

HHS Public Access

Author manuscript Bone Marrow Transplant. Author manuscript; available in PMC 2019 March 19.

Published in final edited form as:

Bone Marrow Transplant. 2019 March ; 54(3): 494-496. doi:10.1038/s41409-018-0334-y.

Donor-specific anti-HLA antibodies in unrelated hematopoietic cell transplantation for non-malignant disorders

Ann Woolfrey¹, Tao Wang², Stephanie J. Lee¹, Michael D. Haagenson³, Ge Chen⁴, Katharina Fleischhauer⁵, John Horan⁶, Katharine Hsu⁷, Michael Verneris⁸, Stephen R. Spellman³, and Marcelo Fernandez-Vina⁴

¹Fred Hutchinson Cancer Research Center, Seattle, WA, USA

²Center for International Blood and Marrow Transplant Research, Medical College of Wisconsin, Milwaukee, WI, USA

³Center for International Blood and Marrow Transplant Research, Minneapolis, MN, USA

⁴Stanford School of Medicine, Stanford, CA, USA

⁵University Hospital Essen, Institute for Experimental Cellular Therapy, Essen, Germany

⁶Emory University, Atlanta, GA, USA

⁷Memorial Sloan Kettering Cancer Center, New York, NY, USA

⁸University of Colorado-Children's Hospital, Aurora, CO, USA

Keywords

Donor specific antibodies; unrelated donors; hematopoietic cell transplantation; graft rejection; survival

To the Editor:

The role of pre-existing donor-specific anti-human leukocyte antigen antibodies (HLA-DSA) in hematopoietic cell transplantation (HCT) is the subject of much debate, reflecting the increasing feasibility of successful transplantation across HLA mismatch barriers. In our study of patients transplanted for non-malignant disease (NMD), HLA-mismatching increased the risk of graft failure and was 2 to 3 fold higher for the compared to patients with malignancies.¹ After adjustment for other factors, the odds ratio for primary or

Conflict of Interest

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

Corresponding author: Stephen R. Spellman, MBS Center for International Blood and Marrow Transplant Research, 500 5th St N, Minneapolis, MN 55401, Phone: 763-406-8334, sspellma@nmdp.org.

Authorship Contributions

AW, MFV, SJL, DT and SRS designed the study; GC performed HLA-DSA testing; TW and MDH performed statistical analyses; AW, MFV, SJL and SRS interpreted analysis, AW and SJL wrote the paper; TW, MDH, GC, KF, JH, KH, DT, MV, SRS and MFV critically reviewed and revised the paper.

The authors have no conflict of interest to declare.

Woolfrey et al.

secondary graft failure for 7/8 and 6/8 matched pairs was 2.81 (1.74–4.54; p<0.0001) and 2.22 (1.26–3.97; p=0.006), respectively.

From the original study of 663 patients with NMD, we tested 236 patients with pretransplant samples for HLA-DSA by solid phase assays utilizing single antigen bead preparations that included detection of IgG antibodies or by complement fixing antibodies based on the C1q binding assay.^{2,3}. HLA-DSA was evaluated by analyzing the reactivity against the mismatched donor antigens determined by IgG or C1q assays; mean fluorescence intensity (MFI) >1,000 was considered positive, MFI >500 and <1,000 was considered potentially positive, and MFI< 500 was considered negative.

The primary outcome tested in the models was primary graft failure; the secondary outcome was overall survival. Donor engraftment was defined as $>500/\mu$ l neutrophils with >5% donor-derived cells within marrow or peripheral blood cell subsets. The univariate and multivariate probabilities of graft failure and survival were evaluated for different cutoffs defining DSA positive. All variables were tested for the affirmation of the proportional hazards assumption, then stepwise forward selection with a threshold of p<0.05 for entry and exit. Center adjustment assumed random effects. Interactions were tested between the explanatory variables and other significant covariates, and none were significant at p<0.05. To adjust for multiple comparisons, p<0.01 was considered significant.

The median age of tested patients was 9 years old (range <1 to 53). Reduced intensity or nonmyeloablative conditioning was used in 48%, most of the patients were given marrow grafts (82%), and most were given either anti-T cell serotherapy (78% ATG, 2% Campath) and/or a T cell depleted graft (44%). The HLA-DSA-positive (MFI>1000) cohort was similar with respect to age at HCT, race, sex, type of NMD, Karnofsky/Lansky score, and year of HCT, however there was a slightly higher proportion of marrow recipients (95% vs 80%, p=.04) when compared to the HLA-DSA-negative cohort. The C1q positive group did not differ from the C1q negative group for these variables. Table 1a shows the distribution of HLA-DSA.

