## Pathophysiological mechanisms explaining poor clinical outcome of older cancer patients with low skeletal muscle mass

<sup>1</sup>Department of Human Movement Sciences, @AgeAmsterdam, Faculty of Behavioural and Movement Sciences, Vrije Universiteit Amsterdam, Amsterdam Movement Sciences, Amsterdam, The Netherlands

<sup>2</sup>University Library, Vrije Universiteit Amsterdam, Amsterdam, The Netherlands

<sup>3</sup>Department of Rehabilitation Medicine, Amsterdam University Medical Center, VU University Medical Center, Amsterdam Movement Sciences, Amsterdam, The Netherlands

<sup>4</sup>Department of Medicine and Aged Care, @AgeMelbourne, The Royal Melbourne Hospital, University of Melbourne, Melbourne, Australia

#### Correspondence

Andrea B. Maier, MD, PhD., Vrije Universiteit Amsterdam, Department of Human Movement Sciences, Van der Boechorststraat 7, 1081 BT Amsterdam, The Netherlands. Email: a.b.maier@vu.nl

#### **Funding information**

This work was supported by the European Union's Horizon 2020 research and innovation programme under the Marie-Sklodowska-Curie grant agreement no. 675003 (PANINI programme) and by an unrestricted grant of the University of Melbourne, Australia, received by Professor Andrea B. Maier. The funders had no role in the design and conduct of the study, data collection and analysis, interpretation of data, writing of the manuscript, or the decision to submit the article for publication.

#### Abstract

Low skeletal muscle mass is highly prevalent in older cancer patients and affects 5% to 89% depending on the type and stage of cancer. Low skeletal muscle mass is associated with poor clinical outcomes such as post-operative complications, chemotherapy toxicity and mortality in older cancer patients. Little is known about the mediating pathophysiological mechanisms. In this review, we summarize proposed pathophysiological mechanisms underlying the association between low skeletal muscle mass and poor clinical outcomes in older cancer patients including a) systemic inflammation; b) insulin-dependent glucose handling; c) mitochondrial function; d) protein status and; e) pharmacokinetics of anticancer drugs. The mechanisms of altered myokine balance negatively affecting the innate and adaptive immune system, and altered pharmacokinetics of anticancer drugs leading to a relative overdosage of anticancer drugs are best-substantiated. The effects of glucose intolerance and circulating mitochondrial DNA as a consequence of low skeletal muscle mass are topics of interest for future research. Restoring myokine balance through physical exercise, exercise mimetics, neuro-muscular activation and adapting anticancer drug dosing on skeletal muscle mass could be targeted approaches to improve clinical outcomes in older cancer patients with low skeletal muscle mass.

#### **KEYWORDS**

aged, cachexia, geriatric oncology, neoplasms, physiopathology, sarcopenia

\*Shared first authorship, both authors contributed equally.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 Scandinavian Physiological Society. Published by John Wiley & Sons Ltd

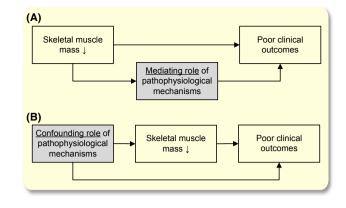
Acta Physiologica

#### 1 | INTRODUCTION

Ageing is associated with loss of skeletal muscle mass<sup>1</sup> and strength.<sup>2</sup> Sarcopenia is diagnosed if muscle mass and strength fall below a certain threshold.<sup>3</sup> Approximately 10% of the older population suffer from sarcopenia,<sup>4</sup> but the prevalence is higher in patients with cancer and other age-related diseases.<sup>5,6</sup> The prevalence rate of sarcopenia in patients with cancer was estimated at 38.6%,<sup>7</sup> varying between 5% and 89% depending on the type and stage of cancer<sup>8</sup> and on the applied diagnostic criteria for sarcopenia.<sup>9-11</sup> In cancer patients, sarcopenia can co-occur with cachexia, which is characterized by severe weight loss and loss of skeletal muscle and adipose tissue.<sup>12</sup> The prevalence of cachexia highly depends on the underlying disease, but between 50% and 80% of patients with advanced malignant cancers are thought to suffer from cachexia.<sup>13</sup> Although considered two separate diseases, the pathophysiology of sarcopenia and cachexia are overlapping, both are multifactorial and include a misbalance between lower protein synthesis and higher protein degradation because of an elevated intracellular inflammation and oxidative stress.14-16

Low skeletal muscle mass is often perceived as a biomarker for deprived fitness and health status, which can lower the resilience to stressors that accompany cancer and cancer treatment.<sup>17-22</sup> Low skeletal muscle mass in cancer patients has been associated with poor clinical outcomes including higher post-operative complication rates,<sup>7,23</sup> higher chemotherapy toxicity,<sup>7,8,23</sup> lower disease-free or progression-free survival<sup>7,8,23</sup> and higher overall mortality,<sup>7,8,23,24</sup> although associations are not considered to be straightforward.<sup>25</sup> Systemic inflammation, insulin-dependent glucose handling and alterations in energy- and protein metabolism and pharmacokinetics have been proposed as pathophysiological mechanisms explaining the association between low skeletal muscle mass and poor clinical outcomes in older patients with cancer.<sup>19,24,26</sup> If and how these mechanisms contribute to clinical outcomes is currently unknown. Understanding the pathophysiological consequences of low skeletal muscle mass on clinical outcomes and disease progression offers new directions for interventions in older cancer patients.

This review provides an overview and discussion of the described pathophysiological mechanisms in the literature that could underlie the association between low skeletal muscle mass and poor clinical outcomes in older cancer patients. We specifically focus on the pathophysiological consequences of low skeletal muscle mass as illustrated in the directed acyclic graph in Figure 1.<sup>27</sup> We will describe: a) the role of skeletal muscle mass to modulate the immune system through cytokines and myokines including the effects of physical activity; b) the influence of low skeletal muscle mass on insulin-dependent glucose handling and c) mitochondrial function; d) the effects on whole-body protein status; e) pharmacokinetics



**FIGURE 1** Pathophysiological mechanisms underlying the association between low skeletal muscle mass and poor clinical outcomes by mediation 1(a) and confounding 1(b) in older cancer patients using directed acyclic graphs

of anticancer drugs. Figure 2 provides an illustrated overview of the scope of the article. The literature search is presented in the Appendix. We conclude by exploring future directions for research and potential interventions that could decrease the risk of poor clinical outcomes in older cancer patients with low skeletal muscle mass.

#### 2 | OVERVIEW OF PATHOPHYSIOLOGICAL MECHANISMS

Figure 3 provides a summarized overview of the mechanisms that potentially play a role in the association between low skeletal muscle mass and poor clinical outcomes in older cancer patients. These mechanisms will be discussed extensively throughout this narrative review.

#### **3** | SYSTEMIC INFLAMMATION

Skeletal muscle fibres are able to actively shape the immune system in both a pro- and anti-inflammatory manner, regulating innate and adaptive immune responses.<sup>28,29</sup> In this way, low skeletal muscle mass directly contributes to chronic low-grade local and systemic inflammation.<sup>22,30,31</sup> Various clinical observational studies showed significant associations between low skeletal muscle mass and higher inflammatory markers, such as a higher neutrophil-tolymphocyte ratio and higher C-reactive protein levels, in older cancer patients.<sup>32-37</sup> This dose-response relation between low skeletal muscle mass and systemic inflammation was independent of cancer stage, age and sex.<sup>37</sup> These inflammatory markers are significantly associated with overall<sup>34,38-40</sup> and cancer-specific<sup>38,39</sup> mortality. Patients with a combination of low skeletal muscle mass and high

#### ACTA PHYSIOLOGICA

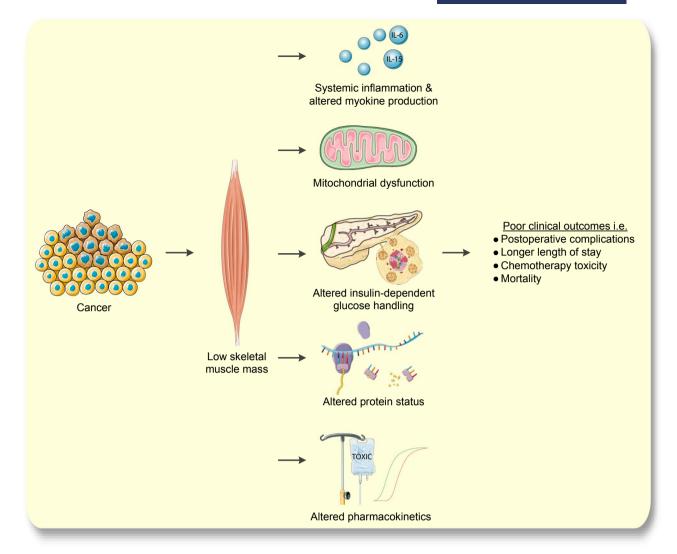
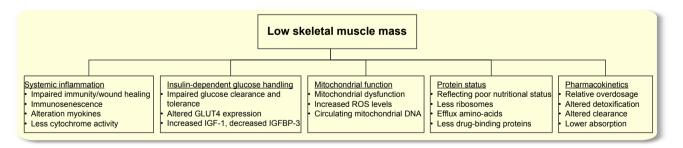


FIGURE 2 Theoretical framework and overview of the scope of the review



**FIGURE 3** Pathophysiological mechanisms and markers potentially underlying the association between low skeletal muscle mass and poor clinical outcomes in older cancer patients

inflammatory markers had higher mortality rates than patients with low skeletal muscle mass and low inflammatory markers.<sup>37,41</sup> Therefore, it remains elusive whether systemic inflammation is an additional or mediating mechanism for poor clinical outcomes in cancer patients. Figure 3 includes a summary of pathophysiological mechanisms caused by systemic inflammation that may explain the association between low skeletal muscle mass and poor clinical outcomes.

