

# Recent Advances on Pickering Emulsions Stabilized by Diverse Edible Particles: Stability Mechanism and Applications

# Wei Li<sup>1†</sup>, Bo Jiao<sup>1,2†</sup>, Sisheng Li<sup>1</sup>, Shah Faisal<sup>1</sup>, Aimin Shi<sup>1</sup>, Weiming Fu<sup>1</sup>, Yiying Chen<sup>1</sup> and Qiang Wang<sup>1,2\*</sup>

**OPEN ACCESS** 

### Edited by:

Jian Zhong, Shanghai Ocean University, China

#### Reviewed by:

Eric Dickinson, University of Leeds, United Kingdom Hongbin Zhang, Shanghai Jiao Tong University, China

> \*Correspondence: Qiang Wang wangqiang06@caas.cn

<sup>†</sup>These authors have contributed equally to this work and share first authorship

#### Specialty section:

This article was submitted to Nutrition and Food Science Technology, a section of the journal Frontiers in Nutrition

Received: 29 January 2022 Accepted: 23 March 2022 Published: 06 May 2022

### Citation:

Li W, Jiao B, Li S, Faisal S, Shi A, Fu W, Chen Y and Wang Q (2022) Recent Advances on Pickering Emulsions Stabilized by Diverse Edible Particles: Stability Mechanism and Applications. Front. Nutr. 9:864943. doi: 10.3389/fnut.2022.864943 <sup>1</sup> Institute of Food Science and Technology, Chinese Academy of Agriculture Sciences/Key Laboratory of Agro-Products Processing, Ministry of Agriculture and Rural Affairs, Beijing, China, <sup>2</sup> College of Food Science and Engineering, Nanjing University of Finance and Economics/Collaborative Innovation Center for Modern Grain Circulation and Safety, Nanjing, China

Pickering emulsions, which are stabilized by particles, have gained considerable attention recently because of their extreme stability and functionality. A food-grade particle is preferred by the food or pharmaceutical industries because of their noteworthy natural benefits (renewable resources, ease of preparation, excellent biocompatibility, and unique interfacial properties). Different edible particles are reported by recent publications with distinct shapes resulting from the inherent properties of raw materials and fabrication methods. Furthermore, they possess distinct interfacial properties and functionalities. Therefore, this review provides a comprehensive overview of the recent advances in the stabilization of Pickering emulsions using diverse food-grade particles, as well as their possible applications in the food industry.

Keywords: Pickering emulsion, food-grade particles, stabilization mechanism, particles shapes, food application

# INTRODUCTION

Pickering emulsions are stabilized by solid or soft nano (micro) particles. They are first discovered by Ramsden (1) in 1903 that the mixture of wax and water could be stabilized by solids, thus forming emulsions. Subsequently, Pickering (2) put forward to stabilize emulsions by using nanoparticles and microparticles, which promoted the progress of emulsion research. Hence, the emulsion stabilized by particles was identified and described as "Pickering emulsion", which can be oil-in-water (O/W), water-in-oil (W/O), water-in-water (W/W) (3), or even multiple (4–6). There are prime differences in the behavior of the low-molecular-weight surfactants and natural polymers, particles with sizes ranging from just several nanometers to micrometers during the preparation and stabilization of the Pickering emulsions. Particles can provide a space barrier between the two immiscible phases to prevent droplets' coalescence (7) and Ostwald ripening in Pickering emulsions (8). Some researchers have provided different stability functions of particles in Pickering emulsion. Tavasoli et al. (9) considered that the capillary pressure due to the

1

anisotropic shapes of solid particles can prevent the interface film drainage. Dickinson et al. (9, 10) presented a fact that the form of a network of particles in the continuous phase can generate a barrier to improving the stability of many gel-like emulsions of moderately high oil volume fraction. These all could provide significantly higher stability.

Initially, Pickering emulsions did not draw enough attention because of the limitation of materials which showed partial wetting in both phases. Subsequently, Pickering emulsion stabilized by inorganic solid nanoparticles such as silica particles (11), polymer lattices (12), and clay (13) received a wide scope of research attention because of the development of chemical engineering and material science (14). Furthermore, it has been applied to some novel areas, such as oil recovery. However, their applications in cosmetic products, food technology, as well as pharmaceutical industries were extremely limited due to their biodegradability and biocompatibility concerns. Recently, with the development of biomacromolecules materials science, varieties of novel particles with adjustable surface wettability and better biocompatibility were used to stabilize Pickering emulsions (15). Particles made of natural edible resources such as polysaccharides (16, 17), protein (18) have been broadly used to stabilize Pickering emulsions to satisfy diverse needs, from food and cosmetic industries to catalysis, tissue engineering, and drug delivery.

The shapes of the particles can be different due to the diverse nature, composition, and structure of these biopolymers. Specifically, the behavior at the interface and the ability to stabilize the emulsions can be governed by the shape of particles (19). Some researchers (20, 21) considered that due to their higher aspect ratio of anisotropic particles, the interfacial layer, the desorption energy value, and the capillary force can all be increased leading to more stable emulsion systems. The research and development of particles with different shapes have become a hotspot. The novel shape of solid particles, including ellipsoids, nanofibrils, nanocages, platedshape, nanotubes, and other irregular shapes (22), could exhibit different stabilization mechanisms of Pickering emulsion. We explicitly dedicated a section summarizing and discussing other mechanisms for stabilization of Pickering emulsions other than the irreversible adsorption behavior of particles at the oil-water interface and the physical obstacle especially the stability mechanism of high internal phase Pickering emulsions (HIPEs). It is also worth noting that, Pickering emulsion catalysis has advanced rapidly in the past decades, some potential applications have been found in the food area. However, no review has been published in the food area contained the application aspect of Pickering emulsion catalysis. To comprehensively review the recent advances on Pickering emulsions stabilized by diverse edible particles, besides the novel points we mentioned above, an integrated overview of the stabilization mechanism and application of Pickering emulsions have been given. Furthermore, an outlook on this topic has been raised as well. We believe this review could provide the latest progress and new insight into the field of Pickering emulsions readers.

# THE STABILIZATION MECHANISM OF PICKERING EMULSIONS

# The Irreversible Adsorption and Robust Physical Barriers of Particles at the Oil-Water Interface

The mechanism of Pickering emulsions is different from emulsions stabilized by traditional surfactants (**Figure 1**) and biopolymers with hydrophilic and hydrophobic two distinct regions. The most common one is that Pickering emulsions are stabilized through robust interface adsorption by particles so it does not need to be amphiphilic (23). The wettability of particles is critical for particles to adsorb at the surface of the droplets and the final stabilization of Pickering emulsions. To understand the difference between them, it is necessary to explain the irreversible adsorption and robust physical barriers of particles at the oilwater interface.

