24-Hour Kinetics of Cardiac Troponin-T Using a "High-Sensitivity" Assay in Thoroughbred Chuckwagon Racing Geldings after Race and Associated Clinical Sampling Guidelines

E. Shields, I. Seiden-Long, S. Massie, and R. Leguillette iD

Background: A "high-sensitivity" cardiac troponin-T (hscTnT) assay recently has been validated for use in horses and is a specific biomarker of myocardial damage. Postexercise release kinetics of cTnT utilizing the hscTnT assay have yet to be established in horses.

Objectives: To determine: (1) cTnT release kinetics in racing Thoroughbreds after a high-intensity 5/8th mile Chuckwagon race; (2) the effects of age on pre- and postrace cTnT concentrations; and (3) sampling guidelines for clinicians evaluating horses presenting after exercise.

Animals: Samples were obtained from 38 Thoroughbred geldings aged 5–16 years before racing and immediately, 2, 3, 4, 6, 12, and 24 hour postrace.

Methods: Prospective, observational study with convenience sampling. A fifth-generation hscTnT assay was used for plasma sample analysis, and concentrations were compared at all time-points. Correlations were determined between cTnT concentrations and age. Biochemistry analysis was performed to assess rhabdomyolysis, renal failure, and exercise-induced dehydration.

Results: All horses with measureable cTnT concentrations had significant postexercise increases in cTnT with a median peak (8.0 ng/L) at 3-hour postrace. All horses had peak postexercise cTnT concentrations 2- to 6-hour postrace \leq the 99th percentile upper reference limit of 23.2 ng/L, after which all cTnT concentrations decreased until returning to baseline by 12–24 hours. There was no correlation over time between cTnT concentrations and age.

Conclusions and Clinical Importance: In racing Thoroughbreds completing short-duration, high-intensity Chuckwagon races, cTnT concentrations are expected to be increased 2- to 6-hour postrace and to decrease by 12-24 hours while remaining ≤ 23.2 ng/L throughout. This study contributes to establishing guidelines for clinical use of the hscTnT assay in exercising horses.

Key words: Cardiomyopathy; Clinical chemistry; Horse; Physiology-exercise.

Ardiac troponins (cTn) are highly specific biomarkers of myocardial cell damage, have been utilized widely in human and veterinary medicine, and play a role in helping to identify cardiac pathology in equids. They are central to the diagnoses of acute myocardial infarctions (AMI in humans), but increases in cTn also are apparent in conditions that result in cardiac stress in the absence of obstructive coronary disease.^{1–4} Meeting the stringent recommendations of the Clinical and Laboratory Standards Institute (CLSI), the National Academy of Clinical Biochemistry (NACB), and the American Society of Veterinary Clinical Pathology (ASVCP), the "high-sensitivity" cTnT (hscTnT) assay recently has been validated for use in horses with total imprecision $\leq 10\%$ at the 99th percentile and measurable cTn analyte concentrations obtained in >50% of the

From the Faculty of Veterinary Medicine (UCVM), University of Calgary, Calgary, Alberta, Canada (Shields, Massie, Leguillette); Faculty of Medicine and Calgary Lab Services (CLS), University of Calgary, Calgary, Alberta Canada (Seiden-Long).

Corresponding author: R. Leguillette, 3330 Hospital Dr. NW Calgary, Alberta T2N 4N1, Canada; e-mail: rleguill@ucalgary.ca

Submitted August 30, 2016; Revised August 28, 2017; Accepted October 11, 2017.

Copyright © 2017 The Authors. Journal of Veterinary Internal Medicine published by Wiley Periodicals, Inc. on behalf of the American College of Veterinary Internal Medicine.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

DOI: 10.1111/jvim.14870

Abbreviations:

AST	aspartate aminotransferase			
ASVCP	American Society of Veterinary Clinical Pathology			
CI	confidence interval			
CK	creatine kinase			
CLSI	Clinical and Laboratory Standards Institute			
cTn-I	cardiac troponin-I			
cTnT	cardiac troponin-T			
CV	coefficient of variation			
ECG	electrocardiogram			
hr	hour			
hscTnT	"high-sensitivity" cardiac troponin-T			
IQR	interquartile range			
LoD	limit of detection			
LoQ	limit of quantitation			
NACB	National Academy of Clinical Biochemistry			
ROC	receiver operating characteristic			
TP	total protein			
URL	upper reference limit			

