

'Community evolution' – laboratory strains and pedigrees in the age of genomics

Matthew J. Dorman¹ and Nicholas R. Thomson^{1,2,*}

Abstract

Molecular microbiologists depend heavily on laboratory strains of bacteria, which are ubiquitous across the community of research groups working on a common organism. However, this presumes that strains present in different laboratories are in fact identical. Work on a culture of *Vibrio cholerae* preserved from 1916 provoked us to consider recent studies, which have used both classical genetics and next-generation sequencing to study the heterogeneity of laboratory strains. Here, we review and discuss mutations and phenotypic variation in supposedly isogenic reference strains of *V. cholerae* and *Escherichia coli*, and we propose that by virtue of the dissemination of laboratory strains across the world, a large 'community evolution' experiment is currently ongoing.

Much of our current knowledge of bacteriology has been founded on the study of laboratory strains that are members of a species of interest. These strains are often deeply characterized, have had their genomes sequenced to completion, and are present in laboratories around the world. Standardizing on a handful of laboratory strains for molecular studies, rather than using environmental or clinical isolates, enables reproducible studies of a single strain's genetics and physiology. This facilitates the stepwise accumulation of knowledge in research – as one group publishes on the physiology of a laboratory strain, another group can build upon that knowledge when they study their own stock of that strain. A large caveat to focusing on limited numbers of lab strains is that just as type strains define bacterial species taxonomically [1], a type strain is not the archetype of a species [2]. Similarly, it can be argued that we know more about our laboratory strains of bacteria at a genetic and biochemical level than we do about a species as a whole [3].

In a recent study, we and our collaborators revived a lyophilized stock of NCTC 30, a *Vibrio cholerae* first isolated in 1916 [4]. We sequenced the genome of this isolate, as well as examining its morphology and phenotype. Although we did not find many previous studies that had examined NCTC 30 [5–7] (Fig. 1), we took comfort in the fact that the cultures with which we worked behaved similarly to the findings reported in previous papers. We observed that NCTC 30 was

phylogenetically distinct from pandemic *V. cholerae*, in agreement with a previous taxonomic study [6]. Concordant with another study, we found that NCTC 30 was less susceptible to ampicillin than NCTC 5395, another *V. cholerae* [7], and we identified a functional β -lactamase gene in the genome of NCTC 30 that we believe confers this phenotype [4]. All of this reassured us that although we were working with a bacterial stock that has been maintained for over 102 years, we were likely to be handling a descendent or close relative of the cultures studied in the past.

However, a striking difference between our results and previous reports was the finding that our stock of NCTC 30 did not express flagella when examined microscopically, and was not motile on soft agar plates [4]. This disagreed with a previous report, which had described NCTC 30 as being flagellated [7]. Moreover, we were able to identify a mutation in our NCTC 30 genome, which we believe is responsible for this phenotype. Since it is wholly possible that spontaneous mutation due to long-term culture led to the loss of flagella in NCTC 30, we confirmed that this mutation is present in the batches of NCTC 30 currently available to purchase from Public Health England Culture Collections. Thus, researchers that now choose to study NCTC 30 ought to be working with a culture with the same genotype and phenotypes as our stock.

Discrepancies between what one 'expects' of a laboratory strain and how it actually behaves *in vitro* are a fascinating,

Received 15 September 2019; Accepted 15 October 2019; Published 20 January 2020

Author affiliations: ¹Wellcome Sanger Institute, Wellcome Genome Campus, Hinxton, Cambridgeshire, CB10 1SA, UK; ²London School of Hygiene and Tropical Medicine, Keppel St, Bloomsbury, London WC1E 7HT, UK.

***Correspondence:** Nicholas R. Thomson, nrt@sanger.ac.uk

Abbreviations: ATCC, American Type Culture Collection; DSMZ, Deutsche Sammlung von Mikroorganismen und Zellkulturen; NCTC, National Collection of Type Cultures; rRNA, Ribosomal RNA.

000869 © 2020 The Authors



This is an open-access article distributed under the terms of the Creative Commons Attribution License.

