

Repeated Remote Ischemic Conditioning Effect on Ankle-brachial Index in Diabetic Patients - A Randomized Control Trial

Abstract

Background: Remote ischemic preconditioning (RIPC) is a phenomenon where a short period of ischemia in one organ protects against further ischemia in the other organs. We hypothesized that RIPC occurring in diabetic patients with ankle brachial index (ABI) between 0.70 and 0.90 were included with peripheral arterial disease, would make the better coronary flow resulted in the increasing ABI. **Materials and Methods:** This randomized clinical trial study was done in the Afshar Cardiovascular Hospital in Yazd between 2013 and 2014. Sixty participants were randomly divided into two groups (intervention and control groups). The intervention group was undergoing RIPC, and the control group was tested without RIPC. RIPC was stimulated by giving three cycles of 5 min of ischemia followed by 5 min of reperfusion of both upper arms using a blood pressure cuff inflated to 200 mm Hg ($n = 30$). This was compared with no RIPC group which consisted of placing a deflated blood pressure cuff on the upper limbs ($n = 30$). **Results:** The mean of ABI level before intervention in the RIPC and control group was 0.82 ± 0.055 and 0.83 ± 0.0603 ($P = 0.347$) respectively, with no significant difference. It was 0.86 ± 0.066 in the RIPC group compared the control 0.83 ± 0.0603 ($P = 0.046$). So levels of ABI were greater after intervention in the RIPC group. The mean of ABI level increase from 0.82 ± 0.05 to 0.86 ± 0.06 in RIPC group ($P = 0.008$). So the intervention group showed a significant increase in ABI. **Conclusions:** RIPC through using a simple, noninvasive technique, composing three cycles of 5 min-ischemia of both upper arms, showing a significant increase in ABI level in diabetic patients.

Keywords: Ankle brachial index, peripheral arterial disease, remote ischemic preconditioning

Introduction

Peripheral arterial disease (PAD) is found in approximately 60% of patients with coronary artery disease.^[1] Coronary heart disease is the leading cause of morbidity and mortality in the worldwide.^[2]

PAD, as an ankle brachial index (ABI) of <0.9 , increases the long-term risk of cardiovascular and cerebrovascular events, and even death from any cause.^[3]

ABI is the ratio of the systolic blood pressure (SBP) measured at the ankle against one measured at the brachial artery; it was described by Winsor^[4] in 1950; this index was initially offered for the noninvasive diagnosis of lower-extremity PAD.^[5,6] In addition, it was shown that the ABI is an indicator of atherosclerosis at other vascular sites and can be seen as a prognostic marker for cardiovascular events, even in the absence of symptoms.^[7,8]

Reperfusion itself can lead to cell damage, which is known as ischemia-reperfusion injury (IRI). Cells in organs other than the heart are very sensitive to the protective effects of IRI of other tissue.^[9-11]

Ischemic preconditioning (IPC) is a phenomenon to avoid IRI in different vascular sites. The controlled repeated of short periods of ischemia could also protect tissues against the IRI.

IPC was first described in 1986 by Murry *et al.*^[12] as increased cellular resistance to myocardial ischemia when the heart is encountered periods of nonlethal ischemia interspersed with reperfusion. In 1993, Przyklenk *et al.*^[13] showed that increased cell resistance to ischemia also occurred in other tissues that were not directly subjected to ischemia. This phenomenon was named remote IPC (RIPC).^[14-29] The remote ischemic conditioning (RIC) stimulus could be noninvasively induced

Najmeh Shahvazian,
Mansour Rafiee,
Masoud
Rahmanian¹,
Seyed Kazem
Razavi-ratki²,
Mohammad Hadi
Farahzadi³

From the Cardiovascular Research Center; Departments of ¹Endocrinology and Metabolism and ²Nuclear Medicine, Shahid Sadoughi University of Medical Sciences and Health Services, Yazd, ³Department of Neuroscience, Faculty of Advanced Technologies in Medicine, Tehran University of Medical Sciences, Tehran, Iran

Address for correspondence:
Dr. Mohammad Hadi Farahzadi,
Department of Neuroscience,
Faculty of Advanced
Technologies in Medicine,
Tehran University of Medical
Sciences, Tehran, Iran.
E-mail: dr.farahzadi@yahoo.
com

Access this article online

Website: www.advbiores.net

DOI: 10.4103/2277-9175.201685

Quick Response Code:



How to cite this article: Shahvazian N, Rafiee M, Rahmanian M, Razavi-ratki SK, Farahzadi MH. Repeated Remote Ischemic Conditioning Effect on Ankle-brachial Index in Diabetic Patients - A Randomized Control Trial. *Adv Biomed Res* 2017;6:28.

