

RESEARCH ARTICLE

Methylation of *MGMT* Is Associated with Poor Prognosis in Patients with Stage III Duodenal Adenocarcinoma

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Abstract

Background

*O*⁶-methylguanine-DNA methyltransferase (*MGMT*) methylation status has not been extensively investigated in duodenal adenocarcinoma (DA). The aim of this study was to evaluate the *MGMT* methylation status and examine its possible prognostic value in patients with stage III DA.

Methods

Demographics, tumor characteristics and survival were available for 64 patients with stage III DA. *MGMT* methylation was detected by using MethyLight. A Cox proportional hazard model was built to predict survival, adjusted for clinicopathological characteristics and tumor molecular features, including the CpG island methylator phenotype (CIMP), microsatellite instability (MSI), and *KRAS* mutations.

Results

MGMT methylation was detected in 17 of 64 (26.6%) patients, and was not correlated with sex, age, tumor differentiation, CIMP, MSI, or *KRAS* mutations. *MGMT* methylation was the only one factor associated with both overall survival (OS) and disease-free survival (DFS) on both univariate and multivariate analyses. In patients treated with surgery alone, *MGMT*-methylated group had worse OS and DFS when compared with *MGMT*-unmethylated group. However, in patients treated with chemotherapy/radiotherapy, outcomes became comparable between the two groups.

Conclusions

Our results demonstrate *MGMT* methylation is a reliable and independent prognostic factor in DAs. Methylation of *MGMT* is associated with poor prognosis in patients with stage III DAs.

from Astex and has licensed biomarkers to Cepheid Inc. She has served as consultant for Ethicon. No potential conflicts of interest were disclosed by the other authors.

Introduction

Primary adenocarcinoma of the duodenum (duodenal adenocarcinoma, DA) was initially described by Hamburger in 1746, comprising less than 1% of all malignant neoplasms of the gastrointestinal tract [1–3]. Because of its rarity, there is an insufficiency of well-designed studies to guide management. In general, DAs have more favorable outcomes compared to other periampullary malignancies and excision is considered the backbone of treatment for patients with localized tumors or limited metastatic disease when feasible. Data regarding the effect of adjuvant chemotherapy/radiotherapy are limited, with no faithful evidence of significant benefit in survival in patients with DAs. A Cochrane review in 2007 failed to find suitable trials eligible for meta-analysis to determine the role of adjuvant chemotherapy in the treatment of adenocarcinoma of the small intestine [4]. Although adjuvant therapy is regularly used in this disease, more studies are needed to evaluate the effectiveness of adjuvant therapy in the management of DAs.

O⁶-methylguanine-DNA methyltransferase (MGMT) is a ubiquitously expressed DNA repair protein, and it removes methyl and chloroethyl groups from the O⁶ position of guanine in a damage reversal reaction. In the absence of MGMT, O⁶-methylguanine in the DNA generates point mutations and DNA double-strand breaks via cellular replication and DNA mismatch repair that trigger cell death by apoptosis [5]. Methylation of the CpG islands located in the promoter region of *MGMT* is primarily responsible for the inactivation of MGMT in several tumor types [6]. Inactivation of MGMT can lead to it subsequently being unable to protect tumors from cytotoxic damage induced by alkylating chemotherapeutics, i.e. methylating and chloroethylating agents, and thus predicts benefit from these chemotherapeutic agents. *MGMT* methylation may also play a prognostic role in various cancers. To our knowledge, there is only one previous study that has described *MGMT* methylation in DAs in a small number of patients and there was no assessment of *MGMT* methylation frequency or prognostic significance [7].

Microsatellite instability (MSI), developing from defects in other mismatch repair genes *MLH1*, *MSH2*, *MSH6*, and *PMS2*, are seen in 18–35% of small bowel adenocarcinomas including DAs [8–10]. MSI along with *KRAS* mutations represent the most common molecular defects in DA [7, 11, 12]. MSI is associated with prognosis in patients with colorectal cancer. Its prognostic value in DAs is worth investigation. *MGMT* methylation seems to favor mutations in cancer-related genes (e.g. *TP53* and *KRAS*). Kim et al. previously showed the association between *MGMT* methylation and *KRAS* G-to-A transition in a group of patients with carcinomas of the extrahepatic bile ducts, ampulla of Vater, and duodenum [7]. Due to the small number of duodenal carcinomas in the previous study, this correlation still needs validation.

The aims of this study were to assess the methylation status of *MGMT* gene in the largest series of stage III DAs reported to date and to establish whether or not methylation of *MGMT* might have prognostic or predictive value in patients with stage III DA.

Material and Methods

Study population

This retrospective cohort study included patients with pathologically confirmed DA who had a surgical resection. Patients were identified from the Johns Hopkins Hospital Oncology Clinical Information System from January 1997 to December 2009 and 155 duodenal adenocarcinomas patients who underwent surgical resection at our institution were identified. Patients who underwent preoperative chemotherapy/radiotherapy, lacked follow-up information or had

missing archival primary tumors or corresponding matched normal samples were excluded. Formalin-fixed, paraffin-embedded (FFPE) tissue blocks of primary tumors and corresponding matched normal samples were collected from 107 patients. Tissue sections from the blocks were then reviewed by an expert gastrointestinal pathologist. After excluding ampullary tumors and low tumor cellularity sections, the remaining 64 stage III cases formed the final study cohort (Table 1). Ascertainment of survival was performed by using the Johns Hopkins electronic health records, the Cancer Registry and mortality was confirmed also within the Social Security Death Index. The Johns Hopkins Hospital Institutional Review Board approved this research protocol.

Analyses of *KRAS* mutations, and microsatellite instability

Genomic DNA was extracted from FFPE tissues. Polymerase chain reaction (PCR) and sequencing targeted for *KRAS* codons 12 and 13 were performed [11, 13].

MSI status was determined using D2S123, D5S346, D17S250, BAT25, and BAT26 [14]. Microsatellite sizes were compared with those of normal adjacent tissue, and tumors with 2 or more of the markers exhibiting instability were classified as MSI-high. Tumors with only one marker exhibiting instability or no markers with instability were classified as MSI-low or microsatellite stable (MSS), respectively.

Bisulfite modification and methylation analysis

Purified DNA (2 μ g) was bisulfite treated and purified using the EZ DNA methylation kit (Zymo Research, Orange, CA) according to the manufacturer's instructions.

A 5-gene signature was used to assess the CpG island methylator phenotype (CIMP) status of the primary tumor tissue: *CACNA1G*, *IGF2*, *NEUROG1*, *RUNX3*, and *SOCS1* [15]. Methylation of these five genes and *MGMT* was quantified by MethyLight, a methylation-specific, probe-based, real-time PCR technique [12, 15, 16]. Alu was used as a normalization control reaction. All CIMP probes utilized a 5' FAM fluorophore, a 3' IBFQ quencher, and an internal ZEN quencher (Integrated DNA Technologies, Coraville, IA). DNA methylation was reported as the percent of methylated reference (PMR) = $100 \times ((\text{methylated reaction}/\text{Alu})_{\text{sample}}/(\text{methylated reaction}/\text{Alu})_{\text{M.SssI-reference}})$ [15]. We classified each marker as methylated when $\text{PMR} \geq 4$. The PMR cut-off levels were set at plus two standard deviations of the average methylation levels observed in normal duodenal mucosa controls. Samples were considered CIMP+ if at least 3 out of the five studied genes were methylated [15].