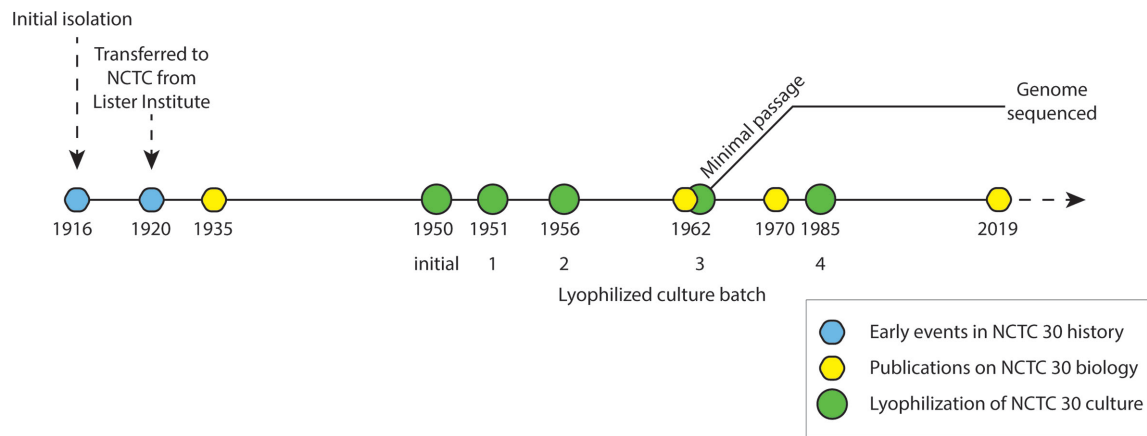


Fig. 1. Overview of the curation of NCTC 30. NCTC 30 was isolated in 1916, transferred to the Lister Institute, and subsequently transferred into the NCTC collection in 1920. The strain was lyophilized in 1950 and four consecutive lyophilized stock batches have been prepared since then. Assorted manuscripts have studied NCTC 30 since its isolation, including [4–7]. Figure drawn using records in the supplementary data of [4].

and yet potentially problematic, aspect of bacteriology. Our study of NCTC 30 meant that we felt it timely to consider the variability of laboratory strains between and within laboratories, and the potentially fascinating community evolution that has been ongoing across these laboratories. Below, we address some recent examples of discrepancies between supposedly isogenic stocks of laboratory strains of bacteria, and between those strains and their reference genome sequence.

V. cholerae researchers typically make use of a small number of reference laboratory strains, which have been shared amongst laboratories worldwide and are used for genetic experiments, phenotypic comparisons between *V. cholerae* biotypes, etc. The initial *V. cholerae* reference genome sequence was determined in 2000 by Heidelberg and colleagues by sequencing the serogroup O1 biotype El Tor strain N16961 [8]. However, differences between a published genome sequence for a laboratory strain and the actual genome of the organism held in a laboratory stock can confound research findings. For instance, Val *et al.* used N16961 as a reference strain for studying the timing of *V. cholerae* chromosomal replication, but found discrepancies between the N16961 reference sequence and their results [9]. In their data, they identified an inversion flanked by rRNA operons in the genome of their N16961 stock relative to that of the reference sequence [9]. They confirmed that the chromosomal orientation in their N16961 cultures was common to N16961 stocks kept by other laboratories, as well as in other *V. cholerae* [9], and speculated that the inversion had occurred in the original stock of N16961 that was sequenced by Heidelberg *et al.* [8, 9].

Similar differences between published reference sequences and the genomes of laboratory strains have been identified in the case of the *V. cholerae* A1552 reference strain, the genome of which has been sequenced at least three times in the last 2 years [10–12]. Kemter *et al.*, in a study of *Vibrio* chromosomal replication termination, used the N16961 reference sequence to design custom microarrays for this work, but found several

inversions between their A1552 strain's sequence and the reference N16961 genome [11]. The isolate sequenced by Kemter *et al.* contained an inversion characteristic of A1552, which was validated using long-read sequencing [11]. The genome sequence was published alongside the paper, and the sequenced strain was deposited in the DSMZ culture collection. Just as we have discussed for NCTC 30, purchase of A1552 from DSMZ ought to provide a researcher with the same bacteria as those characterized previously [11].

Discrepancies between expected and observed phenotypes can also confound genetic experiments in laboratory strains. An example of this was published in 2016, in which seven laboratory stocks of the *V. cholerae* reference strain C6706 were compared for their ability to be naturally transformed [13]. The natural competence of *V. cholerae* can be exploited by researchers to mutagenise *V. cholerae* experimentally [14–16]. Natural competence is induced in *V. cholerae* in response to being grown on chitin [14], and is also regulated by quorum sensing [17–19]. The master regulator of quorum sensing in *V. cholerae*, HapR, positively regulates genes, which encode components of the DNA-uptake apparatus, and negatively regulates expression of the *dns* nuclease gene [17, 20] (summarized in Fig. 2). It is well known in the *V. cholerae* community, for instance, that N16961 cannot be transformed naturally [14], due to a frameshift in the *hapR* gene [21, 22]. In the absence of HapR, *dns* is not repressed under conditions which would otherwise promote natural competence, leading to the degradation of exogenous DNA [20]. *hapR* mutations have been suggested to occur frequently in *V. cholerae* [22], and this has also been suggested to be as a consequence of the intrinsic bias towards collecting virulent isolates [15]; HapR downregulates the expression of cholera toxin and the toxin co-regulated pilus, principal *V. cholerae* virulence determinants (Fig. 2), and thus an isolate lacking functional *hapR* might cause a more acute disease, leading to hospitalization

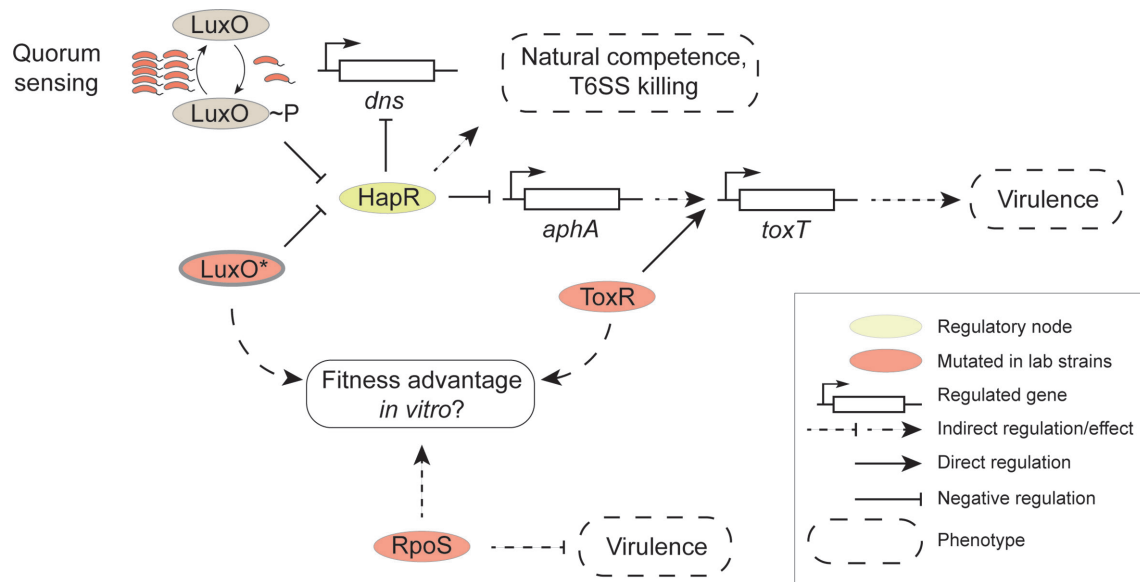


Fig. 2. Laboratory strains of bacteria harbour mutations in several pathways which converge on increasing fitness *in vitro*. Phosphorylated LuxO (LuxO~P) decreases in concentration in *V. cholerae* cells as the population of bacteria grows, measured by quorum sensing. LuxO~P represses HapR; thus, at high-cell density, HapR abundance increases as the concentration of active LuxO~P decreases. HapR influences quorum-dependent phenotypes such as natural competence and virulence, the latter by repressing *aphA*, an activator of virulence gene expression in *V. cholerae* [26]. Mutations in *hapR* and *luxO* therefore influence these phenotypes. Other mutations can prevent the activation of the virulence regulon, such as in *toxR*, or producing constitutively activated LuxO* mimicking low-cell density quorum sensing, thereby forcing the maintenance of HapR repression of the virulence regulon. Similarly, mutations in *rpoS* are reported to repress virulence, and therefore increase the fitness *in vitro* of these mutants relative to wild-type cells.

and the isolation of *V. cholerae* more likely to be added to a strain collection [15].