Patients with low skeletal muscle mass at the time of hospital admission have a doubled risk of nosocomial infections during the first weeks of hospitalization<sup>42</sup> and a higher risk of post-operative complications, requiring inpatient rehabilitation and longer hospital stay.<sup>43</sup> This could potentially be

## ACTA PHYSIOLOGICA

caused by impaired wound healing<sup>44</sup> or by the effect of low skeletal muscle mass on specific muscle function such as breathing or swallowing. For example atrophy and weakness in the diaphragm muscle could lead to respiratory dysfunction. The resulting decreased airflow and inability to fully inflate the lung and cough facilitates the development of pneumonia.45,46 Indeed, muscle wasting in colon-26-bearing mice caused significant atrophy in the diaphragm muscle, which resulted in a lower tidal volume and an inability to increase breathing frequency and tidal volume during a respiratory challenge.<sup>45</sup> Also, a low mass and function of muscles involved in swallowing can lead to dysphagia, which increases the risk of complications such as aspiration pneumonia.47,48 Furthermore, systemic inflammation is known to induce chemotherapy toxicity (see paragraph 'pharmacokinetics of anticancer drugs') and aggravate the already impaired immune function.44,49 Impaired immune function combined with increased inflammatory cytokines contributes to immunosenescence,<sup>29</sup> which could increase the risk of additional complications.<sup>50</sup> Increased tumour aggressiveness,<sup>37</sup> poor treatment response<sup>37</sup> and a higher risk of cancer development<sup>21</sup> have also been attributed to an increased inflammatory status.

# 3.1 | Myokine secretion and physical exercise

Based on the suggestion that skeletal muscle acts as an endocrine organ,<sup>51</sup> one of the predominant, most described and best-substantiated theories is that low skeletal muscle mass results in less myokine production. Myokines are small molecules released by contracting skeletal muscle, which can exert autocrine, paracrine and endocrine effects on other tissues.<sup>51</sup> More than 200 myokines have been discovered so far, but their individual functions are still mostly unknown. Overall, alterations in the balance between myokines and adipokines can negatively influence the innate and adaptive immune system.<sup>52</sup> In the context of exercise and cancer immunology, interleukin (IL)-15 and IL-6 have been studied extensively and modulate the innate and adaptive immune system.<sup>51</sup> IL-15 is involved in the regulation of natural killer cell number and activity and protects natural killer cells from apoptosis.<sup>29,52</sup> IL-15 knockout mice had almost no mature natural killer cells and natural killer cells were destroyed after being transferred into the same knockout mice.<sup>52-54</sup> Lower IL-15 release into the bloodstream as a consequence of low skeletal muscle mass has thus been proposed to lead to lower natural killer cell number and survival,<sup>52</sup> increasing the risk of infectious complications<sup>55</sup> and shortening survival<sup>56</sup> in cancer patients. Moreover, IL-15 is involved in CD8 T-cells homeostasis, the survival of naive T-cells and proliferation of B-cells.<sup>29,57</sup> The first clinical trial in patients with metastatic melanoma or renal cell cancer showed that infusions of IL-15 led to redistribution and hyperproliferation of natural killer cells and CD8 memory T-cells.<sup>57</sup> Although grade 3 toxicities were observed, lower dosages of IL-15 could safely be administered. This allows targeted interventions on myokine infusions to be tested as potential new strategies in anticancer treatment.<sup>57</sup>

IL-6 represents another myokine which is expressed in high levels in skeletal muscle tissue. IL-6 exerts pro-inflammatory effects in response to pathogens, including T-cell recruitment and promoting antibody production from B-cells.<sup>29</sup> As a myokine, IL-6 has been indicated to play an important role in the redistribution and infiltration of natural killer cells, thereby suppressing tumour growth.<sup>58</sup> As low skeletal muscle mass is related to lower levels of IL-6, low muscle mass could inhibit the suppression of tumour progression, worsening the prognosis of cancer patients.<sup>58-60</sup>

Research on the effects of prescribing exercise in oncological patients is rapidly expanding,<sup>61-63</sup> particularly after it was observed that voluntary running in tumour-bearing mice suppressed tumour growth, likely by enabling IL-6-sensitive natural killer cells to infiltrate tumour tissue.58,64 Physical exercise itself was proved to be the crucial factor to evoke the effects of IL-6 on tumour growth, as simply administering an IL-6 injection did not have similar repressing effects.<sup>58</sup> This suggests that likely a combination of currently known and unknown other myokines can explain these adaptations. As such, the main current hypothesis on how exercise prevents and suppresses the development of cancer is that exercise alters the host immune system, via exercise-induced factors (including myokines and other mobilizing serum factors) released in the bloodstream.<sup>65</sup> These positive effects of exercise on immune function have been corroborated in cancer patients, demonstrated by an increase in natural killer cell cytotoxic activity, lymphocyte proliferation and number of granulocytes after chronic aerobic and/or resistance exercise.<sup>66</sup>

Other myokines worth mentioning are IL-8 and myostatin. IL-8 expression is elevated in cancer cachexia<sup>67</sup> and higher IL-8 expression has a signalling role in the tumour microenvironment, it induces angiogenesis and stimulates tumour growth.<sup>67,68</sup> Myostatin, also known as growth differentiation factor 8, negatively correlates with muscle mass. In case of a muscle wasting disease, its expression is increased.<sup>67,69</sup> Myostatin might be secreted from primary tumours, but its precise role in tumour metabolism remains unknown.<sup>69</sup> Myokines such as myonectin, decorin and fibroblast growth factor 21 possibly also link muscle mass to cancer outcome, but are less studied in this context.<sup>67</sup>

Since exercise is accompanied by an increase in the blood concentration of a large number of myokines,<sup>70</sup> other myokines likely contribute as well. Indeed, oncostatin M<sup>65,71</sup> and

irisin<sup>65,72</sup> showed direct anti-proliferative effects on cancer cells in breast cancer cells. Osteonectin, also known as secreted protein acidic and rich in cysteine (SPARC), has similar effects in colon cancer cells.<sup>65,73</sup> Although the function of many myokines remains unknown, some of them have already shown to have therapeutic potential to (in)directly improve clinical outcomes in cancer patients.<sup>64,70</sup> Exerciseinduced alteration in immune function, likely through the secretion of myokines and other mobilizing serum factors, are potentially novel targets and represent promising new directions for treatment options for patients with cancer. Also, exercise mimetics such as musclin are currently receiving a lot of attention in the exercise physiology field. These 'exercise pills' could potentially be of great use for cancer patients who are unable to perform (strenuous) exercise.<sup>74</sup> Non-pharmacological interventions such as neuro-muscular electrical stimulation could also be given to patients who are unable to exercise. Two controlled studies in patients with advanced solid cancers showed that a twelve week program with two sessions of neuro-muscular electrical stimulation per week combined with individualized nutritional support, led to a significantly higher muscle mass and physical performance at the end of the intervention compared to the control group that only received individualized nutritional support.<sup>75,76</sup> These fields of research will likely receive major attention in the coming years.

It should be emphasised that other indirect exercise-induced adaptations may contribute as well to the observed effects. In this review, we highlight the role of skeletal muscle in the pathophysiology of clinical outcome measures, and it is likely that exercise-induced maintenance or increase in muscle mass and oxidative capacity per se contribute to understanding the underlying mechanisms. Indeed, a 16-week high-intensity exercise training intervention in breast cancer patients was found to maintain or increase muscle citrate synthase activity, size and capillarization of both slow-switch (type I) and fast-twitch glycolytic (type II) fibres.<sup>77</sup> Similar as during the process of ageing, mainly fast-twitch glycolytic fibres (type II) are lost during cancer cachexia and may even lead to a fast-to-slow fibre type shift.<sup>78</sup> These alterations were associated with self-reported fatigue, confirming the notion that factors independent of the immune system contribute to improved clinical outcome in cancer patients.<sup>77</sup> Engaging in physical exercise during systemic anticancer treatment can possibly limit the disruption that anticancer drugs cause on molecular signalling pathways.<sup>79</sup> Recent advances have suggested a role for HIF1 $\alpha$  in the development of cancer cachexia and exercise-induced alterations in skeletal muscle function, but more work is needed to fully understand these mechanisms.<sup>80</sup> Clearly, more experimental work is necessary to fully understand the contributing role for exercise in preventing and suppressing development of cancer growth in various types of cancer.