Statistical methods

Differences in categorical variables between study groups were analyzed using χ^2 test or Fisher's exact test. The primary end point for the study was disease-free survival (DFS), defined as the time from surgery to death or recurrence of disease, whichever occurred first. Overall survival (OS) was the secondary end point. Patients without evidence of death or recurrence were censored at last follow-up. Survival was estimated by using the Kaplan-Meier method and log-rank statistics computed to test for differences between survival curves for various prognostic factors. Univariate and multivariate Cox proportional hazard regression models included *MGMT* methylation, sex, age, tumor differentiation, R0 resection, chemoradiation, CIMP, MSI status, and *KRAS* mutations. Results of Cox regression are reported as hazard ratio (HR) with corresponding 95% confidence intervals (CI). All hypotheses tests were two-sided, and results were considered statistically significant for P values < 0.05 . All calculations were performed using SPSS 16.0 software (SPSS Inc, Chicago, IL).

Table 1. Clinicopathological and molecular characteristics of patients and tumors by MGMT methylation status.

Characteristic	All patients (n = 64)	MGMT-U (n = 47)	MGMT-M (n = 17)	P ^a
Chemotherapy/radiotherapy				0.756
No	17 (26.6%)	12 (25.5%)	5 (29.4%)	
Yes	47 (73.4%)	35 (74.5%)	12 (70.6%)	
Sex				0.602
Male	38 (59.4%)	27 (57.4%)	11 (64.7%)	
Female	26 (40.6%)	20 (42.6%)	6 (35.3%)	
Age at surgery				0.144 ^b
< 60	21 (32.8%)	18 (38.3%)	3 (17.6%)	
≥ 60	43 (67.2%)	29 (61.7%)	14 (82.4%)	
Tumor differentiation				0.396
Well/moderate	32 (50.0%)	25 (53.2%)	7 (41.2%)	
Poor	32 (50.0%)	22 (46.8%)	10 (58.8%)	
CIMP				0.111
CIMP-	47 (73.4%)	37 (78.7%)	10 (58.8%)	
CIMP+	17 (26.6%)	10 (21.3%)	7 (41.2%)	
MSI status				1.000 ^b
MSS	49 (76.6%)	36 (76.6%)	13 (76.5%)	
MSI	15 (23.4%)	11 (23.4%)	4 (23.5%)	
KRAS				0.430
Wild-type	39 (60.9%)	30 (63.8%)	9 (52.9%)	
Mutated	32 (32.3%)	17 (36.2%)	8 (47.1%)	

^aMGMT-U versus MGMT-M, χ^2 test unless indicated otherwise

^bFisher's exact test.

Abbreviations: CIMP, CpG island methylator phenotype; MSS, microsatellite stable; MSI, microsatellite instability; U, unmethylated; M, methylated.

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Results

Clinicopathologic characteristics and association with MGMT methylation or MSI status

DNA extraction, MGMT methylation testing by MethyLight, and MSI status testing were successful in all 64 patients. Seventeen patients (26.6%) out of the 64 patients tested were MGMT-methylated (MGMT-M, Table 1). Fifteen patients (23.4%) displayed MSI-high; 9 patients (14.1%) were MSI-low and 40 patients (62.5%) were MSS. Because extensive data indicate that tumors with MSI-low are biologically similar to those exhibiting MSS, both tumors were grouped together and henceforth are referred to as MSS in this study. Among the 17 (26.6%) patients demonstrating the CIMP positive (CIMP+), 7 (41.2%) were MGMT-M as well (Table 1). No correlation between CIMP and MGMT methylation status was observed ($P = 0.111$, Table 1).

Median age at diagnosis of DAs was 64.5 years (64.2 ± 14.3 ; mean \pm SD). MGMT-unmethylated (MGMT-U) and MGMT-M subgroups showed no differences by gender, age, tumor differentiation, CIMP, MSI and KRAS mutation status or the receipt of chemotherapy/radiotherapy between the two groups (Table 1).

MGMT methylation status as a prognostic marker

The mean (SD) follow-up was 42.9 (28.5) months. There were 36 deaths, 24 recurrences, and 42 progressions at the end of follow-up. The median OS was 41.2 months (95% CI, 25.2 to 57.2

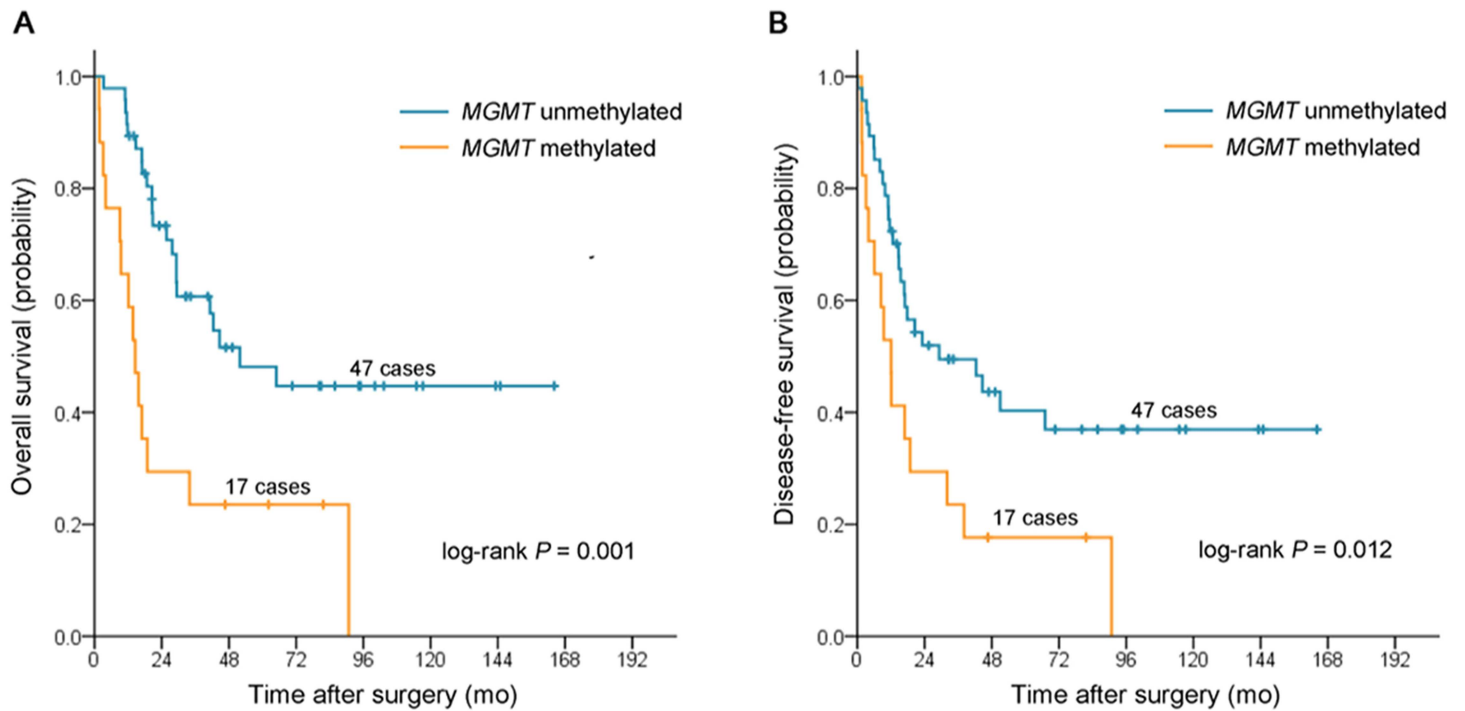


Fig 1. Kaplan-Meier survival estimates between patients with stage III duodenal adenocarcinomas with MGMT methylated and those with MGMT unmethylated. (A) overall survival, (B) disease-free survival.