The strong adsorption is driven by the partial wettability of certain spherical hard solid particles, lead to firm steric hindrance. Therefore, it can prevent the emulsion droplets from flocculation and coalescence by a steric mechanism. Adsorption of particles at the oil-water interface requires partial wetting by both the oil and water phases (24). And the wettability can be measured by contact angles ( $\theta$ ). According to the contact angles ( $\theta$ ) of solid particles, the emulsions are classified into different types. O/W emulsions ( $\theta < 90^\circ$ ) can be formed by particles easily wetted by water, otherwise, W/O emulsions ( $90^\circ < \theta < 180^\circ$ ) are fabricated.

Desorption of particles from the oil-water interface requires overcoming high interfacial energy, so a firm space barrier to prevent droplet coalescence can be formed at the oilwater interface. Thus, Pickering emulsions are endowed with much better stability than traditional emulsions stabilized by surfactants. This is a matter of interfacial energies of the three interfaces: solid-oil, solid-water, and water-oil abbreviated  $\gamma_{s-o}$ ,  $\gamma_{s-w}$ ,  $\gamma_{w-o}$ . O/W emulsions require positive adhesion energy of oil E<sub>Adh</sub>(O/W), and negative spreading coefficient of oil S(O/W) (23), (Equations 1, 2):

$$\mathbf{E}_{\mathrm{Adh}}(\mathbf{O}/\mathbf{W}) = \gamma_{s-w} + \gamma_{w-o} - \gamma_{s-o} > \mathbf{0}$$
(1)

$$\mathbf{S}(\mathbf{O}/\mathbf{W}) = \gamma_{\mathbf{s}-\mathbf{w}} - \gamma_{\mathbf{w}-\mathbf{o}} - \gamma_{\mathbf{s}-\mathbf{o}} < \mathbf{0}$$
(2)

Formation of the W/O emulsion needs the adsorption energy of water,  $E_{Adh}$  (W/O) keeping positive, and S (W/O) remaining negative. When the surface of solid particles is entirely hydrophilic, it will be completely wetted by the water phase. Solid particles will not adsorb at the water-oil interface. They are completely dispersed in the water phase, where  $\theta = 0^{\circ}$ . Similarly, the solid particles are completely wetted by oil, where  $\theta = 180^{\circ}$ . Therefore, emulsions tend to be formed only when particles are partially wetted. Besides Pickering emulsions will be more stable if the contact angle is preferentially close to 90°. One most common practiced methods of the contact angle is the conventional Young's equation ( $\theta_0 = 90^{\circ} \cdot \theta_w$ ) (25). The contact angle in the water phase is  $\theta_w$ , and the contact angle in the oil



phase is  $\theta_0$ , (Equations 3, 4):

$$\cos(\theta_{\mathbf{w}}) = (\gamma_{\mathbf{s}-\mathbf{o}} - \gamma_{\mathbf{s}-\mathbf{w}}) / \gamma_{\mathbf{w}-\mathbf{o}}$$
(3)

$$\cos(\theta_{\mathbf{o}}) = (\gamma_{\mathbf{s}-\mathbf{w}} - \gamma_{\mathbf{s}-\mathbf{o}})/\gamma_{\mathbf{w}-\mathbf{o}}$$
(4)

Since the solid particles for the stabilization of Pickering particles are small, their gravity and buoyancy can be negligible. The detachment energy E required for desorption from the interface of a spherical solid particle can be calculated using the formula (5) below (26):

$$\mathbf{E} = \pi \mathbf{r}^2 \gamma_{\mathbf{w} - \mathbf{o}} (1 - |\cos\theta|)^2 \tag{5}$$

Where r is the radius of solid spherical particles stabilizing the emulsion,  $\gamma_{W-O}$  is the surface tension of oil-water interface, and  $\theta$  is the three-phase contact angles. For instance, in a solid particle with a radius of 10 nm, when the contact angle is 90°, the spontaneous detachment energy required for desorption from the interface enormously exceeds the thermal energy of Brownian motion (27), which also explains why the adsorption of solid particles at the interface is irreversible. Based on the formula, when the contact angle is too close to 0° or 180°, the thermal energy is greater than the detachment energy. As a result, the solid particles cannot permanently adsorb at the interface to stabilize the Pickering emulsion effectively.

### The Capillary Pressure Between Particles

The capillary pressure (**Figure 2A**) between adjacent particles at the interface can prevent thin film formed by particles degradation. The non-spherical particles (rod-like, disk-like, fibers, etc.) may fabricate more stable emulsion systems. One of the reasons has been revealed that anisotropic particles with different geometrical shapes can generate stronger capillary pressure between adjacent particles at the interface (20, 21). The details have been elaborated in Section Pickering Emulsions Stabilized by Different Shapes of Food-Grade Particles.

### The Interfacial Rheological Responses

The interfacial rheological responses are related to the interfacial adsorption and the interaction amongst particles (28) and can affect film (**Figure 2B**) drainage. The interfacial structure may be transformed into the multilayer arrangement, with one or more extra layers of ordered monodisperse hard spheres accommodated in the interdroplet region, where a substantial excess of particles exists in the continuous phase (29). The additional particle layering can lead to an oscillatory potential of mean force and an associated kinetic structural barrier that can protect droplets from coalescence (30). This layering formed by particles that are not adsorbed at the oil-water interface is common, especially in a host of colloidal particles, such as self-assemble entities that are neither fully monodispersed nor closely resembling hard spherical particles.



in a continuous phase.

# The Formation of Particles' Network in the Space Between Droplets

Another important stabilization principle is the intrinsic ability of microgels to act as a versatile thickener agent and colloid stabilizer in an aqueous medium (10). The more stable and highly viscous structure can stabilized HIPEs stabilized by biopolymers due to a self-supporting gel network (**Figure 2C**) (31). In this way, these HIPEs are able to have resistance to good freeze-thaw reversibility and drastic heating. The particles are predominantly playing the role of a "structure agent" rather than necessarily adsorbing at the oil-water interface (32). Xu et al. (33) used bovine serum albumin glycated with galactose possessing much structural integrity and higher refolding ability that can improve the stability of HIPEs compared to native bovine serum albumin. Additionally, Rayner et al. (34) further found that some adsorbed layers of biopolymer particles at the oil-water interface can be made to fuse together into a coherent gel-like lipid-encapsulating layer through post-emulsification thermal processing.

