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Substance P and calcitonin gene-related peptide expression in dorsal root ganglia in sciatic nerve injury rats

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Research Highlights

(1) Substance P and calcitonin gene-related peptide expression in dorsal root ganglia were altered over time in rats with sciatic nerve injury.

(2) Results showed that the expression of the neuropeptides, substance P and calcitonin gene-related peptide, mainly increased in the early stages after sciatic nerve injury. These neuropeptides may possibly serve as an index for evaluating early peripheral nerve injury.

(3) Substance P and calcitonin gene-related peptide are involved in transmitting pain signals and play a role in repair following sciatic nerve injury.

Abstract

The neuropeptides, substance P and calcitonin gene-related peptide, have been shown to be involved in pain transmission and repair of sciatic nerve injury. A model of sciatic nerve defect was prepared by dissecting the sciatic nerve at the middle, left femur in female Sprague Dawley rats. The two ends of the nerve were encased in a silica gel tube. L₅ dorsal root ganglia were harvested 7, 14 and 28 days post sciatic nerve injury for immunohistochemical staining. Results showed that substance P and citonin gene-related peptide expression increased significantly in dorsal root ganglion of rats with sciatic nerve injury. This increase peaked at 7 days, declined at 14 days, and reduced to normal levels by 28 days post injury. The findings indicate that the neuropeptides, substance P and calcitonin gene-related peptide, mainly increased in the early stages after sciatic nerve injury.

Key Words

neural regeneration; peripheral nerve injury; sciatic nerve; dorsal root ganglion; spinal cord; neuropeptides; calcitonin gene-related peptide; substance P; pain; neuroprotection; grants-supported paper; neuroregeneration

INTRODUCTION

Several methods have been utilized to repair peripheral nerve injury^[1-4], but results have not been favorable. Thus, it is important to understand the mechanism of peripheral nerve injury. Neuropeptides are a recently discovered neurotransmitter, with a wide range of bioac-tivities^[2]. A large number of clinical and experimental observations have demonstrated that peripheral nerves participate in the repair of nerve injury by ing various neuropeptides from peripheral nerve endings^[2-4]. In particular, increasing attention has been paid to neu- ropeptide substance P and calcitonin gene- related peptide^[5].

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Author contributions: Fu CM designed the study, evaluated the experiments, wrote the manuscript and conducted statistical analyses. Yu DF and Yang ZH collected experimental data and established the injury model. Yin ZS revised the manuscript. All authors approved the final version of the manuscript.

Conflicts of interest: None declared.

Ethical approval: This study received permission from the Ethics Committee of Animal Experimental Center of Anhui Medical University, China. Author statements: The manuscript is original, has not been submitted to or is not under consideration by another publication, has not been previously published in any language or any form, including electronic, and contains no disclosure of confidential information or authorship/patent application/funding source disputations. Substance P and calcitonin gene-related peptide are the main neuropeptides in peripheral nerve ganglia. They can transmit nociception and function as excitatory transmitters in primary sensory neurons. Furthermore, they anterogradely transmit nociceptive information to the central nervous system and are retrogradely released in local tissues, leading to hyperalgesia^[6].

In addition, substance P and calcitonin generelated peptide are local regulators for mitosis of mesenchymal cells, including bone marrow-derived mesenchymal stem cells, and/or functions of nerve cells^[5]. They can regulate microcirculation in bone, and bidirectionally mediate nociceptive transmission and nerve tissues^[6]. Many studies have emphasized the roles of substance P and calcitonin gene-related peptide in nerve injury, but their expression at different time points remains unclear.

The present study established a rat model of sciatic nerve injury and assessed substance P and calcitonin gene-related peptide expression in dorsal root ganglia at different time points.

RESULTS

Quantitative analysis of experimental animals

A total of 40 rats were obtained, with 30 being used to establish a model of sciatic nerve defect by cutting the middle of the femur. Ten model rats were used for the 7, 14, and 28 day time points. The remaining 10 rats were used as normal controls and sampled at 7 and 14 days following model establishment. All 40 rats were included in the final analysis.