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months), and the median DFS was 18.8 months (95% CI, 5.6 to 32.1 months). In Kaplan-Meier analysis of all patients, MGMT-M was associated with worse OS (log-rank $P = 0.001$; Fig 1A) and DFS (log-rank $P = 0.012$; Fig 1B). The median OS was 51.9 months (95% CI, 22.5 to 81.3 months) vs. 14.5 months (95% CI, 9.7 to 19.3 months), and the median DFS was 29.2 months (95% CI, 0 to 59.7 months) vs. 12.0 months (95% CI, 7.0 to 17.0 months) for patients with MGMT-U tumor vs. MGMT-M tumor, respectively. In univariate models, MGMT-M was associated with worse OS (HR, 3.01; 95% CI, 1.53 to 5.91; $P = 0.001$) and DFS (HR, 2.21; 95% CI, 1.17 to 4.17; $P = 0.014$). This remained statistically significant in multivariate models for OS (HR, 4.25; 95% CI, 2.00 to 9.05; $P = 0.000$) and for DFS (HR, 2.80; 95% CI, 1.43 to 5.48; $P = 0.003$; Table 2).

Adjuvant treatment

Adjuvant treatment with fluorouracil-based chemotherapy/radiotherapy was administered in 47 patients, while 17 patients were treated with surgery alone. There was no significant improvement in OS for patients treated with adjuvant therapy when compared with patients who were not treated (HR, 1.13; 95% CI, 0.51 to 2.51; $P = 0.759$). When comparing DFS, there was no difference based on adjuvant treatment (HR, 0.85; 95% CI, 0.41 to 1.73; $P = 0.648$; Table 2).

In patients treated with surgery alone ($n = 17$), MGMT-M was associated with worse OS (HR, 7.88; 95% CI, 1.83 to 34.00; $P = 0.006$) and DFS (HR, 5.33; 95% CI, 1.40 to 20.30; $P = 0.014$) on univariate analysis. This remained statistically significant in multivariate models for OS (HR, 7.49; 95% CI, 1.04 to 53.84; $P = 0.045$) and OS (HR, 4.11; 95% CI, 1.03 to 16.40; $P = 0.046$). However, no association was observed between MGMT methylation status and both OS (HR, 1.85; 95% CI, 0.84 to 4.11; $P = 0.130$) and DFS (HR, 1.56; 95% CI, 0.74 to 3.30;

Table 2. Univariate and multivariate Cox proportional hazard analysis of overall survival (OS) and disease-free survival (DFS).

Characteristic	Total n	OS				DFS			
		Univariate		Multivariate		Univariate		Multivariate	
		HR (95% CI)	P value	HR (95% CI)	P value	HR (95% CI)	P value	HR (95% CI)	P value
MGMT									
U	47	1.00 (Referent)							
M	17	3.01 (1.53, 5.91)	0.001	4.25 (2.00, 9.05)	0.000	2.21 (1.17, 4.17)	0.014	2.80 (1.43, 5.48)	0.003
Sex									
Male	38	1.00 (Referent)							
Female	26	1.44 (0.75, 2.78)	0.275	1.62 (0.79, 3.35)	0.190	0.98 (0.53, 1.82)	0.950		
Age									
≥60	43	1.00 (Referent)							
<60	21	0.56 (0.26, 1.19)	0.131	0.57 (0.26, 1.27)	0.168	0.79 (0.41, 1.53)	0.488		
Differentiation									
Well/moderately	32								
Poorly	32	1.21 (0.63, 2.33)	0.568			1.54 (0.84, 2.84)	0.163	1.43 (0.77, 2.66)	0.260
R0 resection									
Yes	56								
No	8	1.16 (0.45, 2.98)	0.761			1.11 (0.47, 2.65)	0.807		
Chemoradiation									
Yes	47								
No	17	1.13 (0.51, 2.51)	0.759			0.85 (0.41, 1.73)	0.648		
CIMP									
CIMP-	47								
CIMP+	17	1.61 (0.80, 3.22)	0.180	2.84 (1.28, 6.32)	0.011	1.37 (0.70, 2.68)	0.361		
MSI status									
MSS	49								
MSI	15	0.43 (0.18, 1.04)	0.060	0.18 (0.06, 0.50)	0.001	0.35 (0.15, 0.84)	0.018	0.26 (0.10, 0.64)	0.003
KRAS mutations									
Absent	39								
Present	25	0.78 (0.40, 1.55)	0.482			0.87 (0.47, 1.63)	0.666		

Abbreviations: OS, overall survival; DFS, disease-free survival; HR, hazard ratio; CI, confidence interval; M, methylated; U, unmethylated; CIMP, CpG island methylator phenotype; MSS, microsatellite stable; MSI, microsatellite instability. A backward elimination with threshold of $P = 0.300$ was used to select variables in the final models

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$P = 0.243$; Table 3) in patients treated with chemotherapy/radiotherapy. In Kaplan–Meier analysis, there were also significant differences in survival time distributions between patients with MGMT-M and those with MGMT-U in the group treated with surgery alone (log-rank $P = 0.001$ for OS, Fig 2A; log-rank = 0.006 for DFS, Fig 2B). The median OS was not reached vs. 9.4 months (95% CI, 0 to 25.7 months), and the median DFS was not reached vs. 9.4 months (95% CI, 0 to 25.7 months) for patients with MGMT-U tumor vs. MGMT-M tumor, respectively. No significant differences were found between patients with MGMT-M tumor and those with MGMT-U tumor in the group treated with chemotherapy/radiotherapy (log-rank $P = 0.123$ for OS, Fig 3A; log-rank = 0.239 for DFS, Fig 3B).

Discussion

The present study was designed to better understand the contribution of methylation of MGMT for patients with stage III DAs and to determine its effect in response to fluorouracil-

Table 3. Univariate and multivariate Cox proportional hazard analysis of overall survival (OS) and disease-free survival (DFS) by MGMT methylation and chemotherapy/radiotherapy treatment status.

Characteristic	Total n	OS				DFS			
		Univariate		Multivariate		Univariate		Multivariate	
		HR (95% CI)	P value	HR (95% CI)	P value	HR (95% CI)	P value	HR (95% CI)	P value
Untreated									
MGMT-U	12								
MGMT-M	5	7.88 (1.83, 34.00)	0.006	7.49 (1.04, 53.84)	0.045	5.33 (1.40, 20.30)	0.014	4.11 (1.03, 16.40)	0.046
Treated									
MGMT-U	35								
MGMT-M	12	1.85 (0.84, 4.11)	0.130			1.56 (0.74, 3.30)	0.243		

Abbreviations: OS, overall survival; DFS, disease-free survival; HR, hazard ratio; CI, confidence interval; MGMT-M, MGMT-methylated; MGMT-U, MGMT-unmethylated

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based adjuvant chemotherapy/radiotherapy in a cohort of patients. Our results indicate that, MGMT methylation is a reliable and independent prognostic factor in DAs. MGMT methylation is associated with poor prognosis in patients with stage III DAs. It seems that fluorouracil-based chemotherapy/radiotherapy does not improve outcomes in patients with stage III DAs. However, in the subsets of DAs with MGMT methylation fluorouracil-based chemotherapy/radiotherapy may confer a survival benefit.