populations studied.⁵ Upper reference limits (URL) for the 95th and 99th population distribution percentiles have been defined for apparently healthy horses and Thoroughbred Chuckwagon racing geldings⁵ and plasma concentrations have been described in 2 populations of horses with cardiac damage.⁶ In numerous studies of healthy humans, in laboratory rodents, in sled-dogs, as well as a few studies of horses, increases in cTn concentrations have been identified after exercise.^{7–15} After myocardial injury in humans, loss of cell membrane integrity results in 2 phases of cTn release: a mild-to-moderate increase within 1- to 2-hour postinjury, and a more substantial increase 4- to 6-hour postinjury which may persist for 7-14 days.^{16,17} We are unaware of similar postmyocardial injury cTn release kinetic studies in horses. Similarly, although available in human athletes for hscTnT and hscTnI assays,18,19 no postexercise cTnT kinetics studies have been reported in horses (although 1 previous study utilized an older generation cTnI assay).¹⁵ The mechanisms whereby troponins, for any reason, are liberated into the bloodstream remain incompletely characterized. Even minor increases in cTn above threshold concentrations, however, confer worse prognosis in human patients across a wide spectrum of acute and chronic cardiac and noncardiac disease processes.²⁰⁻²² Increased cTn concentrations after exercise, therefore, can generate clinical concern, and differentiation among normal cTnT reference population concentrations, expected concentrations after a variety of exercise types, and possible pathologic concentrations warrant defining. Interpretation of troponin assay results with respect to sample timing postexercise, for example, must be also judiciously performed. The effects of age on baseline and postexercise troponin concentrations have been evaluated in humans.¹⁸ Another report in horses examined the effects of age on baseline and postexercise troponin-I concentrations.¹¹ In horses, no such correlations among age, baseline cTnT concentrations, and cTnT release after exercise have been explored. Also, postexercise correlations between markers of skeletal muscle injury or kidney function and cTnT concentrations have not been evaluated in horses. In both the human and veterinary medical literature, there is no consensus regarding the prevalence, physiology, or clinical relevance and management of exercise-associated cTn release.

Our specific objectives were to: study the effects of short-duration, maximal-intensity Chuckwagon racing exercise on cTnT plasma concentrations in a population of healthy racing Thoroughbreds by defining postexercise cTnT release kinetics and determine the effects of age on cTnT concentrations pre- and postrace, using a high-sensitivity cTnT assay. We aimed to provide sampling guidelines for clinicians evaluating horses presenting after exercise to aid in distinguishing normal responses of the equine myocardium to high-intensity, short-duration racing exercise from potentially abnormal pathological cardiac processes.

Materials and Methods

Sampling Methods

Population Sampled

Sampling methods were convenience-based at the Calgary Stampede Chuckwagon races over 7 consecutive days. Racing times were recorded, and average speeds calculated over the Chuckwagon racing pattern. Plasma samples were taken from 38 healthy, fit, actively competing Thoroughbred Chuckwagon racing geldings. Horses were competing at this highest level competition for the sport and were deemed by all competitors and

trainers to be in good general health, excellent physical condition, and had no known history of poor performance, or congenital or acquired cardiac abnormalities. A complete physical examination with heart auscultation was performed, and Telemetric electrocardiography (ECG)^a was performed stall-side and recorded for 2 minutes on all horses. Horses with abnormal physical examination findings, cardiac murmur grade $\geq 3/6$, or any arrhythmia other than second-degree atrioventricular block were excluded from the study. All horses included in the study population had no arrhythmia of sinus origin noted. No echocardiogram was performed. In addition, all horses entered to race passed a veterinary lameness examination before competing. Any horse with baseline plasma cTnT concentrations that exceeded the 99th percentile URL⁵ using the hscTnT assay was excluded from data analysis because pre-existing myocardial damage could not be definitively ruled out.

Blood Sampling Timing

Plasma samples were taken when the horses were at rest in the morning before races (baseline), immediately after a 5/8th mile Chuckwagon race, then at 2, 3, 4, 6, 12, and 24 hour postrace.

Blood Sample Handling

Plasma samples were collected by jugular venipuncture in 5-mL lithium-heparinized tubes. Specimens were immediately centrifuged at $2000 \times g$ for 10 minutes, separated, frozen within 90 minutes, and stored at -80°C until batch analyzed. The hscTnT assay,^b a fifth-generation electrochemiluminescence immunoassay, was used on the Cobas-e601 Analyzer^c for analysis of plasma samples, complying with manufacturer's instructions and Calgary Laboratory Services (CLS) internal quality controls systems, and recently validated for use on equine plasma.⁵ Degree of hemolysis in all samples was assessed using the serum indices instrument application on the Roche Cobas-e601 platform^c and no specimen reached the hemolysis threshold for exclusion. Plasma activities of creatine kinase (CK), and aspartate aminotransferease (AST), and concentrations of creatinine, and total proteins (TP) were analyzed at a commercial laboratory.^d The study was approved by the University of Calgary Veterinary Sciences Animal Care Committee.

Postexercise Release Kinetics for High-Sensitivity Cardiac Troponin-T Analyte Concentrations

Postexercise release kinetics for cTnT were determined by assessment of the absolute values of cTnT plasma concentrations at all time-points (prerace, immediately postrace, as well as 2, 3, 4, 6, 12, and 24 hour postrace) using the hscTnT assay.^b

Pre-Exercise and Selected Postexercise Measurements of CK, AST, Creatinine, and TP

Plasma activities of CK and AST, and concentrations of creatinine and TP were measured using prerace (baseline), and 2-hour postrace blood samples. These were used to determine whether horses had pre-existing evidence of rhabdomyolysis or kidney dysfunction, and as indicators of plasma volume change after exercise.

Determination of Effects of Age on Pre- and Postrace hscTnT Concentrations

The possible correlation between cTnT concentration and age was determined. Prerace cTnT analyte concentrations, as well as all postrace cTnT analyte concentrations, were compared to the individuals' ages.

