briggs MD, Hoffman SM, King LM, Olsen AS, Mohrenweiser H, Leroy JG, Mortie GR, Rimoin DL, Lachman RS, Gaines ES, Cekleniak JA, Knowlton RG, Cohn DH: Pseudoachondroplasia and multiple epiphyseal dysplasia due to mutations in the cartilage oligomeric matrix protein gene. Nat Genet 1995, 10:330-336.
- Lohmander LS, Saxne T, Heinegård DK: Release of cartilage oligomeric matrix protein (COMP) into joint fluid after knee injury and in osteoarthritis. Ann Rheum Dis 1994, 53:8-13.
- Jenkins RN, Osborne-Lawrence SL, Sinclair AK, Eddy RL, Byers MG, Shows TB, Duby AD: Structure and chromosomal location of the human gene encoding cartilage matrix protein. J Biol Chem 1990, 265:19624-19631.
- Aszódi A, Bateman JF, Hirsch E, Baranyi M, Hunziker EB, Hauser N, Bösze Z, Fassler R: Normal skeletal development of mice lacking matrilin 1: redundant function of matrilins in cartilage? *Mol Cell Biol* 1999, **19**:7841-7845.
 Okimura A, Okada Y, Makihira S, Pan H, Yu L, Tanne K, Imai K,
- Okimura A, Okada Y, Makihira S, Pan H, Yu L, Tanne K, Imai K, Yamada H, Kawamoto T, Noshiro M, Yan W, Kato Y: Enhancement of cartilage matrix protein synthesis in arthritic cartilage. *Arthritis Rheum* 1997, 40:1029-1036.
- Lorenzo P, Bayliss MT, Heinegård D: A novel cartilage protein (CILP) present in the mid-zone of human articular cartilage increases with age. *J Biol Chem* 1998, 273:23463-23468.
- 35. Lorenzo P, Neame P, Sommarin Y, Heinegård D: Cloning and deduced amino acid sequence of a novel cartilage protein

(CILP) identifies a proform including a nucleotide pyrophosphohydrolase. *J Biol Chem* 1998, **273**:23469-23475.

- Romberger DJ: Fibronectin. Int J Biochem Cell Biol 1997, 29: 939-943.
- Burton-Wurster N, Borden C, Lust G, Macleod JN: Expression of the (V+C)⁻ fibronectin isoform is tightly linked to the presence of a cartilaginous matrix. *Matrix Biol* 1998, 17:193-203.
- Brown RA, Jones KL: The synthesis and accumulation of fibronectin by human articular cartilage. J Rheumatol 1990, 17: 65-72.
- Homandberg GA, Davis G, Maniglia C, Shrikhande A: Cartilage chondrolysis by fibronectin fragments causes cleavage of aggrecan at the same site as found in osteoarthritic cartilage. Osteoarthritis Cart 1997, 5:450-453.
- Grover J, Chen XN, Korenberg JR, Recklies AD, Roughley PJ: The gene organization, chromosome location, and expression of a 55-kDa matrix protein (PRELP) of human articular cartilage. *Genomics* 1996, 38:109-117.
- Bengtsson E, Aspberg A, Heinegård D, Sommarin Y, Spillmann D: The amino-terminal part of PRELP binds to heparin and heparan sulfate. J Biol Chem 2000, 275:40695-40702.
- 42. Grover J, Chen XN, Korenberg JR, Roughley PJ: The structure and chromosome location of the human chondroadherin gene (CHAD). *Genomics* 1997, 45:379-385.
- 43. Hakala BE, White C, Recklies AD: Human cartilage gp-39, a major secretory product of articular chondrocytes and synovial cells, is a mammalian member of a chitinase protein family. J Biol Chem 1993, 268:25803-25810.
- Moss JM, Van Damme MP, Murphy WH, Stanton PG, Thomas P, Preston BN: Purification, characterization, and biosynthesis of bovine cartilage lysozyme isoforms. Arch Biochem Biophys 1997, 339:172-182.
- 45. Recklies AD, White C: Phospholipase A2 is a major component of the salt-extractable pool of matrix proteins in adult human articular cartilage. *Arthritis Rheum* 1991, **34**:1106-1115.