MGMT methylation has been associated with various cancers. Specifically, MGMT methylation was seen in 39–53% of CRCs [17, 18], 11% of gastric cancer [19], 30–38% of lung cancer [20, 21], 34–72% of esophageal cancer [22], 34% of soft tissue sarcomas [23], 58% of breast cancer

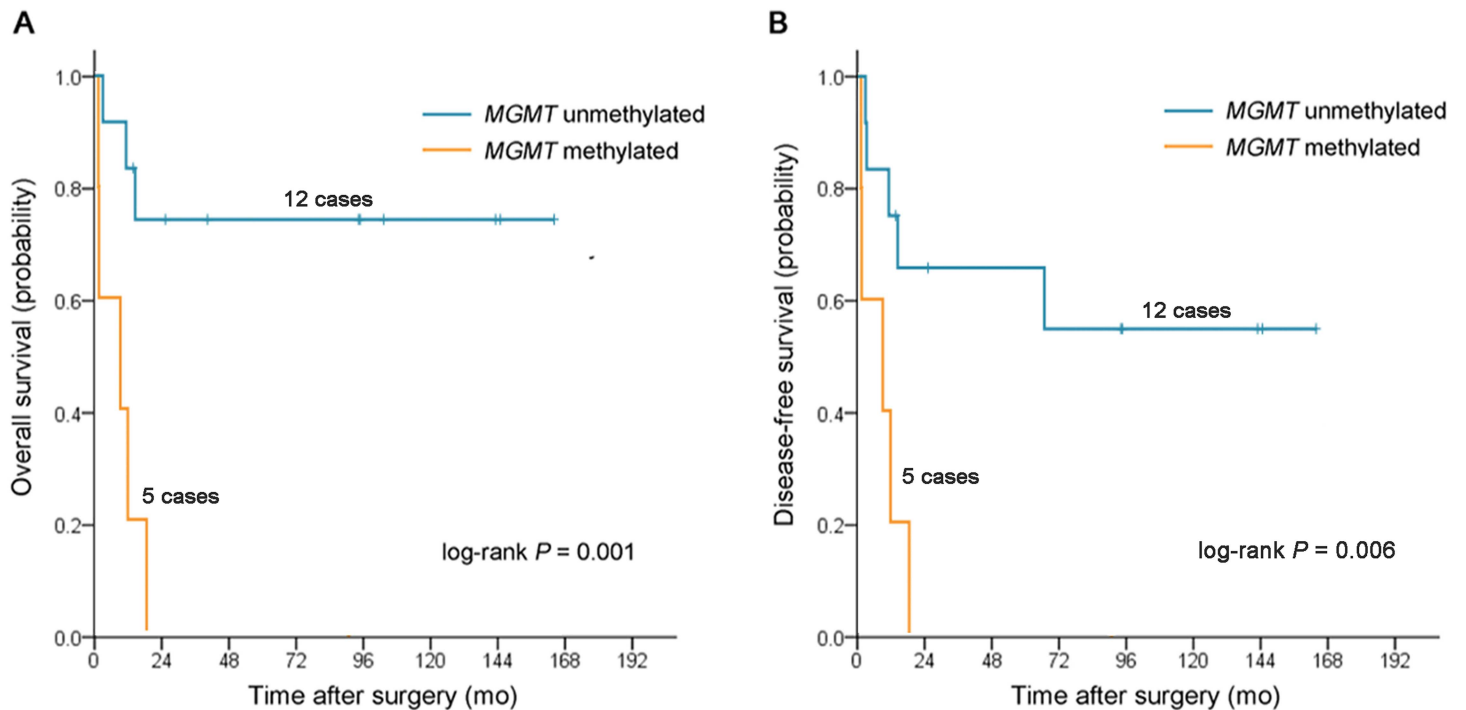


Fig 2. Kaplan-Meier survival estimates between patients with stage III duodenal adenocarcinomas with MGMT methylated and those with MGMT unmethylated in group treated with surgery alone. (A) overall survival, (B) disease-free survival.

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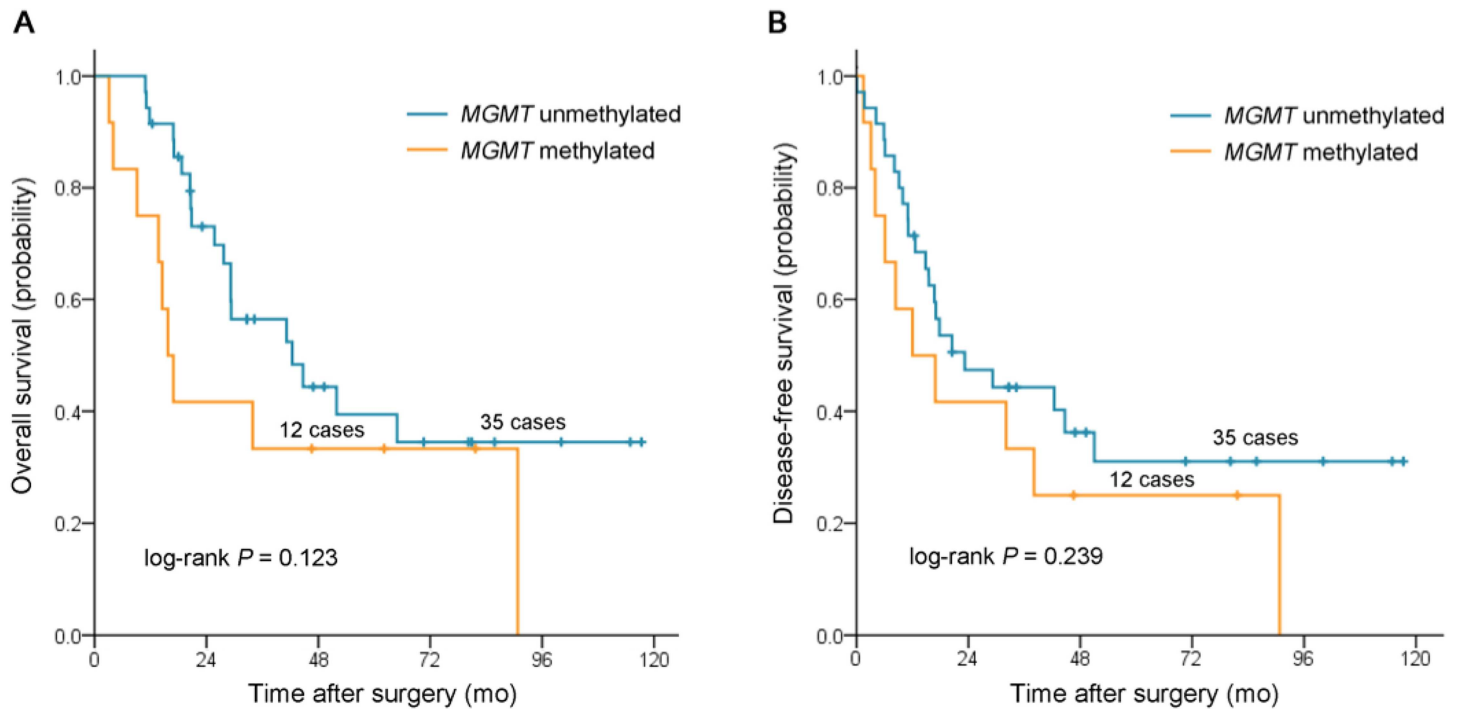


Fig 3. Kaplan-Meier survival estimates between patients with stage III duodenal adenocarcinomas with *MGMT* methylated and those with *MGMT* unmethylated in group treated with fluorouracil-based chemotherapy/radiotherapy. (A) overall survival, (B) disease-free survival.