# PICKERING EMULSIONS STABILIZED BY DIFFERENT SHAPES OF FOOD-GRADE PARTICLES

In recent years, food-grade particles of different shapes have attracted researchers' attention. Zhang et al. (22) summarized protein nanoparticles with different shapes, and various

preparation processes of them are also reviewed. However, many natural polymers other than proteins can be used to prepare particles with various shapes through different modification methods. Common modification ways include octenyl succinic anhydride (OSA) modification of starch, protein modification (enzymatic modification, phosphorylation modification, aldehyde modification, and phenolic modification) (35, 36), and preparation of electrostatic complexes. The particle shapes with different shapes which have various particle densities, desorption energy, and capillary forces between adjacent particles largely affect the stabilization with different principles and application of emulsions (Table 1). Particle shapes can be divided into regular spherical and irregular shapes with anisotropic morphology (25). The causes of different solid particle shapes are related to their sources, properties, structures, and particle preparation methods (22). In addition, particle shape can critically impact the stabilization of emulsion by modifying the wettability behavior of particles and the interactions between adjacent particles.

## **Spherical Solid Particles**

Sphere is one of the most common shapes of food-grade particles, which has been widely used in drug delivery and food products due to its easy preparation and low cost (61). Both undeformable solid spherical particles and porous microspheres belong to common food-grade spherical solid particles, whose diameters are shorter than 400 nm. Among them, the mechanism of undeformable spherical particles stabilizing Pickering emulsions has been described in Section Introduction. Spherical particles are mostly formed by protein, polysaccharide, and other macromolecules crimped by intramolecular forces. The spherical particles formed by a single natural molecule are not regular enough. Hence more extra substances are needed to form complexes to improve the better anti-aggregation properties of solid particles. Shah (39) and Li (40) modified zein and barley gliadin with chitosan to formulate spherical solid particles with obvious anti-aggregation and anti-destabilization ability for embedding active substances respectively. Liu et al. (62) used epigallocatechin gallate (EGCG) connected with zein nanoparticles to form covalent or non-covalent through alkali treatment. Experimental results showed the protein-polyphenol complexes had a smooth surface and reduced emulsion droplet size causing better emulsion stability. Polyphenols alter the hydrophobic and electrostatic interactions among protein molecules, giving higher thermal stability and antioxidant capacity of the complex. Soybean protein is often used to prepare regular spherical solid particles to meet various needs through some processes such as pH shift (63, 64), heating treatment (65, 66), enzymatic hydrolysis (67, 68), denatured agent induction [urea (61, 69), ethanol (70), treatment, etc.], and hydrostatic high-pressure treatment (71). These denatured processes can lead to alteration in protein structure, thus, triggering self-assembly or mutual assembly behavior.

# **Rod-Like Solid Particles**

Regular rod, prolate ellipsoid, and spindle rod are common rod-like shapes of solid particles, often prepared by using cellulose nanocrystals (CNC) with the material of rod-shaped particles, cellulosic colloidal nano-rods (48), etc. These particles have relatively sufficient aspect ratios able to stabilize Pickering emulsions effectively (51). Cellulose is a linear homopolysaccharide composed of repeating units of  $\beta$ -1, 4-Dglucopyranose, and nanocellulose, which refers to nanoscale cellulose with a diameter ranging from 1 to 100 nm (72), which can be obtained from agricultural wastes, such as cotton and wood fibers. Common types of nanocellulose include microfibrillated cellulose (MFC), bacterial nanocellulose (BNC), and nanocrystalline cellulose (NCC) (73). The specific surface area, high strength, low density, and amphiphilic ability of nanocellulose provide conditions for stabilizing Pickering emulsions (52). Solid particles with different shapes can be prepared by using nanocellulose from different sources, and they are mainly rod-shaped (74).

It was found that the principle of stable Pickering emulsion of rod-shaped particles was different from that of regular spherical particles. The adsorption of prolate ellipsoids on the water-oil interface mainly depends on two forces. One is the capillary interactions brought about the interface deformation from the overlap between particles (75). The other is the fluctuationinduced force caused by thermally excited capillary waves arising at fluid interfaces (49). Gurappa et al. (51) found that the interface with controlled surface rheology could be obtained by using shape-induced capillary force as well as packing effects. The capillary force affects the aggregation of bubbles and solid particles at the gas-water interface (76). For slightly heavier particles, the capillarity force comes from interface deformation caused by gravity (75). For exceptionally light particles, although gravity is ignored, capillary action still exists at the interface for electrostatic interaction among particles (77, 78). It may also be because the particles are irregularly spherical solid particles, resulting in different degrees of wetting (79-81). The capillary force generated by the deformation of the interface caused by the small particles can be both mutually attractive and mutually repellant, and the interface deformation gets much larger. Thus, the interaction becomes much stronger (82). Loudet et al. (75) investigated the principle and interface characteristics of Pickering emulsions stabilized by prolate ellipsoids. It was found that both of them showed different aggregation morphology. One was preferentially aligned side by side, the other was preferentially arranged tip to tip, which is due to the different wettability of solid particles' surfaces with different chemical compositions. At the same time, they also found that the interaction force of the ellipsoid at the oil-water interface is 10<sup>5</sup> times higher than the internal energy, while the strong interaction force of the regular sphere was weak or non-existent. Further, the study found that the strong interaction force is anisotropic. Subsequently, Loudet et al. explored the wetting and contact lines of ellipsoid particles (50). It is apparent that the interface is pulled down near the tips of the ellipsoid and pulled up near the middle of the particle (Figure 3A). The contact lines were saddle-shaped, and the deformation increased with the increase of the aspect ratio, while no such phenomenon was found in the contact lines of regular spheres. The deformation of the air-water interface was reported in many pieces of research. The shape of the contact line is determined by the contact angle, but the