Various researchers reported that they were unable to effect natural transformation in C6706, leading to Stutzmann and Blokesch's study [13]. They found that the seven C6706 stocks examined had wild-type *hapR* alleles, but four contained a mutant *luxO* allele predicted to encode a mutant LuxO^{G333S} protein [13]. This correlated with impaired quorum sensing-dependent phenotypes, such as chitin-induced transformation and reduced type VI secretion system-mediated killing of *E. coli* prey bacteria, as well as reduced levels of *hapR* transcription and a concomitant change in the expression of the HapR regulon [13]. They concluded that the circulation of quorum sensing-deficient strains of C6706 necessitated that researchers working with this strain should confirm the *luxO* genotype of their laboratory stocks, if discordant natural transformation phenotypes manifest. They suggested that the presence of the same *luxO* allele in these four stocks meant that the same *luxO* mutant strain had been passed amongst four laboratories [13].

It has been shown that prolonged maintenance of toxigenic *V. cholerae* on agar stabs can select for spontaneous nontoxigenic mutants, which arise due to mutation in the *toxR* promoter [23]. These *toxR* mutants do not express the ToxR regulon, including the cholera toxin, and gain a competitive growth advantage over toxigenic cells *in vitro* [23] (Fig. 2). Likewise, *luxO** mutations producing constitutively active LuxO have

been reported to arise in *Vibrio fischeri* cultures that are maintained in stationary phase for prolonged periods of time [24]. These *luxO** mutants gained a fitness advantage over wild-type *V. fischeri* under the conditions used in the study [24]. The mutant LuxO^{G333S} protein found in C6706 stocks has been shown to mimic constitutively active LuxO [25] (Fig. 2). It is plausible that C6706 *luxO** mutants possess a competitive advantage over their wild-type counterparts, leading to the isolation of such mutants during continuous passage and their subsequent dissemination to laboratories worldwide. It is also possible that these *luxO** mutants express elevated levels of virulence determinants compared to the wild-type C6706 (Fig. 2). Mutations affecting the interactions between LuxO, HapR and AphA directly modulate virulence and cholera toxin expression in *V. cholerae* [26] (Fig. 2), and since C6706 is a lab strain commonly used for studying the regulation of *V. cholerae* virulence, mutants may inadvertently have been selected by researchers that express greater, more detectable, levels of virulence determinants, potentially distorting our view of the evolutionary trajectory of natural isolates.

V. cholerae are not the only bacteria susceptible to lab-to-lab variation in supposedly isogenic strains. Numerous examples of the same phenomenon in *E. coli* have been reported. Although the prototype *E. coli* K-12 was first isolated in 1922, from a convalescent diphtheria patient in Palo Alto, CA, USA [27], its descendants continue to be used by researchers worldwide. In 2006, Hayashi *et al.* [28] reported genome

sequences for two such laboratory strains, MG1655 and W3110, in a manuscript which accompanied the announcement of the Keio mutant collection [29]. The Keio mutants were constructed in BW25113 [30], a strain of *E. coli* derived from W1485, a common ancestor both of MG1655 [27, 31] and W3110 [27, 32]. The pedigree for BW25113 was reported in great detail upon publication of the Keio collection [29], and its genome sequence was subsequently determined [33]. Although an in-depth review of *E. coli* strain pedigrees is beyond the scope of this article, it would be remiss not to mention the work of organizations such as the *E. coli* Genetic Stock Center (CGSC) and Barbara Bachmann, its former curator, in maintaining extensive records of strain provenance, genetic description and pedigree [27, 32, 34]. Hayashi *et al.* reported heterogeneity amongst stocks of W3110 from laboratories across Japan, which they linked chiefly to IS element transposition events, which may have occurred during storage in stabs [28] – it is accepted that genome mutation takes place while *E. coli* is maintained as a stab culture [35]. They speculated that the limited variation that they observed between W3110 and MG1655, despite the two strains having been separated from one another over 50 years prior to their publication, might have been a consequence of using lyophilization as a storage method for these strains [28, 36].

