## 1 SUPPLEMENTARY INFORMATION

- 2 Marine biogenic humic substances control iron biogeochemistry across the Southern Ocean
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## Supplementary methods

Particulate analyses.

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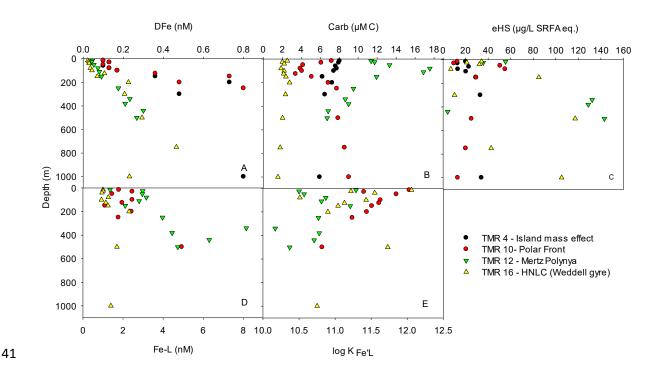
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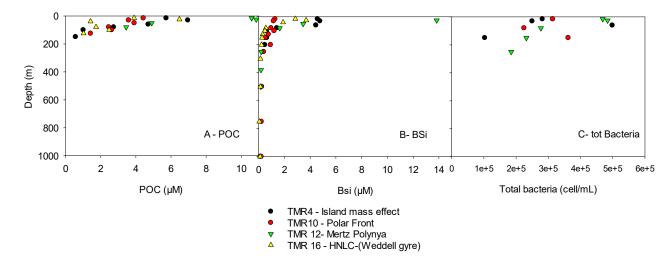
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Microbial abundances. Samples (2 to 5 ml each) were fixed with 1% paraformaldehyde plus 0.05% glutaraldehyde (final concentrations) for 15 min at room temperature and stored at -80°C. For enumeration of heterotrophic prokaryotes (mostly bacteria), thawed samples were stained with SYBRgreen I (Molecular Probes; ~20 μM final concentration) prior to quantification by green fluorescence on a flow cytometer (CyFlow Cube 8, Sysmex Partec). Prokaryotic and eukaryotic picoand nanophytoplankton were also counted on the flow cytometer based on their red and orange autofluorescence<sup>1</sup>. Biogenic silicate (BSi). Seawater samples were collected from the underway seawater supply, filtered on board and then analysed by flow injection following appropriate digestion<sup>2</sup> to yield Bsi concentrations ( $\mu$ M). Particulate organic carbon (POC). Seawater was filtered through pre-combusted (450°C for 8 hr) glass fibre filters (GF-75s; 0.3 μm pore size; Sterlitech) that were then stored in pre-combusted foil envelopes at -80°C pending analysis. In the laboratory, the POC filters were dried at 40°C for 24 hr, then folded into tin capsules that were analysed for C content using a Flash 2000 elemental analyser coupled to a Delta V Plus isotope ratio mass spectrometer in a configuration with a detection limit of 2 μg C. A range of known masses of three in-house standards were analysed alongside the samples, with the resultant relationship of measured standard peak area to mass used to convert sample peaks areas to C mass. POC concentration was calculated by normalizing C mass to seawater volume filtered<sup>3,4</sup>. Phytoplankton. Pigments and CHEMTAX analyses were done as in 5. Chemtax and photosynthetic maximum quantum yield (Fv/Fm) were kindly communicated by C. Robinson.

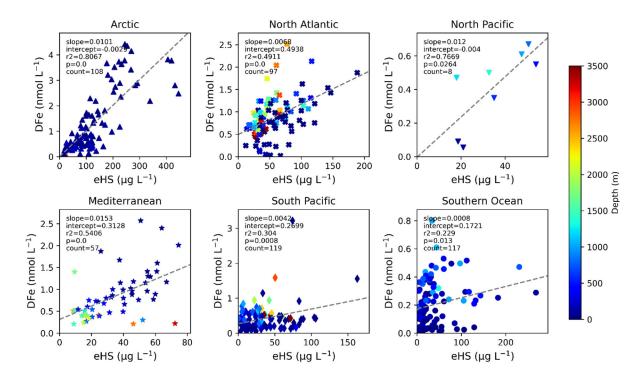
## 40 Supplementary Figures and Tables



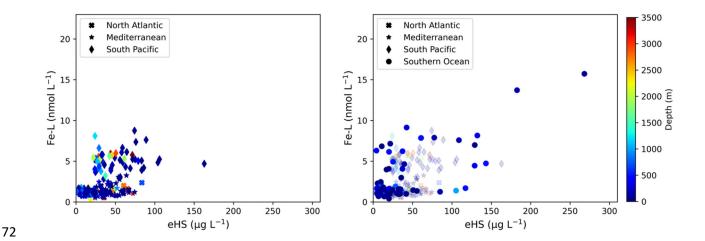
Supplementary Fig. 1. Representation of chemical parameters for contrasted regions of the Southern Ocean. The dissolved Fe (DFe, panel A), hydrolysable carbohydrates (Carb, Panel B), electroactive humic substances (eHS, panel C), Fe-binding ligands (Fe-L, panel D), and the log of the conditional stability constant (log  $K_{Fe'L}$ , panel E) are shown. These regions include area influenced by island mass effect, Polar Front, polynya and typical High Nutrient Low Chlorophyll (HNLC) region of the Southern Ocean. These stations represent a subset of the station sampled during the ACE expedition.



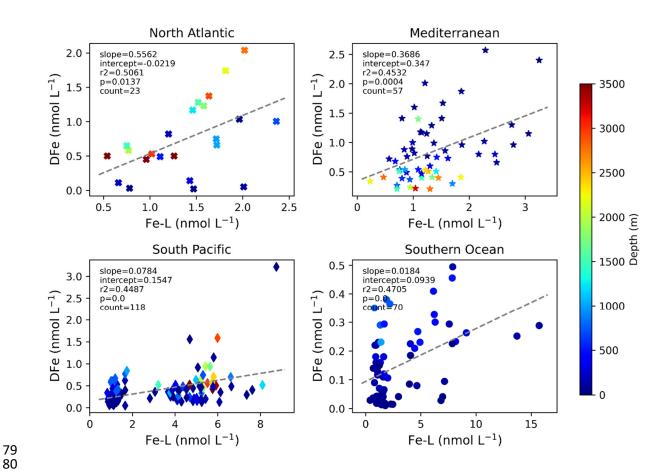
Supplementary Fig. 2. Representation of biological parameters for contrasted regions of the Southern Ocean. The particulate organic carbon (POC, panel A), biogenic silicate (BSi, Panel B), and total heterotrophic bacteria cellular abundance (panel C) are shown. These regions include area influenced by island mass effect, Polar Front, polynya and typical High Nutrient Low Chlorophyll (HNLC) region of the Southern Ocean. These stations represent a subset of the station sampled during the ACE expedition.



**Supplementary Fig. 3.** Relationship between DFe, eHS and depth in different ocean basins. Dara are shown for the Arctic<sup>6,7</sup>; North Atlantic<sup>8,9</sup>; North Pacific<sup>8</sup>; Mediterranean Sea<sup>9,10</sup>; South Pacific<sup>11,12</sup>; and Southern Ocean <sup>8</sup>(this study). Regression parameters are indicated for each of the ocean basin and sea represented.



Supplementary Fig. 4. Relationship between Fe-L and eHS, and depth in different ocean basins. Data are shown for the North Atlantic<sup>8,9</sup>; Mediterranean Sea<sup>9,10</sup>; South Pacific<sup>11,12</sup>; and Southern Ocean <sup>8</sup>(this study). No Arctic and North Pacific iron-binding ligands were available on the selected studies. All Fe-L data were derived using SA as the exchange ligand, except for the Mediterranean Sea where TAC was used.



Supplementary Figure 5. Relationship between DFe and Fe-L in different ocean basins. Dara are shown for the North Atlantic<sup>8,9</sup>; Mediterranean Sea<sup>9,10</sup>; South Pacific<sup>11,12</sup>; and Southern Ocean <sup>8</sup>(this study). No Arctic and North Pacific iron-binding ligands were available on the selected studies.

Regression parameters are indicated for each of the ocean basin and sea represented.

Supplementary Table 1. Antarctic Circumnavigation Expedition (ACE) stations – Cape Town to

Hobart (Leg 1, 20-December-2016 to 19-January-2017, TMR stations 3–7) and Hobart to Punta Arenas

(Leg 2, 22-January-2017 to 22-February-2017, TMR stations 8–20) – onboard the R/V Akademik

Tryoshnikov.

Stn#	TMR#	I	Date	Lat	Long
	16	3	07.01.2017	-50.994	72.04
	19	4	09.01.2017	-53.133	81.721
	20	5	11.01.2017	-54.825	95.684
	21	6	14.01.2017	-53.198	3 118.134
	22	7	17.01.2017	-46.906	5 141.931
	23	8	23.01.2017	-46.393	150.3888
	24	9	25.01.2017	-53.5825	149.2976
	25	10	26.01.2017	-59.6105	148.6399
	26	11	29.01.2017	-67.1036	5 144.9204
	36	12	30.01.2017	-67.1895	145.7209
	43	13	31.01.2017	-65.9932	159.0105
	47	14	04.02.2017	-67.2897	7 163.536
	49	15	05.02.2017	-67.0999	9 167.3602
	56	16	09.02.2017	-71.6993	-143.6983
	68	17	14.02.2017	-68.7486	-99.9592
	71	18	18.02.2017	-63.9632	-66.2425
	72	19	18.02.2017	-62.5089	-68.0376
	73	20	19.02.2017	-59.5999	-67.9187

Supplementary Table 2. Pearson correlations table between selected parameters considering data for the full water column (0-1000m). Selected parameters are depth, dissolved Fe (DFe), Fe-binding ligands (Fe-L) with their conditional stability constant (log  $K_{Fe'L}$ ), hydrolysable carbohydrates (Carb), electroactive humic substances (eHS) and total bacterial cell abundance (Bacttot). Statistically significant (p < 0.05) positive and negative correlations are highlighted in green and red, respectively. Data show coefficient of correlation (R) on top, p value (p) and number of observations at the bottom.

	DFe	Fe-L	log K	Carb	eHS	Bacttot	
depth	0.623	0.0634	-0.0124	-0.213	0.163	-0.262	
	5.45E-27	0.491	0.893	0.000942	0.0944	0.0155	
	238	120	120	239	106	85	
DFe		0.487	-0.0874	-0.296	0.245	-0.291	
		1.65E-08	0.342	0.00000352	0.0119	0.00721	
		120	120	237	105	84	
Fe-L			-0.22	0.0897	0.699	-0.416	
			0.0156	0.33	1.76E-11	0.00451	
			120	120	70	45	
log K				-0.0807		-0.13	
				0.381	0.0287	0.393	
				120	70	45	
Carb					-0.0547	0.395	
					0.578	0.000185	
					106	85	
eHS						-0.397	
						0.00637	
						46	

Supplementary Table 3. Pearson correlations table between selected parameters considering data for the surface water only (0-100m). Selected parameters are dissolved Fe (DFe), Fe-binding ligands (Fe-L) with their conditional stability constant (log K<sub>Fe'L</sub>), hydrolysable carbohydrates (Carb), electroactive humic substances (eHS), particulate organic carbon (POC) biogenic silicate (BSi), and maximum effective quantum yield (Fv/Fm). Flow cytometry derived cellular abundances is shown for total bacteria (totBact), Synechococcus (Syn), Picoeukarotes (Picoeuk 1 and 2), Nanoeukaryotes (Nanoeuk) and Cryptomonas. Phytoplankton classes derived from pigments and CHEMTAX analysis are also shown for Chlorophytes (Chloro), Cryptophytes (Crypto), Cyanobacteria (Cyano), Diatoms (Diat A and B), Dinophytes (Dino), Haptophytes (Hapto8 and 67), Prasinophytes (Pras) and Pelagophytes (Pelago). Statistically significant (p < 0.05) positive and negative correlations are highlighted in green and red, respectively. Data show coefficient of correlation (R) on top, p value (p) and number of observations at the bottom.

	Fe-L los	. r	Carb e	HS Bsi	i PO	r B	acttot S	ovn I	Picoeuk1	Picoeuk2	Nanoeuk	Cryoptomon Ch	oloro C	rypto C	yano	DiatA D	iatB D	Dino H	lapto8	Hapto67	Pras F	Pelago	Fv/Fm
DFe	0.462	-0.251	-0.000496	0.11	-0.123	-0.0276	-0.113	-0.0844	-0.0644	-0.288	0.0235	-0.278	-0.128	0.0322	-0.237	0.00466	-0.0894	-0.17	0.214		-0.0784	-0.165	0.164
Dre	0.000437	0.0673	0.996	0.419	0.234	0.79	0.377	0.518	0.616	0.022	0.0255	0.0287	0.223	0.761	0.0231	0.00400	0.397	0.106	0.0407			0.115	0.104
	54	54	101	56	96	96	63	61	63	63	63	62	92	92	92	92	92	92	92	92	92	92	90
F- 1		0.113	0.197	0.801	0.110	-0.31	0.2	-0.299	0.0053	0.154	0.0122	0.153	-0.135	0.125	0.221	-0.132	0.0005	-0.365	0.0400	5 -0.233	0.00533	0.222	0.510
Fe-L		-0.113			-0.118		-0.2		-0.0853		0.0132				-0.221		-0.0885		0.0406			-0.232	0.518
		0.417	0.154	9.18E-10	0.413	0.027	0.274	0.103	0.643	0.4	0.943	0.41	0.366	0.402	0.135	0.378	0.554	0.0117	0.786			0.117	0.000268
		54	54	39	50	51	32	31	32	32	32	31	47	47	47	47	47	47	47	7 47	47	47	45
log K			-0.112	-0.111	-0.239	-0.231	-0.323	-0.0645	0.0243	-0.151	-0.359	0.0899	-0.147	-0.252	-0.0744	-0.0661	-0.229	0.0816	0.0012	2 -0.18	-0.0667	-0.11	0.0798
			0.421	0.501	0.0946	0.103	0.0712	0.73	0.895	0.408	0.0437	0.631	0.323	0.0876	0.619	0.659	0.122	0.585	0.994	1 0.227	0.656	0.461	0.602
			54	39	50	51	32	31	32	32	32	31	47	47	47	47	47	47	47	7 47	47	47	45
Carb				-0.0297	0.0268	0.101	0.432	0.485	0.47	0.435	0.337	0.254	0.454	-0.0382	0.293	0.07	0.114	0.399	0.19	0.515	0.204	0.314	0.141
				0.828	0.794	0.324	0.000368	0.0000649	0.0000904	0.000333	0.00654	0.045	0.00000479	0.716	0.00444	0.505	0.275	0.000075	0.0687	7 0.00000013	0.0494	0.00216	0.183
				56	97	97	64	62	64	64	64	63	93	93	93	93	93	93	93	93	93	93	91
eHS					-0.0462	-0.174	-0.161	-0.224	-0.336	-0.218	-0.0744	-0.0391	-0.209	-0.0386	-0.186	-0.0585	0.0267	-0.188	-0.099	-0.252	-0.0792	-0.266	0.307
					0.748	0.212	0.38	0.226	0.0602	0.23	0.686	0.835	0.149	0.792	0.201	0.69	0.855	0.195	0.499	0.0802	0.588	0.0647	0.036
					51	53	32	31	32	32	32	31	49	49	49	49	49	49	49	9 49	49	49	47

## **Supplementary References**

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