Revised: 7 July 2021

ORIGINAL ARTICLE

MOLECULAR ECOLOGY WILEY

Fine-scale spatial genetic structure across the species range reflects recent colonization of high elevation habitats in silver fir (*Abies alba* Mill.)

Enikő I. Major¹ | Mária Höhn¹ | Camilla Avanzi² | Bruno Fady³ | Katrin Heer⁴ | Lars Opgenoorth^{5,6} | Andrea Piotti² | Flaviu Popescu⁷ | Dragos Postolache⁷ | Giovanni G. Vendramin² | Katalin Csilléry⁸

¹Department of Botany, Hungarian University of Agronomy and Life Sciences, Budapest, Hungary

²Institute of Biosciences and Bioresources, National Research Council of Italy (IBBR-CNR), Sesto Fiorentino (Firenze), Italy

³Ecology of Mediterranean Forests (URFM), UR629, INRAE, Avignon, France

⁴Conservation Biology, Philipps Universität Marburg, Marburg, Germany

⁵Plant Ecology and Geobotany, Philipps Universität Marburg, Marburg, Germany

⁶Biodiversity and Conservation Biology, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland

⁷National Institute for Research and Development in Forestry "Marin Drăcea", Ilfov County, Romania

⁸Land Change Science, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland

Correspondence

Katalin Csilléry, Evolutionary Genetics Group, Biodiversity and Conservation Biology Unit, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland. Email: katalin.csillery@wsl.ch

Funding information

Schweizerischer Nationalfonds zur Förderung der Wissenschaftlichen Forschung, Grant/Award Number: CRSK-3_190288; Agence Nationale de la Recherche, Grant/Award Number: ANR-12-EBID-0003; German Federal Ministry of Education and Research, Grant/Award Number: 01LC1202A

Abstract

Variation in genetic diversity across species ranges has long been recognized as highly informative for assessing populations' resilience and adaptive potential. The spatial distribution of genetic diversity within populations, referred to as fine-scale spatial genetic structure (FSGS), also carries information about recent demographic changes, yet it has rarely been connected to range scale processes. We studied eight silver fir (Abies alba Mill.) population pairs (sites), growing at high and low elevations, representative of the main genetic lineages of the species. A total of 1,368 adult trees and 540 seedlings were genotyped using 137 and 116 single nucleotide polymorphisms (SNPs), respectively. Sites revealed a clear east-west isolation-by-distance pattern consistent with the post-glacial colonization history of the species. Genetic differentiation among sites ($F_{CT} = 0.148$) was an order of magnitude greater than between elevations within sites ($F_{SC} = 0.031$), nevertheless high elevation populations consistently exhibited a stronger FSGS. Structural equation modelling revealed that elevation and, to a lesser extent, post-glacial colonization history, but not climatic and habitat variables, were the best predictors of FSGS across populations. These results suggest that high elevation habitats have been colonized more recently across the species range. Additionally, paternity analysis revealed a high reproductive skew among adults and a stronger FSGS in seedlings than in adults, suggesting that FSGS may conserve the signature of demographic changes for several generations. Our results emphasize that spatial patterns of genetic diversity within populations provide information about demographic history complementary to non-spatial statistics, and could be used for genetic diversity monitoring, especially in forest trees.

KEYWORDS

climate change, colonization, demography, distribution range, fine-scale spatial genetic structure, forest tree, genetic diversity, reproductive success, sampling

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. *Molecular Ecology* published by John Wiley & Sons Ltd.

[⊥]WILEY-<mark>molecular ecology</mark>

1 | INTRODUCTION

Dispersal capacity of organisms plays a fundamental role in the establishment, persistence and range dynamics of species, especially during environmental changes (Saastamoinen et al., 2018; Travis et al., 2013). Most terrestrial organisms have limited dispersal, which leads to a non-random spatial distribution of genes and genotypes. The short-term consequence of this process is the clustering of related individuals, which is manifested as a decay-of-kinship with distance, and is commonly referred to as fine-scale spatial genetic structure (FSGS) (Epperson, 1995; Rousset, 2000; Sokal & Wartenberg, 1983; Wright, 1943). The long-term consequence of limited dispersal is that populations become subdivided and distant populations become genetically more differentiated than nearby populations; a phenomenon referred to as isolation-by-distance (IBD) in subdivided populations (Kimura & Weiss, 1964; Malécot, 1948; Wright, 1943). Thus, FSGS captures principally the effect of limited dispersal with respect to the past few generations, while, in contrast, IBD reflects deeper demographic time scales. This distinction is important because, ignoring mutations, a drift-dispersal equilibrium can be reached within a few generations on a spatial scale up to one order of magnitude larger than the average parent-offspring distance, while many more generations can be necessary on larger spatial scales (Bradburd & Ralph, 2019; Slatkin, 1993; Vekemans & Hardy, 2004).

IBD is an ubiquitous property of most biological systems, yet this pattern is driven by many ecological and evolutionary factors, and not only variation in dispersal (Duforet-Frebourg & Blum, 2014; Legendre & Fortin, 2010; Rousset, 2000; Slatkin, 1985). Most species evolutionary histories in the northern hemisphere are dominated by recent colonization dynamics from glacial refugia since the Last Glacial Maximum (LGM) (Excoffier et al., 2009; Petit et al., 2008). In these cases, IBD has little to do with the presence or absence of a drift-migration equilibrium (Aguillon et al., 2017; Meirmans, 2012) while patterns of FSGS across the species range could provide complementary information by elucidating the recent demographic history. Indeed, recently expanded populations are expected to more likely show spatially aggregated patterns of related individuals with respect to demographically old and dense populations (De-Lucas et al., 2009; Gapare & Aitken, 2005; Pandey & Rajora, 2012; Torroba-Balmori et al., 2017). However, patterns of FSGS may also differ from one population to another across the species range for reasons other than the demographic age of populations. Differences may reflect natural or human disturbances, such as management or habitat degradation (e.g., Bessega et al., 2016; Dreier et al., 2014; Piotti et al., 2013), fire (Budde et al., 2017), presence or absence of dispersers (Barluenga et al., 2011; Gelmi Candusso et al., 2017; Kling & Ackerly, 2021; Rico & Wagner, 2016), or adaptation to the local environment (e.g., Audigeos et al., 2013; Gauzere et al., 2020; Mosca et al., 2018; Troupin et al., 2006).

Most studies either investigate IBD or FSGS, which impedes the simultaneous inference of long- and short-term demographic signatures on spatial genetic structure. A spatially dense sampling of

non-disturbed populations across the range would be necessary to detect both IBD and FSGS: however, the sampling requirements for inferring FSGS at a given site can be an order of magnitude higher than that for IBD. Further, various ecological and disturbance factors may also affect the optimal size of the sampling area of FSGS studies (Vekemans & Hardy, 2004). Generally speaking, the sampling area should be as small as possible to capture only short-term processes, but it should also contain a sufficiently large number of individuals to precisely estimate the slope of the regression of kinship coefficients against geographic distances or the spatial autocorrelation of allele frequencies (Leblois et al., 2003; Smouse et al., 2008). The level of polymorphism in genetic markers can also vary across the species range, and influence the power of a data set for estimating kinship (Wang, 2016) or the spatial correlograms. The same genetic data that are used to estimate the spatial autocorrelation of allele frequencies may also be used to partially reconstruct the pedigree, which can be used to understand the mechanisms that maintain FSGS, such as the unequal reproductive success among individuals (e.g., Gerzabek et al., 2017; Oddou-Muratorio et al., 2018; Paluch et al., 2019) or sex specific dispersal dynamics (Aguillon et al., 2017).

Silver fir (Abies alba Mill.) is an evergreen coniferous mountain forest tree of Central, Southern and Eastern Europe with slow growth and a generation time of 30 to 40 years (Wolf, 2003). In the context of climate change, silver fir has recently attracted interest for being more drought tolerant than other European dominant forest tree species, such as Norway spruce (Picea abies K.) (Vitali et al., 2017; Vitasse et al., 2019). Fossil data also suggest that silver fir previously survived under much warmer than current temperatures, thus it could have become more adapted to drought (Tinner et al., 2013). There is little fossil evidence for the distribution of silver fir during the climatic oscillations of the Quaternary (past 2.5 Myr BP), but fossil records are common in the Holocene starting from approximately 11.6 kyr BP (Magri et al., 2017). The available data suggests that before the LGM (21 kyr BP) during warm glacial interstadials the species principally remained in refugia in the Italian and the Balkan peninsulas (Tinner et al., 2013). Two main maternal mitochondrial (mt)DNA lineages probably represent these two refugia (Liepelt et al., 2002). The range expansion from these refugia in southern Europe is currently ongoing, which is an exceptional event of the Quaternary history of the species (Willy Tinner, personal communication). The southern part of the Carpathians was probably colonized from the Balkans while the eastern part from the Apennines (Bosela et al., 2016; Gömöry et al., 2012; Liepelt et al., 2002, 2009). The Alps were most probably colonized from the Northern Apennines, but two additional routes have probably contributed, namely from southern France and from the Balkans through the Adriatic coast (Burga & Hussendörfer, 2001; Terhürne-Berson et al., 2004; Liepelt et al., 2009; Cheddadi et al., 2014; Ruosch et al., 2016; Piotti et al., 2017). The Apennines hosted at least three refugial areas during the LGM, whose exact delimitation has been described by Piotti et al. (2017). Silver fir populations from the Pyrenees show a high genetic differentiation from the rest of species range, but belong to the western mtDNA haplotype group, suggesting that Pyrenees have also been

colonized from the Apennines, but earlier then the last glacial cycle (Liepelt et al., 2009).

The main objective of this study was to identify demographic and environmental drivers of FSGS across the species range of silver fir, as well as comparing FSGS across seedling and adult cohorts and assessing population variation in dispersal parameters. Silver fir is a wind pollinated and dispersed, monoecious, mixed-mating but generally outcrossing species (Wolf, 2003), thus we expected to find an overall weak FSGS across the range. We studied natural silver fir populations with no or close-to-nature management, so we assumed that differences in FSGS were due to demographic history and/or environmental constraints/adaptation. We assembled published single nucleotide polymorphism (SNP) marker data for adult trees (Brousseau et al., 2016; Heer et al., 2018; Roschanski et al., 2016) from eight sites sampled across the species range. In particular, the sites represented the two mtDNA haplotype groups and cover all recolonization routes from effective refugia (Liepelt et al., 2002, 2009). Further, two populations from Central Apennines and the Pyrenees that have been isolated from other western populations before the Holocene were included (Terhürne-Berson et al., 2004). At each site, a pair of low and high elevation populations were sampled that most probably have been separated from one another at more recent time scales. This hierarchical sampling design allowed us to disentangle the effects of the distant past, that is, the colonization of Europe from glacial refugia during the Holocene, and those of recent demographic events, such as possible colonization of high elevation habitats during the past few generations (Chauchard et al., 2010; Hernández et al., 2019; Vitasse et al., 2019). We developed a proxy for the effects of distant past using range-wide IBD patterns, and hypothesized that populations close to the supposed refugia are in migration-drift equilibrium and exhibit no FSGS, while populations in recently colonized areas of the species range show some degree of spatial structuring. With respect to elevation, we hypothesized that both recent demographic history and potential effects of environmental adaptation or habitat quality could contribute to differences in FSGS, and predicted stronger FSGS at higher elevations because they represent the colonizing edges of the range (Vitasse et al., 2019). Previous studies on some of these populations showed evidence for adaptation to elevation (Brousseau et al., 2016; Roschanski et al., 2016), and the effect of environmental factors on FSGS have been identified in other Italian silver fir populations (Mosca et al., 2018). Although the scale of our study allowed to disentangle demographic effects at different time scales, the sampling was performed by different research groups, which raised the question whether differences in FSGS across the 16 populations reflect not only biological patterns, but eventual sampling artifacts. To this end, we performed a resampling of our largest populations to assess how different spatial sampling configurations might have influenced our results. Finally, we addressed the question of the maintenance of FSGS across the range. To this end, we estimated the strength of FSGS in seedlings, as well as the distribution of reproductive success in adult trees, and dispersal parameters, such as the selfing rate and

the gametic gene flow. We hypothesized that seedlings would exhibit a stronger FSGS than adults because of limited seed dispersal, especially in recently colonized, non-equilibrium populations.