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[24], and 30–70% of glioblastoma [25, 26]. In this study, we analyzed a large cohort of patients with stage III DAs and showed that *MGMT* methylation existed in 26.6% (17/64) of the tumors.

It was reported that inactivation of *MGMT* by promoter methylation was theoretically associated with the presence of *KRAS* G>A transitions in CRC [27]. Their data suggested that epigenetic silencing of *MGMT* by methylation was strongly associated with, and preceded, G>A mutations in *KRAS* in colorectal tumorigenesis. Some studies proved this possible association in CRCs [28, 29], however, we did not find this link between these two events in DAs ($P = 0.226$; data not shown). This can be secondary to various causes including methodology issues (type of methylation assay, small sample size, intratumor heterogeneity) and most importantly, alternative molecular mechanisms that cause DAs. The concurrence of these epigenetic and genetic lesions in different tumors suggests a more complex relationship between these events. For example, *MGMT* methylation is common [26], but *KRAS* mutations are relatively rare in glioblastoma [30]. Nagy et al. also showed that no conclusions could be drawn with regard to mutation type and methylation in endometrial cancers [31]. In a study of 62 gastric cancer tissue samples, *KRAS* mutations were detected in only one (1.6%) sample and *MGMT* methylation was detected in 13 (21%) samples, and no connection was shown between *KRAS* mutations and *MGMT* methylation [32]. Similar results were shown in a study of 62 soft tissue sarcomas with *MGMT* methylation 33.9% (21/62) and *KRAS* mutations 3.7% (2/62) [23]. In a large cohort study with 1123 CRC, a strong association with *MGMT* methylation was found with *KRAS* mutations both in univariate analysis (OR 2.3, 95% CI 1.7–3.0, $P < 0.0001$) and multivariate analysis (OR 1.9, 95% CI 1.5–2.6, $P < 0.0001$). But on classification of the *KRAS* mutant cancers by mutation type, no association was found between *MGMT* methylation and G>A mutations compared with non-G>A mutations, and in fact frequency of *MGMT*-M and *MGMT*-U tumors was approximately equal for each mutation category [33].

In previous studies, the significance of the correlation between *MGMT* methylation and prognosis of patients was controversial [21, 25, 34–36]. In present study, the impact of *MGMT* methylation on patient survival was assessed by univariate and multivariate analyses. Cox proportional hazard models indicated that methylation of *MGMT* was strongly associated with poor survival in DAs patients.

Despite the absence of prospective randomized data clarifying the role of adjuvant therapy in DAs, the use of adjuvant therapy has increased. Data from the National Cancer Database shows a spread use of adjuvant chemoradiation in small bowel cancers (including 49.1%–58.8% DAs) from 8.1% in 1985 to 22.2% in 2005 ($P < 0.0001$) [37]. In all likelihood, this trend reflects the poor outcome of high-risk dissected DAs, the known efficacy of systemic chemoradiation in the metastatic setting and the significant survival benefit of adjuvant therapy in patients with CRC.

Several studies have individually examined the results of adjuvant therapy after resection of DA. In 1980, Alwmark et al. suggested that chemoradiation might improve the survival of patients with DA [1]. Since then, advances in chemotherapy and radiotherapy have developed, but chemoradiation has commonly been reserved for palliation of DAs. Our institution has previously published a pilot study on 14 patients with node-positive DA who underwent pancreaticoduodenectomy followed by adjuvant fluorouracil-based chemoradiation [38]. This study suggested that adjuvant chemoradiation contributed improved local control compared with historical controls treated with surgery alone (93% vs. 67%), but did not lengthen overall survival (5 year, 44% vs. 43%). However, in this follow up study from our institution of a larger cohort of patients we were unable to reproduce this positive effect of chemoradiation for either local control or OS [39]. Another retrospective study of 103 patients with DA (including 46 stage III DAs) from Massachusetts General Hospital compared patients who underwent resection alone with those who received resection and adjuvant and/or neoadjuvant chemotherapy/chemoradiation and found no marked improvement in OS, or time to recurrence [6]. A similar study of 32 patients with DA from Duke University Medical Center also failed to show a beneficial effect of adjuvant chemoradiation both in terms of OS (44% vs. 57%), disease-free survival (44% vs. 54%) or local control (49% vs. 70%) [40]. In an analysis of 1,611 cases on long-term outcome after resection of DA by utilizing the Surveillance, Epidemiology, and End Results (SEER) database, a large population-based cancer registry showed that the use of radiation was associated with improvements in survival on univariate analysis, but this effect disappeared after controlling for other variable [41].

In this study, we showed that patients treated with adjuvant therapy had similar prognosis to those treated with surgery alone. In patients treated with surgery alone, patients with *MGMT*-M tumor had worse OS and DFS compared with those with *MGMT*-U tumor. However, in patients undergoing adjuvant fluorouracil-based chemotherapy/radiotherapy, outcomes became comparable between patients with *MGMT*-M tumor and those with *MGMT*-U tumor. This might be, to some extent, due to differential responses to chemotherapy/radiotherapy between these two subtypes of tumor. Nevertheless, this phenomenon deserves further investigation. The finding is potentially of great significance, as the addition of adjuvant chemotherapy/radiotherapy in DAs is currently a matter of great debate.

Alkylating agent temozolomide is now the chemotherapeutic agent most regularly used in patients with newly diagnosed glioblastoma. It is well established that *MGMT* methylation is a promising predictor of prolonged prognosis in patients with glioblastoma receiving temozolomide [42, 43]. In a pivotal randomized trial investigating the value of temozolomide added to radiotherapy in patients with glioblastoma, median survival in patients with methylated *MGMT* promoter increased from 15.3 months (95% CI 13.0–20.9) with radiotherapy alone to 21.7 months (17.4–30.4) with radiotherapy and temozolomide (hazard ratio [HR] 0.51, 95% CI

0.31–0.84). However, patients with unmethylated *MGMT* promoter in the tumor showed only a marginal benefit from addition of temozolomide, with a median survival of 12.7 months (95% CI 11.6–14.4) compared with 11.8 months (9.7–14.1) for patients treated with radiotherapy alone (HR 0.69, 95% CI 0.47–1.02) [44]. However, the value of *MGMT* methylation as a prognostic or predictive marker for patients treated with other specific regimens of anticancer agents remains a matter of debate to date. A previous study has shown that CRC patients who received oral fluorouracil-based adjuvant chemotherapy had a low recurrence rate when the tumor revealed methylation in its *MGMT* promoter [45]. Their in vitro study also proved an enhancement of fluorouracil anti-tumor effect for CRC and other malignancies with *MGMT* methylation by controlling the levels of *MGMT* in tumor [46]. It was hypothesized that tumor cells with methylation of *MGMT* are likely to remain in G2/M checkpoint, resulting in increased sensitivity to chemoradiation [47, 48].