Similarly, a recent study by Desroches *et al.* investigated the provenance and evolution of NCTC 86, the Escherich type strain of *E. coli* [37]. In this work, they sequenced cultures of NCTC 86 from stocks at NCTC, the ATCC, DSMZ and Centre de Ressources Biologiques de l'Institut Pasteur, as well as using two additional previously published sequences from NCTC stocks [38, 39] (a third sequence of this isolate was reported in [40] but not included in this analysis). The authors found striking heterogeneity in the genomes of these supposedly identical *E. coli* cultures, ranging from differences in plasmid replicon sequences being present in various stocks, to single-nucleotide polymorphisms and inactivating mutations in mutation-prevention pathways [37]. The authors were able to correlate some of these differences to the records of how NCTC 86 was shared between these culture collections, and from where the sequenced isolates were obtained [37]. A similar study was performed in *Campylobacter jejuni*, where 23 stocks of the laboratory strain NCTC 11168 from across the UK were both sequenced and phenotyped [41]. These *C. jejuni* stocks varied in their growth rates, motility, virulence, and the types and numbers of mutations in their genome, despite ostensibly being the same strain.

Just as certain pathways seem to be commonly mutated in *V. cholerae* laboratory strains (e.g. quorum-sensing pathways), there are mutations common to lab strains of other species. For instance, mutations in *rpoS* are found in the W3110 and NCTC 86 strains discussed above [28, 37]. *rpoS* mutations are known to occur during the process of laboratory adaptation [42, 43], or as a consequence of transferring bacterial strains between laboratories [44]. It is important to note, however, that the increased fitness of *toxR* mutant *V. cholerae* relative to wild-type cells is *rpoS*-independent

[23]. Mutations in *rpoS* can confer a growth advantage to *E. coli* in a laboratory (e.g. grown at 37 °C in Luria–Bertani or minimal media) [45], and contribute to virulence attenuation associated with laboratory strains, such as in the LT2 strain of *Salmonella enterica* serovar Typhimurium [46] (Fig. 2). The gain of fitness conferred by these laboratory-acquired mutations confers a fitness advantage on these mutants relative to the wild-type, and appears to be a theme common to the discussion above. Whether mutations are acquired in quorum sensing [24], virulence regulons [23] or in *rpoS* [42–44], there appears to be a convergence on laboratory culture environments selecting for mutants that have a selective fitness advantage over the wild-type (Fig. 2), just as occurs in *in vitro* artificial evolution experiments [47, 48].

The discussion above illustrates the fact that phenotypic differences between laboratories' strains offer an interesting opportunity to dissect the scientific basis underpinning those differences. This necessitates that researchers are willing to share their isolates with one another, or that they deposit important isolates in culture collections. The use of appropriate preservation and meticulous record curation means that a fellow researcher who orders a bacterial stock reported in a paper, from a culture collection, can be more confident that the organism they receive is that which produced the phenotypes in a previous publication [49]. Moreover, as the genome sequences of strain collections become freely available, the research community will benefit from the availability of biological and bioinformatic resources in which high levels of confidence can be had.