2 | MATERIALS AND METHODS

2.1 | Data sets

We assembled a data set of 1,368 adult trees from previously published data from eight different sites across the species distribution range (Figure 1a and Table 1; Issole (ISS), Lure (LUR), and Vesubie (VES) from Roschanski et al., (2016); Apennines (APE), Fagaras Mountains (FAG), Pyrenees (PYR), and Ventoux (VEN) from Brousseau et al., (2016), and Bavaria (BAV) from Heer et al., (2018)). Each site was sampled at high and low elevations that were subsequently denoted with _H and _L after the population names (Table 1). The number of genotyped adult trees per population ranged from 43 to 250 (Table 1, Figure S1). Additionally, at three sites (APE, BAV and FAG) we sampled and genotyped 540 seedlings with height <1 m and diameter at breast height <4 cm (Table 1, Figure S1). Similar sampling configuration and area were used across all sites, but differences in the terrain and in the population density imposed some differences, especially in the spatial distribution of seedlings. Further, the sites LUR, ISS and VES were considered as replicates of VEN and sampling effort was less important (Table 1), but still a nearly exhaustive sampling of adult trees in a given area appeared suitable for inclusion in this study. As a result, the distance between the two farthest trees ranged from 68 m (ISS H) to 1,397 m (BAV L) (Table 1 and Figure S1). Sampled trees were georeferenced using a GPS device (Garmin Ltd in APE, BAV, ISS, LUR, PYR, VEN and VES; Trimble Juno T41/5 in FAG) or compass and laser distance-meter (Leica Geosystems AG in APE).

All individuals were genotyped using the same set of 267 SNP loci from 175 candidate genes using KASP assays manufactured by LGC Genomics (Roschanski et al., 2013, 2016), with the exception of APE and FAG, where seedlings were genotyped at a subset of 211 loci. Among these, we only used loci with less than 15% missing data, a minor allele frequency >5%, and that did not significantly deviate from the Hardy-Weinberg equilibrium in at least 75% of the populations (Table 2). Further, previous studies identified significant environmental and phenotypic associations at some of the loci (Brousseau et al., 2016; Heer et al., 2018; Roschanski et al., 2016). Here, we excluded only the outlier loci that coded for a nonsynonymous amino acid change. This is because most of these outlier studies were underpowered due to a limited sample size, and, in fact, never identified the same set of loci (Csilléry et al., 2020). Further, even if some of the variants may code for phenotypic traits under selection, their effect sizes are probably too small to matter for estimating kinship at a short spatial scale. Taking together all of the above considerations, we could use a common set of 137 SNPs for the adult trees, and 116 SNPs for the seedlings.



FIGURE 1 (a) Location of the study sites (main map, yellow triangles) and distribution of silver fir (*Abies alba* Mill.) (dark green). Small maps show the position of the high and low elevation populations within each site. Site names are abbreviated using three letter codes; see Table 1 for full names. (b) Genetic clusters across the study sites inferred using a discriminant analysis of principal components (DACP) and the R package adegenet (Jombart, 2008). Clusters are represented in the space defined by the first two linear discriminant (LD) functions. The eight sites were marked with different colors (sites VEN and ISS overlap in the figure). (c) Isolation-by-distance in the studied populations

2.2 | Environmental variables

The habitat at each site was characterized using a combination of climate, soil, and species distribution data. Downscaled historical climate data at 1 km resolution was obtained from the CHELSAcruts time-series (Karger et al., 2017; https://chelsa-climate.org/chels acruts/) from 1901 to 1990. Monthly mean, minimum, and maximum temperatures and monthly precipitation were extracted and used to calculate the annual mean temperature, isothermality, and annual total precipitation using the R package dismo (Hijmans et al., 2017). We calculated only these climatic variables because they have been shown to drive adaptation in silver fir (e.g., Csilléry et al., 2020; Gazol et al., 2015), and we had to limit the number of variables to be tested because of the limited number of observations (16 populations). Soil available water capacity (AWC) was extracted at a 250 m resolution from the SoilGrids data set (Hengl et al., 2017). Measures of tree species composition were derived from the EU-Forest, which is a 1 km resolution tree occurrence database for Europe (Mauri et al., 2017). The occurrences of 14 tree species within areas of 0.3 radius circles (~33 km) around each study populations were extracted.

2.3 | Statistical analyses

First, we assessed genetic diversity parameters and IBD patterns across the species range. We characterized genetic diversity and divergence at the level of sites and populations. Expected heterozygosity ($H_{\rm F}$) and observed heterozygosity ($H_{\rm O}$) were calculated for each site, population and cohort (i.e., adults or seedlings) using SpaGeDi 1.5b (Hardy & Vekemans, 2002), as well as pairwise F_{ST} among sites, between low and high elevation population pairs and between cohorts. Statistical significance of pairwise $F_{\rm ST}$ values was tested by 10,000 permutations. We performed a hierarchical analysis of molecular variance (AMOVA) to determine the partition of the genetic variation within and among sites and populations using the R package pegas (Paradis, 2010). We applied discriminant analysis of principal components (DACP) to identify genetic clusters (Jombart et al., 2010) implemented in the R package adegenet (Jombart, 2008). DACP identifies synthetic variables, so called Linear Discriminants (LD), that differentiate among a priori defined groups (here, populations), while minimizing variation within inferred genetic clusters. We used the function Xval.dapc to find the optimal number of principal components (PCs) to be retained, and find.clusters to find the

					Elevation (m)	Sample size		Max distan	ce (m)
Site ID	Country	Site name	Latitude	Longitude	Low	High	Low	High	Low	High
РУК	France	Ossau Valley (Pyrenees)	42.8550	-0.4578	825	1562	81	82	198	444
VEN	France	Ventoux (Alps)	44.1812	5.27043	995	1340	214	250	242	187
LUR	France	Lure (Alps)	44.1142	5.83912	1410	1628	55	55	82	175
ISS	France	Issole (Alps)	44.0242	6.46244	1108	1585	49	47	92	68
VES	France	Vesubie (Alps)	43.9707	7.36577	1078	1497	43	45	121	62
APE	Italy	Bosco Martese, Ceppo (Central Apennine)	42.6688	13.4322	1375	1600	46 (98)	47 (57)	198	234
BAV	Germany	Bavarian Forest	48.9468	13.4040	770	1120	100 (148)	92 (143)	1397	1031
FAG	Romania	Arges (Fagaras Mountain)	45.4411	24.6947	1070	1410	95 (46)	94 (48)	986	802

- MOLECULAR ECOLOGY - WILEY

optimal number of clusters based on the Bayesian information criterion (BIC). We used the number of populations (16) as the maximum number of clusters. The cross-validation procedure suggested that it was sufficient to include the first 120 PCs. Although the lowest BIC was observed with a grouping in nine clusters, the drop in BIC values was no longer marked after six clusters (Figure S2), where LUR, ISS, and VES belonged to the same "cluster 5" (Figure S2). Since DAPC can be sensitive to sampling density, we also carried out the DAPC analysis using only the two most distant French populations (VEN and VES). Finally, we plotted $F_{sT}/(1-F_{sT})$ against pairwise geographic distances among populations to illustrate the range-wide IBD pattern, but we did not perform a formal Mantel test because the spatial scale was several orders of magnitude larger than the dispersal distance of the species and because the studied populations experienced different long-term demographic dynamics and originated from different glacial refugia (Liepelt et al., 2002, 2009).

Second, FSGS was characterized using two methods, the Sp statistic (Vekemans & Hardy, 2004) and multisite spatial autocorrelation analyses (Smouse et al., 2008). Sp is estimated from a regression of pairwise kinship coefficients against the logarithm of pairwise spatial distances as $-b - log/(1-F_1)$, where b - log is the slope of the regression and F_1 is the average kinship coefficient among individuals of the first distance class (Vekemans & Hardy, 2004). We estimated Sp for each population and each age cohort using the software SpaGeDi 1.5 b. We used the pairwise kinship coefficient (F_{ii}) of Loiselle et al., (1995), both with even distance classes (20 m) and even sample sizes per distance class. The spatial extent of FSGS was estimated as the distance where F_{ii} was significant with the even sample size method, as this option ensured that an adequate number of individuals were placed in each distance class. The statistical significance of F_{ii} , F_1 , and *b*-log was tested using 10,000 permutations of individual locations among individuals. Multisite spatial autocorrelation analyses of allele frequencies were performed using GenAlEx 6.5 (Peakall & Smouse, 2012) using even distance classes (20 m). The statistical significance of the autocorrelation was determined using 999 permutations by randomizing genotypes among distance classes. We also applied the non-parametric heterogeneity test described by Smouse et al., (2008) to test the statistical significance of the spatial autocorrelation patterns observed among populations.