## 4 | INSULIN-DEPENDENT GLUCOSE HANDLING AND TUMOUR GROWTH

Skeletal muscle has a primary role in insulin-mediated glucose metabolism as it is the main target organ of insulin-dependent glucose uptake.<sup>81</sup> In the case of atrophying skeletal muscle, lipids accumulating in muscle tissue can induce glucose intolerance through insulin resistance.82 On the other hand, glucose intolerance and insulin resistance have long been recognized as a manifestation of cancer.<sup>83,84</sup> Insulin resistance was found to be associated with overall and cancer-specific survival<sup>85,86</sup> and post-operative complications.<sup>36</sup> Pathophysiological mechanisms caused by alterations in insulin-dependent glucose handling that may relate to poor clinical outcomes in cancer patients are summarized in Figure 3. Interestingly, the expression of the insulin-regulated glucose transporter, GLUT4, is reported to increase during anticancer drug treatment,<sup>79</sup> but is it unknown what the underlying mechanisms are of these alterations. Since tumour tissue is also known to take up glucose, a lower glucose clearance must be sought in alterations in insulin sensitivity in other organs.87

This whole-body insulin resistance might simply be because of a lower skeletal muscle mass in cachectic cancer patients,<sup>88-90</sup> but other factors likely contribute as well. For instance another mechanism by which low skeletal muscle mass causes and exacerbates insulin resistance is by altering the secretion of insulin sensitivity-regulating myokines.88 Insulin resistance could lead to increased levels of insulin-like growth factor 1 (IGF-1) and decreased levels of insulin-like growth factor-binding protein 3 (IGFBP-3).<sup>91</sup> Higher IGF-1 and lower IGFBP-3 levels are associated with disease progression in patients with prostate cancer.<sup>91-93</sup> Furthermore, a cachexia-related impaired glucose clearance from the blood allows more glucose to become available for uptake in tumour cells.<sup>94</sup> Since tumours often rely on glycolysis for cell survival and proliferation,<sup>94</sup> higher blood glucose levels could accelerate cancer growth and disease progression. Reducing blood glucose levels by caloric restriction or ketogenic diets have recently attracted attention in the literature,<sup>95</sup> with mixed results and opinions.<sup>96</sup> Clearly, such dietary interventions can accelerate the loss of skeletal muscle mass which would not be without consequences. Further research will be needed to clarify the role of low skeletal muscle mass-induced alterations in insulin resistance and insulin-like growth factors in the progression of cancer.

## **5** | MITOCHONDRIAL FUNCTION

A high skeletal mitochondrial function is generally associated with a higher endurance capacity and a lower sense of

## Acta Physiologica

fatigue during submaximal exercise. As a result, an impaired skeletal mitochondrial function can directly explain an increased feeling of fatigue in patients with cancer.<sup>97</sup> Cancer progression, as well as anticancer drugs are both known to negatively affect skeletal muscle mitochondrial function.97 Mitochondrial abnormalities are common in sarcopenia<sup>98,99</sup> and cancer cachexia.<sup>100-103</sup> In Figure 3 the potential pathophysiological mechanisms caused by mitochondrial dysfunction that may affect poor clinical outcomes in cancer patients are highlighted. In particular, disturbed mitochondrial dynamics, mitophagy and an impaired mitochondrial biogenesis are observed in cancer cachexia, all reducing oxidative phosphorylation capacity and increasing reactive oxygen species (ROS) production.<sup>101</sup> These processes likely contribute to the development of muscle wasting in patients with cancer.<sup>100</sup> At the same time, various anticancer drugs are known to non-specifically induce skeletal muscle mitochondrial dysfunction.<sup>104</sup> For instance doxorubicin is known to accumulate inside mitochondria and induces mitochondrial complex I dysfunction, reducing adenosine-5'-triphosphate (ATP) synthesis rates and producing ROS,<sup>102,104</sup> ultimately reducing muscle size and function by DNA damage, protein oxidation and apoptosis.<sup>101</sup> Other chemotherapeutics have similar effects,<sup>97</sup> and can modulate mitochondrial DNA (mtDNA). Clearly, the combination of cancer and current anticancer therapies induces mitochondrial damage and ultimately leads to a vicious circle further deteriorating skeletal muscle mass and function.<sup>101</sup>

More recent evidence hints to an additional role of mitochondria in the pathophysiology of skeletal muscle wasting-induced cancer progression. When mitochondria are defective and are broken down during mitophagy, fragments of mtDNA can be found in the circulation. A high level of circulating mtDNA is linked to a faster cancer progression and poor survival of patients with ovarian cancer.<sup>105,106</sup> It remains unclear whether these mtDNA fragments come from the tumour itself or from non-tumour tissue, although recent evidence hints towards the latter.<sup>107</sup> As skeletal muscle tissue is rich in mitochondria, skeletal muscle wasting might be a source of circulating mtDNA.<sup>108</sup> The underlying molecular mechanism is currently unknown, but two options are plausible. The first one is that a high level of circulating mtDNA serves as a biomarker for high muscle breakdown rates and severe cachexia. Hence, the poor survival rates linked to high circulating mtDNA can be explained by complications because of high muscle breakdown rates. An alternative mechanism is that circulating mtDNA (and other mitochondria-derived molecules) can act as damage-associated molecular pattern (DAMP) molecules and therefore affect distant organ function, including immune function.<sup>109</sup> Circulating mtDNA can activate neutrophil and platelet responses facilitating tumour metastasis and obstructing anti-tumour immunity.<sup>110</sup> This field is vastly unknown and future research will be required to elucidate the underlying mechanisms, clinical contribution and therapeutic potential.

#### 6 | LOW PROTEIN STATUS AND POOR NUTRITIONAL STATUS

An important contributing mechanism to the development of low skeletal muscle mass is protein status alteration. Muscle protein synthesis rate is determined by the overall health status, nutrient availability and physical activity.<sup>111</sup> Low nutrient intake and low levels of muscle activation lead to decreased protein anabolism and increased protein catabolism, which negatively affect skeletal muscle mass in animal models<sup>112</sup> and in human research studies.<sup>111</sup> In case of low skeletal muscle mass and low muscle activation, protein synthesis and function are repressed.<sup>113</sup> The effects of muscle activation are further described in the paragraph on 'myokine secretion and physical activity'. Clinical studies quantifying protein status by albumin levels, have established hypoalbuminaemia to be associated with measures of sarcopenia,114 post-operative complications and longer length of hospital stay.<sup>115,116</sup> As low skeletal muscle mass is also predictive of post-operative complications and overall survival independent of albumin status,<sup>50,55,117</sup> the mediating role of overall protein status in the association between low skeletal muscle mass and poor clinical outcomes in cancer patients is not conclusive. Potential explaining pathophysiological mechanisms are summarized in Figure 3.

It is widely accepted that low protein status is a reflection of a poor nutritional status, which is prognostic for poor clinical outcomes in cancer patients.<sup>50,117,118</sup> Questions have been posed whether serum albumin levels are a proper marker of nutritional status because of the low diagnostic accuracy.<sup>117,119</sup> On the other hand, protein synthesis occurs in the liver where ribosomes are most predominantly present, but also takes place in skeletal muscle fibres.<sup>120</sup> Hence, low skeletal muscle mass is accompanied by fewer ribosomes, leading to lower absolute protein synthesis rates,<sup>121</sup> which might have negative systemic effects and influence clinical outcomes. Another theory is that breakdown of muscle proteins leads to efflux of stored amino acids into the bloodstream,<sup>36</sup> which then becomes available for take-up by the tumour to promote tumour growth.<sup>122</sup> Moreover, low protein status affects the risk of chemotherapy toxicity (see paragraph 'pharmacokinetics of anticancer drugs').