Table 1b shows the lack of association of HLA-DSA with graft failure and survival. Results were similar when HLA-DSA IgG positive and C1q positive (11.5%) were combined for analysis (data not shown). We then used an MFI>5000 as the cutoff value to define a positive HLA-DSA; however, results remained non-significant for an association with graft failure (data not shown).

Several studies have shown a positive HLA-DSA is a potent barrier to hematopoietic stem cell engraftment.^{4–6} A number of factors might explain why HLA-DSA was not found to contribute independently to the risk for graft failure in patients with NMD in our study. These patients largely received marrow grafts and many received ex vivo T cell depleted grafts, both of which are associated with higher rates of graft rejection compared to recipients of T-replete PBSC.^{7,8} Furthermore, reduced intensity conditioning regimens commonly were used. Except for patients with immune deficiencies, most other patients with NMD have stronger immune systems compared to patients with hematologic malignancies who have been treated with cytotoxic chemotherapy. Together these factors

Bone Marrow Transplant. Author manuscript; available in PMC 2019 March 19.

form a milieu in which alloreactive host T cells persist after transplant and may not be counteracted by sufficient donor alloreactivity, leading to graft rejection. In such a setting, the addition of donor-recipient HLA mismatching would further strengthen host alloreactive recommended donor III A might not

responses. Previous sensitization of the host to mismatched donor HLA might not necessarily increase this already heightened reactivity. Finally, specific HLA-DSAs may have different potency but we lumped all positive tests together for analysis.

An alternative explanation for the findings is a lack of power to detect a significant difference. The number of patients that were available for this re-analysis was small and the number with HLA-DSA smaller, which may have reduced the power to detect an effect of HLA sensitization. Additionally, we were not able to examine some other hypotheses, such as class I vs. class II HLA-DSA, or whether there was an association with prior transfusions or disease categories. Therefore, our results should not be interpreted to mean that HLA-DSA testing is not relevant, simply that we were not able to detect a strong association in our cohort. Thus, until larger analyses confirm these results, efforts to optimize all potential factors that could improve engraftment should continue for patients with NMD.

Acknowledgments

The CIBMTR is supported primarily by Public Health Service Grant/Cooperative Agreement 5U24-CA076518 from the National Cancer Institute (NCI), the National Heart, Lung and Blood Institute (NHLBI) and the National Institute of Allergy and Infectious Diseases (NIAID); a Grant/Cooperative Agreement 5U10HL069294 from NHLBI and NCI; a contract HHSH250201200016C with Health Resources and Services Administration (HRSA/ DHHS); two Grants N00014-15-1-0848 and N00014-16-1-2020 from the Office of Naval Research; and grants from *Actinium Pharmaceuticals, Inc.; Alexion; *Amgen, Inc.; Anonymous donation to the Medical College of Wisconsin; Astellas Pharma US; AstraZeneca; Atara Biotherapeutics, Inc.; Be the Match Foundation; *Bluebird Bio, Inc.; *Bristol Myers Squibb Oncology; *Celgene Corporation; Cellular Dynamics International, Inc.; Cerus Corporation; *Chimerix, Inc.; Fred Hutchinson Cancer Research Center; Gamida Cell Ltd.; Genentech, Inc.; Genzyme Corporation; Gilead Sciences, Inc.; Health Research, Inc. Roswell Park Cancer Institute; HistoGenetics, Inc.; Incyte Corporation; Janssen Scientific Affairs, LLC; *Jazz Pharmaceuticals, Inc.; Jeff Gordon Children's Foundation; The Leukemia & Lymphoma Society; Medac, GmbH; MedImmune; The Medical College of Wisconsin; *Merck & Co, Inc.; *Mesoblast; MesoScale Diagnostics, Inc.; *Miltenyi Biotec, Inc.; National Marrow Donor Program; Neovii Biotech NA, Inc.; Novartis Pharmaceuticals Corporation; Onyx Pharmaceuticals; Optum Healthcare Solutions, Inc.; Otsuka America Pharmaceutical, Inc.; Otsuka Pharmaceutical Co, Ltd. - Japan; PCORI; Perkin Elmer, Inc.; Pfizer, Inc; *Sanofi US; *Seattle Genetics; *Spectrum Pharmaceuticals, Inc.; St. Baldrick's Foundation; *Sunesis Pharmaceuticals, Inc.; Swedish Orphan Biovitrum, Inc.; Takeda Oncology; Telomere Diagnostics, Inc.; University of Minnesota; and *Wellpoint, Inc. The views expressed in this article do not reflect the official policy or position of the National Institute of Health, the Department of the Navy, the Department of Defense, Health Resources and Services Administration (HRSA) or any other agency of the U.S. Government.