Received: June, 2015. **Accepted:** September, 2015.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

using a standard blood pressure cuff placed on the upper arm or leg.^[30]

The protective effect by IPC has two windows of protection. The first lasts between 4 and 6 h^[31] and has been named early preconditioning. The second begins at 24 h lasting up to 72 h after the ischemia and reperfusion stimulus.^[32]

Several mechanisms show the phenomenon of RIPC, such as suppression of inflammatory genes, modulation of ATP-sensitive K⁺ channels, nuclear factor kappa beta, nitric oxide synthesis, or free radicals pathways.^[33]

Based on the evidence of RIPC occurring in the tissues, we predicted that RIPC that occurs in patients with PAD and ABI <0.90, making the better coronary flow that increasing ABI. To test this hypothesis, we performed and compared ABI in patients with and without RIPC.

Materials and Methods

Out of 98 investigated patients, 60 individuals were accepted for this randomized clinical trial study conducted in the Afshar Cardiovascular Hospital in Yazd between 2013 and 2014. About 15 patients did not accept the study protocol and 20 patients did not fulfill the inclusion criteria. The written and oral consent was received from all of the participants. This study was approved by the Ethics Committee of the Shahid Sadoughi University of Medical Sciences.

Diabetic patients with ABI between 0.70 and 0.90 were included. Patients with a history of cerebrovascular accident, or glomerular filtration rate lower than 30 ml/min were excluded. Physical examinations showed normal physiology of the upper limbs in all of the investigated participants.

The participants were randomly divided into two groups (intervention and control groups). The intervention group was undergoing RIPC, and the control group was tested without receiving RIPC. Although those colleagues who both measured the ABI and conducted the statistical analysis were not aware of the engaged groups, the patients were not blinded.

At first ABI was measured with blood pressure cuff, sphygmomanometer, and handheld Doppler devices (Summit Doppler L250 ABI); then calculation was performed that divided the higher of the dorsalis pedis or posterior tibial systolic pressure for each ankle by the higher of the two brachial SBP to get the ABI for each leg; next, RIPC was stimulated by giving three cycles of 5 min of ischemia followed by 5 min of reperfusion of both upper arms using a blood pressure cuff inflated to 200 mm Hg ($n = 30$). This was compared to no RIPC, which consisted of placing a deflated blood pressure cuff on the upper limbs ($n = 30$).

The participants were advised to avoid using the following substances, because of their interfering roles during the

process of IPC (RIPC), within 2 h of the test: Cilostazol, sildenafil, dipyridamol, glibenclamide, nicorandyl, phenylephrine, angiotensin-converting enzyme inhibitors, angiotensin receptor blocker II, statins and steroids, caffeine and alcohol.

Statistical analysis

To compare the general characteristics of the two experimental groups, we used the Chi-squared or Fishers' exact test for categorical variables. To compare ABI in each group paired *t*-test and between the two groups, independent *t*-test were used considering the two groups as an independent factor and the ABI as the repeated factor.

Data presented in the form of frequencies, means, and standard deviations. For all of the analyzes, a significance level of $P < 0.05$ was assumed.

Results

Totally 60 patients were analyzed in this study, including 21 males and 39 females ranging from 40 to 74 years old (average 57 years). General patient's characteristics are shown in Table 1. There were no differences between the groups regarding age, gender, hyperlipidemia, hypertension prevalence, and smoking history.

The mean of ABI before and after intervention (RIPC) is shown in Table 2. The mean of ABI before intervention in the RIPC group was 0.82 ± 0.055 in comparison with the control group 0.83 ± 0.0603 ($P = 0.347$), so no significant differences before intervention were seen between groups.