Statistical Analyses

Commercially available software was used for all calculations (Microsoft Excel^e and GraphPad Prism 6.0^t). Distribution of the data was determined by visual inspection of histograms. Normality of the cTnT concentration data was rejected based on D'Agostino-Pearson testing (P < 0.0001). The limit of detection of the hscTnT assay was 3.0 ng/L, and all horses with concentrations reported by the laboratory as <3.0 ng/L were assigned a concentration of 2.9 ng/L. Transformation of the data could not be accomplished because of the number of results below the limit of detection for the assay. Because of the severity of the right-skewed population in this instance, normalization of the data is unreliable with transformation techniques such as Box-Cox transformation. Consequently, visual inspection of the histogram was used to identify potential outliers in the populations tested. The Shapiro-Wilks test confirmed the normality of the CK, AST, creatinine, and TP distributions.

The Friedman Test was applied to the absolute value time course cTnT analyte concentration data to determine any differences in cTnT concentrations at various collection time-points. When a significant main effect was detected it was followed by Dunn's multiple comparisons test. Paired-*t* tests were used to compare CK, AST, creatinine, and TP prerace to 2-hour postrace samples, and results were reported as mean \pm SD. Possible correlation between baseline and postrace maximum cTnT analyte concentrations and age was investigated using Spearman rank correlation. Significance was assigned to *P* values ≤ 0.05 .

Results

Average racing time over the Chuckwagon pattern was 72.7 seconds with average speeds of 13.8 m/s and peak-speeds of 18.6 m/s.

Thirty-seven apparently healthy, actively competing Thoroughbred Chuckwagon racing geldings (ages 5– 16 years; median, 9 years) were included in the study. Another horse (1/38) had cTnT plasma concentrations that exceeded the 99th percentile URL previously published for the hscTnT assay⁵ at all but one time-point, and was removed from the study population before data analysis.

The absolute plasma cTnT analyte concentrations indicated a significant change with time (P < 0.0001; Fig 1). There were no significant differences in cTnT concentrations among prerace, immediately postrace, 12-hour postrace and 24-hour postrace samples (Fig 1). A significant difference was detected for the absolute plasma cTnT concentrations between the prerace, immediately postrace samples, 12, and 24 hour samples versus the 2, 3, 4, and 6 hour postrace concentrations, respectively (P < 0.0001 for all comparisons; Fig 1). Ten to 14 of 37 horses had $a \ge 2$ -fold increase in cTnT concentrations between baseline and 2- to 6-hour postracing time-points (Table 1 and Fig 1). The plasma cTnT analyte median concentration (8.0 ng/L) was the highest at the 3-hour postrace time-point. The median cTnT concentration then decreased back to concentrations not different from baseline at the 12- and 24-hour postrace time-points. Only 2 horses had AST activities above the normal laboratory range for both pre- and 2hour postrace samples. Only 1 horse had CK activities above the normal laboratory range, measured at the



Fig 1. Distributions of cardiac troponin-T across time measured with a high-sensitivity cTnT assay in 37 Chuckwagon racing Thoroughbred geldings. Absolute plasma cTnT concentrations (in ng/L) are shown for 37 Chuckwagon racing Thoroughbred geldings at prerace, immediately postrace, then at 2, 3, 4, 6, 12 and 24 hour postrace time-points. The dotted line represents the upper 99th percentile cTnT concentrations established previously for the cTnT high-sensitivity Roche assay for racing Thoroughbred horses (see⁵). Boxes represent the interquartile range (IQR), whiskers represent 1.5 × the IQR values, and horizontal lines represent medians. Extreme values (\bullet) represent values between 1.5 × IQR and 3 × IQR. Time-point comparisons that showed significant differences (P < 0.0001) as compared to prerace levels are indicated (*).

2-hour postrace time-point. The maximal AST and CK activities recorded were 816 IU/L and 1112 IU/L, respectively. Plasma activity for AST and CK increased mildly (both P < 0.001) between pre- and 2-hour postrace time-points from $296 \pm 130 \text{ IU/L}$ (prerace AST) to 317 \pm 137 IU/L (postrace AST) and from 133 \pm 28 IU/L (prerace CK) to 277 ± 181 IU/L (postrace CK). All horses had creatinine concentrations within the normal laboratory range with the exception of 2 horses at 2-hour postrace plasma (creatinine concentrations: 205 and 198 µmol/L, respectively). One horse had an increased plasma TP concentration prerace (72.0 g/L) and 1 horse had an increased TP concentration 2-hour postrace (71.0 g/L). Whereas plasma TP did not change $(TP = 60.9 \pm 5.8 \text{ g/L}, \text{ prerace}; TP = 60.7 \pm 5.0 \text{ g/L}, 2$ hour postrace; P = 0.85), plasma creatinine concentration increased from $133 \pm 20 \,\mu mol/L$ (prerace) to $153 \pm 21 \ \mu mol/L$ (2-hour postrace; P < 0.001). No correlations were found between prerace baseline or any postrace maximal cTnT analyte concentrations and age (Fig 2).

Discussion

The goal of our study was to determine if high-intensity short-duration Chuckwagon racing exercise induces a significant increase in cTnT concentration in fit, athletic, clinically normal horses.

Postexercise cTnT analyte release kinetics were characterized by a significant increase from baseline, reaching a median peak at 3-hour postracing, and returning to baseline at 12- and 24-hour postracing. Data enabled comparison of plasma cTnT concentrations to age of the horse, and as no such correlations were found, cTnT results do not need to be interpreted with respect

Shields et al

Sampling Time-Point	Proportion of Horses Below hscTnT Assay LoD (3.0 ng/L)	Proportion of Horses >2-fold Prerace cTnT Concentration (ng/L)	Proportion of Horses >99th URL (cTnT 23.2 ng/L)	Absolute plasma cTnT concentrations (in ng/L; Median and IQRs) Using a High-Sensitivity Assay
Prerace	13/37	N/A	0/37	4.0 (2.9–6.5)
Immediately Postrace	8/37	0/37	0/37	4.0 (3.0-8.2)
Postrace 2 hours	2/37	10/37	0/37	7.0 (6.0–13.2) * $P < 0.0001$, # $P < 0.0001$, † $P < 0.0001$, * $P < 0.0001$
Postrace 3 hours	3/37	14/37	0/37	8.0 (6.0–12.7) * $P < 0.0001$, # $P < 0.0001$, † $P < 0.0001$, * $P < 0.0001$
Postrace 4 hours	2/37	12/37	0/37	7.5 (6.0–13.7) * $P < 0.0001$, # $P < 0.0001$, † $P < 0.0001$, * $P < 0.0001$
Postrace 6 hours	2/37	12/37	0/37	7.0 (5.7–13.0) * $P < 0.0001$ # $P < 0.0001$, † $P < 0.0001$, * $P < 0.0001$
Postrace 12 hours	6/37	3/37	0/37	5.0 (3.0-10.0)
Postrace 24 hours	9/37	0/37	0/37	4.0 (3.0–6.5)

 Table 1. Description of plasma cTnT analyte concentrations measured with a high-sensitivity cTnT assay related to Chuckwagon Racing.

Plasma cTnT analyte concentrations in 37 racing-fit, healthy, Thoroughbred geldings related to 5/8th mile Chuckwagon race; proportions of horses calculated prerace, immediately postrace, then at 2, 3, 4, 6, 12 and 24 hour postrace. Symbols indicate different from (*P* values): *Prerace, #Immediately Postrace, †12-hour Postrace, †24-hour Postrace.

to age. Because there are many different cTn assays available, the specific value of the hscTnT assay is described below. The median peak cTnT of 8.0 ng/L in the study population of healthy horses demonstrates that a cTnT cut-off point of 6.6 ng/L, as recently suggested to differentiate between normal horses at rest and those diagnosed with primary myocardial disease,⁶ would not be applicable postracing. Cardiac troponin release from the myocardium in response to exercise is not well understood but has clinical implications in the differentiation between cTn concentrations postrace in healthy horses and those with myocardial damage, as discussed below. Based on the cTnT response to this specific short-duration maximal-intensity regimen of exercise in healthy horses, we propose a clinical sampling protocol whereby cTnT concentrations can be taken any time between 2- and 6-hour postrace, paired with a second plasma sample taken between 12- and 24hour postrace. These measurements can be used to calculate a repeat measure ratio to aid in understanding normal responses of the equine myocardium to racing exercise. Our study describes the kinetics of cTnT after exercise in healthy horses, but recognizes the need to document cTnT concentrations in horses with myocardial disease (both at rest and after exercise) in future studies.

In our study, plasma cTnT concentrations were above both the limit of detection (LoD) and the limit of quantitation (LoQ) of the hscTnT assay in 64.9% of the horses at baseline resting concentration (pre-exercise), and in 83.8% and 75.7% of the population at rest 12and 24-hour postrace, respectively (Table 1). This observation illustrates 1 of the benefits of using a highsensitivity cTn assay that is >10-fold more sensitive as compared to conventional cTn assays,^{23,24} and confirms conforming to the CLSI criteria that high-sensitivity assays must measure cTn concentrations in at least 50% of the population at rest. The ability of the present hscTnT assay to function reliably at these low concentrations (LoQ) is unique when compared to the only cTnI assay similarly validated appropriately by ASVCP and CLSI standards for use in horses.¹¹ Because all assays for cTnT are marketed by 1 manufacturer,^b and instruments are calibrated to the same reference material, analysis, and standardization are beneficially more straightforward than for cTnI. Although the postexercise release peak time-points (2-6 hours) identified in our study using the hscTnT assay generally are similar to the results from a previous study using a conventional cTnI assay,¹⁵ direct comparisons between studies must be made with caution because "all troponin assays are not created equal."25 Increased use of this appropriately validated hscTnT assay in horses in future research will improve the ability to compare the results of 1 study to another.

Our study examined the effects of short-duration, high-intensity Chuckwagon racing on the equine myocardium as measured by a hscTnT assay. The response of the myocardium to exercise has been explored to help elucidate mechanisms for cTnT release associated with exercise that may be distinct from actual myocardial necrosis. The underlying mechanisms of the release of cardiac troponins into the bloodstream observed after high-intensity short-duration exercise remain unclear.²⁶ Based on current human medical literature, exercise is categorized as a condition that induces increases in circulatory cardiac troponins without evidence of AMI resulting in necrosis.²⁷ Several theories, such as cardiac strain, have been proposed and are in good agreement with data obtained under experimental





Fig 2. High-sensitivity cardiac troponin-T plasma concentrations versus horse age in 37 Chuckwagon racing Thoroughbred geldings. (A) Baseline prerace plasma cTnT concentrations (in ng/L) versus age measured with a high-sensitivity cTnT assay are shown for 37 Chuckwagon racing Thoroughbred geldings. (B) Plasma cTnT concentrations (in ng/L) taken 3-hour postrace versus age are shown for 37 Chuckwagon racing Thoroughbred geldings.

conditions.^{28,29} Troponin and exercise physiology studies in humans are mostly available for endurance exercise, but more recently an increased focus on nonendurance, higher-intensity exercise can be noted. Collective research in humans examining nonendurance exercise has demonstrated significant increases in cTn concentrations after standardized treadmill running (30 minute at 85-90% VO_{2max}),²⁶ repetitive sprints (12 × 30-second sprints with set recovery periods in between),³⁰ and rowing (30-minute high-intensity).³¹ In research on horses, limited work has been completed for endurance exercise, ^{12,32,33} and only 1 report examined high-intensity short-duration exercise.¹⁵ The exact duration or intensity of exercise necessary to invoke significant changes in cTn concentrations has not been definitively determined in horses or humans. Seven research studies in horses have evaluated cTn concentrations in normal horses after nonendurance exercise. None of these studies identified significant increases in cTn concentrations partly as a consequence of the study population sizes (15, 28, 6, 24, 26, and 15 horses, respectively)^{13–15,34–36} but also as a result of the use of older generation,¹¹ conventional sensitivity assays that

have a higher LoD.^{37,38} We identified significant increases in cTn, using the hscTnT assay, in horses after high-intensity, short-duration racing exercise.

Although unlikely, the increased cTnT concentrations observed here may not have been solely a result of cardiac muscle release of cTn, but also possibly a result of skeletal muscle damage, impaired renal function or exercise-induced dehydration. Our study did not directly investigate the effects of rhabdomyolysis or decreased renal function on cTnT concentrations after racing exercise in horses, but it was confirmed that the study population was healthy and unaffected by rhabdomyolysis or clinically relevant renal impairment as documented by prerace and 2-hour postrace blood biochemistry analyses.^d Additional sampling time-points for blood biochemistry analyses may have further confirmed these findings, but the CK and AST activities obtained suggest that clinically relevant skeletal muscle damage was unlikely. Excellent cardiac tissue specificity furthermore was confirmed in a previous validation study in which the hscTnT assay reactivity of cardiac muscle was 256.2 times higher than that of skeletal muscle $(2.49 \times 10^7 \text{ and } 9.75 \times 10^4 \text{ ng/g}$ wet weight respectively).⁵ Horses with acute, exercise-induced myopathy have been reported to have similar plasma cTnI concentrations to those the subjects had prior to the onset of myopathy.¹⁵ Collectively, this observation suggests that skeletal muscle breakdown after Chuckwagon racing was unlikely to have had an effect on cTnT concentrations in our study and did not limit interpretation of the results. Similarly, the increases in cTnT detected were unlikely to have been affected by any changes in free water. Indeed, TP (as an approximation of plasma water content) did not change significantly in our study between baseline and 2-hour postrace. This was the first time-point at which a significant increase in cTnT concentration (compared to baseline) was noted. Similar conclusions recently were made regarding the increases in cTnI concentrations identified using a high-sensitivity cTnI assay in horses after endurance racing whereby horses with the highest cTnI concentrations did not appear to have more marked hematologic or biochemical changes compared with the remaining horses.³³

All horses with detectable concentrations (35/37) in our study had an increase in cTnT induced by the racing exercise with an average delta absolute change from prerace baseline to peak 3-hour postrace cTnT concentration of 4.3 ng/L (median, 3.6 ng/L; range, 0– 17 ng/L). Not only do the results indicate significant increases in cTnT, but the magnitude of the increases also suggests a relatively uniform response to the same amount of exercise, which is consistent with findings in exercise physiology of humans whereby exercise protocols similar in intensity and duration are correlated with cTn release kinetics and associated peaks.^{27,39–41}

The upper 99th percentile resting cTnT plasma analyte concentration reported for Thoroughbred Chuckwagon Racing horses, using the hscTnT assay, has been previously established to be <23.2 ng/L.⁵ In our study, all cTnT concentrations measured were <23.2 ng/L, but 2/37 horses had peak cTnT plasma concentrations (23.0 ng/L for both horses) that closely approached the 99th percentile URL (see Table 1, Fig 1), and then decreased to baseline concentrations by 24-hour postexercise in accordance with the remainder of the study population. Departure from the normal expected cTnT kinetics after exercise may be a practical way to differentiate "normal" horses from those that: (1) have preexisting subclinical (i.e. occult) cardiac disease that results in an unexpected response of the myocardium to exercise, or (2) the exercise dose itself induces myocardial damage in that particular individual. Data on cTnT kinetics in horses with myocardial damage, however, are not available, and any persistence in cTnT concentration increases postracing in horses with myocardial damage, as reported in humans after AMI,¹⁷ must be verified. In humans after AMI, loss of cell membrane integrity results in multiple phases of cTn release. There is indeed a relatively long half-life of cTn in serum of \geq 2 hour because of persistent leakage after AMI.¹⁷ In contrast, because of rapid renal elimination, the true half-life of both cTnT and cTnI in circulation is approximately 2 hour in humans,⁴¹ and only cTnI has been determined in horses to have a half-life of 0.47 hour.⁴² If ischemia or other insults to cellular homeostasis fail to induce necrosis (such as has been proposed to be the case after exercise), this short true half-life results in a pattern of cTn kinetics unlike those seen after AMI. In our study, cTnT concentrations returned to baseline after 12 to 24 hour thereby suggesting that racing exercise did not induce myocardial necrosis in these Chuckwagon horses.

The kinetics described above provided the basis for sampling protocol recommendations. In practical situations in which a clinician may be measuring cTnT as part of examination of a horse that has undergone high-intensity, short-duration exercise in the recent past, and pre-exercise (baseline) cTnT concentrations are unavailable, our results suggest that because postracing exercise plasma cTnT concentrations are expected to peak 3-hour postexercise and thus be measured anytime between 2- and 6-hour postexercise because no significant differences were noted among these collection times in our study. Although no horse, at any time, in the our study exceeded the URL for cTnT concentrations (i.e. 23.2 ng/L with the hscTnT assay), similar to results from exercise studies in humans, it may be normal for healthy horses to actually meet or possibly exceed the URL for cTnT concentrations at these peak time-points.^{26,30,31,33,43-45} Also, the cTnT concentrations decreased at, and after, 12-hour postrace in all horses that had concentrations above the LoD of the hscTnT assay. Thus, any horse with a cTnT concentration at 12 or 24 hours postrace, that is, equal to or higher than a concentration observed between 2- and 6-hour postrace may warrant further evaluation for possible cardiac damage. Based on the above, we suggest the following repeat measure plasma cTnT ratio using the hscTnT assay (with values ≤ 1 representing normal horses):

Plasma cTnT sample taken between

$$\frac{12 \text{- to } 24 \text{- h postrace}\left(\frac{\text{ng}}{L}\right)}{\text{Plasma cTnT sample taken between}} \le 1$$
$$2 \text{- to } 6 \text{- h postrace}\left(\frac{\text{ng}}{L}\right)$$

The implication of the increased resting cTn concentrations measured in males vs. females and with increasing age, as confirmed in a study of 1540 people, is currently unknown. Individual factors such as sex and age are important to address for clinical relevance with respect to clinical decision limits (i.e. application of established reference population, URL). In horses, a weak positive age, but not sex, correlation has been corroborated in 1 study of 586 Standardbreds at rest.¹¹ Age correlations also have been explored in subjects undergoing exercise challenges, and in a recent cTnT postexercise study of humans using a high-sensitivity assay, no age correlation was found between age and peak postexercise cTn concentrations. Finally, in our study, no correlations were found between age and baseline or peak postexercise cTnT concentrations.

Limitations of our study included the lack of echocardiography, exercise or postexercise ECG, and possible inclusion of horses with nonphysiological murmurs of grade <3/6, and as such it is possible that horses with mild cardiac impairments were inadvertently included. Fitness levels of horses were not individually evaluated, and therefore, the intensity of racing exercise (e.g. as a percentage of VO_{2max}) may have differed among individuals thereby impacting postexercise cTn concentration kinetics. In addition, conclusions and clinician sampling guideline recommendations are based on a population of horses that underwent a specific racing dose of exercise (short-term, high-intensity), and thus extrapolation to other forms of exercise or equestrian activities must be made with caution. Finally, the results of our study define the response of myocardium to exercise in healthy horses, but the cTnT kinetics and associated peak concentrations in horses with myocardial disease may be different and warrant independent evaluation in future studies.

Conclusions

The adoption of cTn assays for use in veterinary medicine, coupled with concern about the possible association of underlying or exercise-induced myocardial injury and pathological exercise-associated arrhythmias, ^{32,33,37,46,47} has led to numerous studies in horses exploring increases in cTn in exercising populations and after exercise.^{11,12,14,15,32–34,48,49} The development and validation of high-sensitivity cTn assays have allowed further assessment of the response of the equine myocardium to short-duration, high-intensity racing exercise. Our study demonstrates that (1) this exercise induces a significant increase in plasma cTnT from baseline (or 12- to 24-hour postexercise) concentrations between 2- and 6-hour postexercise; (2) cTnT concentration peaks at 3-hour post-xercise; (3) cTnT concentration at 12 or 24 hour postexercise was lower than at 2- to 6-hour postexercise; (4) cTnT concentrations at 12 or 24 hour postexercise were not different than pre-exercise concentrations in any horse. The combination of postrace cTnT measurements using a hscTnT assay taken any time between 2- and 6-hour postrace, paired with a second plasma sample taken at 12 or 24 hours postrace to calculate the repeat measure ratio, and absolute cTnT plasma concentrations that fall under the 99th percentile URL of 23.2 ng/L by 12to 24-hour postrace, are appropriate clinical guidelines to differentiate normal responses of the equine myocardium to high-intensity short-duration racing exercise from potentially pathological cardiac processes that may warrant further evaluation and monitoring. Future studies to determine absolute cTn concentrations during and after exercise above which myocardial damage is indicated, as well as to verify that a cTnT repeat measure ratio ≥ 1 , or single sample cTnT analyte concentration ≥ 23.2 ng/L 12 or 24 hours after racing, may be positively correlated with myocardial damage in a horse are needed. In clinical settings, to prevent misdiagnosis of cardiac pathology in otherwise healthy horses, results of cTn analyte measurements must be analyzed in light of sampling timing in relationship to exercise, and ideally interpreted in conjunction with detailed clinical history and other objective cardiac measurements such as ECG and echocardiographic findings.⁸ Recognition that exercise stimulates release of cTn will help veterinarians make informed clinical decisions about how to use cTn testing appropriately in horses after exercise.

Footnotes

- ^a Televet-100, Engel Engineering, Offenbach, Germany
- ^b Troponin-T hs, Roche Diagnostics, Indianapolis, IN
- ^c Cobas-e601, Roche Diagnostics, Indianapolis, IN
- ^d Idexx Laboratories, Beckman AU680 Chemistry Analyzer, Calgary, AB
- ^e Microsoft Excel, Microsoft Corporation, Redmond, WA
- ^f GraphPad Prism 6.0, GraphPad Prism Software, Inc., La Jolla, CA

Acknowledgments

The study was completed in Calgary, Alberta, Canada, at the Calgary Exhibition and Stampede Grounds Racetrack. The study was supported University of Calgary, Faculty of Veterinary Medicine—Clinical Research Fund. The authors thank contributing owners, The Calgary Exhibition and Stampede, and the Calgary Laboratory Services collaborators for technical assistance.

Conflict of Interest Declaration: Authors declare no conflict of interest.

Off-label Antimicrobial Declaration: Authors declare no off-label use of antimicrobials.

References

1. Ammann P, Maggiorini M, Bertel O, et al. Troponin as a risk factor for mortality in critically ill patients without acute coronary syndromes. J Am Coll Cardiol 2003;41:2004–2009.

2. Pattanshetty DJ, Bhat PK, Aneja A, et al. Elevated troponin predicts long-term adverse cardiovascular outcomes in hypertensive crisis: A retrospective study. J Hypertens 2012;30:2410–2415.

3. Vasile VC, Chai HS, Abdeldayem D, et al. Elevated cardiac troponin T levels in critically ill patients with sepsis. Am J Med 2013;126:1114–1121.

4. Vasile VC, Chai HS, Khambatta S, et al. Significance of elevated cardiac troponin T levels in critically ill patients with acute respiratory disease. Am J Med 2010;123:1049–1058.

5. Shields E, Seiden-Long I, Massie S, et al. Analytical validation and establishment of reference intervals for a 'high-sensitivity' cardiac troponin-T assay in horses. BMC Vet Res 2016;12:104.

6. Van Der Vekens N, Decloedt A, Ven S, et al. Cardiac troponin I as compared to troponin T for the detection of myocardial damage in horses. J Vet Intern Med 2015;29:348–354.

7. Vilela EM, Bastos JC, Rodrigues RP, et al. High-sensitivity troponin after running–a systematic review. Neth J Med 2014;72:5–9.

8. Lippi G, Cervellin G, Banfi G, et al. Cardiac troponins and physical exercise. It's time to make a point. Biochem Med 2011;21:55–62.

9. Nie J, Close G, George KP, et al. Temporal association of elevations in serum cardiac troponin T and myocardial oxidative stress after prolonged exercise in rats. Eur J App Physiol 2010;110:1299–1303.

10. McKenzie EC, Jose-Cunilleras E, Hinchcliff KW, et al. Serum chemistry alterations in Alaskan sled dogs during five successive days of prolonged endurance exercise. J Am Vet Med Assoc 2007;230:1486–1492.

11. Slack J, Boston RC, Soma L, Reef VB. Cardiac troponin I in racing standardbreds. J Vet Intern Med 2012;26:1202–8.

12. Holbrook TC, Birks EK, Sleeper MM, et al. Endurance exercise is associated with increased plasma cardiac troponin I in horses. Equine Vet J Suppl 2006;36:27–31.

13. Nostell K, Haggstrom J. Resting concentrations of cardiac troponin I in fit horses and effect of racing. J Vet Cardiol 2008;10:105–109.

14. Trachsel DS, Schwarzwald CC, Bitschnau C, et al. Atrial natriuretic peptide and cardiac troponin I concentrations in healthy Warmblood horses and in Warmblood horses with mitral regurgitation at rest and after exercise. J Vet Cardiol 2013;15:105–121.

15. Durando M, Reef V, Kline K, et al. Acute effects of short duration, maximal exercise on cardiac troponin I in healty horses. Equine Comp Exerc Physiol 2007;3:217–223.

16. Bleier J, Vorderwinkler KP, Falkensammer J, et al. Different intracellular compartmentations of cardiac troponins and myosin heavy chains: A causal connection to their different early release after myocardial damage. Clin Chem 1998;44:1912–1918.

17. Cardinaels EP, Mingels AM, van Rooij T, et al. Timedependent degradation pattern of cardiac troponin T following myocardial infarction. Clin Chem 2013;59:1083–1090.

18. Tian Y, Nie J, Huang C, et al. The kinetics of highly sensitive cardiac troponin T release after prolonged treadmill exercise in adolescent and adult athletes. J Appl Physiol 2012;113:418–425.

19. Lippi G, Schena F, Dipalo M, et al. Troponin I measured with a high sensitivity immunoassay is significantly increased after a half marathon run. Scand J Clin Lab Invest 2012;72:467–470.

20. Ahmed AN, Blonde K, Hackam D, et al. Prognostic significance of elevated troponin in non-cardiac hospitalized patients: A systematic review and meta-analysis. Ann Med 2014;46:653–663.

21. Latini R, Masson S, Anand IS, et al. Prognostic value of very low plasma concentrations of troponin T in patients with stable chronic heart failure. Circulation 2007;116:1242–1249.

22. Januzzi JL Jr, Filippatos G, Nieminen M, et al. Troponin elevation in patients with heart failure: On behalf of the third Universal Definition of Myocardial Infarction Global Task Force: Heart Failure Section. Eur Heart J 2012;33:2265–2271.

23. Katus HA, Giannitsis E, Jaffe AS, et al. Higher sensitivity troponin assays: Quo vadis? Eur Heart J 2009;30:127–128.

24. Jesse RL. On the relative value of an assay versus that of a test: A history of troponin for the diagnosis of myocardial infarction. J Am Coll Cardiol 2010;55:2125–2128.

25. Christenson RH, Phillips D. Sensitive and high sensitivity next generation cardiac troponin assays: More than just a name. Pathology 2011;43:213–219.

26. Shave R, Ross P, Low D, et al. Cardiac troponin I is released following high-intensity short-duration exercise in healthy humans. Int J Cardiol 2010;145:337–339.

27. Chenevier-Gobeaux C, Bonnefoy-Cudraz E, Charpentier S, et al. High-sensitivity cardiac troponin assays: Answers to frequently asked questions. Arch Cardiovasc Dis 2015;108:132–149.

28. Nunes JP, Macedo F. An analytical triad for the diagnosis of pulmonary embolism. Cardiology 2000;94:264.

29. Nunes JP. Cardiac troponin I in systemic diseases. A possible role for myocardial strain. Rev Port Cardiol 2001;20:785–788.

30. Weippert M, Divchev D, Schmidt P, et al. Cardiac troponin T and echocardiographic dimensions after repeated sprint vs. moderate intensity continuous exercise in healthy young males. Sci Rep 2016;6:24614.

31. Legaz-Arrese A, Lopez-Laval I, George K, et al. Individual variability in cardiac biomarker release after 30 min of high-intensity rowing in elite and amateur athletes. Appl Physiol Nutr Metab 2015;40:951–958.

32. Flethoj M, Schwarzwald CC, Haugaard MM, et al. Left ventricular function after prolonged exercise in equine endurance athletes. J Vet Intern Med 2016;30:1260–1269.

33. Flethoj M, Kanters JK, Haugaard MM, et al. Changes in heart rate, arrhythmia frequency, and cardiac biomarker values in horses during recovery after a long-distance endurance ride. J Am Vet Med Assoc 2016;248:1034–1042.

34. Ducharme NG, Fortier LA, Kraus MS, et al. Effect of a tart cherry juice blend on exercise-induced muscle damage in horses. Am J Vet Res 2009;70:758–763.

35. Naylor RJ, Luis-Fuentes V, Livesey L, et al. Evaluation of cardiac phenotype in horses with type 1 polysaccharide storage myopathy. J Vet Intern Med 2012;26:1464–1469.

36. Buhl R, Peterson EE, Lindholm M, et al. Cardiac arrythmias in standardbreds during and after racing - possible

association between heart size, valvular regurgitations, and arrhythmias. J Equine Vet Sci 2013;33:590–596.

37. Rossi TM, Pyle WG, Maxie MG, et al. Troponin assays in the assessment of the equine myocardium. Equine Vet J 2014;46:270–275.

38. Balmelli C, Meune C, Twerenbold R, et al. Comparison of the performances of cardiac troponins, including sensitive assays, and copeptin in the diagnostic of acute myocardial infarction and long-term prognosis between women and men. Am Heart J 2013;166:30–37.

39. Tian Y, Nie J, George KP, et al. Reproducibility of cardiac biomarkers response to prolonged treadmill exercise. Biomarkers 2014;19:114–120.

40. Klinkenberg LJ, Res PT, van Loon LJ, et al. Strong link between basal and exercise-induced cardiac troponin T levels: Do both reflect risk? Int J Cardiol 2012;158:129–131.

41. Gerhardt W, Katus H, Ravkilde J, et al. Troponin T in suspected ischemic myocardial injury compared with mass and catalytic concentrations of Creatine kinase isoenzyme MB. Clin Chem 1991;37:1405–1411.

42. Kraus MS, Kaufer BB, Damiani A, et al. Elimination halflife of intravenously administered equine cardiac troponin I in healthy ponies. Equine Vet J 2013;45:56–59.

43. Stewart GM, Yamada A, Haseler LJ, et al. Influence of exercise intensity and duration on functional and biochemical perturbations in the human heart. J Physiol 2016;594:3031–44.

44. Lopez-Laval I, Legaz-Arrese A, George K, et al. Cardiac troponin I release after a basketball match in elite, amateur and junior players. Clin Chem Lab Med 2016;54:333–338.

45. Duttaroy S, Thorell D, Karlsson L, et al. A single-bout of one-hour spinning exercise increases troponin T in healthy subjects. Scand Cardiovasc J: SCJ 2012;46:2–6.

46. Boden LA, Charles JA, Slocombe RF, et al. Sudden death in racing thoroughbreds in Victoria, Australia. Equine Vet J 2005;37:269–271.

47. Physick-Sheard PW, McGurrin MK. Ventricular arrhythmias during race recovery in Standardbred Racehorses and associations with autonomic activity. J Vet Intern Med 2010;24:1158–1166.

48. Begg LM, Hoffmann KL, Begg AP. Serum and plasma cardiac troponin I concentrations in clinically normal Thoroughbreds in training in Australia. Austral Vet J 2006;84:336–337.

49. Phillips W, Giguere S, Franklin RP, et al. Cardiac troponin I in pastured and race-training Thoroughbred horses. J Vet Intern Med 2003;17:597–599.