Simply by virtue of disseminating strains to different laboratories, researchers may have unwittingly conducted a natural evolution experiment. Comparing lab strains across different groups could present interesting opportunities to study evolution, or to resolve differences that are observed, such as explaining the presence of C6706 *luxO* mutants in laboratories' strain collections. A fascinating exercise, for instance, might be to attempt to retrieve stocks of NCTC 30 from laboratories worldwide, which might contain their own stocks, and to perform a comparison both phenotypically and genomically of these stocks. If data on the provenance of each isolate was available and could be used to construct a pedigree of how and when NCTC 30 was disseminated to each laboratory, this could be compared to the phylogenetics of the isolate. This might allow for identifying when phenotypes changed and mutations were accumulated over the course of the strain's propagation. Although *V. cholerae* NCTC 30 has inspired this example, and we have discussed *Vibrio cholerae* and *Escherichia coli* predominantly in this paper, precisely the same 'community evolution' experiment is likely to be ongoing in laboratories worldwide. Bachmann [27] lamented in 1996 that 'the derivation of many strains being isolated today can never be traced because of the failure of some laboratories to maintain adequate records of strain constructions'. Now, with the ever-decreasing cost of bacterial genome sequencing, phylogenetic analysis of

sequenced isolates with even partial records of their provenance may enable the recapitulation of pedigrees, and in the absence of records of strain exchange but with the genome sequences of the strains, perhaps to work out ‘who gave what to who’ using a phylogeny, just as in studies of bacterial transmission (e.g. [50]).

Funding information

This work was supported by Wellcome (grant 206194). M.J.D. is supported by a Wellcome Sanger Institute PhD Studentship.

Acknowledgements

We thank our collaborators at NCTC and the members of the Thomson lab for helpful discussions.

Author contributions

M.J.D. and N.R.T. wrote and edited the manuscript.

Conflicts of interest

The authors declare that there are no conflicts of interest.