#### 7 | PHARMACOKINETICS OF ANTICANCER DRUGS

Pharmacokinetics play an important role in patients with cancer since the majority of patients are treated with systemic

therapies such as chemotherapy. Pathophysiological mechanisms caused by an alteration in pharmacokinetics of anticancer drugs because of low skeletal muscle mass that may increase the risk of poor clinical outcomes in cancer patients are highlighted in Figure 3. Over the past decades, dosing of anticancer drugs such as chemotherapy has been based on total body surface area, a constitute of body weight and height.<sup>123</sup> As basing dosage on body surface area did not reduce interpatient variability in drug clearance<sup>124</sup> or the prevalence of dose-limiting toxicity,<sup>125</sup> it has been questioned whether body surface area is the appropriate measure to determine drug dosage. Dosing chemotherapy protocols based on body surface area led to a higher dosage of chemotherapy per kilogram lean body or skeletal muscle mass, which in turn was associated with chemotherapy toxicity.<sup>126,127</sup> The so-called 'overdosage hypothesis' states that basing treatment dosage on body surface area leads to a relative overdosing of treatment in patients with low skeletal muscle mass because of a lower area and volume of distribution of drugs,<sup>26</sup> and has been recalled by many others in the oncological field.<sup>19,22,36,126,127</sup> Therefore, lean body mass has been suggested to be used to individualize treatment dosage. This is of even more importance in hydrophilic agents that are mainly metabolized and distributed in lean tissue.<sup>26</sup> In addition, detoxification pathways of specific chemotherapeutics partly occur in skeletal muscles. For example anthracyclines such as doxorubicin are metabolized in the electron transport chains of mitochondria which are present in high concentrations in skeletal muscle tissue.<sup>104</sup> The level of sequestering of doxorubicin in skeletal muscle influences its systemic availability and rate and amount of detoxification.<sup>128</sup>

Next to the decreased distribution of chemotherapeutics, clearance might be altered in cancer patients with low skeletal muscle mass. Patients with low skeletal muscle mass were found to have a higher area under the curve (AUC) and lower plasma clearance of multiple chemotherapeutics compared to patients with normal skeletal muscle mass.<sup>36,126,127</sup> Patients with low skeletal muscle mass and low clearance also had a higher risk of chemotherapy toxicity.<sup>129,130</sup> On the other hand, the association between skeletal muscle mass and plasma clearance<sup>131-133</sup> and the association between plasma clearance and chemotherapy toxicity<sup>20,132,133</sup> could not always be confirmed. As the current body of literature shows inconsistencies, further research investigating the link between altered clearance and the association between low skeletal muscle mass and poor clinical outcomes in cancer patients is necessary.

Another process of pharmacokinetics is the absorption of anticancer drugs. Low skeletal muscle mass in cancer patients is accompanied by an increase in permeability of the gut barrier, causing a leakage of endotoxins into the systemic circulation evoking a low-grade systemic inflammatory response.<sup>134,135</sup> Moreover, anticancer drugs could cause the tight junctions in the intestinal tissues to become weaker and therewith further induce gut barrier dysfunction.<sup>134</sup> The resulting increase in leakage of anticancer drugs into intestinal tissues and the systemic circulation might increase the risk of toxicity of anticancer drugs.<sup>36,134</sup>

Other roles of how low skeletal muscle mass affects pharmacokinetics are via inflammation and overall protein status. The low-grade inflammatory state that accompanies low skeletal muscle mass leads to a decrease in liver cytochrome activity.<sup>136-138</sup> The resulting lower metabolic capacity of the liver increases the exposure to chemotherapeutics and causes toxicity.<sup>19,20,36,130</sup> Because of a lower skeletal muscle mass, less skeletal muscle proteins might be available for potential protein-binding of chemicals, also increasing exposure to chemotherapeutics and the risk of toxicity.<sup>20,36,130,131,139</sup> In addition, the concentration and activity of dihydropyrimidine dehydrogenase (DPD) are thought to decrease as a consequence of low protein status. Particular chemotherapeutics that are metabolized by DPD, such as 5-fluorouracil, could consequently accumulate in the bloodstream, leading to increased toxicity.<sup>36</sup> Countering an influence of low protein status on pharmacokinetics and risk of toxicity, low protein levels were not associated with more unbound chemotherapeutic in patients with hepatic dysfunction.<sup>140</sup> However, apart from low protein serum levels, low skeletal muscle mass itself could contribute to less drug-binding and higher exposure to anticancer drugs as protein-binding also occurs in skeletal muscle tissue.<sup>131</sup>

Skeletal muscle mass was predominantly measured using bio-impedance analysis derived lean body mass in the aforementioned pharmacokinetics studies. Bio-impedance analysis is considered a valid tool to for the assessment of total body and segmental body composition.<sup>141</sup> As lean body mass not only includes skeletal muscle mass but also organs, bones and inter- and intracellular water, other tissues such as the liver could have also contributed to the absorption, distribution and metabolism of anticancer drugs. However, as clearance of chemotherapeutics cannot be fully explained by liver volume or liver metabolism, skeletal muscle mass is expected to contribute to drug metabolism.<sup>131,142,143</sup>

## 8 | REVERSE CAUSATION: TUMOUR CAUSING SKELETAL MUSCLE DYSFUNCTION

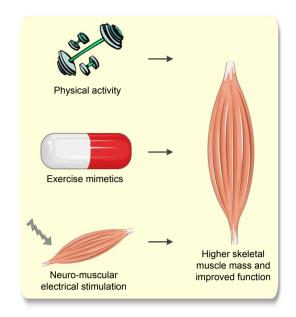
The majority of this review is based on associations only, as longitudinal studies assessing the association between low skeletal muscle mass, pathophysiological mechanisms and poor clinical outcomes in cancer patients are scarce. Thus, mediating roles of pathophysiological mechanisms cannot be substantiated firmly, as they can also reversely affect skeletal muscle mass.<sup>15,16,29</sup> Mutual influence is most likely,<sup>37</sup>

## cta Physiologica

which limits the ability to determine causality. Another factor is the interplay with cancer and anticancer treatment, as both can influence skeletal muscle mass, systemic inflammation, insulin-dependent glucose handling, protein status and pharmacokinetics of anticancer drugs.<sup>22,144</sup> The definition of cachexia incorporates the negative influence of a highdemanding metabolic disease on skeletal muscle mass.<sup>12</sup> As more aggressive tumours have a higher metabolic demand, low skeletal muscle mass could also be an indicator of more aggressive cancers<sup>59,145,146</sup> or of tumour progression.<sup>147</sup> and thus negatively affects clinical outcomes. Moreover, tumourproduced cytokines can lead to a state of inflammation and can increase insulin resistance.<sup>144,146</sup> Protein status is often lower becaus of loss of appetite provoked by anticancer treatment and malnutrition caused by the catabolic state of the body.<sup>148</sup> Although a negative effect of low skeletal muscle mass on clinical outcomes was observed in early-stage cancers when cancer cachexia is not expected,<sup>22,149</sup> these pathophysiological mechanisms cannot be seen separately from the influence of cancer disease activity and the influence of anticancer treatment.<sup>22</sup> It is not clear whether the association between low muscle mass and the risk of poor clinical outcomes is linear or if a critical threshold of muscle mass associates with poor clinical outcomes. Described muscle mass cut-offs to distinguish patients with a low and high risk of poor clinical outcomes are highly variable and have not been validated yet in older cancer patients.

## 9 | LIMITATIONS AND FUTURE DIRECTIONS

This review focussed on low skeletal muscle mass only. Other aspects of sarcopenia and cachexia or the components of muscle failure,<sup>150</sup> that is muscle strength and physical performance, could also be risk factors for poor clinical outcomes in older cancer patients.<sup>151</sup> Although sarcopenia and cachexia are separate diseases, the distinction is very difficult in the presence of cancer as they share a common clinical presentation, that is low muscle mass. In non-longitudinal studies, it is impossible to distinguish whether low skeletal muscle mass is a consequence of agerelated sarcopenia or cancer-related cachexia. The majority of studies defines these diseases based on low muscle mass, whereas other measures including muscle strength, physical performance, weight loss, fat wasting and metabolic state are required to make the distinction.<sup>3,12</sup> Reverse causation has to be kept in mind because of the interplay between skeletal muscle mass, pathophysiology and cancer. In addition, it is important that the pathophysiological mechanisms are likely not separate entities but are probably highly interconnected and interact in their influence on poor clinical outcomes in cancer patients.



**FIGURE 4** Overview of possible therapeutic interventions to reduce the risk of poor clinical outcomes in older cancer patients

The work presented in this review summarized how low skeletal muscle mass might lead to poor clinical outcomes in older cancer patients. The first step in reducing the risk of poor clinical outcomes in older cancer patients would be to prevent the loss of skeletal muscle mass. Inducing myokine production through physical exercise may act as a therapeutic target to prevent or counteract skeletal muscle mass decline<sup>67,152</sup> and may prevent its negative effects on clinical outcomes. Voluntary wheel running in mice was able to preserve skeletal muscle mass during anticancer treatment with cisplatin, whereas mice without training lost more than 20% of their lean body mass.<sup>153</sup> However, while exercise improved skeletal muscle mass in untreated and chemotherapy-treated tumour-bearing mice, it worsened survival in late cachexia stages.<sup>154</sup> Cancer patients with advanced muscle wasting may have passed 'a point of no return' in which exercise can become dysfunctional. For cancer patients who are unable to exercise, alternative administration of myokines such as newly developed exercise mimetics or neuro-muscular electrical stimulation may offer possibilities to reduce the risk of poor clinical outcomes.<sup>74</sup> If muscle deprivation is already present, targeted interventions to prevent the consequent pathophysiological mechanisms from affecting clinical outcome may be beneficial. The ability of exercise training (aerobic and resistance) and nutritional interventions to reduce inflammation and improve immunity,<sup>66,155</sup> reduce oxidative stress and insulin resistance,<sup>155,156</sup> preserve mitochondrial content,<sup>77</sup> and simultaneously preserve or ameliorate skeletal muscle mass and improve clinical outcomes in cancer patients<sup>157</sup> has recently been suggested. Figure 4 provides an overview

acta Physiologica

of possible therapeutic interventions to reduce the risk of poor clinical outcomes as a consequence of low muscle mass in older cancer patients. Future research should focus on gaining insight into causality of muscle wasting and poor clinical outcomes by longitudinal, interventional studies during controlled muscle wasting in animal models. Eventually, this should debouch into specific interventions on these mechanisms to improve clinical outcomes in older patients with cancer.

