

LETTER TO THE EDITOR

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Recurrent *SETD2* mutation in *NPM1*-mutated acute myeloid leukemia

Jiwen Sun¹, Wenjuan Yu^{2,3*} and Xiang Zhang^{2,3*}

Abstract

SETD2 is the only methyltransferase for H3K36me₃, and our previous study has firstly demonstrated that it functioned as one tumor suppressor in hematopoiesis. Consistent with it, *SETD2* mutation, which led to its loss of function, was identified in AML. However, the distribution and function of *SETD2* mutation in AML remained largely unknown. Herein, we integrated *SETD2*-mutated AML cases from our center and literature reports, and found that *NPM1* mutation was the most common concomitant genetic alteration with *SETD2* mutation in AML, with its frequency even higher than *MLL* rearrangement and *AML1-ETO*. Though this result indicated the cooperation of *SETD2* and *NPM1* mutations in leukemogenesis, our functional study showed that *SETD2* was required for the proliferation of *NPM1*-mutated AML cell line OCI-AML3, but not *MLL*-rearranged AML cell line THP-1, via maintaining its direct target *NPM1* expression, which was just opposite to its role of tumor suppressor. Therefore, we speculated that *SETD2* possibly had two different faces in distinct subtypes and stages of AML.

Keywords: *SETD2* mutation, *NPM1* mutation, Acute myeloid leukemia

To the editor

SETD2 has been demonstrated as one tumor suppressor in hematopoiesis [1], and *SETD2* mutation affected AML, in which its distribution remained not fully understood [2]. Herein, we analyzed the *SETD2* mutation in *NPM1*-mutated AML.

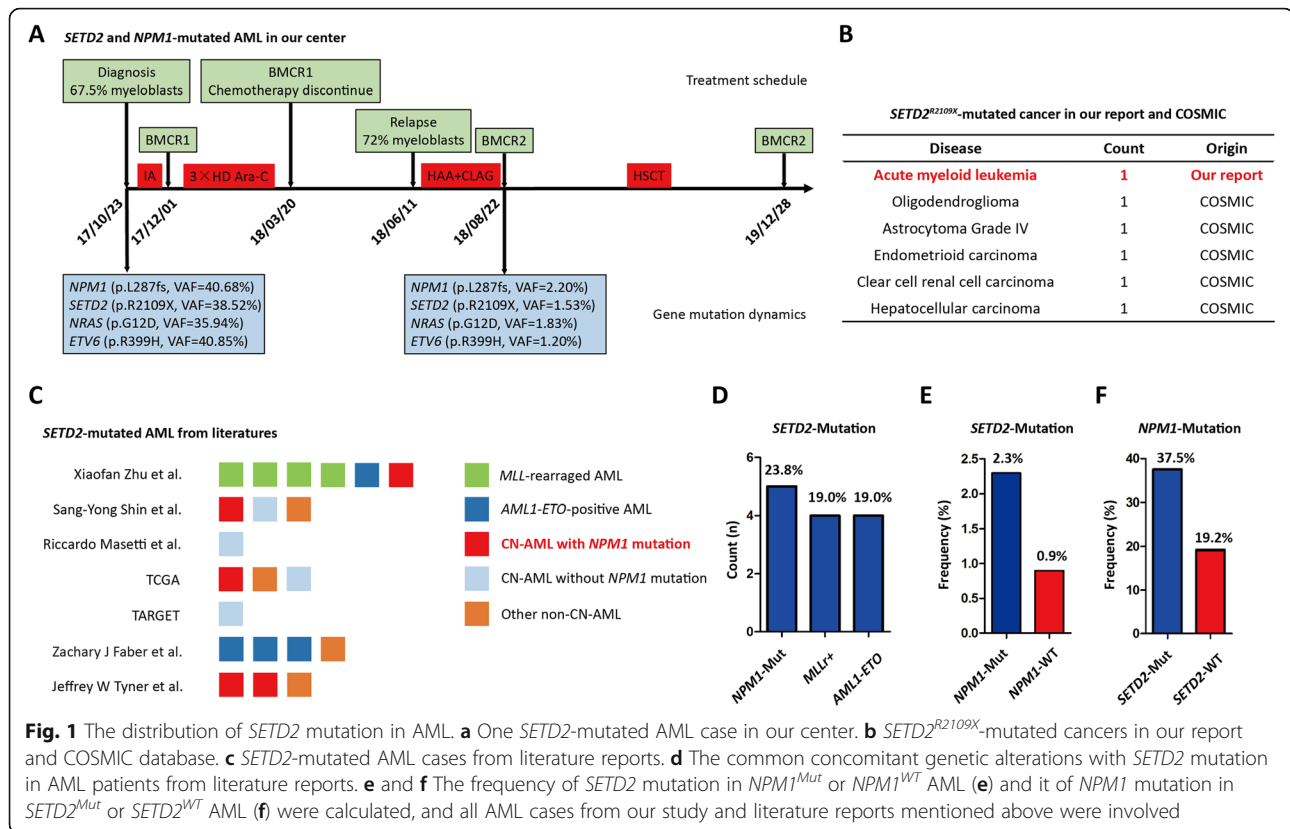
One 36-year-old woman was committed due to abdominal pain and fever for 7 and 3 days, respectively. PB test showed WBC: $52.4 \times 10^9/L$, Hb: 98 g/L, PLT: $48 \times 10^9/L$, circulated blast: 80%. BM examination exhibited 67.5% myeloblasts with the immunophenotype of CD11b-CD13^{dim} + CD14-CD15^{dim} + CD33 + CD34^{partial} + CD35-CD38^{dim} + CD45 + CD64-CD65^{dim} + CD71 + CD117 + CD123^{dim} + HLA-DR-. Though karyotype was normal and *CBF* or *MLL* rearrangements were negative, *NPM1*, *SETD2*, *NRAS* and *ETV6* mutations were identified.

Therefore, AML with mutated *NPM1* was diagnosed. After receiving the operation for co-existed acute appendicitis, she accepted IA regimen as induction therapy, and CR1 was achieved. Subsequently, she received three cycle of medium-dose cytarabine regimen. However, AML relapsed at the 3 months after cessation of chemotherapy, and 72% myeloblasts re-emerged in BM. Due to the early recurrence, she accepted HAA and CLAG regimen successively, and achieved CR2. However, the leukemic clones were not eradicated reflected by persistent above mutations. Therefore, allogeneic semi-compatible HSCT was immediately conducted. As follow-up, CR was still maintained at the 15 months after HSCT (Fig. 1a).

In this patient, *SETD2*^{R2109X} was identified, and it was also found in other malignancies from COSMIC database (Fig. 1b), so *SETD2*^{R2109X} was one driver in cancer. However, *SETD2* deficiency was not sufficient to generate AML, so additional hits were required [1, 3]. Therefore, we reviewed AML studies involving *SETD2* mutation [2, 4–9], and found that *NPM1* mutation

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rather than *MLL* rearrangement or *AML1-ETO* was the most common co-existed genetic alteration of *SETD2* mutation in AML (Fig. 1c-d). To establish their association, we displayed subgroup analysis in above studies, then submitted it Pearson's chi-square test, and calculated OR. Strikingly, *SETD2* and *NPM1* mutations were the concomitant mutation in AML ($P=0.031$; OR = 3.28) (Fig. 1e-f). To address whether *SETD2* mutation mediated drug resistance in AML, we analyzed their therapeutic response to standard chemotherapy. Among 22 *SETD2*-mutated AML patients, the data were available in 11 patients, while CR, PR, and NR was 72.7%, 9.09%, and 18.2%, respectively. Notably, the CR was comparable to it in total AML. Interestingly, all with *NPM1*-mutated AML achieved CR, and two with *MLL*-rearranged AML exhibited NR. Therefore, *SETD2* mutation was possibly not one determinant in drug sensitivity for AML. Furthermore, we analyzed the OS between *SETD2*-mutated and wild-type groups with cBioPortal database [10, 11], but no significance between two groups was found (Additional file 1: Figure S1). Regrettably, the data about EFS were not available.