The mean of ABI after intervention in the RIPC group was 0.86 ± 0.066 in comparison with the control group 0.83 ± 0.0603 ($P = 0.046$). Hence, comparing

Table 1: General characteristics of participants

Variables	Intervention group ($n=30$)	Control group ($n=30$)	<i>P</i>
Age (years)			
Mean±SD	56.57±8.7	55.9±9.39	0.784
Gender (%)			
Female	18 (60)	21 (70)	0.294
Male	12 (40)	9 (30)	
Risk factors (%)			
Hypertension			
Yes	25 (83.3)	23 (76.7)	0.374
No	5 (16.7)	7 (23.3)	
Hyperlipidemia			
Yes	17 (56.7)	16 (53.3)	0.5
No	13 (43.3)	14 (46.7)	
History of smoking			
Yes	7 (23.3)	5 (16.7)	0.374
No	23 (76.7)	25 (83.3)	

Values are expressed as the mean±SD. SD: Standard deviation

Table 2: Effect of RIPC on the ABI in the two experimental groups

Groups	Intervention	Control	P
ABI			
Before (mean±SD)	0.82±0.055	0.83±0.0603	0.347
After (mean±SD)	0.86±0.066	0.83±0.0603	0.046
P	0.008	1	

Values are expressed as the mean±SD. SD: Standard deviation, RIPC: Remote ischemic preconditioning, ABI: Ankle brachial index

the control group, the mean of ABI were greater after intervention in the RIPC group.

The mean of ABI level increased from 0.82 ± 0.05 to 0.86 ± 0.06 in RIPC group ($P = 0.008$). The intervention group showed a significant increase in the ABI in comparison with control group.

Discussion

In this study, we compared the ABI before and after RIPC. Our results showed a significant increase in ABI level in diabetic patients with $0.70 \leq \text{ABI} \leq 0.90$ after RIPC.

Przyklenk *et al.* indicated that small cycles of coronary artery occlusion in dogs maintain the myocardial cells from longer periods of ischemia.^[13] In addition to the cardiac muscle other organs have been shown to reply to the protective impress of RIPC, including the lungs, kidneys, liver, and skeletal muscle.^[34-37]

Hausenloy *et al.* illustrated three 5 min cuff inflations and deflations of a cuff placed on the upper arm to 200 mm Hg, administered before cardiac surgery preoperative myocardial injury (43% less troponin T release) in patients undergoing elective coronary artery bypass grafting surgery.^[38]

Hausenloy and Yellon and Tapuria *et al.* reported that the mechanisms underlying the phenomenon of RIC can be noted as three interrelated events: (1) The initial events occurring in the organs in response to the RIC irritant. The application of brief episodes of RIC (Remote Ischemic Conditioning) to the remote organisms believed to produce endogenous factors which can keep them from injury. (2) The transmission of the protective signal may be multi-factorial blood factor(s), neuronal mechanisms, and systemic responses. (3) The events are occurring in the target organ or tissue which present the protective effect.^[39,40]

RIPC may be mediated by humoral and neurogenic factors consisting of an early phase that starts about 24 h after RIPC and lasts for nearly 48 h.

Several humoral factors involved in RIPC phenomenon, including opioids, nitric oxide, adenosine, catecholamines, bradykinin, tumor necrosis factor-alpha, free radicals, prostaglandins and angiotensin.

Organs exhibit to ischemia have been shown to induct the protein kinase C intracellular pathways resulting in the nuclear translocation of nuclear factor kappa beta and the activation of nitric oxide synthesis. Current mechanisms involved in target organs and tissues in the early and late process of RIPC stay indeterminate; however, some studies have offered roles for mitochondrial KATP channels and neutrophils.

RIPC was also found to provoke neovascularization in the ischemic myocardium by up-regulation of vascular endothelial growth factor gene expression that limit the infarct size.^[41] Notwithstanding its promise in animal studies, the clinical benefit of RIPC in human studies remains controversial.

Administration of KATP channel blockers prevents IPC in healthy volunteers, and IPC phenomenon is mimicked by KATP channel opening drugs.^[42,43] A number of studies proposed that IRI is dependent on increased oxidative stress,^[44,45] making it possible that IPC and RIPC stimulate antioxidant defenses.^[46]

Recent studies have discovered the RIPC that occurs in patients with PAD and low ABI^[47-49] might lead to the elevation of the circulating levels of some anti-inflammatory, vasodilator, or angiogenesis-inducing contents.

Repeated ischemia-reperfusion occurrence caused by physical training could be irritant for intracellular biochemical changes leading to a more valid use of oxygen by the muscle that ameliorated the endothelial function.^[41]

Conclusion

RIPC is a potent innate protective mechanism against ischemic injury. This study demonstrates that RIPC through using a simple, noninvasive technique including three cycles of 5 min of ischemia followed by 5 min of reperfusion of both upper arms showed a significant increase in ABI level in diabetic patients with $0.70 \leq \text{ABI} \leq 0.90$. But the mechanism is not yet fully revealed; however, it has indicated a promising point in the clinical trials. No enough trial has been put forward to illustrate the effect of RIPC to decline the incidence of clinically relevant sequel of IRI. However, more powerful studies are necessary to report the case within the next 3–4 years.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- Hertzer NR, Beven EG, Young JR, O'Hara PJ, Ruschhaupt WF 3rd, Graor RA, *et al.* Coronary artery disease in