Third, we tested the sensitivity of FSGS to sample size and sampling configuration using resampling. For this purpose, we chose VEN_H and VEN_L populations because of their high sample sizes (Table 1). The resampling was performed by applying three strategies: (1) resampling randomly within the sampling area, (2) reducing the size of the sampling area, and (3) resampling along linear transects. The first resampling approach was performed selecting 50 random individuals 20 times using R (R Core Team, 2019). The second resampling approach was carried out in the program QGIS (QGIS Development Team, 2016), where equal-sized (0.45 ha for VEN_H and 0.65 ha for VEN_L), choosing 20 circles with random centres placed within the central area of the population, so that the whole circle could always be inside the population. Finally, the third resampling approach required sampling across 20 linear transects,

Geolocalization of the eight sampling sites, sample sizes, and the maximum distance between sampled adult trees in metres. At each site, a low and a high population was sampled

TABLE 1

II FY-MOLECULAR ECOLOGY

TABLE 2 Genetic diversity and summaries of the fine-scale spatial genetic structure (FSGS) in silver fir across eight sites with a high (_H) and low (_L) elevation populations from each. Statistics are also reported for adult trees, and, at three sites, for seedlings

Populations	H _E	Н _о	HWE (%)	F ₁	b-log	Sp	NS	Extent (m)
PYR_H	0.23	0.22	4	0.021	-0.009***	0.009	108	45
PYR_L	0.23	0.23	10	0.004	-0.002	0.002	416	-
VEN_H	0.29	0.29	5	0.011	-0.006***	0.006	163	41
VEN_L	0.30	0.29	4	0.008	-0.002***	0.002	414	36
LUR_H	0.30	0.30	3	0.016	-0.007**	0.007	134	36
LUR_L	0.29	0.29	1	-0.001	-0.002	0.002	553	-
ISS_H	0.29	0.28	4	0.012	-0.013***	0.013	78	17
ISS_L	0.29	0.29	1	0.002	-0.004*	0.005	222	-
VES_H	0.30	0.30	3	0.001	-0.003	0.003	328	-
VES_L	0.29	0.29	1	-0.005	0.002	-0.001	00	-
APE_H	0.29	0.28	3	0.014	-0.004	0.004	215	28
APE_H seedling	0.28	0.29	3	0.012	-0.007***	0.007	156	36
APE_L	0.29	0.27	5	0.009	-0.002	0.002	413	23
APE_L seedling	0.28	0.28	5	0.015	-0.009***	0.009	109	42
BAV_H	0.32	0.33	1	0.034	-0.004***	0.004	260	121
BAV_H seedling	0.33	0.31	9	0.086	-0.014***	0.014	72	81
BAV_L	0.32	0.33	7	0.013	-0.003***	0.003	319	76
BAV_L seedling	0.32	0.32	5	0.045	-0.008***	0.008	118	87
FAG_H	0.23	0.22	4	-0.010	0.001	-0.001	8	-
FAG_H seedling	0.22	0.24	1	NA	NA	NA	NA	-
FAG_L	0.23	0.22	5	0.042	-0.002	0.002	478	-
FAG_L seedling	0.21	0.21	3	NA	NA	NA	NA	-

 $H_{\rm E}$, expected heterozygosity; $H_{\rm O}$, observed heterozygosity; HWE, percentage of loci that significantly deviated from the Hardy-Weinberg equilibrium (R package pegas, Paradis, 2010); F_1 , mean kinship of the first distance class (0–20 m), *b-log*, regression slope of kinship (*p < .05, **p < .01,

*** p < .001); NA, not available because the spatial clustering was too strong (see Materials and Methods for details); NS, neighbourhood size.

Extent (m) is the extent of FSGS in metres estimated as the distance where Fij was significant with the even sample sizes method.

randomly drawn across each population using the R packages "sf" (Pebesma, 2018), "sp" (Pebesma & Bivand, 2005), and raster (Hijmans et al., 2015). The maximum and minimum coordinates within each population were used to create the transect, and then the transect was rotated 20 times by 9° relative to the centre of the transect. FSGS analyses of the resampled data sets were carried out using an in-house python script that automated SpaGeDi, and using the even sample size option. We tested whether the resampling would have changed our conclusions in terms of the strength and extent of FSGS using one sample t test, assuming that the estimates from the full data sets were the true values. Finally, we assessed the effect of sampling using all populations. For this, we calculated the correlation between summary statistics of FSGS (F1, b-log, Sp) and summaries of the sampling design, such as the area of the sampling site (m^2) , the maximum distance among the furthest individuals (m), and the density of sampling (number of samples/area of the sampling site) using R.

Fourth, we used a structural equation modelling (SEM) approach to identify the potential demographic and environmental drivers of FSGS using the lavaan R package (Rosseel, 2011). Since we had only 16 populations, we were able to test a limited number of variables. To partially circumvent this problem, we defined two models. In the first model, we tested the effect of demography. We included synthetic variables from the DAPC analyses (i.e., the first and second linear discriminant functions, LD1.DAPC and LD2.DAPC) as proxies for long-term demography, and elevation as a binary variable for short-term demography. In the second model, we tested the effect of the environmental variables (temperature, isothermality, precipitation, AWC and species composition). We simplified the occurrence data of 14 tree species using a principal component analysis (PCA) and the prcomp function in R. The first three axes explained 78% of the variance of the original variables, thus we retained only these for the SEM analyses (hereafter PC1.spcomp, PC2.spcomp, and PC3.spcomp). PC1.spcomp was mainly determined by the presence of Acer pseudoplatanus, Betula pendula, Fagus sylvatica, Pinus nigra, Quercus pubescens and Sorbus aucuparia. PC2.spcomp was dominated by the presence of Castanea sativa, Betula pendula, Picea abies, Pinus halepensis and Quercus ilex, while PC3.spcomp was dominated by Acer pseudoplatanus, Castanea sativa, Populus tremula and Quercus robur (Figure S3). All variables except species composition were normalized using the scale function in R. For both models, we started with an initial model where several plausible functional relationships

were hypothesized between *Sp* and explanatory variables. Then, nonsignificant paths and variables were excluded step-by-step until the best explanatory model was found. Based on Grace (2006) and Kline (2015), the best-fit model was selected using the following indices: Chi-square test (p > .05), the comparative fit index (CFI > 0.9), Bayesian information criterion, standardized root mean square residual (SRMR < 0.06) and root mean square error of approximation (RMSEA < 0.05).

Fifth, we tested for the presence and strength of FSGS in seedlings using the same method as described for the adult trees at the three sites where data for seedlings were available (APE, BAV, and FAG). We also tested for the heterogeneity in correlation of allele frequencies between cohorts (Smouse et al., 2008). Seedlings from FAG were excluded from these analyses because regeneration was limited to four patches in FAG_H and one patch in FAG_L (see Figure S1). In order to estimate dispersal parameters, we carried out parentage analysis. We used the likelihood-based approach implemented in the software CERVUS 3.0.7. (Kalinowski et al., 2007; Marshall et al., 1998). We used the "parent-pair" option with unknown sexes and the 95% or "strict" confidence levels. Simulation parameters were set as follows: simulation of 50,000 offspring, individual genotypes at a minimum of 68 loci, and a genotyping error rate of 0.01 (conform with the high precision of the KASP chemistry). The proportion of candidate parents included in the sample had a great influence on the proportion of assigned parent-offspring pairs. In order to choose the most probable value for each population, we performed the analysis with values of 50, 60, 70, 80, and 90% of candidate parents, and recorded the parent-offspring mismatch rates in each. For the final analysis, we used the highest proportion of candidate parents that gave the lowest mismatch rate across the tested values (Figure S4). We used the reconstructed genealogies of the offspring, together with the coordinates of both parents and offspring, to estimate (1) selfing rate; (2) mean variance and maximum reproductive success among all adult trees, defined as the number of gametes produced by each adult; (3) gametic gene flow rate, defined as the number of gametes that originated from outside the sampling area over the total number of gametes sampled; and (4) average parent-offspring distance. We also plotted and visually compared the distribution of individual reproductive success among populations.

3 | RESULTS

3.1 | Genetic diversity and divergence across sites and populations

Genetic diversity was the highest in the Bavarian (BAV) populations ($H_E = 0.32$) and the lowest in the eastern and western range margin (FAG and PYR, $H_E = 0.23$, Table 2) populations. The hierarchical AMOVA revealed that most of the variance in allele frequencies was within populations (82.6%, $F_{ST} = 0.174$, p < .001, Table S1). However, a strong and significant genetic differentiation was detected among

sites (14.8%, $F_{CT} = 0.148$, p < .001). In contrast, the differentiation between high and low elevation populations within sites was low, albeit significant (3.1%, $F_{SC} = 0.0308$, p < .001, Table S1). The most genetically differentiated pairs of sites were also the most geographically distant ones (Figures 1b and c, Table S2): PYR and FAG ($F_{ST} = 0.275$), and PYR and APE ($F_{ST} = 0.216$).

The DAPC analysis revealed that the first linear discriminant function (LD1) ordered the populations from east to west with the exception of PYR, which situated close to the French populations (VEN, LUR, ISS, VES) (Figure 1a). The second linear discriminant function (LD2) mainly separated the population from the Pyrenees (PYR) from the other populations (Figure 1b). The following linear discriminants appeared difficult to be interpreted (Figure S5). Although genetic clustering methods are known to be sensitive to sample size, the same clustering was obtained even after having excluded LUR and ISS, which balanced out the sample size (Figure S5). Finally, a rather clear IBD pattern was found across the species range (Figure 1c).

3.2 | FSGS across sites and populations in adult trees

We found a significant FSGS in eight out of 16 populations based on the *b*-log statistic (Table 2), and in seven populations based on the *r* statistic in the first distance class (0–20 m) (Figure 2). Based on the two tests taken together, populations from the sites BAV, ISS, and VEN, as well as LUR_H and PYR_H showed significant FSGS. Significant *Sp* values ranged from 0.003 to 0.013 (Table 2). The average extent of FSGS was 47 m (range: 17–121 m) across all populations (Table 2). The spatial autocorrelation of allele frequencies did not differ globally among sites based on a test of heterogeneity ($\omega = 145$, p = .64), while, in population-wise comparisons, APE_L differed significantly from several other populations based on the heterogeneity test (Table S3).

3.3 | The effect of sampling scheme on FSGS

With all three methods of resampling, we obtained FSGS correlograms similar to those based on the full data set (Figure 3). Nevertheless, the *Sp* values based on a reduced number of samples did not always reflect the significant *Sp* values observed using the full data set, both in VEN_L and VEN_H (Table 2). While reduced area and linear transect resampling resulted in significant *Sp* values in 83% and 88% of the total number of resampling, respectively, random resampling showed significant FSGS only in 46% of the cases. In terms of the absolute value of *Sp*, all resampling methods provided consistent results with those obtained using the full data set in VEN_H (Figure 3, one-sample *t* tests, *p* > .76; Table S4). In contrast, in VEN_L only random resampling method provided the same results based on the full data set (one-sample *t* tests, *p* = .64), while both reduced area and linear transect resampling overestimated the *Sp*



FIGURE 2 Fine-scale spatial genetic structure (FSGS) across 16 silver fir (*Abies alba* Mill.) populations based on 137 SNPs using the software GenAlEx 6.5 (Peakall & Smouse, 2012). Correlation coefficients (*r*) are plotted against the maximum geographical distance between individuals in each distance class. Shaded areas represent the 95% confidence interval obtained through random shuffling of individual geographic locations (1,000 times). Vertical bars around mean *r* values represent 95% confidence intervals generated by bootstrapping (10,000 times) pairwise comparisons within each distance class. Note that in FAG_L we could not estimate the confidence intervals in the first distance class due to the limited number of individuals

values (Figure 3, p < .001; Table S4). Finally, no correlation was found between the observed F_{ij} , F_1 , *b-log*, or *Sp* values in the 16 populations and the area of the sampling site, the maximum distance among individuals, and the density of sampling (Pearson correlation < 1.59, p > .13).