#### **10** | **CONCLUSIONS**

In the underpinning of the association of low skeletal muscle mass with poor clinical outcomes in older cancer patients, pathophysiology-based mechanisms of altered myokine balance affecting the innate and adaptive immune system and altered pharmacokinetics of anticancer drugs leading to a relative overdosage are best-substantiated. The effects of insulin resistance and circulating mitochondrial DNA as a consequence of low skeletal muscle mass require further exploration. It remains elusive whether these mechanisms are caused by low skeletal muscle mass, and reverse causation should be considered carefully. Developing targeted interventions to restore myokine balance through physical exercise, neuro-muscular electrical stimulation or exercise mimetics and adapting anticancer drug dosing based on skeletal muscle mass, might be targeted approaches to improve clinical outcomes in older cancer patients with low muscle mass.

#### ACKNOWLEDGEMENT

None.

#### **CONFLICTS OF INTEREST**

The authors declare that they have no conflict of interest.

#### ORCID

*Stéphanie M. L. M. Looijaard* https://orcid. org/0000-0001-8170-2677 *Rob C. I. Wüst* https://orcid.org/0000-0003-3781-5177 *Andrea B. Maier* https://orcid.org/0000-0001-7206-1724

#### REFERENCES

- Mitchell WK, Williams J, Atherton P, Larvin M, Lund J, Narici M. Sarcopenia, dynapenia, and the impact of advancing age on human skeletal muscle size and strength; a quantitative review. *Front Physiol.* 2012;3:260.
- Beenakker KG, Ling CH, Meskers CG, et al. Patterns of muscle strength loss with age in the general population and patients with a chronic inflammatory state. *Ageing Res Rev.* 2010;9:431-436.
- Cruz-Jentoft AJ, Bahat G, Bauer J, et al. Sarcopenia: revised European consensus on definition and diagnosis. *Age Ageing*. 2019;48:16-31.

- Shafiee G, Keshtkar A, Soltani A, Ahadi Z, Larijani B, Heshmat R. Prevalence of sarcopenia in the world: a systematic review and meta- analysis of general population studies. *J Diabetes Metab Disord*. 2017;16:21.
- Buford TW, Anton SD, Judge AR, et al. Models of accelerated sarcopenia: critical pieces for solving the puzzle of age-related muscle atrophy. *Ageing Res Rev.* 2010;9:369-383.
- Pacifico J, Geerlings MAJ, Reijnierse EM, Phassouliotis C, Lim WK, Maier AB. Prevalence of sarcopenia as a comorbid disease: a systematic review and meta-analysis. *Exp Gerontol*. 2020;131:110801.
- Pamoukdjian F, Bouillet T, Levy V, Soussan M, Zelek L, Paillaud E. Prevalence and predictive value of pre-therapeutic sarcopenia in cancer patients: a systematic review. *Clin Nutr.* 2018;37:1101-1113.
- Rier HN, Jager A, Sleijfer S, Maier AB, Levin MD. The prevalence and prognostic value of low muscle mass in cancer patients: A review of the literature. *Oncologist.* 2016;21:1396-1409.
- Reijnierse EM, Trappenburg MC, Leter MJ, et al. The impact of different diagnostic criteria on the prevalence of sarcopenia in healthy elderly participants and geriatric outpatients. *Gerontology*. 2015;61:491-496.
- Bijlsma AY, Meskers CG, Ling CH, et al. Defining sarcopenia: the impact of different diagnostic criteria on the prevalence of sarcopenia in a large middle aged cohort. *Age (Dordr)*. 2013;35:871-881.
- Reijnierse EM, Buljan A, Tuttle CSL, et al. Prevalence of sarcopenia in inpatients 70 tos and older using different diagnostic criteria. *Nurs Open*. 2019;6:377-383.
- Muscaritoli M, Anker SD, Argiles J, et al. Consensus definition of sarcopenia, cachexia and pre-cachexia: joint document elaborated by Special Interest Groups (SIG) "cachexia-anorexia in chronic wasting diseases" and "nutrition in geriatrics". *Clin Nutr.* 2010;29:154-159.
- von Haehling S, Anker MS, Anker SD. Prevalence and clinical impact of cachexia in chronic illness in Europe, USA, and Japan: facts and numbers update 2016. *J Cachexia Sarcopenia Muscle*. 2016;7:507-509.
- Bowen TS, Schuler G, Adams V. Skeletal muscle wasting in cachexia and sarcopenia: molecular pathophysiology and impact of exercise training. *J Cachexia Sarcopenia Muscle*. 2015;6:197-207.
- Kim TN, Choi KM. Sarcopenia: definition, epidemiology, and pathophysiology. *J Bone Metab*. 2013;20:1-10.
- Ziaaldini MM, Marzetti E, Picca A, Murlasits Z. Biochemical pathways of sarcopenia and their modulation by physical exercise: a narrative review. *Front Med.* 2017;4:167.
- 17. Anandavadivelan P, Lagergren P. Cachexia in patients with oesophageal cancer. *Nat Rev Clin Oncol.* 2016;13:185-198.
- Antoun S, Baracos VE, Birdsell L, Escudier B, Sawyer MB. Low body mass index and sarcopenia associated with dose-limiting toxicity of sorafenib in patients with renal cell carcinoma. *Ann Oncol.* 2010;21:1594-1598.
- Antoun S, Borget I, Lanoy E. Impact of sarcopenia on the prognosis and treatment toxicities in patients diagnosed with cancer. *Curr Opin Support Palliat Care*. 2013;7:383-389.
- Mir O, Coriat R, Blanchet B, et al. Sarcopenia predicts early dose-limiting toxicities and pharmacokinetics of sorafenib in patients with hepatocellular carcinoma. *PLoS One*. 2012;7:e37563.

## ta Physiologica

- Prado CM, Lieffers JR, McCargar LJ, et al. Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: a population-based study. *Lancet Oncol.* 2008;9:629-635.
- Cespedes Feliciano EM, Kroenke CH, Caan BJ. The obesity paradox in cancer: how important is muscle? *Annu Rev Nutr.* 2018;38:357-379.
- Kazemi-Bajestani SM, Mazurak VC, Baracos V. Computed tomography-defined muscle and fat wasting are associated with cancer clinical outcomes. *Semin Cell Dev Biol.* 2016;54:2-10.
- 24. Shachar SS, Williams GR, Muss HB, Nishijima TF. Prognostic value of sarcopenia in adults with solid tumours: a meta-analysis and systematic review. *Eur J Cancer*. 2016;57:58-67.
- Looijaard SMLM, Meskers CGM, Slee-Valentijn MS, et al. Computed Tomography-based body composition is not consistently associated with outcome in older patients with colorectal cancer. *Oncologist.* 2019.
- Prado CM, Baracos VE, McCargar LJ, et al. Body composition as an independent determinant of 5-fluorouracil-based chemotherapy toxicity. *Clin Cancer Res.* 2007;13:3264-3268.
- Suttorp MM, Siegerink B, Jager KJ, Zoccali C, Dekker FW. Graphical presentation of confounding in directed acyclic graphs. *Nephrol Dial Transplant*. 2015;30:1418-1423.
- Afzali AM, Muntefering T, Wiendl H, Meuth SG, Ruck T. Skeletal muscle cells actively shape (auto)immune responses. *Autoimmun Rev.* 2018;17:518-529.
- Nelke C, Dziewas R, Minnerup J, Meuth SG, Ruck T. Skeletal muscle as potential central link between sarcopenia and immune senescence. *EBioMedicine*. 2019;49:381-388.
- Malietzis G, Aziz O, Bagnall NM, Johns N, Fearon KC, Jenkins JT. The role of body composition evaluation by computerized tomography in determining colorectal cancer treatment outcomes: a systematic review. *Eur J Surg Oncol.* 2015;41:186-196.
- Kalinkovich A, Livshits G. Sarcopenic obesity or obese sarcopenia: A cross talk between age-associated adipose tissue and skeletal muscle inflammation as a main mechanism of the pathogenesis. *Ageing Res Rev.* 2017;35:200-221.
- 32. He WZ, Yang QX, Xie JY, et al. Association of low skeletal muscle index with increased systematic inflammatory responses and interferon γ-induced protein 10 levels in patients with colon cancer. *Cancer Management and Research*. 2018;10:2499-2507.
- Kim EY, Kim YS, Seo JY, et al. The Relationship between Sarcopenia and systemic inflammatory response for cancer cachexia in small cell lung cancer. *PLoS One*. 2016;11:e0161125.
- Lin JX, Lin JP, Xie JW, et al. Prognostic value and association of sarcopenia and systemic inflammation for patients with gastric cancer following radical gastrectomy. *Oncologist*. 2019;24:e1091-e1101.
- Malietzis G, Johns N, Al-Hassi HO, et al. Low muscularity and myosteatosis is related to the host systemic inflammatory response in patients undergoing surgery for colorectal cancer. *Ann Surg.* 2016;263:320-325.
- Hilmi M, Jouinot A, Burns R, et al. Body composition and sarcopenia: The next-generation of personalized oncology and pharmacology? *Pharmacol Ther*. 2019;196:135-159.
- Feliciano EMC, Kroenke CH, Meyerhardt JA, et al. Association of systemic inflammation and sarcopenia with survival in nonmetastatic colorectal cancer: results from the C SCANS Study. *JAMA Oncol.* 2017;3:e172319.