Our results show that *MGMT* methylation is an important prognostic factor in stage III DAs. Our data also suggest a possible role for fluorouracil-based chemotherapy/radiotherapy in management of stage III DAs patients with *MGMT* methylation and *MGMT*-M may also then have a predictive role. Further studies in larger samples will help validate these.

Supporting Information

S1 File. SPSS file for statistical analysis.
(SAV)

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Author Contributions

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Data curation: TF NA.

Formal analysis: TF.

Funding acquisition: TF NA.

Investigation: TF AS FX YL KL WW CLW NA.

Methodology: TF NA.

Project administration: TF NA.

Resources: AS CLW NA.

Software: TF.

Supervision: TF SBB CLW NA.

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References

1. Alwmark A, Andersson A, Lasso A. Primary carcinoma of the duodenum. *Annals of surgery*. 1980; 191(1):13–8. PMID: [7352773](#); PubMed Central PMCID: [PMC1344610](#).
2. Overman MJ, Hu CY, Wolff RA, Chang GJ. Prognostic value of lymph node evaluation in small bowel adenocarcinoma: analysis of the surveillance, epidemiology, and end results database. *Cancer*. 2010; 116(23):5374–82. doi: [10.1002/ncr.25324](#) PMID: [20715162](#).
3. Overman MJ, Hu CY, Kopetz S, Abbruzzese JL, Wolff RA, Chang GJ. A population-based comparison of adenocarcinoma of the large and small intestine: insights into a rare disease. *Annals of surgical oncology*. 2012; 19(5):1439–45. doi: [10.1245/s10434-011-2173-6](#) PMID: [22187121](#); PubMed Central PMCID: [PMC3342860](#).
4. Singhal N, Singhal D. Adjuvant chemotherapy for small intestine adenocarcinoma. The Cochrane database of systematic reviews. 2007;(3):CD005202. doi: [10.1002/14651858.CD005202.pub2](#) PMID: [17636789](#).
5. Christmann M, Verbeek B, Roos WP, Kaina B. O(6)-Methylguanine-DNA methyltransferase (MGMT) in normal tissues and tumors: enzyme activity, promoter methylation and immunohistochemistry. *Biochimica et biophysica acta*. 2011; 1816(2):179–90. doi: [10.1016/j.bbcan.2011.06.002](#) PMID: [21745538](#).
6. Cecchini S, Correa-Gallego C, Desphande V, Ligorio M, Dursun A, Wargo J, et al. Superior prognostic importance of perineural invasion vs. lymph node involvement after curative resection of duodenal adenocarcinoma. *Journal of gastrointestinal surgery: official journal of the Society for Surgery of the Alimentary Tract*. 2012; 16(1):113–20; discussion 20. doi: [10.1007/s11605-011-1704-6](#) PMID: [22005894](#).
7. Kim SG, Chan AO, Wu TT, Issa JP, Hamilton SR, Rashid A. Epigenetic and genetic alterations in duodenal carcinomas are distinct from biliary and ampullary carcinomas. *Gastroenterology*. 2003; 124(5):1300–10. PMID: [12730870](#).
8. Overman MJ, Pozadzides J, Kopetz S, Wen S, Abbruzzese JL, Wolff RA, et al. Immunophenotype and molecular characterisation of adenocarcinoma of the small intestine. *British journal of cancer*. 2010; 102(1):144–50. doi: [10.1038/sj.bjc.6605449](#) PMID: [19935793](#); PubMed Central PMCID: [PMC2813754](#).
9. Laforest A, Aparicio T, Zaanani A, Silva FP, Didelot A, Desbeaux A, et al. ERBB2 gene as a potential therapeutic target in small bowel adenocarcinoma. *European journal of cancer*. 2014; 50(10):1740–6. doi: [10.1016/j.ejca.2014.04.007](#) PMID: [24797764](#).
10. Raghav K, Overman MJ. Small bowel adenocarcinomas—existing evidence and evolving paradigms. *Nature reviews Clinical oncology*. 2013; 10(9):534–44. doi: [10.1038/nrclinonc.2013.132](#) PMID: [23897080](#).
11. Fu T, Guzzetta AA, Jeschke J, Vatapalli R, Dave P, Hooker CM, et al. KRAS G>A mutation favors poor tumor differentiation but may not be associated with prognosis in patients with curatively resected duodenal adenocarcinoma. *International journal of cancer Journal international du cancer*. 2013; 132(11):2502–9. doi: [10.1002/ijc.27910](#) PMID: [23065691](#); PubMed Central PMCID: [PMC3579006](#).
12. Fu T, Pappou EP, Guzzetta AA, Jeschke J, Kwak R, Dave P, et al. CpG island methylator phenotype-positive tumors in the absence of MLH1 methylation constitute a distinct subset of duodenal adenocarcinomas and are associated with poor prognosis. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2012; 18(17):4743–52. doi: [10.1158/1078-0432.CCR-12-0707](#) PMID: [22825585](#); PubMed Central PMCID: [PMC3482463](#).
13. Yachida S, Mudali S, Martin SA, Montgomery EA, Iacobuzio-Donahue CA. Beta-catenin nuclear labeling is a common feature of sessile serrated adenomas and correlates with early neoplastic progression after BRAF activation. *Am J Surg Pathol*. 2009; 33(12):1823–32. Epub 2009/09/12. doi: [10.1097/PAS.0b013e3181b6da19](#) PMID: [19745699](#).
14. Dietmaier W, Wallinger S, Bocker T, Kullmann F, Fishel R, Ruschoff J. Diagnostic microsatellite instability: definition and correlation with mismatch repair protein expression. *Cancer Res*. 1997; 57(21):4749–56. Epub 1997/11/14. PMID: [9354436](#).
15. Weisenberger DJ, Siegmund KD, Campan M, Young J, Long TI, Faasse MA, et al. CpG island methylator phenotype underlies sporadic microsatellite instability and is tightly associated with BRAF mutation in colorectal cancer. *Nat Genet*. 2006; 38(7):787–93. Epub 2006/06/29. ng1834 [pii] doi: [10.1038/ng1834](#) PMID: [16804544](#).
16. Eads CA, Danenberg KD, Kawakami K, Saltz LB, Blake C, Shibata D, et al. MethyLight: a high-throughput assay to measure DNA methylation. *Nucleic Acids Res*. 2000; 28(8):E32. Epub 2000/03/29. gnd033 [pii]. PMID: [10734209](#); PubMed Central PMCID: [PMC102836](#).
17. Nosho K, Kure S, Irahara N, Shima K, Baba Y, Spiegelman D, et al. A prospective cohort study shows unique epigenetic, genetic, and prognostic features of synchronous colorectal cancers. *Gastroenterology*. 2009; 137(5):1609–20 e1-3. doi: [10.1053/j.gastro.2009.08.002](#) PMID: [19686742](#); PubMed Central PMCID: [PMC2859181](#).