3.4 | Demographic and environmental drivers of FSGS in adult trees

The best SEM model including demographic variables explained 47% of the variation in Sp values across the 16 populations (Table S5). Sp was significantly affected by elevation (High/Low), with high elevation populations exhibiting higher Sp values, and by the first discriminant function (LD1.DACP), with western populations having a stronger FSGS than eastern ones (Figure 4a). The best environmental SEM model explained 58% of the variation in Sp values (Table S6). The environmental variables that influenced Sp the most were elevation and isothermality (Figure 4b). Both effects were positive, thus FSGS was stronger in populations at higher elevations and where the diurnal temperatures oscillations were larger compared to seasonal variations. Although with weaker effects, AWC and species composition (PC2.spcomp) also influenced Sp values. Species composition itself was impacted by elevation, AWC, temperature and precipitation, thus these variables also had an indirect effect on FSGS (Figure 4b).

3.5 | Genetic diversity and FSGS in seedlings and dispersal parameters

Genetic diversity in seedlings was overall similar to that observed in adult trees (Table 2). We found weak, but significant genetic divergence between adult and seedling cohorts in APE and FAG, with F_{ST} values ranging from 0.002 to 0.007 (Table S7), but not in BAV. FSGS in the seedling cohort was significant in all populations (APE_H, APE_L, BAV_H, BAV_L), based on both the *b-log* and r statistics (Figure 5, Table 2). Overall, FSGS in seedlings (average Sp = 0.01, Table 2) was stronger than in adults (two sample *t* test, t(18) = -2.87, p = .01). In contrast, the heterogeneity test of spatial autocorrelation between adult and seedling cohorts was not significant in any of the populations (Table S8). Finally, the extent of FSGS in seedlings ranged between 36 and 87 m (average of 62 m), thus slightly higher than in adults.

Parentage analysis revealed a strong reproductive skew in all six populations (Table 3, Figure 6). Only 54%, 25% and 6% of the adults in BAV, APE, and FAG contributed to the generation of sampled

seedlings, respectively. The maximum individual reproductive success was recorded in APE_L, with 18 offspring assigned to a single adult tree (Table 3). The variance in individual reproductive success was similar in BAV_H and BAV_L (1.8) and APE_L (1.7), while lower in APE_H (0.5) and the lowest in FAG_H and FAG_L (0.1). Self-pollination occurred in APE_L and BAV_H and BAV_L, and was as high as 9% in BAV_H. The gametic gene flow rates were the highest in FAG, 87 and 95% in FAG_H and _L, respectively, because we could not identify the parents of most seedlings, and the lowest in BAV_H and BAV_L, but still 41 and 40%, respectively (Table 3). The mean parent-offspring distance varied among the three sites (APE: 40 m, BAV: 72 m, FAG: 130 m), but it was similar between low and high populations (Table 3), and comparable to the extent of FSGS estimated from the adult and seedling data (Table 2).

4 | DISCUSSION

4.1 | Genetic diversity of silver fir populations across the species range

Consistently with its large effective population size, N_{a} , and its wind pollinated outcrossing mating system (Ennos, 2001; Hardy et al., 2005; Vekemans & Hardy, 2004), we found high levels of genetic diversity across the whole range of silver fir (average $H_{\rm F}$ of 0.3; see Table 2). As changes in genetic diversity scale with 2N_e generations (Ellegren & Galtier, 2016), these high levels of genetic diversity most probably still reflect the diversity of refugial populations during the glacial cycles (Petit & Hampe, 2006). Populations from the French Mediterranean Alps (VEN, LUR, ISS, VES) that have been colonized from main effective refugium situated in the northern Apennines exhibited high levels of diversity (Table 2). The population from the central Apennines, APE, also exhibited a relatively high level of diversity (Table 2). This region probably represents a separate, smaller refugium that evolved independently from populations in the northern and southern Apennines (Piotti et al., 2017). In our study, APE exhibited a stronger similarity to the population from the Carpathians, FAG, than to those from the French Mediterranean Alps (Figure 1b), which is in agreement with some other studies (Brousseau et al., 2016; Martínez-Sancho et al., 2021), but our scattered sampling is clearly not suitable to refine the demographic history of silver fir. Genetic diversity was the lowest in the westernmost (Pyrenees, PYR) and the easternmost (Carpathians, FAG) extremes of the distribution range (Figure 1 and Table 1). The low genetic diversity of these populations are most probably the results of genetic drift due to founder effects, long term isolation or population size reduction due to recurring unfavourable environmental conditions (Excoffier



5257

FIGURE 3 Assessment of the sampling configuration on inferences about the fine-scale spatial genetic structure (FSGS) using two silver fir (*Abies alba* Mill.) populations, VEN_H and VEN_L, and the software SpaGeDi 1.5b (Hardy & Vekemans, 2002). Three spatial resampling methods have been tested: random (a), reduced area (b), and linear transect (c). For each of the three methods: Maps illustrate the resampling method and correlograms show the average kinship coefficients plotted against the mean distance between individuals. Full lines indicate the mean of 20 resamplings and dotted lines indicate the lower and higher limits of kinship values. Box plots show the distribution of *Sp* statistics after resampling. Black diamonds on boxplots illustrate the original *Sp* of all samples

et al., 2009). In particular, silver fir populations from the Pyrenees have long been isolated from the rest of the range as suggested by the presence of macroremains older than 15 kyr BP (Terhürne-Berson et al., 2004) and recent genetic study (Scotti-Saintagne et al., 2021). PYR is also the farthest site from the largest effective refugia of the multiple glacial cycles situated in the Apennines. Also, the low diversity in FAG appears plausible since the Carpathians have been colonized during the Holocene from a relatively small refugium situated on the Balkan (Gömöry et al., 2004, 2012; Liepelt et al., 2010; Tinner et al., 2013). In this study, the highest diversity was observed at the German site, BAV, situated north of the Alps. These populations might have also been colonization from the northern Apennines but via another colonization route through the Dinaric Alps (Liepelt et al., 2009). Here, we found that BAV clustered together with FAG and APE (Figure 1b), thus we may speculate that different lineages might have admixed north of the Alps leading to an elevated level of diversity (Petit et al., 2008).

4.2 | FSGS across the species range and its determinants

We found that silver fir was characterized by a moderate and significant FSGS at most parts of the species range (Figure 2, Table 2). Among the previous studies of FSGS in silver fir, Sagnard et al. (2011) and Leonarduzzi et al. (2016) found a weak or absent FSGS in French and Central Italian populations, respectively, while a relatively strong FSGS was reported from populations from the Italian Alps (Mosca et al., 2018) and the Western Carpathians (Paluch et al., 2019). Based on our results together with the above mentioned studies, Sp in silver fir ranges from 0 to 0.013 with a global mean Sp of 0.005 (CI95% [-0.001, 0.01]). Silver fir is characterized by a long generation time, and heavy seeds (Royal Botanic Gardens Kew, 2015) and pollen grains (Eisenhut, 1961), which could contribute to a limited dispersal (Amm et al., 2012), and to the maintenance of FSGS across generations. Further, silver fir often occurs in association with other species such as Norway spruce and European beech, which causes the isolation from conspecifics and the clustering of relatives (Aussenac, 2002; Dobrowolska et al., 2017), thus the maintenance of FSGS.

In this study, we identified both demographic and environmental drivers of FSGS across the species range. Among the demographic proxies, not surprisingly, the most recent demographic event, that is, the supposed colonization of high elevation habitats, left the strongest signal on FSGS (Figure 4a). Nevertheless, the first discriminant function, ordering populations along an east-west biogeographical gradient, also left a detectable signature on FSGS (Figure 4a),

suggesting that FSGS may conserve the signature of demographic events on deep demographic time scales, potentially reaching back to the LGM. These results confirm that demographically younger populations are more likely to exhibit a spatial genetic structuring, i.e. higher elevation and, to some extent, populations derived from the western lineage, were more likely to exhibit a stronger FSGS. Several previous studies have already demonstrated that recently colonized populations exhibit a stronger spatial structure than core populations (e.g., De-Lucas et al., 2009; Gapare & Aitken, 2005; Pandey & Rajora, 2012). This phenomenon can be mediated by the local population size, i.e. the strength of local genetic drift and/ or the population density (Hamrick et al., 1993; Young & Merriam, 1994). Our results are also concordant with independent evidence about recent range expansion of the species during the past decades (Vitasse et al., 2019). While silver fir has been declining in warm and dry areas, especially at the limit of its distribution range and in low elevations and in the Mediterranean (e.g., Dobrowolska et al., 2017; Gazol et al., 2015; Hernández et al., 2019), the species has been reported to expand towards high elevations across the whole range (Chauchard et al., 2010; Hernández et al., 2019; Vitasse et al., 2019). The reasons for this expansion are diverse and complex, and include a decrease of human activities at high elevations and climatic warming since 1970s, which opened new habitats for silver fir (Chauchard et al., 2010). Further, the expansion of silver fir might have also been facilitated by the reintroduction of large predators in the Alps, such wolf and lynx, that in several cases reduced ungulate numbers (Apollonio et al., 2010), thereby the browsing pressure.

In this study, we also searched for environmental drivers of FSGS and found that the model including habitat variables explained 11% more variance in FSGS that the model including only the three demographic proxies (Figure 4a). In contrast, we found that, among habitat variables as well, elevation was the most important driver of Sp, more important than climate or species composition (Figure 4b). Although these results emphasize that differences in Sp values were more likely reflecting demographic effects than differences in the environmental conditions, high elevation populations can also be characterized by harsher environmental conditions, thus we could not exclude that climatic constrains or adaptation might also have contributed to the higher FSGS observed at high elevations. For example, many of our high elevation sites had a high snow cover in winter and were situated on scree slopes (e.g., VEN, LUR, ISS). We also found a positive effect of isothermality, with larger variances in temperatures associated with a higher FSGS. The other climatic variables had an indirect effect on FSGS, through their influence on species composition (Figure 4b). We note that high isothermality was a good indicator of continental climate, thus this effect could also



FIGURE 4 Path diagram of the two final structural equation models (SEM). (a) SEM estimating the causal effects of demography including high/low status and the first two linear discriminants (LDs) from the DAPC analysis on *Sp*. (b) SEM estimating the causal effects of Temperature, Isothermality, Precipitation, soil available water capacity (AWC) and elevation on *Sp* and on species composition (PC2. spcomp) synthetised as the second principal component of all species composition variables. We also hypothesized an indirect effect of the explanatory variables on *Sp* via PC2.spcomp. Lines in blue correspond to a positive effect (or a positive correlation between variables) whereas lines in red correspond to a negative effect or correlation. Colour intensity is positively related to the significance of the relationship between two variables. Standardized path coefficients between variables are indicated on the arrows



FIGURE 5 Fine-scale spatial genetic structure (FSGS) across four silver fir (*Abies alba* Mill.) population pairs, comprising adult and seedling cohorts, using the software GenAlEx 6.5 (Peakall & Smouse, 2012). Correlation coefficients (*r*) were plotted against the maximum geographical distance between individuals in each distance class. Full lines refer to autocorrelation analyses of seedlings and dashed lines refer to autocorrelation analyses of adult trees

simply coincide with the east-west demographic axis. Mosca et al. (2018) studied the role of climatic variables on FSGS in silver fir, and found a weak negative correlation between FSGS and spring precipitation in the Italian Alps. At a local scale, different climatic factors could indeed play a role in the strength of FSGS. However, regional studies can be limited in revealing the drivers of FSGS because the environment of each population is only compared to similar populations from the same region.