- McSorley ST, Black DH, Horgan PG, McMillan DC. The relationship between tumour stage, systemic inflammation, body composition and survival in patients with colorectal cancer. *Clin Nutr.* 2018;37:1279-1285.
- Okugawa Y, Toiyama Y, Yamamoto A, et al. Close relationship between immunological/inflammatory markers and myopenia and myosteatosis in patients with colorectal cancer: a propensity score matching analysis. *JPEN J Parenter Enteral Nutr.* 2019;43:508-515.
- Dolan RD, McSorley ST, Horgan PG, Laird B, McMillan DC. The role of the systemic inflammatory response in predicting outcomes in patients with advanced inoperable cancer: systematic review and meta-analysis. *Crit Rev Oncol Hematol.* 2017;116:134-146.
- Go SI, Park MJ, Song HN, et al. Sarcopenia and inflammation are independent predictors of survival in male patients newly diagnosed with small cell lung cancer. *Support Care Cancer*. 2016;24:2075-2084.
- Cosqueric G, Sebag A, Ducolombier C, Thomas C, Piette F, Weill-Engerer S. Sarcopenia is predictive of nosocomial infection in care of the elderly. *Br J Nutr.* 2006;96:895-901.
- Lieffers JR, Bathe OF, Fassbender K, Winget M, Baracos VE. Sarcopenia is associated with postoperative infection and delayed recovery from colorectal cancer resection surgery. *Br J Cancer*. 2012;107:931-936.
- Smeets BJJ, Brinkman DJ, Horsten ECJ, et al. The effect of myopenia on the inflammatory response early after colorectal surgery. *Nutr Cancer*. 2018;70:460-466.
- Roberts BM, Ahn B, Smuder AJ, et al. Diaphragm and ventilatory dysfunction during cancer cachexia. *Faseb j.* 2013;27:2600-2610.
- Fogarty MJ, Mantilla CB, Sieck GC. Impact of sarcopenia on diaphragm muscle fatigue. *Exp Physiol*. 2019;104:1090-1099.
- Wakabayashi H, Matsushima M, Uwano R, Watanabe N, Oritsu H, Shimizu Y. Skeletal muscle mass is associated with severe dysphagia in cancer patients. *J Cachexia Sarcopenia Muscle*. 2015;6:351-357.
- Makiura D, Ono R, Inoue J, et al. Preoperative sarcopenia is a predictor of postoperative pulmonary complications in esophageal cancer following esophagectomy: a retrospective cohort study. J Geriatr Oncol. 2016;7:430-436.
- Roxburgh CS, Horgan PG, McMillan DC. The perioperative immune/inflammatory insult in cancer surgery: time for intervention? *Oncoimmunology*. 2013;2:e27324.
- Zhou CJ, Zhang FM, Zhang FY, et al. Sarcopenia: a new predictor of postoperative complications for elderly gastric cancer patients who underwent radical gastrectomy. *J Surg Res.* 2017;211:137-146.
- 51. Pedersen BK. Muscles and their myokines. *J Exp Biol.* 2011;214:337-346.
- Lutz CT, Quinn LS. Sarcopenia, obesity, and natural killer cell immune senescence in aging: altered cytokine levels as a common mechanism. *Aging*. 2012;4:535-546.
- Kennedy MK, Glaccum M, Brown SN, et al. Reversible defects in natural killer and memory CD8 T cell lineages in interleukin 15-deficient mice. *J Exp Med*. 2000;191:771-780.
- Koka R, Burkett PR, Chien M, et al. Interleukin (IL)-15R[alpha]deficient natural killer cells survive in normal but not IL-15R[alpha]-deficient mice. *J Exp Med*. 2003;197:977-984.

- 55. Chen XY, Li B, Ma BW, et al. Sarcopenia is an effective prognostic indicator of postoperative outcomes in laparoscopic-assisted gastrectomy. *Eur J Surg Oncol.* 2019;45:1092-1098.
- Iritani S, Imai K, Takai K, et al. Skeletal muscle depletion is an independent prognostic factor for hepatocellular carcinoma. J Gastroenterol. 2015;50:323-332.
- Conlon KC, Lugli E, Welles HC, et al. Redistribution, hyperproliferation, activation of natural killer cells and CD8 T cells, and cytokine production during first-in-human clinical trial of recombinant human interleukin-15 in patients with cancer. *J Clin Oncol*. 2015;33:74-82.
- Pedersen L, Idorn M, Olofsson GH, et al. Voluntary running suppresses Tumor growth through epinephrine- and IL-6dependent NK cell mobilization and redistribution. *Cell Metab.* 2016;23:554-562.
- Lee JS, Kim YS, Kim EY, Jin W. Prognostic significance of CTdetermined sarcopenia in patients with advanced gastric cancer. *PLoS One*. 2018;13:e0202700.
- Tsukioka T, Izumi N, Mizuguchi S, et al. Positive correlation between sarcopenia and elevation of neutrophil/lymphocyte ratio in pathological stage IIIA (N2-positive) non-small cell lung cancer patients. *Gen Thorac Cardiovasc Surg.* 2018;66:716-722.
- Groen WG, van Harten WH, Vallance JK. Systematic review and meta-analysis of distance-based physical activity interventions for cancer survivors (2013–2018): We still haven't found what we're looking for. *Cancer Treat Rev.* 2018;69:188-203.
- Mishra SI, Scherer RW, Snyder C, Geigle PM, Berlanstein DR, Topaloglu O. Exercise interventions on health-related quality of life for people with cancer during active treatment. *Cochrane Database Syst Rev.* 2012.8(5):CD008465
- Faithfull S, Turner L, Poole K, et al. Prehabilitation for adults diagnosed with cancer: a systematic review of long-term physical function, nutrition and patient-reported outcomes. *Eur J Cancer Care (Engl)*. 2019;e13023.
- 64. Hojman P, Gehl J, Christensen JF, Pedersen BK. Molecular mechanisms linking exercise to cancer prevention and treatment. *Cell Metab.* 2018;27:10-21.
- Hwang JH, McGovern J, Minett GM, et al. Mobilizing serum factors and immune cells through exercise to counteract age-related changes in cancer risk. *Exerc Immunol Rev.* 2020;26:80-99.
- Kruijsen-Jaarsma M, Revesz D, Bierings MB, Buffart LM, Takken T. Effects of exercise on immune function in patients with cancer: a systematic review. *Exerc Immunol Rev.* 2013;19:120-143.
- Manole E, Ceafalan LC, Popescu BO, Dumitru C, Bastian AE. Myokines as possible therapeutic targets in cancer cachexia. J Immunol Res. 2018;2018:8260742.
- Waugh DJ, Wilson C. The interleukin-8 pathway in cancer. *Clin Cancer Res.* 2008;14:6735-6741.
- Smith RC, Lin BK. Myostatin inhibitors as therapies for muscle wasting associated with cancer and other disorders. *Curr Opin Support Palliat Care*. 2013;7:352-360.
- Hojman P. Exercise protects from cancer through regulation of immune function and inflammation. *Biochem Soc Trans.* 2017;45:905-911.
- Hojman P, Dethlefsen C, Brandt C, Hansen J, Pedersen L, Pedersen BK. Exercise-induced muscle-derived cytokines inhibit mammary cancer cell growth. *Am J Physiol Endocrinol Metab.* 2011;301:E504-510.
- Gannon NP, Vaughan RA, Garcia-Smith R, Bisoffi M, Trujillo KA. Effects of the exercise-inducible myokine irisin on malignant

and non-malignant breast epithelial cell behavior in vitro. *Int J Cancer*. 2015;136:E197-202.

Acta Physiologica

- Aoi W, Naito Y, Takagi T, et al. A novel myokine, secreted protein acidic and rich in cysteine (SPARC), suppresses colon tumorigenesis via regular exercise. *Gut.* 2013;62:882-889.
- Re Cecconi AD, Forti M, Chiappa M, et al. Musclin, a myokine induced by aerobic exercise, retards muscle atrophy during cancer cachexia in mice. *Cancers*. 2019;11(10):1541.
- Schink K, Gassner H, Reljic D, et al. Y: Assessment of gait parameters and physical function in patients with advanced cancer participating in a 12-week exercise and nutrition programme: a controlled clinical trial. *Eur J Cancer Care (Engl)*. 2020;29:e13199.
- 76. Schink K, Herrmann HJ, Schwappacher R, et al. Effects of wholebody electromyostimulation combined with individualized nutritional support on body composition in patients with advanced cancer: a controlled pilot trial. *BMC Cancer*. 2018;18:886.
- Mijwel S, Cardinale DA, Norrbom J, et al. Exercise training during chemotherapy preserves skeletal muscle fiber area, capillarization, and mitochondrial content in patients with breast cancer. *Faseb j.* 2018;32:5495-5505.
- Ciciliot S, Rossi AC, Dyar KA, Blaauw B, Schiaffino S. Muscle type and fiber type specificity in muscle wasting. *Int J Biochem Cell Biol.* 2013;45:2191-2199.
- Moller AB, Lonbro S, Farup J, et al. Molecular and cellular adaptations to exercise training in skeletal muscle from cancer patients treated with chemotherapy. *J Cancer Res Clin Oncol.* 2019;145:1449-1460.
- Tanaka M, Sugimoto K, Fujimoto T, et al. Preventive effects of low-intensity exercise on cancer cachexia-induced muscle atrophy. *Faseb j.* 2019;33:7852-7862.
- Zierath JR, Krook A, Wallberg-Henriksson H. Insulin action and insulin resistance in human skeletal muscle. *Diabetologia*. 2000;43:821-835.
- Dumas JF, Simard G, Flamment M, Ducluzeau PH, Ritz P. Is skeletal muscle mitochondrial dysfunction a cause or an indirect consequence of insulin resistance in humans? *Diabetes Metab.* 2009;35:159-167.
- Edwards S. Blood sugar tolerance in cancer. J Indiana State Med Assoc. 1919;12:296.
- Jasani B, Donaldson LJ, Ratcliffe JG, Sokhi GS. Mechanism of impaired glucose tolerance in patients with neoplasia. *Br J Cancer*. 1978;38:287-292.
- 85. Boyd DB. Insulin and cancer. Integr Cancer Ther. 2003;2:315-329.
- Orgel E, Mittelman SD. The links between insulin resistance, diabetes, and cancer. *Curr Diab Rep.* 2013;13:213-222.
- 87. Petruzzelli M, Wagner EF. Mechanisms of metabolic dysfunction in cancer-associated cachexia. *Genes Dev.* 2016;30:489-501.
- Moon SS. Low skeletal muscle mass is associated with insulin resistance, diabetes, and metabolic syndrome in the Korean population: the Korea National Health and Nutrition Examination Survey (KNHANES) 2009–2010. *Endocr J.* 2014;61:61-70.
- Srikanthan P, Hevener AL, Karlamangla AS. Sarcopenia exacerbates obesity-associated insulin resistance and dysglycemia: findings from the National Health and Nutrition Examination Survey III. *PLoS One*. 2010;5:e10805.
- 90. Srikanthan P, Karlamangla AS. Relative muscle mass is inversely associated with insulin resistance and prediabetes. Findings from the third National Health and Nutrition Examination Survey. J Clin Endocrinol Metab. 2011;96:2898-2903.

## ta Physiologica

- 91. Pak S, Park SY, Shin TJ, et al. Association of muscle mass with survival after radical prostatectomy in patients with prostate cancer. *J Urol.* 2019;202:525-532.
- 92. Mehta HH, Gao Q, Galet C, et al. IGFBP-3 is a metastasis suppression gene in prostate cancer. *Cancer Res.* 2011;71:5154-5163.
- 93. Shariat SF, Lamb DJ, Kattan MW, et al. Association of preoperative plasma levels of insulin-like growth factor I and insulinlike growth factor binding proteins-2 and -3 with prostate cancer invasion, progression, and metastasis. J Clin Oncol. 2002;20:833-841.
- Smith RL, Soeters MR, Wust RCI, Houtkooper RH. Metabolic flexibility as an adaptation to energy resources and requirements in health and disease. *Endocr Rev.* 2018;39:489-517.
- Caccialanza R, Cereda E, De Lorenzo F, Farina G, Pedrazzoli P. To fast, or not to fast before chemotherapy, that is the question. *BMC Cancer*. 2018;18:337.
- 96. Caccialanza R, Aprile G, Cereda E, Pedrazzoli P. Fasting in oncology: a word of caution. *Nat Rev Cancer*. 2019;19:177.
- 97. Yang S, Chu S, Gao Y, et al. a narrative review of cancer-related fatigue (CRF) and its possible pathogenesis. *Cells*. 2019;8(7):738.
- Calvani R, Joseph A-M, Adhihetty PJ, et al. Mitochondrial pathways in sarcopenia of aging and disuse muscle atrophy. *Biol Chem.* 2013;394:393-414.
- Bua EA, McKiernan SH, Wanagat J, McKenzie D, Aiken JM. Mitochondrial abnormalities are more frequent in muscles undergoing sarcopenia. *J Appl Physiol*. 2002;92:2617-2624.
- Julienne CM, Dumas JF, Goupille C, et al. Cancer cachexia is associated with a decrease in skeletal muscle mitochondrial oxidative capacities without alteration of ATP production efficiency. *J Cachexia Sarcopenia Muscle*. 2012;3:265-275.
- VanderVeen BN, Fix DK, Carson JA. Disrupted skeletal muscle mitochondrial dynamics, mitophagy, and biogenesis during cancer cachexia: a role for inflammation. *Oxid Med Cell Longev*. 2017;2017:3292087.
- Nicolson GL. Mitochondrial dysfunction and chronic disease: treatment with natural supplements. *Integr Med.* 2014;13:35-43.
- Vyas S, Zaganjor E, Haigis MC. Mitochondria and cancer. *Cell*. 2016;166:555-566.
- 104. Sorensen JC, Cheregi BD, Timpani CA, Nurgali K, Hayes A, Rybalka E. Mitochondria: inadvertent targets in chemotherapy-induced skeletal muscle toxicity and wasting? *Cancer Chemother Pharmacol.* 2016;78:673-683.
- Meng X, Schwarzenbach H, Yang Y, et al. Circulating mitochondrial DNA is linked to progression and prognosis of epithelial ovarian cancer. *Transl Oncol.* 2019;12:1213-1220.
- Kalavska K, Minarik T, Vlkova B, et al. Prognostic value of various subtypes of extracellular DNA in ovarian cancer patients. J Ovarian Res. 2018;11:85.
- Weerts MJA, Timmermans EC, van de Stolpe A, et al. Tumorspecific mitochondrial DNA variants are rarely detected in cellfree DNA. *Neoplasia*. 2018;20:687-696.
- 108. Picca A, Lezza AMS, Leeuwenburgh C, et al. Circulating mitochondrial DNA at the crossroads of mitochondrial dysfunction and inflammation during aging and muscle wasting disorders. *Rejuvenation Res.* 2018;21:350-359.
- 109. Rodriguez-Nuevo A, Zorzano A. The sensing of mitochondrial DAMPs by non-immune cells. *Cell Stress*. 2019;3:195-207.
- 110. Singel KL, Grzankowski KS, Khan A, et al. Mitochondrial DNA in the tumour microenvironment activates neutrophils and is

associated with worse outcomes in patients with advanced epithelial ovarian cancer. *Br J Cancer*. 2019;120:207-217.