- peripheral vascular patients. A classification of 1000 coronary angiograms and results of surgical management. *Ann Surg* 1984;199:223-33.
2. Lloyd-Jones D, Adams R, Carnethon M, De Simone G, Ferguson TB, Flegal K, *et al.* Heart disease and stroke statistics-2009 update: A report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation* 2009;119:e21-181.
 3. Bhatt DL, Peterson ED, Harrington RA, Ou FS, Cannon CP, Gibson CM, *et al.* Prior polyvascular disease: Risk factor for adverse ischaemic outcomes in acute coronary syndromes. *Eur Heart J* 2009;30:1195-202.
 4. Winsor T. Influence of arterial disease on the systolic blood pressure gradients of the extremity. *Am J Med Sci* 1950;220:117-26.
 5. Carter SA. Indirect systolic pressures and pulse waves in arterial occlusive diseases of the lower extremities. *Circulation* 1968;37:624-37.
 6. Yao ST, Hobbs JT, Irvine WT. Ankle systolic pressure measurements in arterial disease affecting the lower extremities. *Br J Surg* 1969;56:676-9.
 7. Criqui MH, Langer RD, Fronek A, Feigelson HS, Klauber MR, McCann TJ, *et al.* Mortality over a period of 10 years in patients with peripheral arterial disease. *N Engl J Med* 1992;326:381-6.
 8. Ankle Brachial Index Collaboration, Fowkes FG, Murray GD, Butcher I, Heald CL, Lee RJ, *et al.* Ankle brachial index combined with Framingham Risk Score to predict cardiovascular events and mortality: A meta-analysis. *JAMA* 2008;300:197-208.
 9. Ates E, Genç E, Erkasap N, Erkasap S, Akman S, Firat P, *et al.* Renal protection by brief liver ischemia in rats. *Transplantation* 2002;74:1247-51.
 10. Mallick IH, Yang W, Winslet MC, Seifalian AM. Ischaemic preconditioning improves microvascular perfusion and oxygenation following reperfusion injury of the intestine. *Br J Surg* 2005;92:1169-76.
 11. Tapuria N, Kumar Y, Habib MM, Abu Amara M, Seifalian AM, Davidson BR. Remote ischemic preconditioning: A novel protective method from ischemia reperfusion injury – A review. *J Surg Res* 2008;150:304-30.
 12. Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: A delay of lethal cell injury in ischemic myocardium. *Circulation* 1986;74:1124-36.
 13. Przyklenk K, Bauer B, Ovize M, Kloner RA, Whittaker P. Regional ischemic ‘preconditioning’ protects remote virgin myocardium from subsequent sustained coronary occlusion. *Circulation* 1993;87:893-9.
 14. Przyklenk K. Efficacy of cardioprotective ‘conditioning’ strategies in aging and diabetic cohorts: The co-morbidity conundrum. *Drugs Aging* 2011;28:331-43.
 15. Przyklenk K, Darling CE, Dickson EW, Whittaker P. Cardioprotection ‘outside the box’ – the evolving paradigm of remote preconditioning. *Basic Res Cardiol* 2003;98:149-57.
 16. Przyklenk K, Frelinger AL 3rd, Linden MD, Whittaker P, Li Y, Barnard MR, *et al.* Targeted inhibition of the serotonin 5HT2A receptor improves coronary patency in an *in vivo* model of recurrent thrombosis. *J Thromb Haemost* 2010;8:331-40.
 17. Przyklenk K, Heusch G. Late preconditioning against myocardial stunning. Does aspirin close the “second window” of endogenous cardioprotection? *J Am Coll Cardiol* 2003;41:1195-7.
 18. Przyklenk K, Li G, Simkhovich BZ, Kloner RA. Mechanisms of myocardial ischemic preconditioning are age related: PKC-epsilon does not play a requisite role in old rabbits. *J Appl Physiol* 2003;95:2563-9.
 19. Przyklenk K, Li G, Whittaker P. No loss in the *in vivo* efficacy of ischemic preconditioning in middle-aged and old rabbits. *J Am Coll Cardiol* 2001;38:1741-7.
 20. Przyklenk K, Maynard M, Darling CE, Whittaker P. Pretreatment with D-myo-inositol trisphosphate reduces infarct size in rabbit hearts: Role of inositol trisphosphate receptors and gap junctions in triggering protection. *J Pharmacol Exp Ther* 2005;314:1386-92.
 21. Przyklenk K, Maynard M, Darling CE, Whittaker P. Aging mouse hearts are refractory to infarct size reduction with post-conditioning. *J Am Coll Cardiol* 2008;51:1393-8.
 22. Przyklenk K, Maynard M, Greiner DL, Whittaker P. Cardioprotection with postconditioning: Loss of efficacy in murine models of type-2 and type-1 diabetes. *Antioxid Redox Signal* 2011;14:781-90.
 23. Przyklenk K, Maynard M, Whittaker P. First molecular evidence that inositol trisphosphate signaling contributes to infarct size reduction with preconditioning. *Am J Physiol Heart Circ Physiol* 2006;291:H2008-12.
 24. Przyklenk K, Maynard M, Whittaker P. Reduction of infarct size with D-myo-inositol trisphosphate: Role of PI3-kinase and mitochondrial K(ATP) channels. *Am J Physiol Heart Circ Physiol* 2006;290:H830-6.
 25. Przyklenk K, Undyala VV, Wider J, Sala-Mercado JA, Gottlieb RA, Mentzer RM Jr. Acute induction of autophagy as a novel strategy for cardioprotection: Getting to the heart of the matter. *Autophagy* 2011;7:432-3.
 26. Przyklenk K, Whittaker P. *In vitro* platelet responsiveness to adenosine-mediated “preconditioning” is age-dependent. *J Thromb Thrombolysis* 2005;19:5-10.
 27. Przyklenk K, Whittaker P. Cardioprotection with adenosine: ‘A riddle wrapped in a mystery’. *Br J Pharmacol* 2005;145:699-700.
 28. Przyklenk K, Whittaker P. Adaptation of a photochemical method to initiate recurrent platelet-mediated thrombosis in small animals. *Lasers Med Sci* 2007;22:42-5.
 29. Przyklenk K, Whittaker P. Cardioprotection via adaptation to hypoxia: Expanding the timeline and targets? *Basic Res Cardiol* 2011;106:325-8.
 30. Kharbanda RK, Mortensen UM, White PA, Kristiansen SB, Schmidt MR, Hoschitzky JA, *et al.* Transient limb ischemia induces remote ischemic preconditioning *in vivo*. *Circulation* 2002;106:2881-3.
 31. Rodrigo GC, Samani NJ. Ischemic preconditioning of the whole heart confers protection on subsequently isolated ventricular myocytes. *Am J Physiol Heart Circ Physiol* 2008;294:H524-31.
 32. Das M, Das DK. Molecular mechanism of preconditioning. *IUBMB Life* 2008;60:199-203.
 33. Saes GF, Zerati AE, Wolosker N, Ragazzo L, Rosoky RM, Ritti-Dias RM, *et al.* Remote ischemic preconditioning in patients with intermittent claudication. *Clinics (Sao Paulo)* 2013;68:495-9.
 34. Ates E, Genç E, Erkasap N, Erkasap S, Akman S, Firat P, *et al.* Renal protection by brief liver ischemia in rats. *Transplantation* 2002;74:1247-51.
 35. Harkin DW, Barros D’Sa AA, McCallion K, Hoper M, Campbell FC. Ischemic preconditioning before lower limb ischemia – reperfusion protects against acute lung injury. *J Vasc Surg* 2002;35:1264-73.
 36. Lai IR, Chang KJ, Chen CF, Tsai HW. Transient limb ischemia induces remote preconditioning in liver among rats: The protective role of heme oxygenase-1. *Transplantation* 2006;81:1311-7.
 37. Addison PD, Neligan PC, Ashrafpour H, Khan A, Zhong A,

