Briefings in Bioinformatics, 17(3), 2016, 505-516

doi: 10.1093/bib/bbv063 Advance Access Publication Date: 5 August 2015 Paper

Reasoning over genetic variance information in causeand-effect models of neurodegenerative diseases

Mufassra Naz, Alpha Tom Kodamullil and Martin Hofmann-Apitius

Corresponding author: Martin Hofmann-Apitius. Head of the Department of Bioinformatics, Fraunhofer Institute for Algorithms and Scientific Computing (SCAI), 53754 Sankt Augustin, Germany. E-mail: martin.hofmann-apitius@scai.fraunhofer.de

Abstract

The work we present here is based on the recent extension of the syntax of the Biological Expression Language (BEL), which now allows for the representation of genetic variation information in cause-and-effect models. In our article, we describe, how genetic variation information can be used to identify candidate disease mechanisms in diseases with complex aetiology such as Alzheimer's disease and Parkinson's disease. In those diseases, we have to assume that many genetic variants contribute moderately to the overall dysregulation that in the case of neurodegenerative diseases has such a long incubation time until the first clinical symptoms are detectable. Owing to the multilevel nature of dysregulation events, systems biomedicine modelling approaches need to combine mechanistic information from various levels, including gene expression, microRNA (miRNA) expression, protein–protein interaction, genetic variation and pathway. OpenBEL, the open source version of BEL, has recently been extended to match this requirement, and we demonstrate in our article, how candidate mechanisms for early dysregulation events in Alzheimer's disease can be identified based on an integrative mining approach that identifies 'chains of causation' that include single nucleotide polymorphism information in BEL models.

Key words: BEL model; Alzheimer's disease; genetic variants; GWAS; causal reasoning; cause-and-effect

Systems biology models and genetic variation: two separate worlds

Barabási *et al.* [1] assert 'given the functional interdependencies between the molecular components in a human cell, a disease is rarely a consequence of an abnormality in a single gene, but reflects the perturbations of the complex intracellular and intercellular network'.

Genome-wide genetic association studies (GWAS) have become a useful and frequently used tool for discovering genetic variants as a disease risk [2]. However, for complex traits and phenotypes, interpretation of association data largely benefits from available prior biological and environmental knowledge, spanning over multiple scientific disciplines [3]. In human genetics, several strategies were developed and implemented to determine the effect of single nucleotide polymorphisms (SNPs), particularly, for the analysis of genotyping data. The limitation of many of these algorithms is that they can predict only either to have no effect or to have negative effect on clinical readouts and endpoints. However, the spectrum of possible biological effects caused by genetic variants is much wider, and thus, methods are required to predict also potential gain, loss or even modification of gene function [4]. Moreover, most of the algorithms can predict only variant effects on individual proteins [5], and machine learning supervised and semisupervised approaches are being used to predict the effect of deleterious SNPs [4]. Generally, GWA studies are used to establish links between genotypes and phenotypes through

Submitted: 2 April 2015; Received (in revised form): 9 July 2015

© The Author 2015. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Mufassra Naz is a researcher at Fraunhofer SCAI and PhD student at the University of Bonn. Her main research interests are functional interpretation of genetic variants, causal algorithms and neurodegenerative diseases.

Alpha Tom Kodamullil is a researcher at Fraunhofer SCAI and PhD student at the University of Bonn. Her research work focuses on automatic expansion of computable disease models by developing reasoners.

Martin Hofmann-Apitius is Professor for Applied Life Science Informatics at the University of Bonn. He is also the Head of the Department of Bioinformatics at Fraunhofer Institute SCAI.

identifying the differences (and commonalities) between thousands of individuals. These approaches work as black boxes and make use of statistical and machine learning approaches that require huge data sets.

To reveal the functional context at the molecular level, substantial knowledge about the genes involved, their expression at RNA and protein level, the time when they are expressed and in which tissue and in which organ, is required. Regulation of gene expression is mediated through genetic regulatory systems, which are controlled by complex interaction networks involving DNA, RNA, proteins and small molecules. These regulatory networks involve many components linked to each other by positive and negative feedback loops, and a deterministic understanding of their dynamics is hard to attain owing to rapidly increasing complexity. Therefore, specialized methods and computer software are essential, for the modelling and simulation of genetic regulatory networks [6].

Systems biology is the systemic contextual representation and modelling of a plurality of discrete observations. In systems biology, modelling is a representation of disease high-level concepts in a unified and comprehensive network that can help to identify the differential subnetworks by comparing it with a network representing the healthy state [1, 7–10]. Building blocks of systems biology models, such as signalling pathways, metabolic systems and gene regulation networks, are already widely used in computational biology. Comprehensive disease models, however, are going way beyond these comparably well-understood functional modules. One of the explicit goals of systems biomedicine is to use a generalized model of disease to assess the parameters from high-throughput data of a single patient, to generate 'personalized models' that predict disease progression and treatment responses [11, 12].

In systems biology, there are several 'entry points' to generate initial networks: protein-protein interaction, metabolic networks and signalling pathways have been widely used to model biological processes [7]. In the past decade, however, new modelling approaches have been developed [13]. For pharmacogenomics, these networks represent complex relationships between drugs and targets. The diseasome [14] is a diseasegene and drug-target (protein) network, where disease information is associated with a gene and drugs are linked to proteins by drug-target associations [15].

Despite the complexity of regulatory networks, attempts at unravelling the impact of genetic variation on regulatory networks have been addressed by a number of groups. Leiserson et al. [16], Carter et al. [17] and Atias et al. [18] have worked on network approaches to scrutinize the genetic risks for human disease. They have developed methodology that allows to detect causal genes within disease-associated loci by network analysis, and to ascertain causal paths from allele to disease through intermediate molecular phenotypes [16-18]. Trynka et al. [19] proposed new approaches on the interpretation of transcriptional regulation effects to estimate the involvement of variant alleles in common diseases. They suggested that most of the causal complex trait variants have regulatory roles with cell type specificity, by interconnecting GWAS data with genome-wide chromatin assays results. They emphasized the importance of cell-type-specific regulatory context and highlighted the value of the inclusion of epigenomics information [19].

Sahni et al. [20] questioned the strong bias in the literature towards coding variant effects on protein–DNA, protein–RNA and protein–protein interactions. He proposed to put more emphasis on effects outside the protein centric scope of functional assessment, to understand the impact of genetic variants on specific interactions; for instance, mechanisms safeguarding protein folding and stability [20].

Types of genetic variation information relevant for systems biomedicine

If genetic variation is to be included in a systems biology model of disease, we need to assess the biological impact of a SNP or a mutation. Dependent on the way (the 'mode-of-action') how a SNP or a mutation exerts its biological impact, we can distinguish several classes ('types') of SNPs. In this section, we identify and discuss the different functional categories that can be distinguished as 'mode-of-SNP-action' classes (see Table 1).

Genetic variants on coding regions

The risk associated with non-synonymous genetic variants can be easily translated into a change in protein structure or function owing to a change in the amino acid sequence. It can modify amino acid composition, or truncate the protein sequence by causing an early stop codon [21]. Synonymous genetic variants do not alter the codon sequence. However, synonymous genetic risk variants can still impact protein function by modulating translation rates with direct consequences to protein folding [22]. For example, rs1045642 SNP slows down the rate of translation of the MDR1 mRNA and impacts protein folding [23]. Exon splicing enhancers or silencers are typically 6-8 consecutive nucleotide sequences in an exon region. Where, SNP can also result in deleterious intron retention or exon skipping, and translate the protein isoform [24-27]. For example, rs1800693 SNP affects the splicing of the TNFRSF1A mRNA, leading to translate an isoform [28].