Loss of *SETD2* function accelerated the progression of *MLL*-rearranged or *AML1-ETO*-positive AML, but whether it was the same in *NPM1*-mutated AML remained unknown. Herein, we displayed shRNA-mediated *SETD2* knockdown, which simulated its loss of

function caused by *SETD2* frame-shift or nonsense mutation, in *NPM1*-mutated AML cell line OCI-AML3 and *MLL*-rearranged AML cell line THP-1. Interestingly, *SETD2* knockdown impaired the proliferation of OCI-AML3 but not THP-1 cells (Fig. 2a-d). Furthermore, the proliferative defect of OCI-AML3 was caused by increased cell apoptosis (Fig. 2e) and cell cycle arrested at G1/G0 phase (Fig. 2f). It has been reported that the viability of OCI-AML3 relied on the function of *NPM1* mutation [12], while *NPM1* expression was regulated by the transcriptional activation mark, H3K36me3, which indicated by ChIP-Seq in the HSPCs of *Mll-af9*-positive leukemia (Fig. 2g) [13]. Consistently, we demonstrated that *NPM1* and its direct targets *MEIS*, *HOXA9* were significantly down-regulated in *SETD2* knockdown OCI-AML3 cells (Fig. 2h-i). Therefore, our results indicated that *SETD2* knockdown-mediated OCI-AML3 proliferation inhibition was possibly attributed to *NPM1* down-regulation.

The detailed role of *SETD2* mutation in *NPM1*-mutated AML remained mysterious. Theoretically, *SETD2* and *NPM1* mutations probably cooperated in leukemogenesis. However, our results showed that *SETD2* was required for the maintenance of OCI-AML3. To our knowledge, two possibilities existed: firstly, *SETD2* mutation played different roles in the initiation and maintenance of *NPM1*-mutated AML; secondly,