| Morphology         | Materials                    | Particles size   | Emulsion type | Application  | References |
|--------------------|------------------------------|--|---------------|--|------------|
| Spherality         | Green tea polyphenol         | 100–400 nm diameter  | O/W           | Enhance the oxidability of emulsion  | (37)       |
|                    | Rice peptide                 | 357.8 nm diameter  | O/W           | Improve oxidative stability of<br>emulsion   | (38)       |
|                    | Zein-chitosan                | _  | 0/W           | Encapsulation of VD <sub>3</sub>   | (39)       |
|                    | Hordein-chitosan             | 594.9 nm diameter  | O/W           | Encapsulation of active<br>substances and replacing<br>existing food system  | (40)       |
|                    | Zein-pullulan complex        | 147.4 nm diameter without pullulan;<br>594.9 nm diameter with pullulan   | O/W           | Encapsulation and releasing of<br>active substances  | (41)       |
|                    | Luteolin                     | 125.6 $\pm$ 24.7 nm diameter   | O/W           | Enhance the oxidability of pine nut oil  | (42)       |
|                    | Amylopectin-xyloglucan       | 184 $\pm$ 7 nm diameter  | O/W           | _  | (43)       |
|                    | Zein                         | About 200 nm diameter  | _             | _  | (44)       |
|                    | nanocellulose                | $\leq$ 150 nm diameter   | O/W           | Encapsulation of curcumin and coumarin   | (45)       |
| Rod-like           | Cellulose nanocrystals       | several microns length $30.61 \pm 6.67  \text{nm}$ width   | O/W           | Improving the storage stability of<br>emulsion against high<br>temperature and sheering  | (46)       |
|                    | Cellulose nanocrystals       | The needle-like morphology with<br>about 200 nm length and 10 nm<br>diameter<br>CNCs modified by BSA with<br>500 nm length and 30 nm width | O/W           | Broden the application of food<br>and drugs in the central nervous<br>system   | (47)       |
|                    | Cellulose colloidal nanorods | 185 nm length; 13–160 aspect radio   |               | _  | (48)       |
| Ellipsoid          | Cellulose nanocrystals       | _  | 0/W           | _  | (49)       |
|                    | Polystyrene                  | Long axis 11–45 μm<br>Short axis 4.5–9 μm  | O/W           | _  | (50)       |
| Long ellipsoid     | Polystyree                   | Aspect radio 1:9   | 0/W           | _  | (51)       |
| Spindle            |                              | Aspect radio 1:6   | W/O           |  |            |
| Nanofibrils        | Nanocellulose                | 20 nm average diameter, 2.0 $\pm$ 0.5 nm length, aspect radio 100  | O/W           | Food storage, food emulsifier,<br>active food packing, wound<br>healing, surface biocides and<br>antibacterial cleaning products | (52)       |
|                    | Nanocellulose                | 200 nm length, 8 nm width  | 0/W           | Application in food and drugs  | (53)       |
|                    | Nanocrystals                 | _  | O/W           | Replacing of existing food system  | (52)       |
| Nanocages          | E2 protein                   | 25 nm diameter with 12 openings of about 5 nm  | O/W           | Encapsulation of new food and<br>cosmetics   | (54)       |
| Plate-Shaped       | wool keratin                 | _  | O/W and W/O   | Carrying new drugs and<br>cosmetics  | (55, 56)   |
| Nanotubes          | α-lac-protein                | 20–30 nm diameter; 0.5–1 μm<br>length  | O/W           | Releasing flavor fatty acid  | (57)       |
| Surface depression | Spray drying soy protein     | 20–60 µm   | O/W           | -  | (58)       |
| Spongy             | Plant spore                  | 20–60 µm   | 0/W           | -  | (59)       |
| Flaky              | Phytosterol                  | 30–35 µm   | O/W           | _  | (60)       |

contact angle of the ellipsoid is not fixed. The contact angle at the tip of ellipsoid particles is smaller than that in the middle. Liu et al. (47, 75) first stabilized emulsion with food-grade cellulose nanocrystals (CNCs), shaped like short needles (about 200 nm long and 10 nm wide), and modified the CNCs with different concentrations of bovine serum protein (BSA about 500 nm long and 30 nm wide). This kind of particle can decrease the strong hydrophilicity of pure CNCs effectively. It was found that HIPEs could be stabilized by CNCs which are covered with BSA. It can form a bridge, and the hardness and microstructure of HIPEs could be adjusted by changing the concentration of CNCs.

## Nanofibrils

In addition to the above mentioned rod-like particles, nanocellulose can also be made into nanofibrils particles (**Figure 3B**). The most significant difference between nanofibrils



and rod-like particles is that the former solid particles have a much larger aspect ratio, forming strong entangled and disordered network structures (83), which provides suitable conditions for stabilizing Pickering emulsion. Winuprasith et al. (52) extracted MFC from mangosteen rind to stabilize O/W Pickering emulsions and found that all emulsions remained stable at any concentration of MFC in the experiment, for the fact that at the oil-water interface the MFC may occur as single, dispersed fibers as well as in networks (84). Souza et al. (85) stabilized Pickering emulsions to prevent different essential oils (cinnamon, cardamom, and sapwood) volatilization with cellulose nanofiber (CNF) because CNF had the greatest coverage, the strongest network structure around cardamom oil droplets. In summary, nanofibrils particles tend to form three-dimensional networks structures on the surface of oil droplets and improve the mechanical properties of the emulsion. Besides, an elastic monolayer structure will be produced (86).

# Nanocages

At present, cage-like food-grade particles are mostly newtype nanocages with hollow interiors and porous wall surfaces prepared with protein. Nanocages are similar to spheres so the stability mechanism of nanocages resembles that of spherical particles (22). Sarker et al. (54, 87) prepared dodecahedral hollow protein nanocages using E2 protein to stabilize pH-responsive Pickering emulsions and proved that the cage-shaped solid particles show assembled structure of approximately 25 nm in diameter with 12 openings of about 5 nm each (**Figure 3C**).

# **Plate-Shaped and Nanotubes**

Due to the complicated structure of the protein, it is difficult to prepare disk solid particles (**Figure 3D**). Hikima et al. (55, 56) prepared Pickering emulsions with disk-like wool keratin nanoparticles by alkaline hydrolysis.  $\alpha$ -lac-protein ( $\alpha$ -lac) nanotubes (NTs, **Figure 3E**) formed by self-assembly of partially hydrolyzed  $\alpha$ -lac-protein ( $\alpha$ -lac) peptides were applied as an immobilization carrier for lipases (57), and to stabilize O/W Pickering emulsion by lipase-nanotube, which improved lipase activity, promoted milk fat hydrolysis and increased fatty acid release. In this way, it enhanced the flavor of the cheese.

# **Other Irregular Shapes**

In addition to the common shapes mentioned above, particles can also be shaped as spore particles (59), scaly (60), and so on. Binks et al. (59) investigated the potential of spore particles with rough surface structure and the presence of a Y-shaped marking (**Figure 3F**) of Lycopodium clavatum solely stabilizing Pickering emulsion with oils varying polarity, which can form





clusters and chains at the oil-water interface. Liu et al. (60) prepared phytosterol colloidal particles showing (**Figure 3G**) gellike would provide a novel candidate for stabilization and delivery system for active substances.

# EXTERNAL FACTORS AFFECTING PICKERING EMULSION STABILITY

The effectiveness of particle emulsifiers' ability is related to their wettability, so it is strongly influenced by the hydrophobicity of particles, therefore, it can influence the type of Pickering emulsion. In most cases, changing the surface chemistry with small molecules or polymers through chemical anchoring and physical adsorption can alter the wettability of particles. Nevertheless, the interaction between particles and amphiphiles renders particles' wettability tunable in *in situ* modification. Xiao et al. (88) summarized tailoring the wettability by surface chemistry and surface roughness for Pickering emulsions, emphasizing the effects of surface roughness. In addition to the wettability of particles, external factors also have a critical influence on the stability of emulsions, such as the pH of the emulsion, particle concentration, ionic strength, the

corresponding particle size, droplet size of emulsion, the type and fraction of oil phase.

# рΗ

The pH of the system alters the surface charge of particles, hence affecting the electrostatic interaction among them, regulating the adsorption behavior as well as the space barrier formed at the oil-water interface. Therefore, the emulsion droplets exhibit dispersion or flocculation with the different adsorption behaviors and space barriers of particles (89, 90). Xiao et al. (91) stabilized Pickering emulsions with grafted carboxymethyl maize starch (CMS) nanoparticles showing pHresponsive properties. After imparted with amphipathic 2-(dimethylamine) ethyl methacrylate (DMAEMA) CMS exhibited different hydrophobicity at different pH values. Under acidic conditions, the protonated tertiary amine groups could decrease leading to a significant decrease in electrostatic interactions, hydrogen bonding, covalent bonding, and van der Waals forces. This resulted in particles separating from the two-phase interface into the oil phase. So the emulsions were unstable under acidic conditions. On the contrary, the nanoparticles can stabilize emulsions at pH 10. Li et al. (92), explored the influence of pH on the edible mayonnaise-like Pickering emulsions. They

found the maximum droplet size did not appear at pI, but at pH 5.5, which could be due to the  $\theta_{O/W}$  of 131.9  $\pm$  0.6, which allowed the protein particles more easily to be wetted by the oil phase. Jiao et al. (93) found at pH 3.0, the droplet size is larger (5-50  $\mu$ m,) than that at pH 9.0 (1-20  $\mu$ m). Besides, at pH 9.0, the diameter of the peanut protein isolate microgel particles was around 270 nm and a larger interfacial area can be coated by them causing a smaller droplet size. Chitosan is the only natural positively charged polysaccharide, which can crosslink with the biomacromolecules such as proteins to prepare protein-chitosan particles due to its unique electrical properties free amino and hydroxyl groups carried along the backbone of the composite will be protonated or not by adjusting pH value of the system. Therefore, its adsorption quantity at the oil-water interface can be changed to realize the reversible process of emulsification-demulsification easily. Liu et al. (94) applied chitosan interaction to prepare pH-responsive Pickering emulsion. At pH > 6, chitosan nanoparticles or micrometersized floccular precipitates were formed and adsorbed on the interface to stabilize the emulsion. While at pH < 6, chitosan dissolved in water, and demulsification occurred (Figure 4A). Li et al. (95) prepared gliadin-chitosan nanoparticles (GCNPs) with three different structures, including primary complexation, soluble complexes, and coacervates by facile pH alteration. At pH 7.0, the coacervates type was prepared, owing to the decrease of the charge between the complexes and the weakening of the repulsive force. Droplet coalescence could be better prevented by the force on account of the highest viscoelastic and solid-like properties.

# The Concentration of Food-Grade Particles

The concentration of solid particles can influence the stability of the Pickering emulsion by affecting the droplet size of the emulsions, which is similar to the effect of surfactants on the stability of the traditional emulsions (96). It is generally considered that the final drop size of the Pickering emulsion virtually has nothing to do with the concentration of solid particles in excess (particles-rich regime at a high level) (97), which depended on the stirring intensity. However, if the concentration of solid particles keeps not very high (particlespoor regime, at a low level) the newly formed emulsion droplets only can be partly covered by solid particles. A number of droplets progressively combined with each other leading to the form of fast coalescence and large droplets. As a result, the final drop size is related to the concentration of solid particles. In contrast, with the increase in the concentration, more particles can adsorb at the interface to form a single or multi-layer structure, which prevents the coalescence of emulsion droplets (98, 99), thus stabilizing the emulsion. This is consistent with the finding of Wachira et al. (100). They proved that smaller oil droplet size could be caused by higher chitosan particle concentration, which increased larger surface area that reduced the collisions as well as movement between droplets. Concentration of particles can not only affect droplet size but also affect the structure of porous materials used to stabilize Pickering emulsions. Porous materials could be fabricated by the Pickering emulsion template method (93) whose thickness was able to tailor with various concentrations of the particles. At low particle concentration, the wall of the porous materials was fragile and shrunk. With increasing the concentration of the particles, the pore structure would be relatively uniform, and more stable emulsions could be obtained.

# Ionic Strength

Salt in the aqueous phase of the emulsion will produce an electrostatic shielding effect on the particles, leading to a change of particle surface charges and adsorption patterns at the interface, thus, changing the stability of the emulsion. Xiao (101), De Folter (102), and Liu (103) used kafirin nanoparticles, waterinsoluble protein zein colloidal particles, and soybean protein to stabilize O/W emulsions, respectively. They found that within a certain range, with the increase of ionic strength of the system, the average droplet size decreased, and the emulsions tended to be more stable. The phenomenon is due to the shielding effect of ionic concentration within a certain range, which can enhance the interaction among particles, leading to the aggregation and migration of particles to the oil-water interface. Thus, the stability of the emulsion was improved. Addition to the alteration of the adsorption behaviors of particles at the oil-water interface, the ionic strength can also alter the hydrophobicity of particles. The ζ-potential of pea protein isolate microgel particle would decrease by increasing the concentration of NaCl, which may trigger the hydrophobic group to be embedded into protein curl, resulting in lower hydrophobicity (92).

# The Size of Solid Particles and Emulsion Droplets

The particle size of solid particles also influences the stability of the emulsions. Theoretically, a harder space obstacle can be formed with a decrease in particle size. So, it can make the solid particles adsorbed pretty closely at the water-oil interface (104), preventing the coalescence among droplets and the instability of emulsion. However, some studies have shown that particle size kept too large or too small may have an adverse effect on the stability of the emulsion. Ge et al. (105) found that the wettability of particles may be changed with the change in particle size. The Pickering emulsions stabilized by sweet potato and corn starch with a diameter ranging from 100–220 nm had much better stability than the particles with a diameter either less than 100 nm or larger than 220 nm. Additionally, the decrease in the size of droplets caused by increasing gel particles strength leads to maintain the stability of emulsions.

# **Oil Type and Volume Fraction**

The type of oil used for the preparation of Pickering emulsion and the radio of the dispersed and continuous phase play important roles in the stability of the emulsion. The oil type determines the interfacial tension of the droplets and the interactions among the particles on the two-phase interface can also be influenced. Tsuji et al. (106) reported that poly (Nisopropylacrylamide) (PNIPAM)-carrying particles tend to form O/W Pickering emulsions with varieties of oil types such as hexadecane, heptane, and trichloroethylene. This kind of oil phase all possesses the work of adhesion (Wa) ranging from 43 to 65 mJ/m<sup>2</sup> related to the surface tension of water, O/W Pickering emulsions are preferably formed. The principle can be extended to food-grade emulsions. Zhang et al. (107) stabilized O/W Pickering emulsions, hexadecane/H<sub>2</sub>O, and decane/H<sub>2</sub>O Pickering emulsions, with poly (sodiump-styrenesulfonate) bush (PS@PSS), by ultrasonic power. The microscope images showed that droplets were significantly bigger and more homogeneous in hexadecane than in decane. When the oil fraction increased from 0.2 to 0.6, the emulsions type changed from O/W to W/O which was stabilized by the water-insoluble phytosterols (60) (**Figure 4B**).

# Temperature

The emulsifying temperature also has effects on food-grade Pickering emulsions. The nanoparticles had poor thermal stability, however, the thermal stability of Pickering emulsions improved with the application of the triple emulsifier (108). In the actual preparation process, the final stability of the emulsion is the result of the interaction of various factors.

# **APPLICATIONS IN THE FOOD FIELD**

There have been some stable food-grade emulsions and they can be a potential candidate for promising functions. In the past few years, Pickering emulsions for their superior performance have been reported in cosmetics, pharmacy, tissue engineering, and the food field (15, 109–111). Especially, the edible particles are able to meet the demand for a "clean label", These particles can act as either the direct stabilizer of Pickering emulsions or indirect materials fabricated by them.

# **Pickering Emulsion for Functional Active Substances Encapsulation**

Pickering emulsion is a good encapsulation agent, which can improve the storage stability of functional active substances and improve their bioavailability. Currently, it has been widely used to carry functional active substances in various fields (112). Based on Pickering emulsion, microcapsules and microspheres are commonly prepared to explore the utilities of a delivery system for bioactive compounds. By emulsifying, active ingredients can be firstly encased in the droplets. The interfaces of the droplets are further stabilized through interfacial assembly, deposition, and the shelled microcapsules can be fabricated by interfacial polymerization (113). According to the hydrophilicity or hydrophobic of functional activities, they can be encapsulated and delivered by Pickering emulsion of different types (O/W or W/O). Curcumin is a hydrophobic active substance extracted from turmeric rhizomes (114), which has a series of pharmacological activities such as anti-inflammatory, antibacterial, and antiulcer activities (115-117). However, the use of curcumin has been limited by low physicochemical stability and water solubility (118). Therefore, a wide variety of encapsulation delivery systems should be applied (118, 119) to improve its stability and solubility (Figure 5A). Shah et al. (120) prepared chitosan-tripolyphosphate nanoparticles (CS-TPP) by ionic gelation techniques to stabilize O/W Pickering emulsion, encapsulated curcumin, and evaluated the stability and

release kinetics of curcumin based on spectrophotometry. The results showed that only 14% of curcumin was degraded within 24 h, which was significantly optimized compared with Leung et al. (121) used sodium dodecyl sulfate (SDS) encapsulation of curcumin, which degraded 40% in 20 h. In addition, the emulsions can also improve the stability of curcumin in acid, alkali, and salt existed environment. In addition to preparing nano-particle stabilized Pickering emulsions by ionic gelation techniques, Lv et al. (122) prepared whey protein isolate (WPI) gel by hydrostatic high-pressure treatment (HHP) for loading and wrapping curcumin. Since WPI could form a compact solidlike gel at pH 5, it has been verified by simulated gastrointestinal digestion experiments in vitro that the release rate of curcumin at pH 5 was slower than that at pH 6.0. On the other hand, because the non-thermal means were used to prepare gelation, it can effectively avoid the generation of undesirable odor. Li et al. (95) prepared Pickering emulsions stabilized by wheat gliadinchitosan nanoparticles with three structures by changing pH and confirmed that the complex formed by wheat gliadin-chitosan and curcumin could effectively reduce lipid oxidation through accelerating thermal storage experiments. Hydrophilic activities such as lutein and anthocyanins can be delivered by W/O Pickering emulsion encapsulated in the water phase. Li et al. (123) stabilized Pickering emulsion gels by octenylsuccinate quinoa starch (OSQS) with different interface activities and structures by adjusting the oil phase volume fraction. This emulsion was used to deliver, and the retention rate of lutein could keep 55.38% after 31 days of storage. Furthermore, multiple Pickering emulsions due to their function of isolation, protection, and targeted release have been used to encapsulate active ingredients to improve their stability to light and oxygen (124). Beicht et al. (125) fabricated multiple emulsions by fish gelatin (FG), whey protein isolate (WPI), and dodecyltrimethylammonium bromide (DTAB), which can also improve the stability of lutein obviously. Wang et al. (126) formulated water-in-oil-in-water (W/O/W) emulsions to encapsulate trans-resveratrol in the internal water phase and oil phase together with an encapsulation efficiency of  $99.97 \pm 0.001\%$ .

# Pickering Emulsion for Existing Food System Replacing

In recent years, the consumption of margarine, shortening, and so on showed a steady growth trend with the pursuit of diverse food and the demand for various special fats. However, most of the edible oil has to be commercially produced by the hydrogenation process, which produced a large number of trans-fatty acids (TFA). It is reported (127, 128) that TFA can increase the incidence of myocardial infarction and coronary heart disease. Therefore, it is crucial to find substitutes for partially hydrogenated vegetable oils. Jiao (93) et al. stabilized a high internal phase Pickering emulsion solely by peanut protein microgel particle whose high viscosity is similar to that of cream, which is expected to become the substitution for margarine containing partially hydrogenated oils. Li et al. (92) fabricated edible egg-free mayonnaise-like Pickering emulsion by pea protein isolate microgels, which showed similar properties

to commercial mayonnaise. Especially, the thixotropy recovery rate can achieve nearly 100%. Typical effects on ζ-potential, the droplet size of the emulsion, such as pH, NaCl, and sucrose, were explored. This is the first study to scale up the processing of mayonnaise-like Pickering emulsion manufacturing at a pilotplant scale. Besides many researchers (91, 129-132) also have made mayonnaise-like Pickering emulsion based on yolk-free materials. Romoscanu et al. (133) prepared a kind of lowviscosity liquid and oleogels based on the protein-template method. It avoided the hydrogenation process and thus TFA would not be produced. Besides, the oleogels had a viscoelastic and crystal structure similar to that of solid fat, which would play a great potential role in the food industry. The study found that (134) nanocellulose can change the rheological properties of liquid food and cellulose nanofiber (CNFs) because of its high transverse and longitudinal. Under the condition of low concentration, nanofibrils can form a gel (135). Meanwhile, the quality and structure strength of the gel can be changed (136). The gel strength will increase, and the gel will keep transparent with surfactants added to it. Conversely, the gel strength will decrease when the surfactant concentration gets high. Velásquez-Cock et al. (137) substituted fat with CNFs to improve the melting physical characteristics of ordinary fat ice cream and found that the melting rate of ordinary ice cream could be reduced with the increase of CNFs content. When the CNFs content reached the maximum, the melting rate decreased by 50%. However, the melting rate of the lowfat ice cream was significantly reduced only when the CNFs content reached 0.3%. This is related to water retention and lipid retention of hydroxyl and amino groups in the mixed system (138). Wang et al. (139) reported a facial method where Pickering emulsion-template was used to stabilize the Pickering emulsion via zein/chitosan colloidal particles (ZCCP) which can transform the liquid oil into solid fat, potential antibacterial and antioxidant properties.

# Pickering Emulsion for Food Packaging Film Preparation

Plastic products have brought great convenience to people's daily life and production-manufacturing since their appearance, but they also produced inevitable negative effects. So far, only 9% of plastic in the world can be recycled, and most of the plastic in the environment is not biodegradable, posing a great threat to animals, plants, and humans. Recently, the use of edible films prepared by using polysaccharides, proteins, and lipids instead of plastic products has attracted considerable attention. Compared with traditional edible films, emulsion films combine the advantages of hydrocolloid (polysaccharide or protein) and lipophilic lipids, which can be used as an effective control barrier for flavor and moisture (140). Among them, Pickering films are synthesized by adding emulsifiers to the lipid in combination with polysaccharides or proteins forming homogeneous emulsions to form Pickering films (141). Konjac glucomannan (KGM) has been considered as a promising material to fabricate Pickering films for its biocompatibility as well non-toxicity (142). Liu et al. (143) investigated the emulsion particle size that played an important role in the properties of KGM-based films. The properties of KGM-based films were improved by incorporating Pickering emulsions with various particle sizes. Some researchers prepared Pickering film encapsulation volatiles by combining polylactic acid film with nanocellulose (144) forming a complementary film. Soluble soybean protein-polysaccharide (SSPS) linked with acid-soluble soybean protein (ASSP) (145) was also used to prepare food film (**Figure 5B**). The membrane has good barrier performance of moisture and oxygen. Furthermore, it has been proved that incorporating essential oil, bioactive compounds, and other functional composition into polysaccharide-or protein-based films could be a promising method to improve the antimicrobial and antioxidant abilities of the films (146).

# Pickering Emulsion for Catalytic Efficiency Improving

The conventional biphasic system (BS) exhibits poor catalysis efficiency, due to the low contact area between the immiscible phases, thus limiting the reaction and production. Pickering emulsions as the carrier of catalytic reaction can be a promising strategy to preferably improve the efficiency of the catalysis reaction, which not only has high catalytic efficiency and selectivity but also is conducive to the recovery and utilization of solid particles and catalysts. It is generally recognized that the interface is significant for enzyme activity because the reaction often occurs at the oil-water interface in Pickering emulsion catalytic systems. Generally, for Pickering emulsion catalysis, it is not necessary to add external force for mechanical mixing, and no traditional surfactant is used in classical emulsions, thus adding particles to divide one phase into droplets within micrometers in another immiscible phase. In this way, as the numerous droplets act as micro-compartments, a dramatically large interface area can be provided, thus enhancing the encounter of the enzyme and substrate. At the same time, because of the existence of the microdroplets, the transfer distance between the catalyst and reactant can be shortened (Figure 5C). Nardello-Rataj et al. (147) defined the Pickering emulsion catalysis (PEC) as two types, Pickering Assisted Catalysis (PAC) as well as Pickering Interfacial Catalysis (PIC). For PAC, the catalysts can be confined in the organic or aqueous phase not be used to stabilize Pickering emulsions. For PIC, Pickering emulsions will be stabilized by catalysts combined with solid particles. In recent years, PEC has been applied in the food field although the particles are not foodgrade (118, 148-153). At present, there exist Pickering emulsions stabilized by edible particles used in the food field to improve the catalytic efficiency of lipases, such as hydrolysis, esterification, and deacidification. The wettability of chitosan was changed in situ modification, which can lead to the interaction triggered by lipase between the free fatty acid and the nanogel particles (118). In this experiment, genipin was used as a cross-linking reagent for lacking physiological toxicity. After 13 batches, the activity of the Pickering emulsion can still remain at 55% of the initial system. For generating free fatty acids, the emulsion would undergo an inversion of phase thus facilitating the separation

of products and reactants. Another study (57) used partially hydrolyzed  $\alpha$  -whey protein ( $\alpha$ -LAC) peptides to form  $\alpha$ -whey protein nanotubes (NTs) and immobilized lipase to improve the catalytic activity of lipase. It was found that free fatty acids

# **CONCLUSION AND OUTLOOK**

hydrolyzed by free lipase.

hydrolyzed by lipase-NTS were 1.5 times as much as those

Food-grade particles stabilized Pickering emulsion has the advantages of environmental friendly, biocompatibility, stable property. Based on their natural properties and feasible modification, they have been widely used in food science. Either stabilizing mechanism or food application to Pickering emulsions stabilized by spherical particles has been mature and widely applied. However, there still exist some challenges in implementing edible Pickering emulsions in commercial applications. In addition, it is expected that understanding materials and applications from the state-of-art technology will enable researchers to modify and design more innovative formulations, which could be more beneficial for exploring food production and processes. Several types of studies should be further explored in the future. (1) Compared with spherical food-grade particles, the mechanism and application of Pickering emulsions stabilized by non-spherical particles need further exploration. The researchers are still required to identify how the nature, application, and stability are related to the shape of particles. More accurate approaches should be adopted to study the interfacial behavior of these irregular shape particles. Without particles adsorbing at the interface of two phases, the interfacial layers affected by the order of addition are worth investigating which can cause diverse interfacial structures (154). (2) Due to the high sensitivity and low bioavailability of bioactive compounds, Pickering emulsions stabilized by hybrid biopolymeric particles have attracted researchers' attention. But there is still a lack of detailed research on the toxicity and allergy of biopolymers although they exhibit a lot of advantages in the food field (155). Besides, the encapsulation efficiency and loading capacity of the Pickering emulsions should be considered. Thus, it is also important to carry out in vivo studies to validate their biological activities. (3) To attain excellent sensorial pleasure, the

# REFERENCES

- Ramsden W. Separation of solids in the surface-layers of solutions and 'suspensions' (observations on surface-membranes, bubbles, emulsions, and mechanical coagulation)—preliminary account. *Proc R Soc Lond.* (1903) 72:156–64. doi: 10.1098/rspl.190 3.0034
- Pickering SU. Cxcvi.—emulsions. J Chem Soc Trans. (1907) 91:2001– 21. doi: 10.1039/CT9079102001
- Hossain KMZ, Deeming L, Edler KJ. Recent progress in pickering emulsions stabilised by bioderived particles. RSC Adv. (2021) 11:39027– 44. doi: 10.1039/D1RA08086E
- Nonomura Y, Kobayashi N, Nakagawa N. Multiple pickering emulsions stabilized by microbowls. *Langmuir*. (2011) 27:4557– 62. doi: 10.1021/la2003707

oral tribology and sensory perception of the Pickering emulsion will also increasingly deserve attention. Generally speaking, for consumers, the rheological and tribological properties of emulsion play a prime role in sensory perception during oral processing (131). So, it is urgent to improve the rheological and tribological properties to improve the palatability of food-based Pickering emulsion. (4) Although we have a relatively clear understanding of the mechanism of HIPEs, the applications of HIPEs still need to be explored, such as desired porous materials. The porous materials can be applied to medical and tissue engineering for moderate and natural crosslinking ways. Besides, the porous materials are to expected modify and design functional materials with exquisite structure and responsiveness. The construction of the porous scaffolds is expected to control the release of drugs and bioactives. (5) There are urgent needs for commercially viable plant-scale manufacturing techniques and commercially available food products based on Pickering emulsions. In practice, matching techniques in lab-scale to real plant-scale processes or equipment is very challenging. (6) More edible particles rather than inorganic solid particles should be explored and applied to form Pickering emulsion catalysis system to achieve high enzyme activity and recovery efficiency in food industry. These edible particles will provide more sustainable and safer for food products.

# **AUTHOR CONTRIBUTIONS**

WL and BJ: conceptualization and writing—original draft. SL: data curation. SF: grammar. WF and YC: polish. AS and QW: supervision. QW: visualization. All authors read and approved the submitted version of the manuscript.

# FUNDING

This research was funded by the National Natural Science Foundation of China (U21A20270 and 32172149), China Postdoctoral Science Foundation (2020M680777 and 2021T140717), and the Agricultural Science and Technology Innovation Program of the Chinese Academy of Agricultural Sciences (CAAS-ASTIP-2020-IFST).

- Maeda H, Okada M, Fujii S, Nakamura Y, Furuzono T. Pickeringtype water-in-oil-in-water multiple emulsions toward multihollow nanocomposite microspheres. *Langmuir*. (2010) 26:13727–31. doi: 10.1021/ la102529d
- Jia H, Dai JJ, Miao LC, Wei X, Tang HT, Huang P, et al. Potential application of novel amphiphilic janus-Sio<sub>2</sub> nanoparticles stabilized O/W/O emulsion for enhanced oil recovery. *Colloids And Surfaces A-Physicochemical And Engineering Aspects*. (2021) 622:126658. doi: 10.1016/j.colsurfa.2021.126658
- Dickinson E. Use of nanoparticles and microparticles in the formation and stabilization of food emulsions. *Trends Food Sci Technol.* (2012) 24:4– 12. doi: 10.1016/j.tifs.2011.09.006
- Sarkar A, Zhang S, Holmes M, Ettelaie R. Colloidal Aspects of Digestion of Pickering Emulsions: Experiments and Theoretical Models of Lipid Digestion Kinetics. *Adv Colloid Interface Sci.* (2019) 263:195– 211. doi: 10.1016/j.cis.2018.10.002

- Tavasoli S, Liu Q, Jafari SM. Development of pickering emulsions stabilized by hybrid biopolymeric particles/nanoparticles for nutraceutical delivery. *Food Hydrocoll.* (2022) 124:107280. doi: 10.1016/j.foodhyd.2021. 107280
- Dickinson E. Biopolymer-based particles as stabilizing agents for emulsions and foams. *Food Hydrocoll.* (2017) 68:219– 31. doi: 10.1016/j.foodhyd.2016.06.024
- Binks BP. Macroporous silica from solid-stabilized emulsion templates. Adv Mater. (2002) 14:1824–7. doi: 10.1002/adma.200290010
- Binks BP, Rodrigues JA. Double inversion of emulsions by using nanoparticles and a di-chain surfactant. Angewandte Chemie International Edition. (2007) 46:5389–92. doi: 10.1002/anie.200700880
- Yan N, Masliyah JH. Characterization and Demulsification of Solids-Stabilized Oil-in-Water Emulsions Part 2. Demulsification by the addition of fresh oil colloids and surfaces a: physicochemical and engineering aspects. (1995) 96:243–52. doi: 10.1016/0927-7757(94)03059-9
- Zhang T, Davidson A, Bryant SL, Huh C. Nanoparticle-Stabilized Emulsions for Applications in Enhanced Oil Recovery. In: SPE Improved Oil Recovery Symposium (Tulsa, OK). (2010). doi: 10.2118/129885-MS
- Wu J, Ma G-H. Recent studies of pickering emulsions: particles make the difference. