Inflammatory myofibroblastic tumor

Sangeeta Palaskar, Supriya Koshti, Mahesh Maralingannavar, Anirudha Bartake

Abstract

Inflammatory myofibroblastic tumor is an uncommon lesion of unknown cause. It encompasses a spectrum of myofibroblastic proliferation along with varying amount of inflammatory infiltrate. A number of terms have been applied to the lesion, namely, inflammatory pseudotumor, fibrous xanthoma, plasma cell granuloma, pseudosarcoma, lymphoid hamartoma, myxoid hamartoma, inflammatory myofibrohisticcytic proliferation, benign myofibroblatoma, and most recently, inflammatory myofibroblastic tumor. The diverse nomenclature is mostly descriptive and reflects the uncertainty regarding true biologic nature of these lesions. Recently, the concept of this lesion being reactive has been challenged based on the clinical demonstration of recurrences and metastasis and cytogenetic evidence of acquired clonal chromosomal abnormalities. We hereby report a case of inflammatory pseudotumor and review its inflammatory versus neoplastic behavior.

Keywords: ALK-1, myofibroblasts, neoplastic, reactive

Introduction

Inflammatory myofibroblastic tumor (IMT) is an uncommon lesion of unknown cause. It encompasses a spectrum of myofibroblastic proliferation along with varying amount of inflammatory infiltrate. A number of terms have been applied to the lesion, namely, inflammatory pseudotumor, fibrous xanthoma, plasma cell granuloma, pseudosarcoma, lymphoid hamartoma, myxoid hamartoma, inflammatory myofibrohistiocytic proliferation, benign myofibroblatoma, and most recently, inflammatory myofibroblastic tumor. The diverse nomenclature is mostly descriptive and reflects the uncertainty regarding its true biologic nature of these lesions.^[1]

IMT was first observed in lungs and described by Bunn in 1939. It was named as IMT by Umiker *et al.* because it mimics malignant neoplasm clinically, radiologically and histopathologically. Various pathogenetic backgrounds have been proposed as initiating factors such as reactive, infections, autoimmune and neoplastic processes, but the etiology of most remains unknown. Recently, the concept of this lesion being reactive has been challenged based on

Department of Oral Pathology, Sinhgad Dental College and Hospital, Pune, Maharashtra, India

Correspondence: Dr. Sangeeta Palaskar, Department of Oral Pathology, Sinhgad Dental College and Hospital, Pune – 411 041, India. E-mail: palaskarsangeeta@gmail.com

Access this article online	
Quick Response Code:	
	Website: www.contempclindent.org
	DOI: 10.4103/0976-237X.91787

the clinical demonstration of recurrences and metastasis and cytogenetic evidence of acquired clonal chromosomal abnormalities.^[1,2] The lungs, liver and gastrointestinal tract (GIT) are the most common sites for IMT. In the head and neck, it has been reported in epiglottis, endolarynx, parapharyngeal spaces, maxillary sinus, submandibular region and oral cavity.^[3]

IMT in the maxillofacial region is exceptionally rare, and it is often mistaken as malignancy. The diagnosis is still difficult and based on the histologic examination of the lesions.^[4] In the oral cavity, IMTs have been reported in multiple locations like gingiva, tongue, hard palate, mandible, buccal mucosa and submandibular salivary gland.^[5,6]

Clinically, they are painless, indurated mass or swelling of relatively short duration or following specific symptoms related to the site of origin. They have no age preferences, but the affected patients tend to be children and young adults.^[7]

Computed tomography (CT) scan and magnetic resonance imaging (MRI) of IMTs in the head and neck region might be nonspecific and often suggest infiltrative growth, aggressive malignant lesion or granulomatous disease.^[7]

IMTs of head and neck are generally benign lesions and usually cured by radical excision, steroids, irradiation and/or chemotherapy. CO2 laser is a new modality of treatment.^[7]

Case Report

A 19-year-old male patient reported to Sinhgad Dental College and Hospital (SDCH) with a complaint of swelling over right side of face since 3 months. On examination, diffuse extraoral swelling was found on right side of the face [Figure 1]. The swelling was firm on palpation, overlying skin was normal and it was mildly tender in nature. Intraoral examination revealed an oval mobile swelling extending from 44 region to the angle of the mandible in the buccal vestibule, which was nontender and not fixed, and the overlying mucosa was normal. Incisional biopsy was taken and the H and E stained section showed non-encapsulated mass composed of connective tissue stroma with diffuse and focal infiltration of lymphocytes, plasma cells and histiocytes. Focal areas of inflammatory cells were seen gathered around blood vessels and also forming germinal centers. Spindle-shaped cells were seen interspersed in the connective tissue [Figure 2a and b]. Based on the findings, a diagnosis of IMT was rendered. The lesion was surgically excised and subjected to immunohistochemical staining which showed strong vimentin positivity [Figure 3a and b], smooth muscle actin (SMA) positivity in spindle cells [Figure 3c] and focal positivity for Anaplastic Lymphoma Kinase ALK-1 [Figure 3d] in the spindle cells. The diagnosis of IMT (lymphoplasmacytic variant) was confirmed.

Discussion

Lymphocytes, plasma cells, histiocytes, fibroblasts and myofibroblasts are the basic components of IMT, present in variable proportions. Four basic histologic patterns are commonly seen as follows:

- a. Dominant lymphoplasmacytic infiltrate;
- b. Dominant lymphohistiocytic infiltrate;
- c. Young and active myofibroblastic process and
- d. Predominantly collagenized process with lymphocytic infiltrate.



Figure 1: Pre-operative photograph showing diffuse extraoral swelling

The lymphoplasmacytic IMT consists of a mature lymphoid and plasma cell infiltrate with germinal centers, hence the name given is plasma cell granuloma.^[3]

Lymphohistiocytic IMT most commonly resembles an infectious process as foamy histiocytes are predominant.^[3]

The young and active IMT has a densely cellular fascicular and storiform pattern resembling fibrous histiocytoma except for the inflammatory infiltrate or nodular fascitis.^[3]

Collagenized IMT is less cellular and resembles a desmoid tumor but with a prominent inflammatory infiltrate. A zonation/maturation effect may be observed. Progression of patterns may also be seen in some longstanding cases, necessitating multiple procedures.^[3]

Recently, the concept of IMT being a benign reactive lesion has been challenged owing to clinical demonstration of recurrences as high as 37%, the presence of regional metastases and cytogenetic evidence of acquired clonal chromosomal abnormality. However, the issue of reactive versus neoplastic pathogenesis of this lesion remains unsolved.^[7]

There are cases of IMT that clinically and radiologically

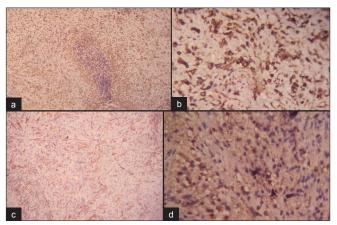


Figure 3: (a) Focal ALK-1 positivity seen in spindle cells (\times 100); (b) SMA positivity seen in spindle cells (\times 100); (c) vimentin positivity in spindle cells (\times 100); (d) vimentin positivity seen in spindle cells (\times 400)

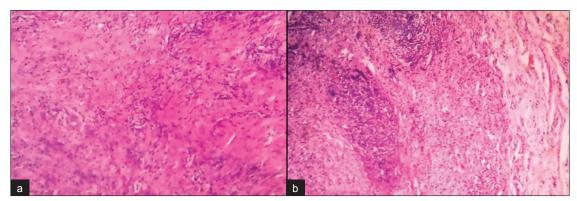


Figure 2: (a) Spindle cell proliferation (×100); (b) inflammatory cells forming germinal centers (×100)



Figure 4: Postoperative photograph

may simulate malignant tumors, but the bland histologic appearance of proliferating spindle cell and plasma cell rich inflammatory background is characteristic. Cellularity, mitotic counts and extent of inflammation do not appear to be prognostic. In contrast, some authors state that at least a subset of IMTs represent true neoplasm and found that cytological atypias, presence of ganglion like cells, p53 expression and DNA aneuploidy may be useful for identifying tumors that are more likely to pursue an aggressive clinical behavior with recurrences and malignant transformation.^[7]

The neoplastic behavior is reflected by some cytogenetic and immunofluoroscence aberrations, especially chromosome 2p23 involving the Anaplastic Lymphoma Tyrosine Kinase (ALK) receptor and its fusion with clathrin heavy chains.^[8] This abnormality has been detected in up to 50% of soft tissue IMTs. Other abnormalities in t(2, 17), (p23, q23), tropomyosin 4 (TPM 4), TPM 3, t(p25, p23), cysteinyl tRNA synthetase and Ran binding protein also have been identified in IMTs. It appears that ALK-1 expression is highly specific for IMT, but it is not 100% sensitive, depending to some extent on the site of origin. ALK-1 negative IMTs are morphologically indistinguishable from ALK-1 positive cases. No clinical, morphological or prognostic difference is found associated with ALK-1 status of the IMT. Nodular fascitis and desmoid fibromatoses do not express ALK-1.^[7,9]

Cytogenetic and molecular studies point to the possibility that some subsets of IMTs are in fact true monoclonal neoplasms, and are also invasive and locally recurrent. Metastatic forms of abdominal, mediastinal and paranasal sinus IMTs have also been reported. In contrast to the latter, there are others who believe that the reported cases are multicentric IMTs and not a true metastasis.^[9]

It is suggested that inflammatory mediators such as cytokines and interleukin-1 (IL-1) are released in response to an insult causing proliferation of fibroblasts, leaky and pro-coagulant endothelium and extravasation of polymorphous cellular infiltrate into the extracellular spaces.^[4] Several triggers for its development have been described, including smoking,

Contemporary Clinical Dentistry | Oct-Dec 2011 | Vol 2| Issue 4

minor trauma, post-adenoidectomy and infection; however, the exact etiology is not known [Figure 4].^[1,3]

Some cases have been reported as having bacterial etiology and Epstein–Barr virus (EBV). Few large pathological series exist, but there is no apparent age, ethnic or geographic predilection.^[1,2] The list of infectious agents is expanding, such as *Mycobacterium*, actinomycetes, *Nocardia*, mycoplasm, *Escherichia coli*, *Klebsiella*, *Bacillus sphaericus*, *Pseudomonas*, HIV, *Helicobacter pyroli*, EBV, human herpes virus 8 (HHV-8).^[7,9,10]

The D/D of IMT comprises low-grade myofibroblastic sarcoma as well as a long list of benign, reactive or neoplastic spindle cell lesions, such as leiomyoma, solitary fibrous tumor, spindle cell carcinoma, nodular fascitis and peripheral nerve sheath tumor.^[2]

Mitotic figures and plump spindle cells are present, but nuclear pleomorphism and apoptosis are not usually seen in IMT. Their presence should lead one to consider an inflammatory sarcomatous process or spindle cell carcinoma. IMT may overlap, in terms of biologic potential and histology, with the entity described as a "low-grade inflammatory fibrosarcoma." The plasmacytic and histiocytic component of IMT may raise the possibility of infections such as syphilis and atypical mycobacterium. In such cases, special stains (acid fast bacillus, silver stain) are warranted. Autoimmune diseases (lupus erythematosus, rheumatoid arthritis) can cause intense laryngeal inflammation, and hence appear as an inflammatory pseudotumor.^[2]

The diagnosis of nodular fascitis, fibrous histiocytoma and fibromatosis can come to the mind when examining IMT. Perhaps IMT is also related to these processes, but features like storiform pattern, lack of necrosis, and pronounced chronic inflammatory component are inconsistent with those processes. Overlapping histologic features between IMT, nodular fascitis and fibrous histiocytoma corroborate the place of these entities in the pathologic spectrum between reactive and neoplastic process.^[2]

In IMTs, spindle cells lack the cytologic atypia and nuclear hyperchromasia of sarcomas. The immunohistochemistry is usually utilized to confirm the myofibroblastic phenotype of the tumor spindle cells which are typically reactive to vimentin (99%), SMA (92%), muscle-specific actin (89%), and desmin (69%). Spindle cells may be focally positive to epithelial markers such as cytokeratin, epithelial membrane antigen (EMA; 36%) and CD68 (25%). IMTs are typically negative for myoglobin and S100 protein. Only few splenic and liver IMTs, particularly those in immunosuppressed patients, were found to be positive to EBV-latent membrane protein (LMP) and HHV-8.^[7,9] IL-6 promotes proliferation of fibroblasts, and both IL-1 and IL-6 promote differentiation of B cells. Major cellular sources of IL-1 and IL-6 are monocytes and macrophages which are constant constituents of IMT.^[11] It is quite common for patients with IMT to undergo multiple biopsy procedures to establish a diagnosis. On biopsy evaluation, IMT appears to be a diagnosis of exclusion. The polymorphous appearance of IMT reflects its variable etiology or its shifting histology during the course of the disease.^[7,12]

In general, IMT follows benign course with favorable outcome after radical local excision. However, invasive, locally recurrent and metastatic forms have been reported in abdomen, mediastinum and paranasal sinuses..^[7,12]

Conclusion

In our case, there was a strong positivity of vimentin and SMA in the spindle-shaped cells, while a focal faint positivity of ALK-1 was seen. Moreover, the absence of cytological atypia and nuclear hyperchromasia in the spindle-shaped cells prompted the diagnosis of IMT (lymphoplasmacytic variant). The lesion was surgically excised and there was no sign of recurrence after 6 months of follow-up.