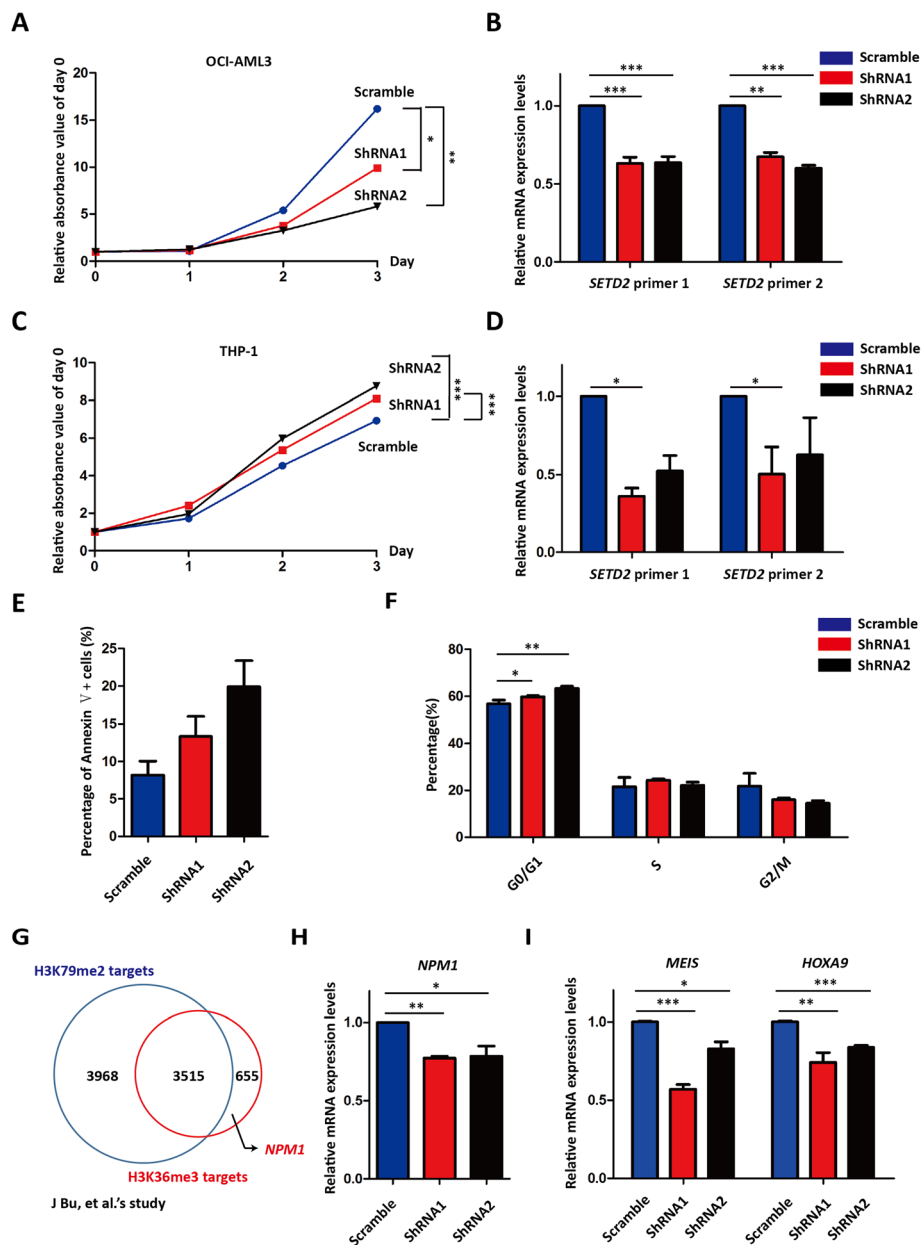


Fig. 2 *SETD2* was required for the maintenance of *NPM1*-mutated AML cell line OCI-AML3. **a** and **b** The proliferation (**a**) and *SETD2* expression (**b**) of scramble and *SETD2* knockdown OCI-AML3 cells. **c** and **d** The proliferation (**c**) and *SETD2* expression (**d**) of scramble and *SETD2* knockdown THP-1 cells. **e** Annexin-V staining for detecting cell apoptosis in OCI-AML3 cells. **f** PI staining for cell cycle analysis in OCI-AML3 cells. **g** *NPM1* has been demonstrated as one direct target of H3K36me3 in the literature report. **h** and **i** The expression of *NPM1* (**h**) and its direct downstream targets, *MEIS1* and *HOXA9* (**i**), was analyzed in scramble and *SETD2* knockdown OCI-AML3 cells. ***, $P < 0.001$; **, $P < 0.01$; *, $P < 0.05$; T test was used for each graph

additional genetic alteration influenced *SETD2* function in *NPM1*-mutated AML. Therefore, further investigations were needed in the future.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40364-020-00243-y>.

Additional file 1: Figure S1. The OS of *SETD2*- wild type and mutated AML patients from the summary of TCGA, TARGET, and OHSU studies.

Abbreviations

AL: Acute leukemia; ALL: Acute lymphoblastic leukemia; AML: Acute myeloid leukemia; BM: Bone marrow; ChIP-Seq: Chromatin immunoprecipitation-sequencing; CLAG: Cladribine, cytarabine plus granulocyte colony-stimulating factor; CR: Complete remission; EFS: Event-free survival duration; H3K36me3: Tri- methylated histone 3 lysine 36; HAA: Homoharringtonine, aclacinomycin, plus cytarabine regimen; HB: Hemoglobin; HSCT: Hematopoietic stem cell transplantation; HSPCs: Hematopoietic stem progenitor cells; IA: Idarubicin plus cytarabine regimen; MDS: Myelodysplastic syndrome; NR: No response; OR: Odds ratio; OS: Overall survival duration; PB: Peripheral blood; PLT: Platelet; PR: Partial remission; WBC: White blood cell

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Authors' contributions

X.Z. designed the experiments. W.-J. Y. collected and integrated clinical materials. J.-W. S. displayed the experiments. X. Z. integrated and analyzed all the data. X. Z. wrote the manuscript. J.-W. S. and W.-J. Y. revised the manuscript. The authors read and approved the final manuscript.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate

This study was approved by the ethical review committees of the First Affiliated Hospital to Zhejiang University School of Medicine.

Consent for publication

Written informed consent was obtained from this patient.

Competing interests

The authors declare that they have no competing interests.

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