Genetic variants on non-coding regions

Model gene system studies have revealed that local DNA interactions between regulatory sites and genes are important for transcriptional control. Such regulatory interactions, in mammals, can take place over significant chromosomal distances up to an entire mega-base (1 Mb) [29]. Genetic risk variants are frequent on non-coding sequences [30]. Post-GWAS studies have revealed the capacity of these genetic risk variants to regulate gene expression by modulating cis-regulatory machineries through mechanisms involving DNA methylation [31], transcription factor binding [32], chromatin looping [33] or microRNA (miRNA) recruitment [34]. If SNPs occur within transcriptional regulatory regions, like transcription factor binding sites, CpG islands and miRNAs, they may modify the binding affinity of the regions, remove recognition sites or create new binding sites for other regulatory proteins. All of these modifications can lead to alterations in the level, timing and localization of gene expression [35].

DNA methylation

DNA methylation means addition of methyl groups to a cytosine nucleotide, which is basically part of a CpG dinucleotide [36]. DNA hyper-methylation near transcription start sites of tumour suppressor genes associates with their silencing [37].

Transcription factor binding to regulatory elements

Across the genome, transcription factors bind to thousands of regulatory elements, including promoters (directly upstream of their target genes) and cis-regulatory elements such as

Types of generic variation morniation relevant for systems biometicine						
DNA regions		Functional categories	Functional consequences			
1.	Coding regions	1. Non-synonymous genetic variants	Change in protein structure or function due to a change in the amino acidsequence or protein sequence truncation			
		2. Synonymous genetic variants	Modulating translation rates with direct consequences to pro- tein folding			
		3. Exon splicing enhancers or silencers	Translate the protein isoform by deleterious intron retention or exon skipping			
2.	Non-coding regions	1. DNA methylation	Associates with genes silencing			
		2. Transcription factor binding to regulatory elements	Can change transcription factor binding to DNA that leads to differential target gene expression			
		 Chromatin loop bridging the enhancers and promoters 	Can alter the DNA affinity for looping factors and chromatin interactions, which regulates gene expression			
		4. MiRNAs	Can affect gene functionality: (i) by transcription of primary transcript, (ii) by pri-miRNA and pre-miRNA processing and (iii) by effecting miRNA-miRNA interaction			
		5. lncRNAs	Can modify highly conserved lncRNA tertiary structure that can affect chromatin regulator's interactions			

Table 1. Types of genetic variation information relevant for systems biomedicine: DNA regions with functional categories and consequences

enhancers, insulators and silencers [38]. Genetic risk variants					
located within promoter regions can also change transcription					
factor binding to DNA, leading to differential target gene expres-					
sion [39, 40]. For example, expression of the α -globin gene locus					
is affected by a genetic variant associated with the a-thalassae-					
mia blood disorder [39]. Enhancers are commonly targeted by					
those genetic variants of risk-associated loci that map to DNA					
recognition motifs, bound by transcription factors. These gen-					
etic variants can modulate the chromatin affinity for transcrip-					
tion factors and consequently gene expression [33, 41–46, 47].					
For example, the rs12740374 SNP, which is associated with a					
lower level of plasma low-density lipoprotein cholesterol, in-					
creases the expression level of the SORT1 (Sortilin 1) gene by					
increasing the binding affinity of the C/EBP (CCAAT enhancer-					
binding protein) transcription factor to chromatin [33].					

Types of genetic variation information relevant for systems biomedicine

Chromatin loop bridging the enhancers and promoters

Genetic risk variants can modulate chromatin loop formation; it can alter the DNA affinity for looping factors, which results in allele-specific chromatin loop formation. The human genome is structured in a three-dimensional architecture, which is thought to regulate a diverse set of DNA-template processes [47–52]. This facilitates regulatory elements, like promoters and enhancers, to interact physically through long-range chromatin loops, or chromatin interactions, to regulate gene expression [53, 54]. This has been shown for the rs12913832 SNP, which resides in an enhancer 21 Kb upstream of the OCA2 (Oculocutaneous albinism II) pigment gene [55]. Over the past decade, the development of chromosome conformation capture (3C) technology has initiated several 3D studies on regulatory chromatin loops, but what has been done until now is far from exhaustive. If a minor fraction of these potential regulatory elements participate in chromatin looping, then most of the genomic interactions have yet to be characterized again, because many such loops appear to be tissue-specific [56-58], which makes their comprehensive analysis appear even more disconcerting [59].

MicroRNAs

miRNAs target mRNAs by recognizing their complementary sequences mainly in 3' untranslated regions (3'UTRs). miRNAs largely function as post-transcriptional repressors. They recruit RNA-induced silencing complex to their target mRNAs, leading to mRNA degradation or translation repression [60]. They can regulate the translation of hundreds of genes through sequence-specific binding to mRNA [61]. SNP variants, linked with miRNAs, can affect gene functionality with three different ways: (i) by transcription of primary transcript, (ii) by primary miRNA (pri-miRNA) and precursor miRNA (pre-miRNA) processing and (iii) by effecting miRNA-miRNA interaction [62]. For example, rs10065172, a Crohn's disease-associated SNP, lies within the 3'UTR of the IRGM (immunity-related GTPase M) gene and alters the complementary target sequence of miRNA-196 [36].

Long non-coding RNAs

Long non-coding RNAs (lncRNAs) are found across intergenic regions of the human genome [63]. They can interact with chromatin regulators for their recruitment by chromatin [64, 65], a process that relies on a highly conserved lncRNA tertiary structure, which can be changed by genetic risk variants [66]. Kim et al. [67] described enhancer RNAs (eRNAs), a new class of noncoding RNAs, formed from polymerase II-bound enhancers. The level of expression of eRNAs is positively correlated with the expression of neighbouring coding genes [67] Genetic variants in enhancer sequences can modify Transcription Factor (TF) binding, resulting in 'improper' gene expression and eventually susceptibility to diseases [68, 69]. The micropeptides, called small pri-peptides, are also expressed from the lncRNA-pri and direct the proteolytic cleavage or other modifications of target proteins or transcription factors [70].

Expression quantitative trait loci

Studying the association between genetic variation and gene expression offers a straightforward way to begin the complicated task of connecting risk variants to their putative target genes [71]. Networks created using gene expression data from patient samples can be exploited to bridge GWAS results with an underlying disease mechanism, as exemplified in the autism spectrum disorder [72]. Genetic variation associated with gene expression, known as expression quantitative trait loci (eQTL), can identify the target genes of risk loci [73–77]. Polymorphism situated in DNA regulatory elements can alter the gene transcript frequency. Thus, as a quantitative trait locus, gene transcript frequency can be determined with substantial power [78, 79]. Brem et al. [80] published the first genome-wide study of gene expression in 2002. Stranger and Raj [81] reviewed the genetics of human variation and diversity in eQTLs. These eQTL data are dynamic with great specificity for different tissues and environmental perturbations.