- Moses M, *et al.* Noninvasive remote ischemic preconditioning for global protection of skeletal muscle against infarction. *Am J Physiol Heart Circ Physiol* 2003;285:H1435-43.
38. Hausenloy DJ, Mwamure PK, Venugopal V, Harris J, Barnard M, Grundy E, *et al.* Effect of remote ischaemic preconditioning on myocardial injury in patients undergoing coronary artery bypass graft surgery: A randomised controlled trial. *Lancet* 2007;370:575-9.
 39. Hausenloy DJ, Yellon DM. Remote ischaemic preconditioning: Underlying mechanisms and clinical application. *Cardiovasc Res* 2008;79:377-86.
 40. Tapuria N, Kumar Y, Habib MM, Abu Amara M, Seifalian AM, Davidson BR. Remote ischemic preconditioning: A novel protective method from ischemia reperfusion injury – A review. *J Surg Res* 2008;150:304-30.
 41. Stewart KJ, Hiatt WR, Regensteiner JG, Hirsch AT. Exercise training for claudication. *N Engl J Med* 2002;347:1941-51.
 42. Cleveland JC Jr, Meldrum DR, Cain BS, Banerjee A, Harken AH. Oral sulfonyleurea hypoglycemic agents prevent ischemic preconditioning in human myocardium. Two paradoxes revisited. *Circulation* 1997;96:29-32.
 43. Klepzig H, Kober G, Matter C, Luus H, Schneider H, Boedeker KH, *et al.* Sulfonylureas and ischaemic preconditioning; a double-blind, placebo-controlled evaluation of glimepiride and glibenclamide. *Eur Heart J* 1999;20:439-46.
 44. Clarke SJ, Khaliulin I, Das M, Parker JE, Heesom KJ, Halestrap AP. Inhibition of mitochondrial permeability transition pore opening by ischemic preconditioning is probably mediated by reduction of oxidative stress rather than mitochondrial protein phosphorylation. *Circ Res* 2008;102:1082-90.
 45. Weinbrenner C, Schulze F, Sárváry L, Strasser RH. Remote preconditioning by infrarenal aortic occlusion is operative via delta1-opioid receptors and free radicals *in vivo* in the rat heart. *Cardiovasc Res* 2004;61:591-9.
 46. Yellon DM, Downey JM. Preconditioning the myocardium: From cellular physiology to clinical cardiology. *Physiol Rev* 2003;83:1113-51.
 47. Blann AD, Belgore FM, McCollum CN, Silverman S, Lip PL, Lip GY. Vascular endothelial growth factor and its receptor, Flt-1, in the plasma of patients with coronary or peripheral atherosclerosis, or Type II diabetes. *Clin Sci (Lond)* 2002;102:187-94.
 48. Makin AJ, Chung NA, Silverman SH, Lip GY. Vascular endothelial growth factor and tissue factor in patients with established peripheral artery disease: A link between angiogenesis and thrombogenesis? *Clin Sci (Lond)* 2003;104:397-404.
 49. Findley CM, Mitchell RG, Duscha BD, Annex BH, Kontos CD. Plasma levels of soluble Tie2 and vascular endothelial growth factor distinguish critical limb ischemia from intermittent claudication in patients with peripheral arterial disease. *J Am Coll Cardiol* 2008;52:387-93.