References

- Poh CF, Priddy RW, Dahlman DM. Intramandibular inflammatory myofibroblastic tumour: A true neoplasm or reactive lesion? Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2005;100:460-6.
- Volker HU, Scheich M, Holler S, Strobel P, Hagen R, Hermenlink HK, et al. Differential diagnosis of laryngeal spindle cell carcinoma and inflammatory myofibroblastic tumour: Report of two cases with similar morphology. Diagn Pathol 2007;2:1-7.
- 3. Margaret S, Silloo BK, Gnepp DR. Nonsquamous pathology of the larynx, hypopharynx, and trachea. In: Gnepp DR, editor. Diagnostic

surgical pathology of the head and neck. 4th ed. New York: W.B. Saunders Company; 2001. p. 287-8.

- Oh JM, Yim JH, Joon BW, Choi BJ, Lee DW, Kwon YD. Inflammatory pseudotumour in the mandible. J Craniofac Surg 2008;19:1552-3.
- Kujima M, Nakamura S, Itoh H, Suchi T Masawa N. Inflammatory pseudoumour of the submandibular gland: Report of a case presenting with autoimmune disease like manifestation. Arch Pathol Lab Med 2001;125:1095-7.
- Montgomery E, Speight PM, Fisher C. Myofibromas presenting in the oral cavity: A series of 9 cases. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2000;89:343-8.
- Al-Sindi KA, Al-Shehabi MH, Al-Khalifa SA. Inflammatory myofibroblastic tumour of paranasal sinuses. Saudi Med J 2007;28:623-7.
- Nikitakis NG, Brooks JK, Frankel BF, Papadimitriou JC, Sauk JJ. Inflammatory myofibroblastic tumour of oral cavity: Review of literature and presentation of an ALK positive case. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2004;98:197-8.
- Coffin CM, Patel A, Perkins S, Elentioba-Jhonson, Perlaman E, Griffin CA. ALK -1 and p80 expression and chromosomal rearrangements involving 2p23 in IMT. Mod Pathol 2001;14:569-76.
- 10. Gomez-Roman JJ. Human herpes virus 8 genes are expressed in pulmonary IMT. Am J Surg Pathol 2001;25:624-9.
- Coffin CM, Watterson J, Priest JR, Dehener LP. Extrapulmonary inflammatory myofibroblastic tumour (inflammatory pseudotumour): A clinicopathologic and immunohistochemical study of 84 cases. Am J Surg Pathol 1995;19:859-72.
- 12. Deshingkar SA, Tupkari JV, Barpande SR. Inflammatory myofibroblastic tumour of the maxilla. J Oral Maxillofac Pathol 2007;11:76-9.

How to cite this article: Palaskar S, Koshti S, Maralingannavar M, Bartake A. Inflammatory myofibroblastic tumor. Contemp Clin Dent 2011;2:274-7.

Source of Support: Nil. Conflict of Interest: None declared.

Author Help: Online submission of the manuscripts

Articles can be submitted online from http://www.journalonweb.com. For online submission, the articles should be prepared in two files (first page file and article file). Images should be submitted separately.

1) First Page File:

Prepare the title page, covering letter, acknowledgement etc. using a word processor program. All information related to your identity should be included here. Use text/rtf/doc/pdf files. Do not zip the files.

2) Article File:

The main text of the article, beginning with the Abstract to References (including tables) should be in this file. Do not include any information (such as acknowledgement, your names in page headers etc.) in this file. Use text/rtf/doc/pdf files. Do not zip the files. Limit the file size to 1024 kb. Do not incorporate images in the file. If file size is large, graphs can be submitted separately as images, without their being incorporated in the article file. This will reduce the size of the file.

3) Images:

Submit good quality color images. Each image should be less than **4096 kb (4 MB)** in size. The size of the image can be reduced by decreasing the actual height and width of the images (keep up to about 6 inches and up to about 1800 x 1200 pixels). JPEG is the most suitable file format. The image quality should be good enough to judge the scientific value of the image. For the purpose of printing, always retain a good quality, high resolution image. This high resolution image should be sent to the editorial office at the time of sending a revised article.

4) Legends:

Legends for the figures/images should be included at the end of the article file.