The ENCODE project: identification of genomic functional elements

The ENCODE project has delivered an incredible compilation of genetic functional elements of the human genome [82]. As most of the SNPs detected in GWAS data belong to non-coding regions of the genome, usage of ENCODE regulatory elements to reinterpret GWAS data sets might be a valuable approach [83]. Undoubtedly, structural genomic variation are more influential and systemic than the smaller scale variations; however, any framework or methodology used to predict genetic variant effects needs to contribute for both small- and large-scale variations [13]. If possible, it should be able to predict the level in which coding or non-coding genetic variants individually or collectively have a functional impact on biology, ranging from relevant protein function or expression to the perturbation of entire networks. It can help us to annotate the massive amount of resequencing data meaningfully without having to test the effects of all variants experimentally [13].

Thus, now it is the time to move ahead from merely bio-statistical approaches for GWAS data interpretations to a more comprehensive approach that can be acquainted with genegene and gene-environment interactions, along with the complexity of the relationship between genotype and phenotype [84].

The need to integrate genetic variant information in systems biomedicine models

Currently, GWAS variance data interpretation has become a bottleneck in the progression of mapping and exploring complex diseases. For example, multiple genes have been associated with Amyotrophic lateral sclerosis in GWAS data, but there is no clear perspective of involved pathways and mechanisms that would emerge from the available high-throughput data, by taking multiple rare variants into account [85].

Substantial research for several complex diseases has been conducted to unravel causal mechanisms underlying their disease aetiology. Often this type of research is multidisciplinary, using research studies spreading over a wide range of time and length scales. Consequently, a disease model representing disease aetiology may have many modules and interactions. Such a disease model would provide a nice template for the interpretation of the functional consequences of genetic variation [86].

One of the obvious questions is of course, which methodology can help in interpretation of GWAS data, when most of the SNPs have small effects on disease susceptibility [87]. There is lack of efficient and reliable algorithms as well as appropriate multi-scale modelling methodology, to evaluate the huge number of interdependent data from GWAS [5]. One way to reduce the combinatorial complexity of GWAS data is, to reduce the dimensionality of genetic variation data by taking a priori knowledge about functional relationships between genes and proteins into account. Formalized knowledge about causal and correlative relationships in systems biology models provides a good starting point for that dimension reduction. So far, there have been only few serious efforts to predict how these genetic variants would collectively be effective for specific phenotypes [88, 89].

Systems biology modelling language syntax adaptations

A massive amount of data for molecular interactions and pathways are stored in online databases. Moreover, experimental data are accumulating rapidly, and correspondingly, the demand for exchange of data to allow analysis and comparison of larger data sets is intensifying. Thus, there is a need for representation of data in standardized formats. Comparisons and evaluations of modern systems biology modelling languages show [90,91] that XML is a remarkable and easy-to-use format for systems biology information representation. Here, we compare the recent updates with the standard XML-based representation formats for exchange of data.

The Resource Description Framework

The Resource Description Framework (RDF) model [92] is based on the idea of making statements about resources. A RDF statement, also called a triple in RDF terminology, is an association of the form (subject, predicate, object). RDF Schema [93] and the Web Ontology Language [94] are used to explicitly represent the meanings of the resources described on the Web and how they are related. These specifications, called ontologies, describe the semantics of classes and properties used in Web documents. These ontologies should be linked to a top-level ontology to enable knowledge sharing and reuse [94]. Unfortunately, each bioontology seems to be built as an independent piece of information, which does not enable the sharing and reuse of knowledge and complicates data integration [95]. Moreover, various sources of biological data must be combined to obtain a full picture and to build new knowledge. However, a large majority of current databases does not use a uniform way to name biological entities. As a result, a same biomedical object is frequently associated with different names.

Systems Biology Markup Language

Systems Biology Markup Language (SBML) [96–99] was designed by the Systems Biology Workbench Development group. The purpose of SBML is to model biochemical reaction networks, comprising cell signalling, gene regulation and metabolic pathways. In SBML, 'Species' is used as a notation to represent the interactors, while reaction, modelling a transformation, transport or binding to represent interaction. Each reaction is allowed to interact with three predefined interactors i.e. reactant, product and modifier [100]. An SBML model encodes a reaction network as pathway. Mathematical relations are also available for reactions. References to other sources and extra information can be added only in the annotation field. Currently, the representation of parts of molecules is not possible [101].

The Proteomics Standards Initiative Molecular Interaction XML format

The Proteomics Standards Initiative Molecular Interaction XML format (PSI MI) [102] is designed by the Proteomics Standards Initiative, which is an initiative of the Human Proteome Organization. The main purpose of the initiative is to standardize proteomics data representation to facilitate data exchange, comparison and verification. The format is projected for exchange of protein-protein interaction data [102]. PSI MI is structured around an entry. It is not anticipated to be a pathway [101]. Links to publications and databases are possible, but a representation of relationships through mathematical equations and an inheritance is not available [101].

The Biological Pathway Exchange

The Biological Pathway Exchange (BioPAX) format is designed by the BioPAX working group [103, 104]. The main purpose of this standard is to introduce a unified framework for sharing pathway information. BioPAX offers more explicit use of relations between concepts than SBML and PSI MI. It is defined as ontology of concepts with attributes [104]. However, reasoning and integration of data increases its computational complexity [101]. A specific data type is available for pathway representation, but mathematical equations underlying the relations are not possible.

CellML

CellML Model Repository [105] contains biochemical pathway models that have been published in peer-reviewed articles or expressed in SBML [106]. CellML [107] and the CellML Model Repository are part of the IUPS Physiome Project [108]. The CellML Model Repository contains models describing a wide range of biological processes [109]. It uses mathematical descriptions of biological systems and adds semantic meaning by annotating elements by ontologies and constrained vocabularies [109]. It is also precise, and thus, the association between dependent and independent species is implicit rather than explicit. However owing to this generality and explicit nature, complexity is increased, especially for software developers, and consequently, there are a few tools that can read and write CellML [110].

Biological Expression Language

Biological Expression Language (BEL) is a highly expressive, triple-based knowledge representation language for the representation of knowledge about causal and correlative relationships [111]. Several groups in academia and pharma are already applying BEL in various areas including biological network analysis, disease modelling, understanding drug efficacy and toxicity, mechanisms for drug sensitivity and resistance and other research and development-related projects. A suite of software components called the BEL Framework provides tools that are required to create, compile, assemble and deliver computable knowledge models to BEL-aware applications [111].

BEL represents complex biological content as simplified, formalized, computable semantic triples that provide the ability to use and reuse experimental observations. BEL can also be used for next-generation sequencing applications, like gene expression profiling and genome annotation data, by using Reverse Causal Reasoning (RCR) algorithm to get mechanistic insights into the high-throughput data, which could be complementary to the result of analysis using pathway gene set. BEL has many utility tools such as a dedicated Cytoscape plug-in for network visualization, algorithms of causal reasoning (RCR) for understanding disease mechanism by identifying up-stream and down-stream controllers, electronic workbook integration, BELto-RDF translation, text mining in BEL and nano-publication concepts [112]. BEL has the potential to impact scientific literature, by introducing computable expressions in scientific publishing, which could be integrated efficiently into existing knowledge environment [113]. Moreover, these causal-reasoning models can provide a valuable addition to the biologists to interpret the gene expression data [114]. By using these models, Huang *et al.* [115] has proposed a data-driven method, Correlation Set Analysis, to detect active regulators in disease by integrating co-expression analysis and literature-derived causal relationships [115].

Reasoning over genetic variance information integrated in disease networks: concepts and strategies

A key task in genetic variants interpretation, to understand the phenotypic consequences, lies in the ability to predict the molecular-level mechanistic consequences of gene polymorphisms and mutations.

As a consequence, systems biomedicine modelling approaches need to combine mechanistic information from various levels, including gene expression, miRNA expression, protein–protein interaction, genetic variation and pathway information. OpenBEL, the open source version of BEL, has recently been extended to match this requirement. With the extended syntax, the new version of BEL 2.0 is now enabled for encoding genetic variants in biomedical models. The last release of the BEL syntax proposes a representation for different genetic variant types, for example, <substitution>, <insertion>, <deletion> and <intergenic>, by introducing new variant functions for DNA, RNA and protein levels.

In this version, the variant (<expression>) function can be used as an argument within a gene(), rna(), microRNA() or protein() to indicate a sequence variant of the specified level. The variant() function takes Human Genome Variation Society (HGVS) variant description expression, e.g. for a substitution, insertion, or deletion of variants. The extended BEL syntax is supposed to support reasoning over cause-effect models that include genetic variation information.

Representation of variant at protein level

Effects of genetic variants located on coding region or splice site, if expressed at protein level, can be represented through protein-level functions. Protein-level variants representation is purposed to see the genetic variants with their relevancy to protein, like their location on the protein sequence and effect on the protein structure (see Table 2).

Representation of variant across DNA/RNA

To see the genetic variants' impact at DNA/RNA level, proteinlevel variants can also be expressed by DNA/RNA-level functions. Whereas, genetic variants located on non-coding regions (like intergenic or intronic) can only be represented through DNA/RNAlevel functions, which are designed to see the genetic variants with their relevancy to genome or gene expression (see Table 3).

Integration of genetic variation information in BEL models of Alzheimer's disease: enhanced functional interpretation of complex SNP patterns

As a support of this review, here we demonstrate an example to highlight this promising approach, by integrating genetic

Variant categories	Variant() function in protein
Reference allele	p(HGNC:CFTR, var(=))
Unspecified variant	p(HGNC:CFTR, var(?))
Substitution variant	p(REF:NP_000483.3, var(p.Gly576Ala))
Deletion variant	p(REF:NP_000483.3, var(p.Phe508del))
Frameshift variant	p(REF:NP_000483.3,
(HGVS short description)	var(p.Thr1220Lysfs))
Frameshift variant	p(REF:NP_000483.3,
(HGVS long description)	var(p.Thr1220Lysfs*7))

Table 2. Representation of different genetic variant categorie	s with
variant functions at proteins level in BEL (2.0V)	

Table 3. Representation of genetic variants across DNA/RNA with the reference of chromosomal or mRNA position in BEL (2.0V)

Level categories	var() function at different genetic levels	
DNA—SNP	g(SNP:rs113993960,	
	var(delCTT))	
DNA—chromosome	g(REF:NC_000007.13,	
	var(g.117199646_117199648delCTT))	
DNA—coding sequence	g(REF:NM_000492.3,	
	var(c.1521_1523delCTT))	
RNA—coding sequence	r(REF:NM_000492.3,	
	var(c.1521_1523delCTT))	
RNA—RNA sequence	r(REF:NM_000492.3,	
	var(r.1653_1655delcuu))	

variant information into an Alzheimer's disease (AD) BEL model.

We have recently published the AD BEL model [116]. This model has 4052 nodes and 9926 edges, and it was generated by extracting relevant knowledge from the specific biomedical literature. The AD BEL model comprises disease-associated genes, protein–protein interactions, miRNAs, bioprocesses and pathways. To integrate disease-specific genetic variant information into AD BEL model, genetic data are retrieved from GWAS databases and the biomedical literature using text-mining methods. The AD BEL model was enriched with AD-SNP-associated data, after annotating functional impact of these genetic variants using the ENSEMBL variant database.

Subsequently, these genetic variants were prioritized, according to their functional consequences. Then we mapped them to the AD BEL model to identify subnetworks with SNPs that display a substantial biological impact. To complete the functional impact assessment for these variants, we have excavated the biomedical literature to analyse the role of these SNPs in the context of age of onset of AD and specifically in the endocytosis pathway.

The early endosome is the first vacuolar compartment in the context of EP, where enlarged early endosomes are identified as the earliest neuro-pathologic features to develop in the early onset of AD. In sporadic AD, endosomal enlargement adds to an average 2.5-fold larger total endosomal volume per neuron, suggesting a significant increase in endocytic activity. It is the site of internalization and initial processing of amyloid precursor protein (APP) and apolipoprotein E, two significant proteins in AD aetiology [117–119]. Here we focus on the internalization of APP based on the functional role of SNPs.

AD is mainly characterized by the deposition of insoluble amyloid beta peptides 42 (A β 42) in the brain, which cannot be easily removed through the blood-brain barrier. In healthy brain, APP is processed by ADAM10, which produces soluble amyloid beta peptide 40 (A β 40), whereas in the non-amyloidogenic pathway, APP is proteolytically processed by BACE and γ secretase to generate A β 42 peptides. A SNP rs514049, linked to the ADAM10 gene, may perturb the normal processing of APP to produce soluble A β 40, as rs514049 is associated with lower level of CSF APP α in AD [120]. BACE1 and BACE2 are associated with γ -secretase complex proteins. Moreover, a SNP rs3754048, with allele G, in the promoter of APH1A gene, might alter the binding ability of YY1 transcription factor, resulting in an increased level of APH1A and γ -secretase activity to facilitate A β 42 generation [121].

All these players in the non-amyloidogenic pathway are trans-membrane proteins, which traffic through the endocytic pathway [122], where these proteins are internalized from the plasma membrane and recycled back to the surface (as in early endosomes and recycling endosomes), or, alternatively, sorted to degradation (as in late endosomes and lysosomes [123, 124]. However, BACE1 is a genetically significant gene with a number of high-ranked AD-associated SNPs. It is also evident that APP and BACE1 are up-regulated in AD. Moreover, experimental evidence suggested that at the cell surface, APP and BACE1 strongly interact and co-localize and are being internalized together into early endosomes, where both proteins remain colocalized and produce amyloid- β . This evidence confirms that endocytosis may be an important step for amyloid- β production [125]. This can be again supported by the association of genetic variants linked with the trafficking proteins in the EP.

As shown in Figure 1, there are two branches of EP: firstly, clathrin-mediated endocytosis (CME), and secondly, retromermediated endocytosis. In the CME pathway, various proteins such as CLTC, PICALM, DNM2, EPS15 and BIN1 modulate APP transport for its further internalization, subsequent Aß generation and further processing in lysosomes, which is required for neurotransmission and signal transduction. CLTC is a major protein component of coated vesicles and coated pits in CME pathway [126]. These specialized organelles are involved in the intracellular trafficking of receptors and endocytosis of a variety of macromolecules including APP with the help of additional accessory proteins such as PICALM, EPS1, DNM2, EGF and its substrate EPS15. PICALM encodes a clathrin assembly protein, which recruits CLTC and AP2, and regulates the size of the clathrin vesicle at neuromuscular junction, whereas an intronic PICALM SNP, rs588076, is associated with allelic expression of a PICALM isoform [127]. Stable DNM2 recruitment during CME correlates well with CLTC lifetime [128], while a risk allele at rs892086 is associated with reduced expression of DNM2 mRNA in the hippocampus in AD patients compared with non-demented controls [129]

On the other hand, the EP is also regulated by retromer, which transports APP from early endosomes to trans-Golgi network (TGN) and released outside cell mainly by retromer complex (VPS35, VPS29, VPS26), SORL1, SNX3, SNX1, WASH complex (KIAA1033) and so on [130]. SORL1 protein belongs to type-I trans-membrane, which is expressed in neurons and plays a critical role in the intracellular transport and in APP processing. SORL1 binds to the retromer complex and works as an adaptor protein for APP trafficking from endosomes to TGN. It is observed that SORL1 levels are reduced in AD-diseased brain, while overexpression of it redistributes APP to the Golgi apparatus; thus, the placement and interaction time of APP and



Figure 1. In this diagram, we present a flowchart that depicts an abstracted BEL subnetwork derived from the original AD BEL Model. This flowchart represents causal relationships between genes and genetic variants for the EP components. Gene symbols written in the textboxes with red outline are showing association with the GWAS identified SNPs for AD. A colour version of this figure is available at BIB online: http://bib.oxfordjournals.org.

BACE1 is reduced in the early endosomes, which will reduce the amount of A β 42. SNX3 mediates recruitment of cargo-selective retromer complex in association with VPS35 [130]. Recent studies have shown that SNX3 and RAB7A are also required for proper recruitment of the cargo-selective complex. Constitutively active RAB7A Q67L mutant is overexpressed, resulting in displacement of the cargo-selective complex [131]. The cargo-selective retromer subcomplex (VPS35–VPS29–VPS26) recruits the WASH complex (KIAA1033), which mediates the production of branched actin networks on the surface of endosomes. The cargo-selective retromer complex together with SNX27 and the WASH complex operates in the endosome-tocell surface recycling of receptors and proteins.

Integration of genetic variation information enhances the evidence base for shared pathophysiology pathways in neurodegenerative diseases

Parkinson's Disease (PD) and AD may share pathophysiological mechanisms and—as a consequence—may share some of their molecular aetiology. To identify evidence that would speak for shared pathophysiology between AD and PD, we systematically analysed genetic variation information that is common between AD and PD and that can be mapped to putatively shared pathways. We have selected the common SNPs from AD and PD GWAS data, and mapped them to gene annotation. Then we searched diseased BEL models to identify the functional impacts of these genes on AD, PD or neurodegenerative diseases (see Table 4).

Conclusion

Given the complexity of neurodegenerative diseases and the limited accessibility to experimental tissues of brain, we need new strategies to integrate data-driven and knowledge-driven approaches to unravel the mechanism behind these complex diseases. Disease networks based on the systems biology models, comprising various interacting molecules such as genes, proteins and bioprocesses, succeeded in integrating most of the available data. In this review, we tried to recapitulate all the major breakthroughs, which demonstrated the collective capturing of disease-related knowledge, modelling it as a system. In addition, we have revisited the major studies around identification of genetic variants and prioritizing these variants based on statistical analysis.

So far, disease networks could not easily accommodate information on genetic variation. We have introduced a novel methodology based on BEL, which enables us to integrate genetic variation information into a disease network. We developed a strategy to analyse the functional consequences of SNPs based

Common SNPs in AD and PD	Gene	AD	PD	NDD
rs931977 (Intronic)	ERG2	EGR2 targeted by mAChRs (muscarinic acetylcholine re- ceptors), which is associated with cognitive functions, synaptic plasticity and memory EGR2 also associated with apoptosis	-	EGR2 is involved in myelin- ation of peripheral nerves
rs2672893 (Intronic)	RPTOR	RPTOR is downstream of MTOR and is expressed highly in AD hippocampus RPTOR activates of PI3K-Akt pathway	Alpha-synuclein reduced the activa- tion of AMPK target RPTOR	_
rs6488270 (Intergenic)	Downstream_variant for: TMEM52B Upstream_variant for: GABARAPL1	_	GABARAPL1 plays role in development and homeostasis of the mouse brain	GABARAPL1 presents a regulated tissue expres- sion and is the most highly expressed gene among the family in the central nervous system
rs4742095 (Intergenic)	Upstream_variant for: CD274 PLGRKT	PD1/PD-L1 (CD274) pathway have role in neuroinflamma- tion of AD PD1/PD-L1 (CD274) pathway is associated with IL-10 production	-	 PLGRKT is regulating plasminogen activation, which plays a key role in regulating catecholaminergic neurosecretory cell function PLGRKT is also involved in macrophage recruitment in the inflammatory response
rs1984129 (Intergenic)	Downstream_variant	-	-	PLGRKT is believed to have role in plasminogen bind- ing and cell migration LRRC8B is implicated in proliferation and activa-
	Upstream_variant for: LRRC8B			tion of lymphocytes and monocytes
rs10515758 (Intergenic)	Downstream_variant for: EBF1 Upstream_variant for: CLINT1	-	-	EBF1 have role in axonal pathfinding CLINT1 interacts with cla- thrin, the adapter protein AP-1 and phosphoinositi- des. This protein may be involved in the formation of clathrin coated ves- icles and trafficking be- tween the TGN and endosomes
rs6810871 (Intergenic)	Downstream_variant for: FAM114A1, TMEM156 Upstream_variant for: KLHL5, TLR6	-	-	FAM114A1 plays a role in neuronal cell development FAM114A1 expressed in dentate gyrus, the hippo- campus, the cerebellum and the olfactory bulb

Table 4. A list of common SNPs/genes in AD and PD with their possible role in the disease context specifically for AD and PD and generally for neurodegenerative diseases (NDD)

on their location in the genome and an interpretation of their putative role in a network model. Currently using the capabilities of extended BEL version, we have developed the AD BEL models together with genetic variants with their DNA, RNA or protein position, variant type and associated allele, which can be used to better understand the role of SNPs in a disease context and tried to predict its consequences based on the functional context provided by the network model.

Although BEL provides certain powerful algorithms like RCR, which allows identifying upstream controllers of an observed

effect, there are still limitations to overcome to enable reasoning over genetic variants. It is obvious, that we need to develop more sophisticated algorithms for reasoning over genetic variant information in network models, by integrating the functional impact of genetic variants on genes in the disease context. One route to go to refine that algorithm is based on machine learning approaches to train a model with the established knowledge of functionally identified genetic variants for different complex diseases. That model will then be applied to neurodegenerative diseases to overcome the deficiency of genetic variant evidential data in this area.

Key Points

- Systems biomedicine modelling approaches need to combine various types of mechanistic details to address multilevel nature of disease dysregulation processes.
- This work represents genetic variation information integration in cause-and-effect models to identify candidate disease mechanisms in diseases with complex aetiology.
- It is an integrative mining approach that identifies 'chains of causation' with reasoning over genetic information in BEL models.
- It exemplifies a new strategy to integrate data-driven and knowledge-driven approaches to unravel the mechanism of complex diseases.

Supplementary Data

Supplementary data are available online at http://bib. oxfordjournals.org/.

Funding

The research leading to these results has received support from the EU/EFPIA Innovative Medicines Initiative Joint Undertaking under AETIONOMY grant agreement n°115568, resources of which are composed of financial contribution from the European Union's Seventh Framework Programme (FP7/2007-2013) and EFPIA companies' in kind contribution.

References

- Barabási AL, Gulbahce N, Loscalzo J. Network medicine: a network-based approach to human disease. Nat Rev Genet 2011;12:56–68.
- Frazer KA, Ballinger DG, Cox DR, et al. A second generation human haplotype map of over 3.1 million SNPs. Nature 2007;449:851–61.
- 3. Williamson R. The molecular genetics of complex inherited diseases. Br J Cancer Suppl 1988;9:14–16.
- Cardoso JG, Andersen MR, Herrgård MJ, et al. Analysis of genetic variation and potential applications in genome-scale metabolic modeling. Front Bioeng Biotechnol 2015;3:13.
- Zhao N, Han JG, Shyu CR, et al. Determining effects of nonsynonymous SNPs on protein-protein interactions using supervised and semi-supervised learning. PLoS Comput Biol 2014;10(5):e1003592.
- de Jong H. Modeling and simulation of genetic regulatory systems: a literature review. J Comput Biol 2002;9(1):67–103.

- 7. Vidal M, Cusick ME, Barabási AL. Interactome networks and human disease. *Cell* 2011;**144**:986–98.
- Sonnenschein N, Golib Dzib JF, Lesne A, et al. A network perspective on metabolic inconsistency. BMC Syst Biol 2012;6:41.
- 9. Ideker T, Krogan NJ. Differential network biology. Mol Syst Biol 2012;8:565.
- 10. Hütt MT. Understanding genetic variation—the value of systems biology. Br J Clin Pharmacol 2014;77(4):597–605.
- 11. Kuepfer L. Towards whole-body systems physiology. Mol Syst Biol 2010;**6**:409.
- Kühn A, Lehrach H. The 'virtual patient' system: modeling cancer using deep sequencing technologies for personalized cancer treatment. J Verbr Lebensm 2012;7:55–62.
- 13. Goh KI, Cusick ME, Valle D, et al. The human disease network. Proc Natl Acad Sci USA 2007;**104**:8685–90.
- Wysocki K, Ritter L. Diseasome: an approach to understanding gene-disease interactions. Annu Rev Nurs Res 2011;29:55– 72.
- 15. Yildirim MA, Goh KI, Cusick ME, et al. Drug-target network. Nat Biotechnol 2007;25:1119–26.
- Leiserson MD, Eldridge JV, Ramachandran S, et al. Network analysis of GWAS data. Curr Opin Genet Dev 2013;23(6):602– 10.
- 17. Carter H, Hofree M, Ideker T. Genotype to phenotype via network analysis. *Curr Opin Genet Dev* 2013;**23**(6):611–21.
- Atias N, Istrail S, Sharan R. Pathway-based analysis of genomic variation data. Curr Opin Genet Dev 2013;23(6):622–6.
- Trynka G, Raychaudhuri S. Using chromatin marks to interpret and localize genetic associations to complex human traits and diseases. *Curr Opin Genet Dev* 2013;23(6):635–41.
- Sahni N, Yi S, Zhong Q, et al. Edgotype: a fundamental link between genotype and phenotype. Curr Opin Genet Dev 2013;23(6):649–57.
- Smyth DJ, Cooper JD, Bailey R, et al. A genome-wide association study of nonsynonymous SNPs identifies a type 1 diabetes locus in the interferon-induced helicase (IFIH1) region. Nat Genet 2006;38(6):617–19
- 22. Zhang X, Bailey SD, Lupien M. Laying a solid foundation for Manhattan—'setting the functional basis for the post-GWAS era'. Trends Genet 2014;**30**(4):140–9.
- Komar AA. Genetics. SNPs, silent but not invisible. Science 2007;315(5811):466–7.
- Matlin AJ, Clark F, Smith CW. Understanding alternative splicing: towards a cellular code. Nat Rev Mol Cell Biol 2005;6(5):386–98.
- Blencowe BJ. Exonic splicing enhancers: mechanism of action, diversity and role in human genetic diseases. Trends Biochem Sci 2000;25(3):106–10.
- Cartegni L, Chew SL, Krainer AR. Listening to silence and understanding nonsense: exonic mutations that affect splicing. Nat Rev Genet 2002;3(4):285–98.
- Fairbrother WG, Yeh RF, Sharp PA, et al. Predictive identification of exonic splicing enhancers in human genes. Science 2002;297(5583):1007–13.
- Gregory AP, Dendrou CA, Attfield KE, et al. TNF receptor 1 genetic risk mirrors outcome of anti-TNF therapy in multiple sclerosis. Nature 2012;488(7412):508–11.
- Holwerda S, de Laat W. Chromatin loops, gene positioning, and gene expression. Front Genet 2012;3:217.
- 30. Frazer KA, Murray SS, Schork NJ, et al. Human genetic variation and its contribution to complex traits. Nat Rev Genet 2009;**10**(4):241–51.

- Docherty SJ, Davis OS, Haworth CM, et al. A genetic association study of DNA methylation levels in the DRD4 gene region finds associations with nearby SNPs. Behav Brain Funct 2012;8:31.
- Sribudiani Y, Metzger M, Osinga J, et al. Variants in RET associated with Hirschsprung's disease affect binding of transcription factors and gene expression. *Gastroenterology* 2011;140(2):572–82.e2.
- Wright JB, Brown SJ, Cole MD. Upregulation of c-MYC in cis through a large chromatin loop linked to a cancer risk-associated single-nucleotide polymorphism in colorectal cancer cells. Mol Cell Biol 2010;30(6):1411–20.
- Brest P, Lapaquette P, Souidi M, et al. A synonymous variant in IRGM alters a binding site for miR-196 and causes deregulation of IRGM-dependent xenophagy in Crohn's disease. Nat Genet 2011;43(3):242–5.
- Bandele OJ, Wang X, Campbell MR, et al. Human single-nucleotide polymorphisms alter p53 sequence-specific binding at gene regulatory elements. Nucleic Acids Res 2011;39(1):178– 89.
- 36. Jones PA, Baylin SB. The epigenomics of cancer. Cell 2007;**128**(4):683–9.
- Ong CT, Corces VG. Enhancer function: new insights into the regulation of tissue-specific gene expression. Nat Rev Genet 2011;12(4):283–93.
- De Gobbi M, Viprakasit V, Hughes JR, et al. A regulatory SNP causes a human genetic disease by creating a new transcriptional promoter. Science 2006;312(5777):1215–17.
- Huang Y, Yang H, Borg BB, et al. A functional SNP of interferon-gamma gene is important for interferon-alphainduced and spontaneous recovery from hepatitis C virus infection. Proc Natl Acad Sci USA 2007;104(3):985–90.
- Cowper-Sal Lari R, Zhang X, Wright JB, et al. Breast cancer risk-associated SNPs modulate the affinity of chromatin for FOXA1 and alter gene expression. Nat Genet 2012;44:1191–8.
- Harismendy O, Notani D, Song X, et al. 9p21 DNA variants associated with coronary artery disease impair interferongamma signalling response. Nature 2011;470(7333):264–8.
- Bauer DE, Kamran SC, Lessard S, et al. An erythroid enhancer of BCL11A subject to genetic variation determines fetal hemoglobin level. Science 2013;342(6155):253–7.
- Musunuru K, Strong A, Frank-Kamenetsky M, et al. From noncoding variant to phenotype via SORT1 at the 1p13 cholesterol locus. Nature 2010;466(7307):714–19.
- 44. Tuupanen S, Turunen M, Lehtonen R, et al. The common colorectal cancer predisposition SNP rs6983267 at chromosome 8q24 confers potential to enhanced Wnt signaling. Nat Genet 2009;41(8):885–90.
- Pomerantz MM, Ahmadiyeh N, Jia L, et al. The 8q24 cancer risk variant rs6983267 shows long-range interaction with MYC in colorectal cancer. Nat Genet 2009;41(8):882–4.
- 46. Zhang X, Cowper-Sal lari R, Bailey SD, et al. Integrative functional genomics identifies an enhancer looping to the SOX9 gene disrupted by the 17q24.3 prostate cancer risk locus. *Genome Res* 2012;**22**(8):1437–46.
- 47. Bickmore WA. The spatial organization of the human genome. Annu Rev Genomics Hum Genet 2013;14:67–84.
- Fraser P, Bickmore W. Nuclear organization of the genome and the potential for gene regulation. Nature 2007;447(7143):413–17.
- Gibcus JH, Dekker J. The hierarchy of the 3D genome. Mol Cell 2013;49(5):773–82.
- Misteli T. Beyond the sequence: cellular organization of genome function. Cell 2007;128(4):787–800.

- Roix JJ, McQueen PG, Munson PJ, et al. Spatial proximity of translocation-prone gene loci in human lymphomas. Nat Genet 2003;34(3):287–91.
- Sanyal A, Lajoie BR, Jain G, et al. The long-range interaction landscape of gene promoters. Nature 2012;489(7414):109–13.
- 53. Li G, Ruan X, Auerbach RK, *et al.* Extensive promoter-centered chromatin interactions provide a topological basis for transcription regulation. *Cell* 2012;**148**(1-2):84–98.
- Visser M, Kayser M, Palstra RJ. HERC2 rs12913832 modulates human pigmentation by attenuating chromatin-loop formation between a long-range enhancer and the OCA2 promoter. *Genome Res* 2012;22(3):446–55.
- Tolhuis B, Palstra RJ, Splinter E, et al. Looping and interaction between hypersensitive sites in the active beta-globin locus. Mol Cell 2002;10(6):1453–65.
- Lanzuolo C, Roure V, Dekker J, et al. Polycomb response elements mediate the formation of chromosome higherorder structures in the bithorax complex. Nat Cell Biol 2007;9(10):1167–74.
- 57. Spilianakis CG, Flavell RA. Long-range intrachromosomal interactions in the T helper type 2 cytokine locus. Nat *Immunol* 2004;5(10):1017–27.
- Sexton T, Bantignies F, Cavalli G. Genomic interactions: chromatin loops and gene meeting points in transcriptional regulation. Semin Cell Dev Biol 2009;20(7):849–55.
- 59. Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell 2009;**136**(2):215–33.
- Bartel B. MicroRNAs directing siRNA biogenesis. Nat Struct Mol Biol 2005;12(7):569–71.
- Ryan BM, Robles AI, Harris CC. Genetic variation in microRNA networks: the implications for cancer research. Nat Rev Cancer 2010;10(6):389–402.
- Brest P, Lapaquette P, Souidi M, et al. A synonymous variant in IRGM alters a binding site for miR-196 and causes deregulation of IRGM-dependent xenophagy in Crohn's disease. Nat Genet 2011;43(3):242–5.
- Guttman M, Amit I, Garber M, et al. Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. Nature 2009;458(7235):223–7.
- Rinn JL, Kertesz M, Wang JK, et al. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. Cell 2007;129(7):1311–23.
- Tsai MC, Manor O, Wan Y, et al. Long noncoding RNA as modular scaffold of histone modification complexes. Science 2010;**329**(5992):689–93.
- Shen LX, Basilion JP, Stanton VP Jr. Single-nucleotide polymorphisms can cause different structural folds of mRNA. Proc Natl Acad Sci USA 1999;96(14):7871–6.
- Kim TK, Hemberg M, Gray JM, et al. Widespread transcription at neuronal activity-regulated enhancers. Nature 2010;465(7295):182–7.
- Musunuru K, Strong A, Frank-Kamenetsky M, et al. From noncoding variant to phenotype via SORT1 at the 1p13 cholesterol locus. Nature 2010;466:714–19.
- Harismendy O, Notani D, Song X, et al. 9p21 DNA variants associated with coronary artery disease impair interferon-γ signalling response. Nature 2011;470:264–8.
- Kondo T, Plaza S, Zanet J, et al. Small peptides switch the transcriptional activity of Shavenbaby during Drosophila embryogenesis. Science 2010;329(5989):336–9.
- Califano A, Butte AJ, Friend S, et al. Leveraging models of cell regulation and GWAS data in integrative network-based association studies. Nat Genet 2012;44(8):841–7.

- Voineagu I, Wang X, Johnston P, et al. Transcriptomic analysis of autistic brain reveals convergent molecular pathology. Nature 2011;474(7351):380–4.
- Dimas AS, Deutsch S, Stranger BE, et al. Common regulatory variation impacts gene expression in a cell type-dependent manner. Science 2009;325(5945):1246–50.
- Grisanzio C, Werner L, Takeda D, et al. Genetic and functional analyses implicate the NUDT11, HNF1B, and SLC22A3 genes in prostate cancer pathogenesis. Proc Natl Acad Sci USA 2012;109(28):11252–7.
- Li Q, Seo JH, Stranger B, et al. Integrative eQTL-based analyses reveal the biology of breast cancer risk loci. Cell 2013;152(3):633–41.
- 76. Pomerantz MM, Shrestha Y, Flavin RJ, et al. Analysis of the 10q11 cancer risk locus implicates MSMB and NCOA4 in human prostate tumorigenesis. PLoS Genet 2010;6(11):e1001204.
- 77. Nicolae DL, Gamazon E, Zhang W, et al. Trait-associated SNPs are more likely to be eQTLs: annotation to enhance discovery from GWAS. PLoS Genet 2010;6(4): e1000888
- Schadt EE, Monks SA, Drake TA, et al. Genetics of gene expression surveyed in maize, mouse and man. Nature. 2003;422(6929):297–302.
- Morley M, Molony CM, Weber TM, et al. Genetic analysis of genome-wide variation in human gene expression. Nature 2004;430(7001):743–7.
- Brem RB, Yvert G, Clinton R, et al. Genetic dissection of transcriptional regulation in budding yeast. Science 2002;296(5568):752–5.
- Stranger BE, Raj T. Genetics of human gene expression. Curr Opin Genet Dev 2013;23(6):627–34.
- The ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. Nature 2012;488:57–74.
- Ecker JR, Bickmore WA, Barroso I, et al. Genomics: ENCODE explained. Nature 2012;489:52–5.
- Moore JH, Asselbergs FW, Williams SM. Bioinformatics challenges for genome-wide association studies. *Bioinformatics* 2010;26:445–55.
- Kiernan MC, Vucic S, Cheah BC, et al. Amyotrophic lateral sclerosis. Lancet 2011;377:942–55.
- Thomas PD, Mi H, Swan GE, et al. Pharmacogenetics of nicotine addiction and treatment consortium. a systems biology network model for genetic association studies of nicotine addiction and treatment. *Pharmacogenet Genomics* 2009;**19**(7):538–51.
- Bader JS. The relative power of SNPs and haplotype as genetic markers for association tests. *Pharmacogenomics* 2001;2:11–24.
- Burga A, Lehner B. Predicting phenotypic variation from genotypes phenotypes and a combination of the two. Curr Opin Biotechnol 2013;24, 803–9.
- Lehner B. Genotype to phenotype: lessons from model organisms for human genetics. Nat Rev Genet 2013;14:168– 78.
- 90. Achard F, Vaysseix G, Barillot E. XML, bioinformatics and data integration. *Bioinformatics* 2001;17:115–25.
- McEntire R, Karp P, Abernethy N, et al. An evaluation of ontology exchange languages for bioinformatics. Proc Int Conf Intell Syst Mol Biol 2000;8:239–50.
- Bratt S. Toward a Web of data and programs. Local to Global Data Interoperability - Challenges and Technologies. 2005, pp.124, 128, 20–24 June 2005.