Expression of substance P and calcitonin gene-related peptide in L_5 dorsal root

ganglia from rats with sciatic nerve injury Immunohistochemistry revealed substance P-positive products in dorsal root ganglion of normal rats stained lightly yellow or dark yellow, and were mainly distributed in the cytoplasm. After model establishment for 7 days, substance P-positive products were stained brown and seen mainly in intercellular substance. Staining at this time point gradually darkened. At 14 days, the dark yellow substance P-positive products mainly distributed in loose connective tissue. By 28 days, the expression of substance P-positive products was similar to levels seen in normal rats (Figure 1).

Changes in calcitonin gene-related peptide expression were similar to those seen for substance P. More specifically, in normal rats, calcitonin gene-related peptide-positive products stained darkly yellow in dorsal root ganglion. At 7 days after model establishment, positive products stained brown and were mainly distributed intercellularly, at a higher density than substance P-positive products. At 14 days, calcitonin gene-related peptide-positive products were mainly distributed in loose connective tissue and were stained dark yellow or brown. By 28 days, the expression of calcitonin gene-related peptide products was similar to levels seen in normal rats (Figure 2).

Changes in intensity of substance P and calcitonin gene-related peptide expression in L_5 dorsal root ganglia from rats with sciatic nerve injury

Semi-quantitative analysis using a medical image analysis system showed that the intensity of substance P and calcitonin generelated peptide expression remained unchanged in the dorsal root ganglia of normal rats at 7 and 14 days after model establishment. Compared with the normal control substance Ρ and calcitonin group, gene-related peptide expression was significantly increased in the model group at 7 and 14 days (P < 0.05). In addition, the intensity of substance P and calcitonin gene-related peptide expression was significantly higher at 7 days compared with 14 days in the model group (P < 0.05), but no significant difference was found between 7 and 14 days in the normal control group (P > 0.05; Table 1).

Changes in absorbance of substance P and calcitonin gene-related peptide staining in L_5 dorsal root ganglia from rats with sciatic nerve injury Quantitative analysis of immunohistochemistry using the JEDA-801D Morphology Image Analysis System showed that the absorbance of substance P and calcitonin gene-related peptide in dorsal root ganglia reached a peak value in rats with sciatic nerve injury at 7 days post injury, slightly declined at 14 days, and was restored to normal level by 28 days (Figure 3).

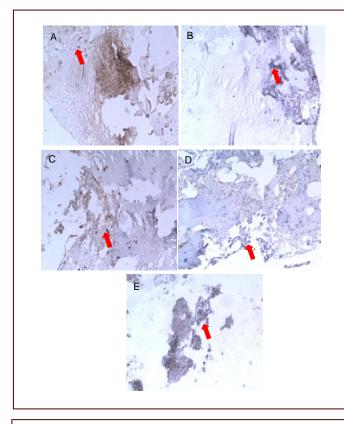


Figure 1 Effects of sciatic nerve defects on substance P expression in L_5 dorsal root ganglia of rats (immunohistochemical staining, streptavidin-biotin complex, x 400).

Light microscopy showed that substance P-positive products (arrows) stained dark yellow or brown, mainly distributing in fibrous tissue.

(A) Substance P-positive products were dense and darkly stained in the dorsal root ganglion of normal rats at 7 days.

No significant difference was found in staining or distribution of substance P-positive products between 14 (B) and 7 days (A) in the dorsal root ganglion of normal rats.

(C) Substance P-positive products stained darker in rats with sciatic nerve injury than in normal rats and expression was significantly increased at 7 days.

Expression of substance P was reduced and staining became lighter in rats with sciatic nerve injury at 14 days (D) compared with 7 days (C), but was still higher than in normal rats.

(E) Expression of substance P in rats with sciatic nerve injury was similar to that in normal rats at 28 days.

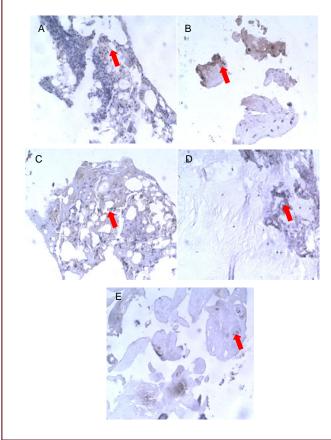


Figure 2 Effects of sciatic nerve defect on calcitonin gene-related peptide expression in L_5 dorsal root ganglia of rats (immunohistochemical staining, streptavidin-biotin complex, \times 400).

Light microscopy showed that calcitonin gene-related peptide-positive products (arrows) stained dark yellow or brown, mainly distributing in fibrous tissue.

(A) Calcitonin gene-related peptide-positive products were dense and darkly stained in the dorsal root ganglion of normal rats at 7 days.

No significant difference was found in staining or distribution of calcitonin gene-related peptide-positive products between 14 (B) and 7 days (A) in the dorsal root ganglion of normal rats.

(C) Calcitonin gene-related peptide-positive products stained darker and significantly increased in rats with sciatic nerve injury than in normal rats at 7 days.

Expression of calcitonin gene-related peptide was reduced and the staining became lighter in rats with sciatic nerve injury at 14 days (D) compared with 7 days (C), but remained higher than in normal rats.

(E) Expression of calcitonin gene-related peptide in rats with sciatic nerve injury was similar to that in normal rats at 28 days.

Table 1 (Comparison of the intensity of substance P and
calcitonin (gene-related peptide expression between groups

Group	Substance P	Calcitonin gene-related peptide	
7 days post injury			
Normal control	1.18±0.32	1.65±0.25	
Model	2.02±0.31 ^a	2.58±0.23 ^a	
14 days post injury			
Normal control	0.89±0.33	1.45±0.18	
Model	1.59±0.11 ^{ab}	2.13±0.25 ^{ab}	
14 days post injury Normal control	0.89±0.33	1.45±0.18	

Results are expressed as mean \pm SD of 10 rats from the model group and five rats from the normal control group at each time point. Four high-power fields (x 400) were randomly selected from each section. ^a*P* < 0.05, *vs.* normal control (two-sample *t*-test); ^b*P* < 0.05, *vs.* 7 days after model establishment (one-way analysis of variance, least significant difference test for paired comparison). Light yellow-stained particles in cytoplasm were identified as positive expression: (+), light yellow, 1 point; (++), dark yellow, 2 points; (+++), brown, 3 points.

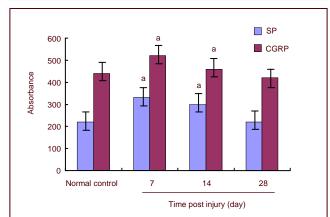


Figure 3 Absorbance of substance P (SP) and calcitonin gene-related peptide (CGRP) in L_5 dorsal root ganglia of rats with sciatic nerve injury.

Data are expressed as mean \pm SD of 10 rats from the normal control and model group at each time point (mean value of all rats at 7 and 14 days in normal control group). Immunohistochemistry results were quantitatively analyzed using the JEDA-801D Morphology Image Analysis System. ^a*P* < 0.05, *vs.* normal control group (two-sample *t*-test).

DISCUSSION

Role of the neuropeptides substance P and calcitonin gene-related peptide in the early stages of peripheral nerve injury

Peripheral nerve injury is axonal injury of nerve cells, and restoration depends on axonal regeneration. Post peripheral nerve injury, Wallerian degeneration occurs in distal neurites, axoplasmic transport is blocked, and local inflammatory reactions result in proliferation of various types of cells to form a tissue barrier. These processes are accompanied by neuronal apoptosis in dorsal root ganglia. Thus, successful regeneration of injured peripheral nerves mainly depends on central motor neuron viability and the microenvironment that allows nerve growth^[7].

After peripheral nerve injury, motor end-plates and sensory corpuscles disappear due to degeneration and denervation, hindering peripheral nerve regeneration^[8]. Recent evidence indicates that motor end-plates and sensory corpuscles can regenerate^[9-10]. However, little is known about neuropeptide changes in this environment. Substance P and calcitonin gene-related peptide are known to mediate nociception^[5]. Alterations in substance P and calcitonin gene-related peptide activity reflect the function of the posterior horn of the spinal cord, therefore neural regeneration post peripheral nerve injury represents restoration of posterior horn function^[11]. If this regeneration does not occur, the function of the posterior horn may not be completely recovered^[12]. The present study assessed the changes in substance P and calcitonin gene-related peptide in dorsal root ganglia of rats with left sciatic nerve defects. Results showed that positive substance P and calcitonin gene-related peptide expression in dorsal root ganglion gradually increased at 7 days post injury. As substance P and calcitonin gene-related peptide can transmit nociceptive information in an anterograde fashion to the central nervous system^[5], increased release of substance P and calcitonin gene-related peptide provides a microenvironment for neural regeneration. Thus, broken nerves can heal well at this time^[6]. In the present study, expression of substance P and calcitonin gene-related peptide was higher at 14 days post sciatic nerve injury compared with the normal control group, consistent with previous results^[13-14]. By 28 days, expression of substance P and calcitonin gene-related peptide gradually decreased and returned to normal level, supporting previous clinical denervation studies^[3]. Thus, we conclude that substance P and calcitonin gene-related peptide play an important role in the early stages of neural regeneration in dorsal root ganglia. The protective effects of substance P and calcitonin gene-related peptide after peripheral nerve injury may be associated with injured neurons, autoregulation in the central nervous system and sprouting of lateral buds^[14-16].

Role of substance P and calcitonin gene-related peptide in the metabolism of the nervous system

Substance P is a member of the tachykinin family, produced from a polyprotein precursor after differential splicing of the preprotachykinin A gene, and consists of 11 amino acids linked by 10 peptide bonds^[16]. Tachykinin receptors include the G-protein-coupled neurokinin-1, neurokinin A (NK2) and neurokinin B (NK3), with neurokinin-1 being the most sensitive to substance P^[16]. When the receptor binds ligand it promotes hydrolysis of phosphinositides, a protein coupling agent, producing inositol 1,4,5-trisphosphate, which increases intracellular Ca2+ concentration, alters membrane potential, and results in neurophysiologic effects^[16]. Substance P has high affinity for the neurokinin-1 receptor, and therefore can increase viability and proliferation of nerve cells. Calcitonin gene-related peptide is extensively distributed in the nervous system, and copied from the same gene as calcitonin. It is located at 11p15.4, is 7.6 kb, and produces calcitonin gene-related peptide mRNA after transcription. Calcitonin gene-related peptide is known as an effective vasodilator, and induces local ischemia/ necrosis of nerve tissues with inflammatory factors. In addition, it plays a role in neural regeneration^[17]. Calcitonin gene-related peptide can bind receptors on nerve cells to promote cyclic adenosine monophosphate formation, insulin-like growth factor mRNA expression, and synthesis of insulin-like growth factor polypeptide to improve nerve cell activity. Calcitonin gene-related peptide can also inhibit differentiation of mouse bone marrow-derived mesenchymal stem cells into nerve cells^[18]. Vasodilation caused by substance P and calcitonin gene-related peptide is mainly based on non-endothelium-dependent vasodilation of vascular smooth muscle^[19-20]. Substance P directly acts at vascular endothelial cells, resulting in rapid and transient vasodilation, in addition to mast cell degranulation, histamine release and inflammatory reaction regulation^[19]. Calcitonin gene- related peptide can promote angiogenesis and increase blood flow to local nerve tissues^[5].

Significance of substance P and calcitonin gene-related peptide expression in dorsal root ganglia of rats with sciatic nerve injury

Although the mechanism surrounding peripheral nerve injury remains uncertain, some conclusions have been formulated^[7]. Gleichmann *et al* ^[21] found that after peripheral nerve lesion a transient increase in stromal cell-derived factor-1beta mRNA expression was observed, indicating that stromal cell-derived factor-1beta may play a role in neural regeneration. An early study established a bilateral sciatic nerve injury model in rats and injected Hoechst-labeled bone marrow stromal stem cells into L₄ dorsal root ganglion^[22]. Results showed that the bone marrow stromal stem cells only migrated to L₃, L₅, and L₆ ganglion that innervated injured sciatic nerves. This observation indicates that stem cells could promote neural regeneration. In another model of peripheral neuropathy, Schwann cell secreted erythropoietin was the most effective endogenous mechanism of preventing axonal degeneration^[23]. Furthermore, erythropoietin can protect cortical neurons from HIV-1/gp120-induced apoptosis^[24]. In the present study, we established a left sciatic nerve model in rats and observed substance P and calcitonin gene-related peptide expression in dorsal root ganglia. Results indicated that substance P and calcitonin gene-related peptide expression significantly increased at 7 and 14 days post injury, demonstrating that these neuropeptides are involved in the metabolism of injured nerves^[16-18]. In addition, substance P and calcitonin gene-related peptide expression reached peak levels at 7 days post injury, gradually declined at 14 days and returned to normal levels by 28 days post injury. This suggests that substance P and calcitonin gene-related peptide are mainly expressed in the early stages after injury, and may be useful as a sensitive index to estimate the early stages of peripheral nerve injury. Furthermore, Substance P and calcitonin gene-related peptide are derived from a sensory neuropeptide. They can protect nerves against further injury and potentiate normal growth or reconstruction of nerves as well as biomechanical alignment^[25-26].

MATERIALS AND METHODS

Design

A randomized, controlled, animal study.

Time and setting

Experiments were performed at the Animal Experimental Center of Anhui Medical University, China from November 2011 to June 2012.

Materials

A total of 40 clean grade, healthy, female, Sprague Dawley rats, aged 4 weeks, weighting 180–230 g, were purchased from the Animal Center of Anhui Medical University [No. SCXK (Wan) 2011-001]. Animal protocols were conducted in accordance with the *Guidance Suggestions for the Care and Use of Laboratory Animals*, issued by the Ministry of Science and Technology of China in 2006^[27].

Methods

Establishment of sciatic nerve defect model

Rats were anesthetized by intraperitoneal injection of 10% chloral hydrate (3 mL/kg), and an oblique incision was made at the posterior femur to expose the left sciatic nerve. The sciatic nerve was dissected out, and cut off at the mid femur. Both broken ends were encased in steri-

lized 10-mm silica gel tubes (Supratec, Attendorn, Germany) to a depth of 1.5 mm to create a 7-mm defect. The distal and proximal ends were anastomosed with the tube using 10-0 microsuture line^[28]. The incision was then sutured closed.

Sampling of L₅ dorsal root ganglion

Samples of dorsal root ganglion were harvested at 7, 14 and 28 days post injury (7 and 14 days for the normal control group). Rats were anesthetized by intraperitoneal injection of 10% chloral hydrate and placed in a supine position. The left sciatic nerve was exposed, and L_5 dorsal root ganglia connected with the vertebral canal were harvested, fixed in 4% paraformaldehyde, dehydrated, cleared, embedded in wax, and sectioned (40 µm) using a RM201644820 histotome (Leica, Solms, Germany).

Immunohistochemical staining

Paraffin sections were baked at 60°C for 2 hours, dewaxed, hydrated, and mixed with freshly prepared 3% H₂O₂ for 10 minutes. Sections were then washed three times with distilled water, and antigens were retrieved using a microwave. Sections were subsequently cooled, washed twice with PBS and incubated with normal goat serum blocking solution at room temperature for 20 minutes. Following this blocking step, sections were incubated with primary antibody (rabbit anti-substance P and calcitonin gene-related peptide polyclonal antibody; 1:200; Abcam, Cambridge, UK) at room temperature for 2 hours, and biotinlabeled mouse anti-rabbit IgG (1:100; Abcam) at room temperature for a further 2 hours. Three washes with PBS were conducted between each step. Sections were then incubated with streptavidin-biotin complex (Abcam) at room temperature for 20 minutes, washed four times with PBS and then visualized with diaminobenzidine (Abcam). Staining was finally observed using a light microscope [Motic B5; Motic (Xiamen) Electric Group Co., Ltd., Xiamen, Fujian Province, China].

Image analysis

Images were loaded into a Medical Image Analysis System (Cambridge Instrument, Cambridge, UK) using Motic B5 microscopic imaging system [Motic (Xiamen) Electric Group Co., Ltd.]. Four high-power fields (x 400) were randomly selected from each section. Immunohistochemical staining results were identified^[29]: (+), light yellow, 1 point; (++), dark yellow, 2 points; (+++), brown, 3 points. Images were analyzed using the JEDA-801D Morphology Image Analysis System (Cambridge Instrument) and mean absorbance was calculated.

Statistical analysis

Data were analyzed using SPSS version 13.0 (SPSS, Chicago, IL, USA) and were expressed as mean \pm SD. Intergroup differences were compared utilizing twosample *t*-test, and intragroup differences at different time points were compared using one-way analysis of variance. Between-group comparison was conducted using least significant difference test. A value of *P* < 0.05 was considered statistically significant.

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