References

- Lapage SP, Sneath PHA, Lessel EF, Skerman VBD, Seeliger HPR *et al.* *International Code of Nomenclature of Bacteria: Bacteriological Code*. Washington, DC: ASM Press; 1990 Revision.
- Kyrpides NC, Hugenholtz P, Eisen JA, Woyke T, Göker M *et al.* Genomic encyclopedia of bacteria and archaea: sequencing a myriad of type strains. *PLoS Biol* 2014;12:e1001920.
- Hobman JL, Penn CW, Pallen MJ. Laboratory strains of *Escherichia coli*: model citizens or deceitful delinquents growing old disgracefully? *Mol Microbiol* 2007;64:881–885.
- Dorman MJ, Kane L, Domman D, Turnbull JD, Cormie C *et al.* The history, genome and biology of NCTC 30: a non-pandemic *Vibrio cholerae* isolate from World War One. *Proc Biol Sci* 2019;286:20182025.
- Gardner AD, Venkatraman KV. The antigens of the cholera group of vibrios. *J Hyg* 1935;35:262–282.
- Colwell RR. Polyphasic taxonomy of the genus *Vibrio*: numerical taxonomy of *Vibrio cholerae*, *Vibrio parahaemolyticus*, and related *Vibrio* species. *J Bacteriol* 1970;104:410–433.
- Davis GH, Park RW. A taxonomic study of certain bacteria currently classified as *Vibrio* species. *J Gen Microbiol* 1962;27:101–119.
- Heidelberg JF, Eisen JA, Nelson WC, Clayton RA, Gwinn ML *et al.* DNA sequence of both chromosomes of the cholera pathogen *Vibrio cholerae*. *Nature* 2000;406:477–483.
- Val M-E, Marbouty M, de Lemos Martins F, Kennedy SP, Kemble H *et al.* A checkpoint control orchestrates the replication of the two chromosomes of *Vibrio cholerae*. *Sci Adv* 2016;2:e1501914.
- Matthey N, Drebes Dörr NC, Blokesch M. Long-read-based genome sequences of pandemic and environmental *Vibrio cholerae* strains. *Microbiol Resour Announc* 2018;7.
- Kemter FS, Messerschmidt SJ, Schallopp N, Sobetzko P, Lang E *et al.* Synchronous termination of replication of the two chromosomes is an evolutionary selected feature in *Vibrionaceae*. *PLoS Genet* 2018;14:e1007251.
- Allué-Guardia A, Echazarreta M, Koenig SSK, Klose KE, Eppinger M. Closed genome sequence of *Vibrio cholerae* O1 El Tor Inaba strain A1552. *Genome Announc* 2018;6:e00098–18.
- Stutzmann S, Blokesch M. Circulation of a quorum-sensing-impaired variant of *Vibrio cholerae* strain C6706 masks important phenotypes. *mSphere* 2016;1.
- Meibom KL, Blokesch M, Dolganov NA, Wu C-Y, Schoolnik GK. Chitin induces natural competence in *Vibrio cholerae*. *Science* 2005;310:1824–1827.
- Blokesch M. A quorum sensing-mediated switch contributes to natural transformation of *Vibrio cholerae*. *Mob Genet Elements* 2012;2:224–227.
- Marvig RL, Blokesch M. Natural transformation of *Vibrio cholerae* as a tool—optimizing the procedure. *BMC Microbiol* 2010;10:155.
- Lo Scudato M, Blokesch M. The regulatory network of natural competence and transformation of *Vibrio cholerae*. *PLoS Genet* 2012;8:e1002778.
- Lo Scudato M, Blokesch M. A transcriptional regulator linking quorum sensing and chitin induction to render *Vibrio cholerae* naturally transformable. *Nucleic Acids Res* 2013;41:3644–3658.
- Suckow G, Seitz P, Blokesch M. Quorum sensing contributes to natural transformation of *Vibrio cholerae* in a species-specific manner. *J Bacteriol* 2011;193:4914–4924.
- Blokesch M, Schoolnik GK. The extracellular nuclease Dns and its role in natural transformation of *Vibrio cholerae*. *J Bacteriol* 2008;190:7232–7240.
- Zhu J, Miller MB, Vance RE, Dziejman M, Bassler BL *et al.* Quorum-sensing regulators control virulence gene expression in *Vibrio cholerae*. *Proc Natl Acad Sci U S A* 2002;99:3129–3134.
- Joelsson A, Liu Z, Zhu J. Genetic and phenotypic diversity of quorum-sensing systems in clinical and environmental isolates of *Vibrio cholerae*. *Infect Immun* 2006;74:1141–1147.
- Paul K, Ghosh A, Sengupta N, Chowdhury R. Competitive growth advantage of nontoxic mutants in the stationary phase in archival cultures of pathogenic *Vibrio cholerae* strains. *Infect Immun* 2004;72:5478–5482.
- Kimbrough JH, Stabb EV. Antisocial *luxO* Mutants provide a stationary-phase survival advantage in *Vibrio fischeri* ES114. *J Bacteriol* 2016;198:673–687.
- Jung SA, Chapman CA, Ng W-L. Quadruple quorum-sensing inputs control *Vibrio cholerae* virulence and maintain system robustness. *PLoS Pathog* 2015;11:e1004837.
- Kovacikova G, Skorupski K. Regulation of virulence gene expression in *Vibrio cholerae* by quorum sensing: HapR functions at the *aphA* promoter. *Mol Microbiol* 2002;46:1135–1147.
- Bachmann BJ. Derivations and genotypes of some mutant derivatives of *Escherichia coli* K-12. In: *Escherichia coli and Salmonella typhimurium Cellular and Molecular Biology*. ASM Press 1996.
- Hayashi K, Morooka N, Yamamoto Y, Fujita K, Isono K *et al.* Highly accurate genome sequences of *Escherichia coli* K-12 strains MG1655 and W3110. *Mol Syst Biol* 2006;2:2006.0007.
- Baba T, Ara T, Hasegawa M, Takai Y, Okumura Y *et al.* Construction of *Escherichia coli* K-12 in-frame, single-gene knockout mutants: the Keio collection. *Mol Syst Biol* 2006;2:2006.0008.
- Datsenko KA, Wanner BL. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc Natl Acad Sci U S A* 2000;97:6640–6645.
- Guyer MS, Reed RR, Steitz JA, Low KB. Identification of a sex-factor-affinity site in *E. coli* as gamma delta. *Cold Spring Harb Symp Quant Biol* 1981;45 Pt 1:135–140.
- Bachmann BJ. Pedigrees of some mutant strains of *Escherichia coli* K-12. *Bacteriol Rev* 1972;36:525–557.
- Grenier F, Matteau D, Baby V, Rodrigue S. Complete genome sequence of *Escherichia coli* BW25113. *Genome Announc* 2014;2:e01038-14.
- Berlyn MB, Letovsky S. Genome-related datasets within the *E. coli* genetic stock center database. *Nucleic Acids Res* 1992;20:6143–6151.
- Faure D, Frederick R, Włoch D, Portier P, Blot M *et al.* Genomic changes arising in long-term stab cultures of *Escherichia coli*. *J Bacteriol* 2004;186:6437–6442.
- Barratt RW, Tatum EL. A simplified method of lyophilizing microorganisms. *Science* 1950;112:122–123.
- Desroches M, Royer G, Roche D, Mercier-Darty M, Vallenet D *et al.* The odyssey of the ancestral *Escherichia coli* strain through culture collections: an example of allopatric diversification. *mSphere* 2018;3:e00553–17.

38. Dunne KA, Chaudhuri RR, Rossiter AE, Beriotto I, Browning DF *et al*. Sequencing a piece of history: complete genome sequence of the original *Escherichia coli* strain. *Microb Genom* 2017;3:mgen000106.
39. Méric G, Hitchings MD, Pascoe B, Sheppard SK. From Escherich to the *Escherichia coli* genome. *Lancet Infect Dis* 2016;16:634–636.
40. Khetrupal V, Mehershahi KS, Chen SL. Complete genome sequence of the original *Escherichia coli* isolate, strain NCTC86. *Genome Announc* 2017;5:e00243–17.
41. Pascoe B, Williams LK, Calland JK, Meric G, Hitchings MD *et al*. Domestication of *Campylobacter jejuni* NCTC 11168. *Microb Genom* 2019;5.
42. Liu B, Eydallin G, Maharjan RP, Feng L, Wang L *et al*. Natural *Escherichia coli* isolates rapidly acquire genetic changes upon laboratory domestication. *Microbiology* 2017;163:22–30.
43. Bleibtreu A, Clermont O, Darlu P, Glodt J, Branger C *et al*. The *rpoS* gene is predominantly inactivated during laboratory storage and undergoes source-sink evolution in *Escherichia coli* species. *J Bacteriol* 2014;196:4276–4284.
44. Spira B, de Almeida Toledo R, Maharjan RP, Ferenci T. The uncertain consequences of transferring bacterial strains between laboratories - *rpoS* instability as an example. *BMC Microbiol* 2011;11:248.
45. Zambrano MM, Siegele DA, Almirón M, Tormo A, Kolter R. Microbial competition: *Escherichia coli* mutants that take over stationary phase cultures. *Science* 1993;259:1757–1760.
46. Wilmes-Riesenberg MR, Foster JW, Curtiss R. An altered *rpoS* allele contributes to the avirulence of *Salmonella typhimurium* LT2. *Infect Immun* 1997;65:203–210.
47. Blount ZD, Borland CZ, Lenski RE. Historical contingency and the evolution of a key innovation in an experimental population of *Escherichia coli*. *Proc Natl Acad Sci U S A* 2008;105:7899–7906.
48. Good BH, McDonald MJ, Barrick JE, Lenski RE, Desai MM. The dynamics of molecular evolution over 60,000 generations. *Nature* 2017;551:45–50.
49. Boundy-Mills K, Hess M, Bennett AR, Ryan M, Kang S *et al*. The United States culture collection network (USCCN): enhancing microbial genomics research through living microbe culture collections. *Appl Environ Microbiol* 2015;81:5671–5674.
50. Coll F, Harrison EM, Toleman MS, Reuter S, Raven KE *et al*. Longitudinal genomic surveillance of MRSA in the UK reveals transmission patterns in hospitals and the community. *Sci Transl Med* 2017;9:eaak9745.

Five reasons to publish your next article with a Microbiology Society journal

1. The Microbiology Society is a not-for-profit organization.
2. We offer fast and rigorous peer review – average time to first decision is 4–6 weeks.
3. Our journals have a global readership with subscriptions held in research institutions around the world.
4. 80% of our authors rate our submission process as 'excellent' or 'very good'.
5. Your article will be published on an interactive journal platform with advanced metrics.

Find out more and submit your article at microbiologyresearch.org.