4.3 | Limitations of this study: The effect of sampling

Sampling is a key issue in all IBD and FSGS studies, where separating biological patterns from sampling artifacts can be challenging (Legendre & Fortin, 2010; Meirmans, 2012; Smouse et al., 2008; Vekemans & Hardy, 2004). This study had to face this challenge in particular because it was aimed at understanding both range-scale

and population-scale processes, as well as their interaction. At the range-scale, although our sites represented most genetic lineages, several parts of the range were underrepresented (Figure 1). Thus, it is possible that we did not have sufficient power to tease apart the effects of demography and the environment. Measures of FSGS are also sensitive to several aspects of sampling, such as sampling area, configuration, and density, which makes among-population comparisons particularly challenging (Smouse et al., 2008; Vekemans & Hardy, 2004). Although the global test of heterogeneity in spatial autocorrelation of allele frequencies did not show evidence for variation in FSGS across the range, we argue that this result appeared rather conservative given the strong population differences in Sp values and their clear link to demography (Table 2, Figure 4). Unfortunately, little information is available about the performance of this test across a large number of populations. We found that a resampling experiment on our own data can be recommended for understanding the limitations of a data set. The main conclusion from our experiment was that when tree density was low (VEN_L),

MAJOR ET AL.

MOLECULAR ECOLOGY – W

TABLE 3 Summaries of the parentage assignment per population using CERVUS 3.0.7. (Kalinowski et al., 2007; Marshall et al., 1998). The mean reproductive success is the mean number of offspring per adult tree. The variance in reproductive success is the variance of the number of offspring across all adult trees. The gametic gene flow rate was estimated as the number of gametes that originated from outside the sampling area (i.e., number of gametes for which no parent was found within the population) over the total number of gametes sampled (i.e., twice the number of seedlings) (Valbuena-Carabaña et al., 2005)

Summary level	Dispersal parameter	APE _H	APE _L	BAV _H	BAV _L	FAG _H	FAG _L
Seedlings	Both parents identified (trios) (%)	8	18	35	33	0	0
	One parent identified (%)	28	43	48	54	10	26
	Self pollination (%)	0	3	9	4	0	0
Adults	With offspring (parents) (%)	23	26	49	59	3	8
	Mean reproductive success	0.5	1.7	1.8	1.8	0.1	0.1
	Variance in reproductive success	3.0	18.4	7.3	5.5	0.1	0.2
	Maximum reproductive success	11	18	11	11	3	3
Populations	Gametic gene flow rate	0.77	0.60	0.41	0.40	0.95	0.87
	Mean parent-offspring distance (m)	44	35	77	66	119	141

FIGURE 6 (a) Maps of all sampled adults (triangles with size proportional to the diameter at breast height) and seedlings (dots) and parent-offspring relationships (black arrows) inferred using CERVUS 3.0.7. (Kalinowski et al., 2007; Marshall et al., 1998) at three silver fir (*Abies alba* Mill.) sampling sites, each represented by its high (_H) and low (_L) elevation populations. (b) The distribution of reproductive success (i.e., the number identified seedling per adult tree within populations) in the same six populations with matching colour codes



5259

WII FY-MOLECULAR ECOLOGY

the reduced area or linear transect sampling led to an overestimation of *Sp* values (Figure 3b and c). In contrast, the FSGS analyses appeared robust to any kind of resampling when there was a high tree density (VEN_H, Figure 3). Nevertheless, when we considered the observed data from all 16 populations, we found no evidence for a consistently higher *Sp* in populations where the sampling area was reduced, such as LUR, ISS or VES, and we found a strong and significant FSGS in BAV, where the sampling density was low (Figure S1, Table 2, Table S4). Finally, we acknowledge that the sampling error of *Sp* can be very high (see error bars on Figure 3), so we reiterate the warnings of previous studies that large sample size and several populations across the range are needed to draw robust conclusions (Smouse et al., 2008; Vekemans & Hardy, 2004).

4.4 | The maintenance of FSGS across generations

In small and isolated populations, elevated inbreeding levels might reduce fitness of subsequent generations and threaten forest resilience in the long term (González-Díaz et al., 2017; Restoux et al., 2008). Comparing FSGS between life stages (e.g., adult and juvenile cohorts) may identify when populations have undergone disturbances and help understand the intrinsic mechanisms underlying patterns of FSGS (Segelbacher et al., 2010). Here, we found that Sp was significantly stronger in the seedling cohorts compared to adults (Figure 2 and 5, Table 2). This result probably reflects seed shadows around adult trees (González-Martínez et al., 2002; Ng et al., 2004; Troupin et al., 2006). Parentage analyses also supported the existence of seed shadows, especially at high elevations: estimates of the gametic gene flow were consistently higher in the higher elevation populations than in the lower ones (Table 3). This result is in line with our hypothesis that higher elevation populations were more recently colonized, but may also suggest populations at low or intermediate elevations act as continuous sources of gene flow for higher elevation populations (Herrera & Bazaga, 2008). In BAV, seedlings mainly originated from parent trees within the sampling area and the proportion of selfing was also high (Table 3). This result could reflect the low silver fir density in Bavaria, where the proportion of Norway spruce (Picea abies K.) is the highest among all studied sites. In contrast, at the high density APE site, regeneration to a large extent relied on pollen and seed immigration (Table 3). A similar effect of stand density on the outcrossing rate has been documented in Mediterranean silver fir stands (Restoux et al., 2008). In FAG, even though seedlings occurred in spatially aggregated regeneration spots, we were able to identify the parents of only a few seedlings (Table 3). Since we cannot exclude the possibility that parent trees have been cut, estimates of the dispersal parameters from FAG should be treated with caution. To conclude, the most recently colonized populations from Bavaria had the strongest FSGS at the seedling stage as well as the highest selfing rates. Thus this population could be less resilient to environmental stress in comparison to the other studied populations.

The average number of seedlings per adult tree at our sites varied between 0.1 and 1.8 seedlings, with a single tree having 18 seedlings

at the most extreme case in APE (Figure 6). Such pronounced reproductive skew is commonly observed in trees (e.g., Chybicki & Burczyk, 2013; Oddou-Muratorio et al., 2018), and can be driven by genetic or micro-environmental differences. For example, Avanzi et al. (2020) showed that in Norway spruce, trees that were able to maintain high-growth rates despite environmental growth limitations had the highest reproductive success. The assessment of reproductive success also greatly depends on the ontological stage. Indeed, it is well known that in perennials, the shape of the size frequency distribution changes greatly in populations through time, particularly when the density is high (e.g., Dodd & Silvertown, 2000; Herrera & Jovani, 2010). Gerzabek et al. (2017) studied maternal reproductive success across time in a oak stand, and found a marked inequality in reproductive success at the time of seedling emergence, that was considerably reshuffled by later seedling mortality. In our study, not all age classes were present at all sites, suggesting that the date of mast years and selective seedling mortality could have contributed to differences in the estimated reproductive success among sites (Davi et al., 2016). Overall it appears from our study that estimates of dispersal parameters, and also of FSGS in seedlings, are only informative for the FSGS of the next generation of adult trees when an extensive sampling of seedlings across several age classes is performed.

4.5 | FSGS as an indicator for monitoring genetic diversity?

Over the past decade, several studies and reviews have documented the apparent resilience of widespread tree species to the genetic consequences of habitat fragmentation and disturbance (Bacles & Jump, 2011; Kramer et al., 2008). Indeed, forest trees have several mechanisms to overcome genetic diversity loss and fitness reduction. For example, long distance pollen dispersal may assure connectivity among populations (Kremer et al., 2012), or overlapping generations could slow down the loss of genetic diversity (Austerlitz et al., 2000). In an idealized Wright-Fisher population the amount of neutral genetic diversity is determined by the effective population size, N_{a} , and its historical fluctuations (Ellegren & Galtier, 2016). However, in outcrossing wind-pollinated species with large N_{a} , such as many forest trees, monitoring the average genetic diversity can be uninformative because changes in the genetic diversity scale with $2N_{o}$ generations. It is often argued that populations with a higher genetic diversity may resist better or recover more quickly from a disturbance (Sgrò et al., 2011; Wernberg et al., 2018), and the average genetic diversity (Porth & El-Kassaby, 2014) or contemporary N_o (Hoban et al., 2020, 2021) are commonly proposed indices for diversity monitoring (but see also a critique by Fady & Bozzano, 2020). In this study, we found no significant difference in the average genetic diversity between low and high elevation populations, but we did in terms of FSGS. Other authors have also reported that forest management and disturbance regimes left a signature on FSGS but not on the average level of genetic diversity (Leclerc et al., 2015; Paffetti et al., 2012; Piotti et al., 2013; Sjölund & Jump, 2015). The analysis of the spatial distribution of

genetic diversity, that is, FSGS, allows to estimate the local equivalent of N_e , the neighbourhood size (*NS*) (Wright, 1943) as 1/*Sp* (Table 2). Thus, *NS* can be a useful indicator for genetic diversity monitoring in forest trees as it is more sensitive to recent demographic changes and disturbances than non-spatial genetic diversity indices.

ACKNOWLEDGEMENTS

EIM is supported by the Hungarian State doctoral programme implemented at the Hungarian University of Agronomy and Life Sciences, School of Horticultural Science. KC was supported by a Swiss National Science Foundation grant (CRSK-3_190288) while working on this manuscript. CA, AP and GGV have been partially supported by resources available from the Italian Ministry of University and Research (FOE-2019) under the project "Climate Change" (CNR DTA.AD003.474). For genotyping the seedlings, the authors acknowledge the financial support of the ERAnet BiodivERsA project TipTree (ANR-12-EBID-0003 granted to LO, BF and GGV) and the German Federal Ministry of Education and Research (01LC1202A granted to LO). For field and laboratory work in Romania we acknowledge the National Research Programme NUCLEU, Romania Ministry of Education and Research. We thank the Bavarian Forest National Park for supporting the fieldwork in Germany; the team of the INRAE UEFM in Avignon for sample collection and Anne Roig for laboratory work in France; the Gran Sasso e Monti della Laga National Park for logistic support, Cristina Leonarduzzi for help during the fieldwork, and Catia Boggi and Ilaria Spanu for technical assistance and laboratory work in Italy; the team of INCDS Simeria for fieldwork in Romania and Ana lordan for laboratory work. Open Access Funding provided by Lib4RI Library for the Research Institutes within the ETH Domain Eawag Empa PSI and WSL.