- Phillips BE, Hill DS, Atherton PJ. Regulation of muscle protein synthesis in humans. *Curr Opin Clin Nutr Metab Care*. 2012;15:58-63.
- 112. Glass DJ. Signaling pathways perturbing muscle mass. *Curr Opin Clin Nutr Metab Care*. 2010;13:225-229.
- 113. Gordon BS, Kelleher AR, Kimball SR. Regulation of muscle protein synthesis and the effects of catabolic states. *Int J Biochem Cell Biol*. 2013;45:2147-2157.
- 114. van Atteveld VA, Van Ancum JM, Reijnierse EM, Trappenburg MC, Meskers CGM, Maier AB. Erythrocyte sedimentation rate and albumin as markers of inflammation are associated with measures of sarcopenia: a cross-sectional study. *BMC Geriatr.* 2019;19:233.
- 115. Truong A, Hanna MH, Moghadamyeghaneh Z, Stamos MJ. Implications of preoperative hypoalbuminemia in colorectal surgery. *World J Gastrointest Surg.* 2016;8:353-362.
- 116. Kim S, McClave SA, Martindale RG, Miller KR, Hurt RT. Hypoalbuminemia and clinical outcomes: what is the mechanism behind the relationship? *Am Surg.* 2017;83:1220-1227.
- 117. Sharma P, Zargar-Shoshtari K, Caracciolo JT, et al. Sarcopenia as a predictor of overall survival after cytoreductive nephrectomy for metastatic renal cell carcinoma. *Urol Oncol.* 2015;33(339):e317-323.
- Visser M, Kritchevsky SB, Newman AB, et al. Lower serum albumin concentration and change in muscle mass: the Health, Aging and Body Composition Study. *Am J Clin Nutr.* 2005;82:531-537.
- Seres DS. Surrogate nutrition markers, malnutrition, and adequacy of nutrition support. *Nutr Clin Pract*. 2005;20:308-313.
- Figueiredo VC, McCarthy JJ. Regulation of Ribosome Biogenesis in Skeletal Muscle Hypertrophy. *Physiology (Bethesda)*. 2019;34:30-42.
- 121. Dworzak F, Ferrari P, Gavazzi C, Maiorana C, Bozzetti F. Effects of cachexia due to cancer on whole body and skeletal muscle protein turnover. *Cancer*. 1998;82:42-48.
- 122. Mayers JR, Torrence ME, Danai LV, et al. Tissue of origin dictates branched-chain amino acid metabolism in mutant Krasdriven cancers. *Science*. 2016;353:1161-1165.
- 123. Du Bois D, Du Bois EF. A formula to estimate the approximate surface area if height and weight be known. Arch Intern Med. 1916;17:863-871.
- 124. Baker SD, Verweij J, Rowinsky EK, et al. Role of body surface area in dosing of investigational anticancer agents in adults, 1991–2001. J Natl Cancer Inst. 2002;94:1883-1888.
- 125. Sawyer M, Ratain MJ. Body surface area as a determinant of pharmacokinetics and drug dosing. *Invest New Drugs*. 2001;19:171-177.
- 126. Gérard S, Bréchemier D, Lefort A, et al. Body composition and anti-neoplastic treatment in adult and older subjects - a systematic review. J Nutr Health Aging. 2016;20:878-888.
- Hopkins JJ, Sawyer MB. A review of body composition and pharmacokinetics in oncology. *Expert Rev Clin Pharmacol*. 2017;10:947-956.
- Fabris S, MacLean DA. Skeletal muscle an active compartment in the sequestering and metabolism of doxorubicin chemotherapy. *PLoS One.* 2015;10:e0139070.
- 129. Narjoz C, Cessot A, Thomas-Schoemann A, et al. Role of the lean body mass and of pharmacogenetic variants on the pharmacokinetics and pharmacodynamics of sunitinib in cancer patients. *Invest New Drugs*. 2015;33:257-268.

Acta Physiologica

- 130. Massicotte MH, Borget I, Broutin S, et al. Body composition variation and impact of low skeletal muscle mass in patients with advanced medullary thyroid carcinoma treated with vandetanib: results from a placebo-controlled study. *J Clin Endocrinol Metab.* 2013;98:2401-2408.
- Prado CM, Lima IS, Baracos VE, et al. An exploratory study of body composition as a determinant of epirubicin pharmacokinetics and toxicity. *Cancer Chemother Pharmacol.* 2011;67:93-101.
- 132. Wong AL, Seng KY, Ong EM, et al. Body fat composition impacts the hematologic toxicities and pharmacokinetics of doxorubicin in Asian breast cancer patients. *Breast Cancer Res Treat*. 2014;144:143-152.
- 133. Williams GR, Deal AM, Shachar SS, et al. The impact of skeletal muscle on the pharmacokinetics and toxicity of 5-fluorouracil in colorectal cancer. *Cancer Chemother Pharmacol*. 2018;81:413-417.
- Klein GL, Petschow BW, Shaw AL, Weaver E. Gut barrier dysfunction and microbial translocation in cancer cachexia: a new therapeutic target. *Curr Opin Support Palliat Care*. 2013;7:361-367.
- Puppa MJ, White JP, Sato S, Cairns M, Baynes JW, Carson JA. Gut barrier dysfunction in the Apc(Min/+) mouse model of colon cancer cachexia. *Biochim Biophys Acta*. 2011;1812(12):1601–1606.
- 136. Alexandre J, Rey E, Girre V, et al. Relationship between cytochrome 3A activity, inflammatory status and the risk of docetaxel-induced febrile neutropenia: a prospective study. *Ann Oncol.* 2007;18:168-172.
- 137. Kacevska M, Robertson GR, Clarke SJ, Liddle C. Inflammation and CYP3A4-mediated drug metabolism in advanced cancer: impact and implications for chemotherapeutic drug dosing. *Expert Opin Drug Metab Toxicol*. 2008;4:137-149.
- 138. Cvan Trobec K, Kerec Kos M, Trontelj J, et al. Influence of cancer cachexia on drug liver metabolism and renal elimination in rats. *J Cachexia Sarcopenia Muscle*. 2015;6:45-52.
- 139. Huillard O, Mir O, Peyromaure M, et al. Sarcopenia and body mass index predict sunitinib-induced early dose-limiting toxicities in renal cancer patients. *Br J Cancer*. 2013;108:1034-1041.
- 140. Miller AA, Murry DJ, Owzar K, et al. Phase I and pharmacokinetic study of sorafenib in patients with hepatic or renal dysfunction: CALGB 60301. *J Clin Oncol.* 2009;27:1800-1805.
- 141. Ling CH, de Craen AJ, Slagboom PE, et al. Accuracy of direct segmental multi-frequency bioimpedance analysis in the assessment of total body and segmental body composition in middle-aged adult population. *Clin Nutr.* 2011;30:610-615.
- 142. Gusella M, Toso S, Ferrazzi E, Ferrari M, Padrini R. Relationships between body composition parameters and fluorouracil pharmacokinetics. *Br J Clin Pharmacol.* 2002;54:131-139.
- 143. Ensminger WD, Rosowsky A, Raso V, et al. A clinical-pharmacological evaluation of hepatic arterial infusions of 5-fluoro-2'deoxyuridine and 5-fluorouracil. *Can Res.* 1978;38:3784-3792.
- Honors MA, Kinzig KP. The role of insulin resistance in the development of muscle wasting during cancer cachexia. J Cachexia Sarcopenia Muscle. 2012;3:5-11.
- Dodson S, Baracos VE, Jatoi A, et al. Muscle wasting in cancer cachexia: clinical implications, diagnosis, and emerging treatment strategies. *Annu Rev Med.* 2011;62:265-279.

- 146. Cho Y, Kim JW, Keum KC, Lee CG, Jeung HC, Lee IJ. Prognostic significance of sarcopenia with inflammation in patients with head and neck cancer who underwent definitive chemoradiotherapy. *Front Oncol.* 2018;8:457.
- 147. Cushen SJ, Power DG, Teo MY, et al. Body composition by computed tomography as a predictor of toxicity in patients with renal cell carcinoma treated with sunitinib. *Am J Clin Oncol.* 2017;40:47-52.
- 148. Gao S, Wang D. Sarcopenia as an independent prognostic factor in patients following surgery for gallbladder cancer. *International Journal of Clinical and Experimental Medicine*. 2018;11:877-883.
- 149. Caan BJ, Meyerhardt JA, Kroenke CH, et al. Explaining the Obesity Paradox: The Association between Body Composition and Colorectal Cancer Survival (C-SCANS Study). *Cancer Epidemiol Biomarkers Prev.* 2017;26:1008-1015.
- 150. Suetta C, Maier AB. Is muscle failure a better term than sarcopenia? *J Cachexia Sarcopenia Muscle*. 2019;10:1146-1147.
- 151. Versteeg KS, Blauwhoff-Buskermolen S, Buffart LM, et al. Higher muscle strength is associated with prolonged survival in older patients with advanced cancer. *Oncologist.* 2018;23:580-585.
- 152. Piccirillo R. Exercise-induced myokines with therapeutic potential for muscle wasting. *Front Physiol*. 2019;10:287.
- Hojman P, Fjelbye J, Zerahn B, et al. Voluntary exercise prevents cisplatin-induced muscle wasting during chemotherapy in mice. *PLoS One.* 2014;9:e109030.
- 154. Ballaro R, Beltra M, De Lucia S, et al. Moderate exercise in mice improves cancer plus chemotherapy-induced muscle wasting and mitochondrial alterations. *Faseb j.* 2019;33:5482-5494.
- 155. Winters-Stone KM, Wood LJ, Stoyles S, Dieckmann NF. The effects of resistance exercise on biomarkers of breast cancer prognosis: a pooled analysis of three randomized trials. *Cancer Epidemiol Biomarkers Prev.* 2018;27:146-153.
- 156. Bruno E, Roveda E, Vitale J, et al. Effect of aerobic exercise intervention on markers of insulin resistance in breast cancer women. *Eur J Cancer Care*. 2018;27:e12617.
- 157. Pin F, Couch ME, Bonetto A. Preservation of muscle mass as a strategy to reduce the toxic effects of cancer chemotherapy on body composition. *Curr Opin Support Palliat Care*. 2018;12:420-426.

#### SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Looijaard SMLM, te Lintel Hekkert ML, Wüst RCI, Otten RHJ, Meskers CGM, Maier AB. Pathophysiological mechanisms explaining poor clinical outcome of older cancer patients with low skeletal muscle mass. *Acta Physiol*. 2021;231:e13516. https://doi.org/10.1111/apha.13516