- Dupré J. The Disorder of Things: Metaphysical Foundations of the Disunity of Science. Cambridge, MA. US, Harvard University Press, 1993.
- 94. Bodenreider O, Stevens R. Bio-ontologies: current trends and future directions. Brief Bioinform 2006;7:256–74.
- Soldatova LN, King RD. Are the current ontologies in biology good ontologies? Nat Biotechnol 2005;23:1095–8.
- 96. The Systems Biology Markup Language (SBML), http://sbml. org/
- Finney A. Systems biology markup language (SBML) Level 3: proposal: multi-component species features. Proposal manuscript. March 2004 (April 2004).
- Finney A., Hucka M. Systems biology markup language (SBML) Level 2: structures and facilities for model definitions. 2003 (April 2004).
- Hucka M, Finney A, Sauro HM, et al. The systems biology markup language (SBML): a medium for representation and exchange of biochemical network models. *Bioinformatics* 2003;19:524–31.
- 100. Strömbäck L, Lambrix P. Representations of molecular pathways: an evaluation of SBML, PSI MI and BioPAX. Bioinformatics 2005;21(24):4401–7.
- 101. Ruebenacker O, Moraru II, Schaff JC, *et al*. Integrating BioPAX pathway knowledge with SBML models. *IET* Syst Biol 2009;**3**(5):317–28.
- 102. Hermjakob H, Montecchi-Palazzi L, Bader G, et al. The HUPO PSI's molecular interaction format—a community standard for the representation of protein interaction data. Nat Biotechnol 2004;**22**:177–83.
- 103. Biological Pathways Exchange, http://www.biopax.org
- 104.BioPAX Working Group. BioPAX—biological pathways exchange language. 2004. Level 1, Version 1.0 Documentation.
- 105. CellML Model Repository, http://www.cellml.org/models
- 106. Le Novere N, Bornstein B, Broicher A, et al. BioModels database: a free, centralized database of curated, published, quantitative kinetic models of biochemical and cellular systems. Nucleic Acids Res 2006;**34**:D689–91.
- 107. Lloyd C.M., Halstead MD, Nielsen PF. CellML: its future, present and past. Prog Biophys Mol Biol 2004;**85**:433–50.
- 108. Hunter P, Nielsen P. A strategy for integrative computational physiology. Physiology (Bethesda) 2005;20:316–25.
- 109. Lloyd CM, Lawson JR, Hunter PJ, et al. The CellML model repository. Bioinformatics 2008;24(18):2122–3.
- 110. Sauro HM, Bergmann FT. Standards and ontologies in computational systems biology. Essays Biochem 2008;45:211–22.
- 111. OpenBEL: Biological Expression Language (BEL), http://www. openbel.org/bel-expression-language
- 112. Catlett NL, Bargnesi AJ, Ungerer S, *et al.* Reverse causal reasoning: applying qualitative causal knowledge to the interpretation of high-throughput data. *BMC Bioinformatics* 2013;**14**:340.
- 113. Slater T. Recent advances in modeling languages for pathway maps and computable biological networks. *Drug Discov Today* 2014;**19**(2):193–8.
- 114. Chindelevitch L, Ziemek D, Enayetallah A, et al. Causal reasoning on biological networks: interpreting transcriptional changes. Bioinformatics 2012;**28**(8):1114–21.
- 115. Huang CL, Lamb J, Chindelevitch L, et al. Correlation set analysis: detecting active regulators in disease populations using prior causal knowledge. BMC Bioinformatics 2012;13:46.
- 116. Kodamullil AT, Younesi E, Naz M, et al. Computable causeand-effect models of healthy and Alzheimer's disease states and their mechanistic differential analysis. Alzheimers Dement 2015:pii: S1552-5260(15)00083-7.

- 117. Nixon RA. Endosome function and dysfunction in Alzheimer's disease and other neurodegenerative diseases. *Neurobiol Aging* 2005;**26**(3):373–82.
- 118. Cataldo AM, Barnett JL, Pieroni C, et al. Increased neuronal endocytosis and protease delivery to early endosomes in sporadic Alzheimer's disease: neuropathologic evidence for a mechanism of increased beta-amyloidogenesis. J Neurosci 1997;17(16):6142–51.
- 119. Cataldo AM, Peterhoff CM, Troncoso JC, et al. Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: differential effects of APOE genotype and presenilin mutations. Am J Pathol 2000;**157**(1):277–86.
- 120. Bekris LM, Galloway NM, Millard S, et al. Amyloid precursor protein (APP) processing genes and cerebrospinal fluid APP cleavage product levels in Alzheimer's disease. Neurobiol Aging 2011;32(3):556.e13–23.
- 121. Qin W, Jia L, Zhou A, et al. The -980C/G polymorphism in APH-1A promoter confers risk of Alzheimer's disease. Aging Cell 2011;**10**(4):711–19.
- 122. Choy RW, Cheng Z, Schekman R. Amyloid precursor protein (APP) traffics from the cell surface via endosomes for amyloid β (A β) production in the trans-Golgi network. Proc Natl Acad Sci USA 2012;**109**(30):E2077–82.
- 123. Decourt B, Mobley W, Reiman E, et al. Recent perspectives on APP, secretases, endosomal pathways and how they

influence Alzheimer's related pathological changes in down syndrome. J Alzheimers Dis Parkinsonism 2013; Suppl 7:002.

- 124. Grant BD, Donaldson JG. Pathways and mechanisms of endocytic recycling. Nat Rev Mol Cell Biol 2009;10(9):597–608.
- 125. Kinoshita A, Fukumoto H, Shah T, et al. Demonstration by FRET of BACE interaction with the amyloid precursor protein at the cell surface and in early endosomes. *J Cell Sci* 2003;**116**(Pt 16):3339–46.
- 126. McMahon HT, Boucrot E. Molecular mechanism and physiological functions of clathrin-mediated endocytosis. Nat Rev Mol Cell Biol 2011;12(8):517–33.
- 127. Parikh I, Medway C, Younkin S, et al. An intronic PICALM polymorphism, rs588076, is associated with allelic expression of a PICALM isoform. *Mol Neurodegener* 2014;**9**:32.
- 128. Grassart A, Cheng AT, Hong SH, et al. Actin and dynamin2 dynamics and interplay during clathrin-mediated endocytosis. J Cell Biol 2014;205(5):721–35.
- 129. Aidaralieva NJ, Kamino K, Kimura R, et al. Dynamin 2 gene is a novel susceptibility gene for late-onset Alzheimer disease in non-APOE-epsilon4 carriers. J Hum Genet 2008;53(4):296–302.
- 130. Seaman MN. The retromer complex—endosomal protein recycling and beyond. J Cell Sci 2012;125(Pt 20):4693–702.
- 131. Vardarajan BN, Bruesegem SY, Harbour ME, et al. Identification of Alzheimer disease-associated variants in genes that regulate retromer function. *Neurobiol Aging* 2012;**33**(9):2231.e15–30.