AUTHOR CONTRIBUTIONS

Katalin Csilléry, Mária Höhn, Bruno Fady and Giovanni G. Vendramin designed the study. Katalin Csilléry, Bruno Fady, Giovanni G. Vendramin, and Lars Opgenoorth raised funds. Lars Opgenoorth and Katrin Heer collected the seedling data in Germany (site BAV), Andrea Piotti and Camilla Avanzi in Italy (site APE), and Dragos Postolache and Flaviu Popescu in Romania (site FAG). Enikő I. Major performed all analysis with help from Katalin Csilléry. Enikő I. Major and Katalin Csilléry wrote the first draft, and all authors contributed to improve the final manuscript.

OPEN RESEARCH BADGES

This article has earned an Open Data badge for making publicly available the digitally-shareable data necessary to reproduce the reported results. The data is available at https://doi.org/10.5061/ dryad.7d7wm37vm and https://doi.org/10.5281/zenodo.5080520.

DATA AVAILABILITY STATEMENT

The following data have been made available on Dryad (https:// doi.org/10.5061/dryad.7d7wm37vm): Georeferenced adult and seedling SNP genotype data, coordinates used for the resampling study in VEN, the diameter at breast height (DBH) of adult trees (sites APE, BAV and FAG), individual coordinates of the resampled data sets (site VEN), occurrence of 14 tree species at the study sites derived from the EU-Forest database, input variables for the SEM analysis (Sp values, raw climatic variables from CHELSAcruts, AWC, elevation, tree species composition PCs, variables derived from the DAPC analysis), R script for the DACP analysis, R script for the SEM analyses, R script for the transect resampling have been placed in Zenedo (https://doi.org/10.5281/ zenodo.5080520).

ORCID

Enikő I. Major [©] https://orcid.org/0000-0003-4012-9856 Mária Höhn [®] https://orcid.org/0000-0002-8587-8271 Camilla Avanzi [®] https://orcid.org/0000-0003-1632-5304 Bruno Fady [®] https://orcid.org/0000-0003-2379-7617 Katrin Heer [®] https://orcid.org/0000-0002-1036-599X Lars Opgenoorth [®] https://orcid.org/0000-0003-0737-047X Andrea Piotti [®] https://orcid.org/0000-0002-3324-5325 Dragos Postolache [®] https://orcid.org/0000-0001-5250-8752 Giovanni G. Vendramin [®] https://orcid.org/0000-0001-9921-7872 Katalin Csilléry [®] https://orcid.org/0000-003-0039-9296

REFERENCES

- Aguillon, S. M., Fitzpatrick, J. W., Bowman, R., Schoech, S. J., Clark, A. G., Coop, G., & Chen, N. (2017). Deconstructing isolation-by-distance: The genomic consequences of limited dispersal. *PLoS Genetics*, 13(8), e1006911.
- Amm, A., Pichot, C., Dreyfus, P., Davi, H., & Fady, B. (2012). Improving the estimation of landscape scale seed dispersal by integrating seedling recruitment. *Annals of Forest Science*, 69(7), 845–856.
- Apollonio, M., Andersen, R., & Putman, R. (2010). European ungulates and their management in the 21st century. Cambridge University Press.
- Audigeos, D., Brousseau, L., Traissac, S., Scotti-Saintagne, C., & Scotti, I. (2013). Molecular divergence in tropical tree populations occupying environmental mosaics. *Journal of Evolutionary Biology*, 26(3), 529-544.
- Aussenac, G. (2002). Ecology and ecophysiology of circum-Mediterranean firs in the context of climate change. *Annals of Forest Science*, 59(8), 823–832.
- Austerlitz, F., Mariette, S., Machon, N., Gouyon, P. H., & Godelle, B. (2000). Effects of colonization processes on genetic diversity: differences between annual plants and tree species. *Genetics*, 154(3), 1309–1321.
- Avanzi, C., Heer, K., Buentgen, U., Labriola, M., Leonardi, S., Opgenoorth, L., & Piotti, A. (2020). Individual reproductive success in Norway spruce natural populations depends on growth rate, age and sensitivity to temperature. *Heredity*, 124(6), 685–698.
- Bacles, C. F., & Jump, A. S. (2011). Taking a tree's perspective on forest fragmentation genetics. *Trends in Plant Science*, *16*(1), 13–18.
- Barluenga, M., Austerlitz, F., Elzinga, J. A., Teixeira, S., Goudet, J., & Bernasconi, G. (2011). Fine-scale spatial genetic structure and gene dispersal in *Silene latifolia*. *Heredity*, 106(1), 13–24.
- Bessega, C., Pometti, C. L., Ewens, M., Saidman, B. O., & Vilardi, J. C. (2016). Fine-scale spatial genetic structure analysis in two Argentine populations of *Prosopis alba* (Mimosoideae) with different levels of ecological disturbance. *European Journal of Forest Research*, 135(3), 495–505.

II FY-MOLECULAR ECOLOGY

- Bosela, M., Popa, I., Gömöry, D., Longauer, R., Tobin, B., Kyncl, J., & Büntgen, U. (2016). Effects of post-glacial phylogeny and genetic diversity on the growth variability and climate sensitivity of European silver fir. *Journal of Ecology*, 104(3), 716-724.
- Bradburd, G. S., & Ralph, P. L. (2019). Spatial Population Genetics: It's About Time. Annual Review of Ecology, Evolution, and Systematics, 50(1), 427–449.
- Brousseau, L., Postolache, D., Lascoux, M., Drouzas, A. D., Källman, T., Leonarduzzi, C., Liepelt, S., Piotti, A., Popescu, F., Roschanski, A. M., Zhelev, P., Fady, B., & Vendramin, G. G. (2016). Local Adaptation in European Firs Assessed through Extensive Sampling across Altitudinal Gradients in Southern Europe. *PLoS One*, 11(7), e0158216.
- Budde, K. B., González-Martínez, S. C., Navascués, M., Burgarella, C., Mosca, E., Lorenzo, Z., Zabal-Aguirre, M., Vendramin, G. G., Verdú, M., Pausas, J. G., & Heuertz, M. (2017). Increased fire frequency promotes stronger spatial genetic structure and natural selection at regional and local scales in *Pinus halepensis* Mill. *Annals of Botany*, 119(6), 1061–1072.
- Burga, C. A., & Hussendörfer, E. (2001). Vegetation history of Abies alba Mill. (silver fir) in Switzerland-pollen analytical and genetic surveys related to aspects of vegetation history of Picea abies (L.) H. Karsten (Norway spruce). Vegetation History and Archaeobotany, 10(3), 151–159.
- Chauchard, S., Beilhe, F., Denis, N., & Carcaillet, C. (2010). An increase in the upper tree-limit of silver fir (*Abies alba* Mill.) in the Alps since the mid-20th century: A land-use change phenomenon. *Forest Ecology and Management*, 259(8), 1406–1415.
- Cheddadi, R., Birks, H. J. B., Tarroso, P., Liepelt, S., Gömöry, D., Dullinger, S., Meier, E. S., Hülber, K., Maiorano, L., Laborde, H. (2014). Revisiting tree-migration rates: Abies alba (Mill.), a case study. Vegetation History and Archaeobotany, 23(2), 113–122. http://dx. doi.org/10.1007/s00334-013-0404-4
- Chybicki, I. J., & Burczyk, J. (2013). Seeing the forest through the trees: comprehensive inference on individual mating patterns in a mixed stand of *Quercus robur* and *Q. petraea*. Annals of Botany, 112(3), 561–574.
- Csilléry, K., Ovaskainen, O., Sperisen, C., Buchmann, N., Widmer, A., & Gugerli, F. (2020). Adaptation to local climate in multi-trait space: evidence from silver fir (*Abies alba Mill.*) populations across a heterogeneous environment. *Heredity*, 124(1), 77–92.
- Davi, H., Cailleret, M., Restoux, G., Amm, A., Pichot, C., & Fady, B. (2016). Disentangling the factors driving tree reproduction. *Ecosphere*, 7(9), e01389.
- De-Lucas, A. I., González-Martínez, S. C., Vendramin, G. G., Hidalgo, E., & Heuertz, M. (2009). Spatial genetic structure in continuous and fragmented populations of *Pinus pinaster* Aiton. *Molecular Ecology*, 18(22), 4564–4576.
- Dobrowolska, D., Bončina, A., & Klumpp, R. (2017). Ecology and silviculture of silver fir (*Abies alba* Mill.): a review. *Journal of Forest Research*, 22(6), 326-335.
- Dodd, M. E., & Silvertown, J. (2000). Size-specific fecundity and the influence of lifetime size variation upon effective population size in *Abies balsamea. Heredity*, 85(6), 604–609.
- Dreier, S., Redhead, J. W., Warren, I. A., Bourke, A. F., Heard, M. S., Jordan, W. C., & Carvell, C. (2014). Fine-scale spatial genetic structure of common and declining bumble bees across an agricultural landscape. *Molecular Ecology*, 23(14), 3384–3395.
- Duforet-Frebourg, N., & Blum, M. G. B. (2014). Nonstationary patterns of isolation-by-distance: Inferring measures of local genetic differentiation with bayesian kriging. *Evolution*, 68(4), 1110–1123.
- Eisenhut, G. (1961). Untersuchungen über die Morphologie und Ökologie der Pollenkörner heimischer und fremdländischer Waldbäume, Vol. 15. Verlag nicht ermittelbar.
- Ellegren, H., & Galtier, N. (2016). Determinants of genetic diversity. Nature Reviews Genetics, 17(7), 422–433.

- Ennos, R. A. (2001). Inferences about spatial processes in plant populations from the analysis of molecular markers. *Integrating Ecology and Evolution in a Spatial Context*, 45–71. https://ci.nii.ac.jp/naid/10027 154496.
- Epperson, B. K. (1995). Fine-scale spatial structure: correlations for individual genotypes differ from those for local gene frequencies. *Evolution*, 49(5), 1022–1026.
- Excoffier, L., Foll, M., & Petit, R. J. (2009). Genetic consequences of range expansions. Annual Review of Ecology, Evolution, and Systematics, 40, 481–501.
- Fady, B., & Bozzano, M. (2020). Effective population size does not make a practical indicator of genetic diversity in forest trees. *Biological Conservation*, 253, 108904.
- Gapare, W. J., & Aitken, S. N. (2005). Strong spatial genetic structure in peripheral but not core populations of Sitka spruce [*Picea sitchen*sis (Bong.) Carr.]. *Molecular Ecology*, 14(9), 2659–2667. https://doi. org/10.1111/j.1365-294X.2005.02633.x.
- Gauzere, J., Klein, E. K., Brendel, O., Davi, H., & Oddou-Muratorio, S. (2020). Microgeographic adaptation and the effect of pollen flow on the adaptive potential of a temperate tree species. *New Phytologist*, 227(2), 641–653.
- Gazol, A., Camarero, J. J., Gutiérrez, E., Popa, I., Andreu-Hayles, L., Motta, R., & Carrer, M. (2015). Distinct effects of climate warming on populations of silver fir (*Abies alba*) across Europe. *Journal of Biogeography*, 42(6), 1150–1162.
- Gelmi-Candusso, T. A., Heymann, E. W., & Heer, K. (2017). Effects of zoochory on the spatial genetic structure of plant populations. *Molecular Ecology*, 26(21), 5896–5910.
- Gerzabek, G., Oddou-Muratorio, S., & Hampe, A. (2017). Temporal change and determinants of maternal reproductive success in an expanding oak forest stand. *Journal of Ecology*, 105(1), 39–48.
- Gömöry, D., Longauer, R., Liepelt, S., Ballian, D., Brus, R., Kraigher, H., Parpan, V. I., Parpan, T. V., Paule, L., Stupar, V. I., & Ziegenhagen, B. (2004). Variation patterns of mitochondrial DNA of Abies alba Mill. in suture zones of postglacial migration in Europe. Acta Societatis Botanicorum Poloniae, 73(3), 203–206.
- Gömöry, D., Paule, L., Krajmerová, D., Romšáková, I., & Longauer, R. (2012). Admixture of genetic lineages of different glacial origin: A case study of Abies alba Mill. in the Carpathians. Plant Systematics and Evolution, 298(4), 703–712.
- González-Díaz, P., Jump, A. S., Perry, A., Wachowiak, W., Lapshina, E., & Cavers, S. (2017). Ecology and management history drive spatial genetic structure in Scots pine. *Forest Ecology and Management*, 400, 68–76.
- González-Martínez, S. C., Gerber, S., Cervera, M. T., Martínez-Zapater, J. M., Gil, L., & Alía, R. (2002). Seed gene flow and fine-scale structure in a Mediterranean pine (*Pinus pinaster* Ait.) using nuclear microsatellite markers. *Theoretical and Applied Genetics*, 104(8), 1290–1297.
- Grace, J. B. (2006). Structural equation modeling and natural systems. Cambridge University Press.
- Hamrick, J. L., Murawski, D. A., & Nason, J. D. (1993). The influence of seed dispersal mechanisms on the genetic structure of tropical tree populations. *Vegetatio*, 107-108(1), 281–297.
- Hardy, O. J., Maggia, L., Bandou, E., Breyne, P., Caron, H., Chevallier, M.-H., Doligez, A., Dutech, C., Kremer, A., Latouche-Hallé, C., Troispoux, V., Veron, V., & Degen, B. (2005). Fine-scale genetic structure and gene dispersal inferences in 10 Neotropical tree species. *Molecular Ecology*, 15(2), 559–571.
- Hardy, O. J., & Vekemans, X. (2002). SPAGeDI: A versatile computer program to analyse spatial genetic structure at the individual or population levels. *Molecular Ecology Notes*, *2*(4), 618–620.
- Heer, K., Behringer, D., Piermattei, A., Bässler, C., Brandl, R., Fady, B., Jehl, H., Liepelt, S., Lorch, S., Piotti, A., Vendramin, G. G., Weller, M., Ziegenhagen, B., Büntgen, U., & Opgenoorth, L. (2018). Linking dendroecology and association genetics in natural populations: Stress

responses archived in tree rings associate with SNP genotypes in silver fir (*Abies alba* Mill.). *Molecular Ecology*, 27(6), 1428–1438.

- Hengl, T., Mendes de Jesus, J., Heuvelink, G. B. M., Ruiperez Gonzalez, M., Kilibarda, M., Blagotić, A., Shangguan, W., Wright, M. N., Geng, X., Bauer-Marschallinger, B., Guevara, M. A., Vargas, R., MacMillan, R. A., Batjes, N. H., Leenaars, J. G. B., Ribeiro, E., Wheeler, I., Mantel, S., & Kempen, B. (2017). SoilGrids250m: Global gridded soil information based on machine learning. *PLoS One*, *12*(2), e0169748.
- Hernández, L., Camarero, J. J., Gil-Peregrín, E., Sánchez, M. Á. S., Cañellas, I., & Montes, F. (2019). Biotic factors and increasing aridity shape the altitudinal shifts of marginal Pyrenean silver fir populations in Europe. Forest Ecology and Management, 432, 558–567.
- Herrera, C. M., & Bazaga, P. (2008). Adding a third dimension to the edge of a species' range: altitude and genetic structuring in mountainous landscapes. *Heredity*, 100(3), 275–285.
- Herrera, C. M., & Jovani, R. (2010). Lognormal distribution of individual lifetime fecundity: insights from a 23-year study. *Ecology*, 91(2), 422-430.
- Hijmans, R. J., Phillips, S., Leathwick, J., & Elith, J. (2017). Package "dismo". Circles, 9(1), 1-68. https://doi.org/10.1016/b978-0-12-809633-8.02390-6
- Hijmans, R. J., Van Etten, J., Cheng, J., Mattiuzzi, M., Sumner, M., & Greenberg, J. A. (2015). Package 'raster'. R package, 734.
- Hoban, S., Bruford, M., Jackson, J. D. U., Lopes-Fernandes, M., Heuertz, M., Hohenlohe, P. A., & Laikre, L. (2020). Genetic diversity targets and indicators in the CBD post-2020 Global Biodiversity Framework must be improved. *Biological Conservation*, 248, 108654.
- Hoban, S., Paz-Vinas, I., Aitken, S., Bertola, L. D., Breed, M. F., Bruford, M. W., & Laikre, L. (2021). Effective population size remains a suitable, pragmatic indicator of genetic diversity for all species, including forest trees. *Biological Conservation*, 253, 108906.
- Jombart, T. (2008). adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics*, 24(11), 1403–1405.
- Jombart, T., Devillard, S., & Balloux, F. (2010). Discriminant analysis of principal components: a new method for the analysis of genetically structured populations. BMC Genetics, 11(1), 1–15.
- Kalinowski, S. T., Taper, M. L., & Marshall, T. C. (2007). Revising how the computer program CERVUS accommodates genotyping error increases success in paternity assignment. *Molecular Ecology*, 16(5), 1099–1106.
- Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W., Zimmermann, N. E., Linder, H. P., & Kessler, M. (2017). Data Descriptor: Climatologies at high resolution for the earth's land surface areas Background & Summary.
- Kimura, M., & Weiss, G. H. (1964). The stepping stone model of population structure and the decrease of genetic correlation with distance. *Genetics*, 49(4), 561.
- Kline, P. (2015). A handbook of test construction (Psychology Revivals). Introduction to psychometric design. Routledge.
- Kling, M. M., & Ackerly, D. D. (2021). Global wind patterns shape genetic differentiation, asymmetric gene flow, and genetic diversity in trees. Proceedings of the National Academy of Sciences, 118(17), e2017317118.
- Kramer, A. T., Ison, J. L., Ashley, M. V., & Howe, H. F. (2008). The paradox of forest fragmentation genetics. *Conservation Biology*, 22(4), 878–885.
- Kremer, A., Ronce, O., Robledo-Arnuncio, J. J., Guillaume, F., Bohrer, G., Nathan, R., & Schueler, S. (2012). Long-distance gene flow and adaptation of forest trees to rapid climate change. *Ecology Letters*, 15(4), 378–392.
- Leblois, R., Estoup, A., & Rousset, F. (2003). Influence of mutational and sampling factors on the estimation of demographic parameters in a "continuous" population under isolation by distance. *Molecular Biology and Evolution*, 20(4), 491–502.
- Leclerc, T., Vimal, R., Troispoux, V., Périgon, S., & Scotti, I. (2015). Life after disturbance (I): changes in the spatial genetic structure of

Jacaranda copaia (Aubl.) D. Don (Bignonianceae) after logging in an intensively studied plot in French Guiana. *Annals of Forest Science*, 72(5), 509–516.

- Legendre, P., & Fortin, M. J. (2010). Comparison of the Mantel test and alternative approaches for detecting complex multivariate relationships in the spatial analysis of genetic data. *Molecular Ecology Resources*, 10(5), 831–844.
- Leonarduzzi, C., Piotti, A., Spanu, I., & Vendramin, G. G. (2016). Effective gene flow in a historically fragmented area at the southern edge of silver fir (Abies alba Mill.) distribution. *Tree Genetics & Genomes*, 12(5), 1–14.
- Liepelt, S., Bialozyt, R., & Ziegenhagen, B. (2002). Wind-dispersed pollen mediates postglacial gene flow among refugia. Proceedings of the National Academy of Sciences of USA, 99(22), 14590–14594.
- Liepelt, S., Cheddadi, R., de Beaulieu, J. L., Fady, B., Gömöry, D., Hussendörfer, E., Konnert, M., Litt, T., Longauer, R., Terhürne-Berson, R., & Ziegenhagen, B. (2009). Postglacial range expansion and its genetic imprints in *Abies alba* (Mill.) - A synthesis from palaeobotanic and genetic data. *Review of Palaeobotany and Palynology*, 153(1–2), 139–149.
- Liepelt, S., Mayland-Quellhorst, E., Lahme, M., & Ziegenhagen, B. (2010). Contrasting geographical patterns of ancient and modern genetic lineages in Mediterranean Abies species. *Plant Systematics and Evolution*, 284(3), 141–151.
- Loiselle, B. A., Sork, V. L., Nason, J., & Graham, C. (1995). Spatial genetic structure of a tropical understory shrub, *Psychotria officinalis* (Rubiaceae). American Journal of Botany, 82(11), 1420–1425.
- Magri, D., Di Rita, F., Aranbarri, J., Fletcher, W., & González-Sampériz, P. (2017). Quaternary disappearance of tree taxa from Southern Europe: Timing and trends. *Quaternary Science Reviews*, 163, 23–55.
 Malécot, G. (1948). Les Mathématiques de l'Hérédité. Masson.
- Malecot, G. (1948). Les Mathematiques de l'Herealte. Masson.
- Marshall, T. C., Slate, J., Kruuk, L. E. B., & Pemberton, J. M. (1998). Statistical confidence for likelihood-based paternity inference in natural populations. *Molecular Ecology*, 7(5), 639–655.
- Martínez-Sancho, E., Rellstab, C., Guillaume, F., Bigler, C., Fonti, P., Wohlgemuth, T., & Vitasse, Y. (2021). Post-glacial re-colonization and natural selection have shaped growth responses of silver fir across Europe. *Science of the Total Environment*, 779, 146393.
- Mauri, A., Strona, G., & San-Miguel-Ayanz, J. (2017). EU-Forest, a highresolution tree occurrence dataset for Europe. *Scientific Data*, 4(1), 1–8.
- Meirmans, P. G. (2012). The trouble with isolation by distance. *Molecular Ecology*, 21(12), 2839–2846.
- Mosca, E., Di Pierro, E. A., Budde, K. B., Neale, D. B., & González-Martínez, S. C. (2018). Environmental effects on fine-scale spatial genetic structure in four Alpine keystone forest tree species. *Molecular Ecology*, 27(3), 647–658.
- Ng, K. K. S., Lee, S. L., & Koh, C. L. (2004). Spatial structure and genetic diversity of two tropical tree species with contrasting breeding systems and different ploidy levels. *Molecular Ecology*, 13(3), 657-669.
- Oddou-Muratorio, S., Gauzere, J., Bontemps, A., Rey, J. F., & Klein, E. K. (2018). Tree, sex and size: Ecological determinants of male vs. female fecundity in three *Fagus sylvatica* stands. *Molecular Ecology*, 27(15), 3131–3145.
- Paffetti, D., Travaglini, D., Buonamici, A., Nocentini, S., Vendramin, G. G., Giannini, R., & Vettori, C. (2012). The influence of forest management on beech (*Fagus sylvatica* L.) stand structure and genetic diversity. *Forest Ecology and Management*, 284, 34–44.
- Paluch, J., Zarek, M., & Kempf, M. (2019). The effect of population density on gene flow between adult trees and the seedling bank in Abies alba Mill. European Journal of Forest Research, 138(2), 203–217.
- Pandey, M., & Rajora, O. P. (2012). Higher fine-scale genetic structure in peripheral than in core populations of a long-lived and mixedmating conifer-eastern white cedar (*Thuja occidentalis* L.). BMC Evolutionary Biology, 12(1), 1–14.

WILEY-MOLECULAR ECOLOGY

- Paradis, E. (2010). pegas: an R package for population genetics with an integrated-modular approach. *Bioinformatics*, *26*(3), 419–420.
- Peakall, R., & Smouse, P. E. (2012). GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research-an update. *Bioinformatics* (Oxford, England), 28(19), 2537–2539.
- Pebesma, E. (2018). simple Feature for R: Standardized Support for Spatial Vector Data. *The R Journal*, 10(1), 446–439.
- Pebesma, E., & Bivand, R. S. (2005). Classes and Methods for Spatial Data: the sp Package. https://cran.r-project.org/.
- Petit, R. J., & Hampe, A. (2006). Some Evolutionary Consequences of Being a Tree. Annual Review of Ecology, Evolution, and Systematics, 37(1), 187–214.
- Petit, R. J., Hu, F. S., & Dick, C. W. (2008). Forests of the past: a window to future changes. *Science*, 320(5882), 1450–1452.
- Piotti, A., Leonardi, S., Heuertz, M., Buiteveld, J., Geburek, T., Gerber, S., & Vendramin, G. G. (2013). Within-population genetic structure in beech (Fagus sylvatica L.) stands characterized by different disturbance histories: does forest management simplify population substructure? *PLoS One*, 8(9), e73391.
- Piotti, A., Leonarduzzi, C., Postolache, D., Bagnoli, F., Spanu, I., Brousseau, L., Urbinati, C., Leonardi, S., & Vendramin, G. G. (2017). Unexpected scenarios from Mediterranean refugial areas: disentangling complex demographic dynamics along the Apennine distribution of silver fir. Journal of Biogeography, 44(7), 1547–1558.
- Porth, I., & El-Kassaby, Y. A. (2014). Assessment of the genetic diversity in forest tree populations using molecular markers. *Diversity*, 6(2), 283–295.
- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data. *Genetics*, 155(2), 945–959.
- QGIS Development Team (2016). *QGIS geographic information system*. Open Source Geospatial Foundation Project.
- R Core Team (2019). R: A language and environment for statistical computing. R Foundation for Statistical Computing. https://www.R-proje ct.org/.
- Restoux, G., Silva, D. E., Sagnard, F., Torre, F., Klein, E., & Fady, B. (2008). Life at the margin: the mating system of Mediterranean conifers. *Web Ecology*, 8(1), 94–102.
- Rico, Y., & Wagner, H. H. (2016). Reduced fine-scale spatial genetic structure in grazed populations of *Dianthus carthusianorum*. *Heredity*, 117(5), 367–374.
- Roschanski, A. M., Csilléry, K., Liepelt, S., Oddou-Muratorio, S., Ziegenhagen, B., Huard, F., Ullrich, K. K., Postolache, D., Vendramin, G. G., & Fady, B. (2016). Evidence of divergent selection for drought and cold tolerance at landscape and local scales in *Abies alba* Mill. in the French Mediterranean Alps. *Molecular Ecology*, 25(3), 776-794.
- Roschanski, A. M., Fady, B., Ziegenhagen, B., & Liepelt, S. (2013). Annotation and Re-Sequencing of Genes from De Novo Transcriptome Assembly of Abies alba (Pinaceae). Applications in Plant Sciences, 1(1), 1200179.
- Rosseel, Y. (2011). lavaan: an R package for structural equation modeling and more Version 0.4-9 (BETA).
- Rousset, F. (2000). Genetic differentiation between individuals. *Journal* of Evolutionary Biology, 13(1), 58–62.
- Royal Botanic Gardens Kew (2015). *Seed Information Database (SID)*. Version 7.1. Available from: http://data.kew.org/sid/.
- Ruosch, M., Spahni, R., Joos, F., Henne, P. D., Van der Knaap, W. O., & Tinner, W. (2016). Past and future evolution of Abies alba forests in Europe-comparison of a dynamic vegetation model with palaeo data and observations. *Global Change Biology*, 22(2), 727-740.
- Saastamoinen, M., Bocedi, G., Cote, J., Legrand, D., Guillaume, F., Wheat, C. W., & Baguette, M. (2018). Genetics of dispersal. *Biological Reviews*, 93(1), 574–599.
- Sagnard, F., Oddou-Muratorio, S., Pichot, C., Vendramin, G. G., & Fady, B. (2011). Effects of seed dispersal, adult tree and seedling density on the spatial genetic structure of regeneration at fine temporal and spatial scales. *Tree Genetics & Genomes*, 7(1), 37–48.

- Scotti-Saintagne, C., Boivin, T., Suez, M., Musch, B., Scotti, I., Fady, B. (2021). Signature of mid-Pleistocene lineages in the European silver fir (Abies alba Mill.) at its geographic distribution margin. *Ecology* and Evolution, 00, 1–16. http://dx.doi.org/10.1002/ece3.7886
- Segelbacher, G., Cushman, S. A., Epperson, B. K., Fortin, M. J., Francois, O., Hardy, O. J., Holderegger, R., Taberlet, P., Waits, L. P., & Manel, S. (2010). Applications of landscape genetics in conservation biology: Concepts and challenges. *Conservation Genetics*, 11(2), 375–385.
- Sgrò, C. M., Lowe, A. J., & Hoffmann, A. A. (2011). Building evolutionary resilience for conserving biodiversity under climate change. *Evolutionary Applications*, 4(2), 326–337.
- Sjölund, M. J., & Jump, A. S. (2015). Coppice management of forests impacts spatial genetic structure but not genetic diversity in European beech (*Fagus sylvatica* L.). Forest Ecology and Management, 336, 65–71.
- Slatkin, M. (1985). Gene flow in natural populations. Annual Review of Ecology and Systematics, 16(1), 393–430.
- Slatkin, M. (1993). Isolation by distance in equilibrium and nonequilibrium populations. *Evolution*, 47(1), 264–279.
- Smouse, P. E., Peakall, R., & Gonzales, E. (2008). A heterogeneity test for fine-scale genetic structure. *Molecular Ecology*, 17(14), 3389–3400.
- Sokal, R. R., & Wartenberg, D. E. (1983). A test of spatial autocorrelation analysis using an isolation-by-distance model. *Genetics*, 105(1), 219–237.
- Terhürne-Berson, R., Litt, T., & Cheddadi, R. (2004). The spread of *Abies* throughout Europe since the last glacial period: combined macrofossil and pollen data. *Vegetation History and Archaeobotany*, 13(4), 257–268.
- Tinner, W., Colombaroli, D., Heiri, O., Henne, P. D., Steinacher, M., Untenecker, J., Vescovi, E., Allen, J. R. M., Carraro, G., Conedera, M., Joos, F., Lotter, A. F., Luterbacher, J., Samartin, S., & Valsecchi, V. (2013). The past ecology of *Abies alba* provides new perspectives on future responses of silver fir forests to global warming. *Ecological Monographs*, 83(4), 419–439.
- Torroba-Balmori, P., Budde, K. B., Heer, K., Gonzalez-Martinez, S. C., Olsson, S., Scotti-Saintagne, C., & Heuertz, M. (2017). Altitudinal gradients, biogeographic history and microhabitat adaptation affect fine-scale spatial genetic structure in African and Neotropical populations of an ancient tropical tree species. *PLoS One*, *12*(8), e0182515.
- Travis, J. M. J., Delgado, M., Bocedi, G., Baguette, M., Bartoń, K., Bonte, D., Boulangeat, I., Hodgson, J. A., Kubisch, A., Penteriani, V., Saastamoinen, M., Stevens, V. M., & Bullock, J. M. (2013). Dispersal and species' responses to climate change. *Oikos*, 122(11), 1532–1540.
- Troupin, D., Nathan, R., & Vendramin, G. G. (2006). Analysis of spatial genetic structure in an expanding *Pinus halepensis* population reveals development of fine-scale genetic clustering over time. *Molecular Ecology*, 15(12), 3617–3630.
- Valbuena-Carabaña, M., González-Martínez, S. C., Sork, V. L., Collada, C., Soto, A., Goicoechea, P. G., & Gil, L. (2005). Gene flow and hybridisation in a mixed oak forest (*Quercus pyrenaica Willd. and Quercus petraea* (Matts.) Liebl.) in central Spain. *Heredity*, 95(6), 457–465.
- Vekemans, X., & Hardy, O. J. (2004). New insights from fine-scale spatial genetic structure analyses in plant populations. *Molecular Ecology*, 13(4), 921–935.
- Vitali, V., Büntgen, U., & Bauhus, J. (2017). Silver fir and Douglas fir are more tolerant to extreme droughts than Norway spruce in southwestern Germany. *Global Change Biology*, 23(12), 5108–5119.
- Vitasse, Y., Bottero, A., Rebetez, M., Conedera, M., Augustin, S., Brang, P., & Tinner, W. (2019). What is the potential of silver fir to thrive under warmer and drier climate? *European Journal of Forest Research*, 138(4), 547–560.
- Wang, J. (2016). Pedigrees or markers: Which are better in estimating relatedness and inbreeding coefficient? *Theoretical Population Biology*, 107, 4–13.

Wernberg, T., Coleman, M. A., Bennett, S., Thomsen, M. S., Tuya, F., & Kelaher, B. P. (2018). Genetic diversity and kelp forest vulnerability to climatic stress. *Scientific Reports*, 8(1), 1–8.

Wright, S. (1943). Isolation by distance. Genetics, 28(2), 114.

Young, A. G., & Merriam, H. G. (1994). Effects of forest fragmentation on the spatial genetic structure of Acer saccharum Marsh. (sugar maple) populations. *Heredity*, 72(2), 201–208.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Major, E. I., Höhn, M., Avanzi, C., Fady, B., Heer, K., Opgenoorth, L., Piotti, A., Popescu, F., Postolache, D., Vendramin, G. G., & Csilléry, K. (2021). Fine-scale spatial genetic structure across the species range reflects recent colonization of high elevation habitats in silver fir (*Abies alba* Mill.). *Molecular Ecology*, 30, 5247–5265. <u>https://doi.</u> org/10.1111/mec.16107