18. Kohonen-Corish MR, Daniel JJ, Chan C, Lin BP, Kwun SY, Dent OF, et al. Low microsatellite instability is associated with poor prognosis in stage C colon cancer. *Journal of clinical oncology: official journal of the American Society of Clinical Oncology*. 2005; 23(10):2318–24. doi: [10.1200/JCO.2005.00.109](https://doi.org/10.1200/JCO.2005.00.109) PMID: [15800322](https://pubmed.ncbi.nlm.nih.gov/15800322/).
19. Hibi K, Sakata M, Yokomizo K, Kitamura YH, Sakuraba K, Shirahata A, et al. Methylation of the MGMT gene is frequently detected in advanced gastric carcinoma. *Anticancer research*. 2009; 29(12):5053–5. PMID: [20044616](https://pubmed.ncbi.nlm.nih.gov/20044616/).
20. Liu Y, Lan Q, Siegfried JM, Luketich JD, Keohavong P. Aberrant promoter methylation of p16 and MGMT genes in lung tumors from smoking and never-smoking lung cancer patients. *Neoplasia*. 2006; 8(1):46–51. doi: [10.1593/neo.05586](https://doi.org/10.1593/neo.05586) PMID: [16533425](https://pubmed.ncbi.nlm.nih.gov/16533425/); PubMed Central PMCID: PMC1584289.
21. Brabender J, Usadel H, Metzger R, Schneider PM, Park J, Salonga D, et al. Quantitative O(6)-methylguanine DNA methyltransferase methylation analysis in curatively resected non-small cell lung cancer: associations with clinical outcome. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2003; 9(1):223–7. PMID: [12538473](https://pubmed.ncbi.nlm.nih.gov/12538473/).
22. Zhao JJ, Li HY, Wang D, Yao H, Sun DW. Abnormal MGMT promoter methylation may contribute to the risk of esophageal cancer: a meta-analysis of cohort studies. *Tumour biology: the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014. doi: [10.1007/s13277-014-2276-3](https://doi.org/10.1007/s13277-014-2276-3) PMID: [25015189](https://pubmed.ncbi.nlm.nih.gov/25015189/).
23. Kim JI, Suh JT, Choi KU, Kang HJ, Shin DH, Lee IS, et al. Inactivation of O6-methylguanine-DNA methyltransferase in soft tissue sarcomas: association with K-ras mutations. *Human pathology*. 2009; 40(7):934–41. doi: [10.1016/j.humpath.2009.01.005](https://doi.org/10.1016/j.humpath.2009.01.005) PMID: [19356788](https://pubmed.ncbi.nlm.nih.gov/19356788/).
24. Fumagalli C, Della Pasqua S, Bagnardi V, Cardillo A, Sporchia A, Colleoni M, et al. Prevalence and clinicopathologic correlates of O(6)-methylguanine-DNA methyltransferase methylation status in patients with triple-negative breast cancer treated preoperatively by alkylating drugs. *Clinical breast cancer*. 2014; 14(4):285–90. doi: [10.1016/j.clbc.2014.02.010](https://doi.org/10.1016/j.clbc.2014.02.010) PMID: [24709436](https://pubmed.ncbi.nlm.nih.gov/24709436/).
25. Jesien-Lewandowicz E, Jesionek-Kupnicka D, Zawlik I, Szybka M, Kulczycka-Wojdala D, Rieske P, et al. High incidence of MGMT promoter methylation in primary glioblastomas without correlation with TP53 gene mutations. *Cancer genetics and cytogenetics*. 2009; 188(2):77–82. doi: [10.1016/j.cancergencyto.2008.09.015](https://doi.org/10.1016/j.cancergencyto.2008.09.015) PMID: [19100509](https://pubmed.ncbi.nlm.nih.gov/19100509/).
26. Stupp R, Hegi ME, Gorlia T, Erridge SC, Perry J, Hong YK, et al. Cilengitide combined with standard treatment for patients with newly diagnosed glioblastoma with methylated MGMT promoter (CENTRIC EORTC 26071–22072 study): a multicentre, randomised, open-label, phase 3 trial. *The Lancet Oncology*. 2014; 15(10):1100–8. doi: [10.1016/S1470-2045\(14\)70379-1](https://doi.org/10.1016/S1470-2045(14)70379-1) PMID: [25163906](https://pubmed.ncbi.nlm.nih.gov/25163906/).
27. Esteller M, Toyota M, Sanchez-Cespedes M, Capella G, Peinado MA, Watkins DN, et al. Inactivation of the DNA repair gene O6-methylguanine-DNA methyltransferase by promoter hypermethylation is associated with G to A mutations in K-ras in colorectal tumorigenesis. *Cancer research*. 2000; 60(9):2368–71. PMID: [10811111](https://pubmed.ncbi.nlm.nih.gov/10811111/).
28. Nagasaka T, Goel A, Notohara K, Takahata T, Sasamoto H, Uchida T, et al. Methylation pattern of the O6-methylguanine-DNA methyltransferase gene in colon during progressive colorectal tumorigenesis. *International journal of cancer Journal international du cancer*. 2008; 122(11):2429–36. doi: [10.1002/ijc.23398](https://doi.org/10.1002/ijc.23398) PMID: [18240147](https://pubmed.ncbi.nlm.nih.gov/18240147/); PubMed Central PMCID: PMC2851179.
29. Rosty C, Young JP, Walsh MD, Clendenning M, Walters RJ, Pearson S, et al. Colorectal carcinomas with KRAS mutation are associated with distinctive morphological and molecular features. *Modern pathology: an official journal of the United States and Canadian Academy of Pathology, Inc*. 2013; 26(6):825–34. doi: [10.1038/modpathol.2012.240](https://doi.org/10.1038/modpathol.2012.240) PMID: [23348904](https://pubmed.ncbi.nlm.nih.gov/23348904/).
30. Zhang Y, Kim J, Mueller AC, Dey B, Yang Y, Lee DH, et al. Multiple receptor tyrosine kinases converge on microRNA-134 to control KRAS, STAT5B, and glioblastoma. *Cell death and differentiation*. 2014; 21(5):720–34. doi: [10.1038/cdd.2013.196](https://doi.org/10.1038/cdd.2013.196) PMID: [24440911](https://pubmed.ncbi.nlm.nih.gov/24440911/); PubMed Central PMCID: PMC3978301.
31. Nagy E, Gajjar KB, Patel II, Taylor S, Martin-Hirsch PL, Stringfellow HF, et al. MGMT promoter hypermethylation and K-RAS, PTEN and TP53 mutations in tamoxifen-exposed and non-exposed endometrial cancer cases. *British journal of cancer*. 2014; 110(12):2874–80. doi: [10.1038/bjc.2014.263](https://doi.org/10.1038/bjc.2014.263) PMID: [24853176](https://pubmed.ncbi.nlm.nih.gov/24853176/); PubMed Central PMCID: PMC4056065.
32. Wu M, Semba S, Oue N, Ikehara N, Yasui W, Yokozaki H. BRAF/K-ras mutation, microsatellite instability, and promoter hypermethylation of hMLH1/MGMT in human gastric carcinomas. *Gastric cancer: official journal of the International Gastric Cancer Association and the Japanese Gastric Cancer Association*. 2004; 7(4):246–53. doi: [10.1007/s10120-004-0300-9](https://doi.org/10.1007/s10120-004-0300-9) PMID: [15616773](https://pubmed.ncbi.nlm.nih.gov/15616773/).
33. Hawkins NJ, Lee JH, Wong JJ, Kwok CT, Ward RL, Hitchins MP. MGMT methylation is associated primarily with the germline C>T SNP (rs16906252) in colorectal cancer and normal colonic mucosa. *Modern pathology: an official journal of the United States and Canadian Academy of Pathology, Inc*. 2009; 22(12):1588–99. doi: [10.1038/modpathol.2009.130](https://doi.org/10.1038/modpathol.2009.130) PMID: [19734844](https://pubmed.ncbi.nlm.nih.gov/19734844/).

34. Iwagami S, Baba Y, Watanabe M, Shigaki H, Miyake K, Ishimoto T, et al. LINE-1 hypomethylation is associated with a poor prognosis among patients with curatively resected esophageal squamous cell carcinoma. *Annals of surgery*. 2013; 257(3):449–55. doi: [10.1097/SLA.0b013e31826d8602](https://doi.org/10.1097/SLA.0b013e31826d8602) PMID: [23023202](https://pubmed.ncbi.nlm.nih.gov/23023202/).
35. Shima K, Morikawa T, Baba Y, Nosho K, Suzuki M, Yamauchi M, et al. MGMT promoter methylation, loss of expression and prognosis in 855 colorectal cancers. *Cancer causes & control: CCC*. 2011; 22(2):301–9. doi: [10.1007/s10552-010-9698-z](https://doi.org/10.1007/s10552-010-9698-z) PMID: [21140203](https://pubmed.ncbi.nlm.nih.gov/21140203/); PubMed Central PMCID: [PMC3278857](https://pubmed.ncbi.nlm.nih.gov/PMC3278857/).
36. Baumann S, Keller G, Puhlinger F, Napieralski R, Feith M, Langer R, et al. The prognostic impact of O6-Methylguanine-DNA Methyltransferase (MGMT) promoter hypermethylation in esophageal adenocarcinoma. *International journal of cancer Journal international du cancer*. 2006; 119(2):264–8. doi: [10.1002/ijc.21848](https://doi.org/10.1002/ijc.21848) PMID: [16477636](https://pubmed.ncbi.nlm.nih.gov/16477636/).
37. Bilimoria KY, Bentrem DJ, Wayne JD, Ko CY, Bennett CL, Talamonti MS. Small bowel cancer in the United States: changes in epidemiology, treatment, and survival over the last 20 years. *Annals of surgery*. 2009; 249(1):63–71. doi: [10.1097/SLA.0b013e31818e4641](https://doi.org/10.1097/SLA.0b013e31818e4641) PMID: [19106677](https://pubmed.ncbi.nlm.nih.gov/19106677/).
38. Swartz MJ, Hughes MA, Frassica DA, Herman J, Yeo CJ, Riall TS, et al. Adjuvant concurrent chemoradiation for node-positive adenocarcinoma of the duodenum. *Archives of surgery*. 2007; 142(3):285–8. doi: [10.1001/archsurg.142.3.285](https://doi.org/10.1001/archsurg.142.3.285) PMID: [17372054](https://pubmed.ncbi.nlm.nih.gov/17372054/).
39. Poultsides GA, Huang LC, Cameron JL, Tuli R, Lan L, Hruban RH, et al. Duodenal adenocarcinoma: clinicopathologic analysis and implications for treatment. *Annals of surgical oncology*. 2012; 19(6):1928–35. doi: [10.1245/s10434-011-2168-3](https://doi.org/10.1245/s10434-011-2168-3) PMID: [22167476](https://pubmed.ncbi.nlm.nih.gov/22167476/); PubMed Central PMCID: [PMC3663711](https://pubmed.ncbi.nlm.nih.gov/PMC3663711/).
40. Kelsey CR, Nelson JW, Willett CG, Chino JP, Clough RW, Bendell JC, et al. Duodenal adenocarcinoma: patterns of failure after resection and the role of chemoradiotherapy. *International journal of radiation oncology, biology, physics*. 2007; 69(5):1436–41. doi: [10.1016/j.ijrobp.2007.05.006](https://doi.org/10.1016/j.ijrobp.2007.05.006) PMID: [17689032](https://pubmed.ncbi.nlm.nih.gov/17689032/).
41. Cloyd JM, Norton JA, Visser BC, Poultsides GA. Does the Extent of Resection Impact Survival for Duodenal Adenocarcinoma? Analysis of 1,611 Cases. *Annals of surgical oncology*. 2014. doi: [10.1245/s10434-014-4020-z](https://doi.org/10.1245/s10434-014-4020-z) PMID: [25160736](https://pubmed.ncbi.nlm.nih.gov/25160736/).
42. Weller M, Felsberg J, Hartmann C, Berger H, Steinbach JP, Schramm J, et al. Molecular predictors of progression-free and overall survival in patients with newly diagnosed glioblastoma: a prospective translational study of the German Glioma Network. *Journal of clinical oncology: official journal of the American Society of Clinical Oncology*. 2009; 27(34):5743–50. doi: [10.1200/JCO.2009.23.0805](https://doi.org/10.1200/JCO.2009.23.0805) PMID: [19805672](https://pubmed.ncbi.nlm.nih.gov/19805672/).
43. Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJ, Janzer RC, et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. *The Lancet Oncology*. 2009; 10(5):459–66. doi: [10.1016/S1470-2045\(09\)70025-7](https://doi.org/10.1016/S1470-2045(09)70025-7) PMID: [19269895](https://pubmed.ncbi.nlm.nih.gov/19269895/).
44. Hegi ME, Diserens AC, Gorlia T, Hamou MF, de Tribolet N, Weller M, et al. MGMT gene silencing and benefit from temozolomide in glioblastoma. *The New England journal of medicine*. 2005; 352(10):997–1003. doi: [10.1056/NEJMoa043331](https://doi.org/10.1056/NEJMoa043331) PMID: [15758010](https://pubmed.ncbi.nlm.nih.gov/15758010/).
45. Nagasaka T, Sharp GB, Notohara K, Kambara T, Sasamoto H, Isozaki H, et al. Hypermethylation of O6-methylguanine-DNA methyltransferase promoter may predict nonrecurrence after chemotherapy in colorectal cancer cases. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2003; 9(14):5306–12. PMID: [14614014](https://pubmed.ncbi.nlm.nih.gov/14614014/).
46. Murakami J, Lee YJ, Kokeguchi S, Tsujigiwa H, Asaumi J, Nagatsuka H, et al. Depletion of O6-methylguanine-DNA methyltransferase by O6-benzylguanine enhances 5-FU cytotoxicity in colon and oral cancer cell lines. *Oncology reports*. 2007; 17(6):1461–7. PMID: [17487405](https://pubmed.ncbi.nlm.nih.gov/17487405/).
47. Chalmers AJ, Ruff EM, Martindale C, Lovegrove N, Short SC. Cytotoxic effects of temozolomide and radiation are additive- and schedule-dependent. *International journal of radiation oncology, biology, physics*. 2009; 75(5):1511–9. doi: [10.1016/j.ijrobp.2009.07.1703](https://doi.org/10.1016/j.ijrobp.2009.07.1703) PMID: [19931733](https://pubmed.ncbi.nlm.nih.gov/19931733/).
48. Yan L, Donze JR, Liu L. Inactivated MGMT by O6-benzylguanine is associated with prolonged G2/M arrest in cancer cells treated with BCNU. *Oncogene*. 2005; 24(13):2175–83. doi: [10.1038/sj.onc.1208250](https://doi.org/10.1038/sj.onc.1208250) PMID: [15735757](https://pubmed.ncbi.nlm.nih.gov/15735757/).