References

*Corporate Members

- Horan J, Wang T, Haagenson M, Spellman SR, Dehn J, Eapen M, et al. Evaluation of HLA matching in unrelated hematopoietic stem cell transplantation for nonmalignant disorders. Blood. 2012;120:2918–2924. [PubMed: 22829628]
- Pei R, Lee JH, Shih NJ, Chen M, Terasaki PI. Single human leukocyte antigen flow cytometry beads for accurate identification of human leukocyte antigen antibody specificities. Transplantation. 2003;75:43–9. PubMed PMID: . [PubMed: 12544869]
- Chen G, Sequeira F, Tyan DB. Novel C1q assay reveals a clinically relevant subset of human leukocyte antigen antibodies independent of immunoglobulin G strength on single antigen beads. Hum Immunol. 2011;72:849–58. doi:10.1016/j.humimm.2011.07.001 Epub 2011 Jul 18. PubMed PMID: . [PubMed: 21791230]

Bone Marrow Transplant. Author manuscript; available in PMC 2019 March 19.

- Spellman S, Bray R, Rosen-Bronson S, Haagenson M, Klein J, Flesch S, et al. The detection of donor directed, HLA-specific alloantibodies in recipients of unrelated hematopoietic cell transplantation is predictive of graft failure. Blood. 2010;115:2704–2708. [PubMed: 20089963]
- Ciurea SO, Thall PF, Wang X, Wang SA, Hu Y, Cano P, et al. Donor-specific anti-HLA Abs and graft failure in matched unrelated donor hematopoietic stem cell transplantation. Blood. 2011;118:5957–5964. [PubMed: 21967975]
- Ruggeri A, Rocha V, Masson E, Labopin M, Cunha R, Absi L, et al. Impact of donor-specific anti-HLA antibodies on graft failure and survival after reduced intensity conditioning-unrelated cord blood transplantation. Haematologica. 2013;98:1154–1160. [PubMed: 23242594]
- Anasetti C, Logan BR, Lee SJ, Waller EK, Weisdorf DJ, Wingard JR, et al. Peripheral-blood stem cells versus bone marrow from unrelated donors. New Eng J Med. 2012;367:1487–1496. [PubMed: 23075175]
- Margolis D, Camitta B, Pietryga D, Keever-Taylor C, Baxter-Lowe LA, Pierce K, et al. Unrelated donor bone marrow transplantation to treat severe aplastic anaemia in children and young adults. Brit J Haem. 1996; 94:65–72.

Table 1a.

Incidence and mean fluorescence intensity of positive and potentially positive anti donor HLA-specific antibodies (N=236)

	Positive - N (%)	MFI mean (range)	Potentially positive - N(%)	MFI mean (range)
IgG	10 (4.2%)	6451 (1032–13076)	16 (6.8%)	654 (518–909)
C1q	8 (3.4%)	7686 (1036–19673)	3 (1.3%)	836 (639–966)

Abbreviations: Immunoglobulin G (IgG); mean fluorescence intensity (MFI)

Table 1b.

Results of univariate and multivariate modeling testing the association of donor specific antibodies with various outcomes. Univariate estimates at 1 year, multivariate HR (95%) CI and p-values are shown.

Endpoints	HLA-DSA IgG Positive ¹ / Potentially Positive ² vs. Negative	HLA-DSA IgG Positive vs. Potentially Positive / Negative	C1q Positive ³ / Potentially Positive ⁴ vs. Negative	C1q Positive vs. Potentially Positive / Negative
Graft failure	13% vs. 12%, 0.75 (0.23–	10% vs. 12%, 0.72 (0.10–	18% vs. 12%, 1.42 (0.34–	25% vs 11%, 2.19 (0.52–
	2.47), 0.63	5.28), 0.75	5.95), 0.63	9.17), 0.28
Overall survival	42% vs. 52%, 1.20 (0.70–	30% vs 52%, 1.34 (0.62–	27% vs 52%, 1.40 (0.68–	13% v. 59%, 2.07 (0.94–
	2.05), 0.50	2.88), 0.45	2.88), 0.36	4.56), 0.071

GVHD, graft-versus-host disease; HLA-DSA, donor specific anti-HLA antibody

¹IgG positive HLA-DSA: HLA-A=3, -B=1, -C=1, -DPB1=6 (MFI >1000)

²IgG potentially positive HLA-DSA: HLA-A=1, -B=1, -C=2, -DQB1=1, -DPB1=11 (MFI 500–1000)

³C1q positive HLA-DSA: HLA-A=4, -DPB1=4 (MFI>1000)

⁴C1q potentially positive HLA-DSA: HLA-C=1, -DPB1=2 (MFI 500-1000)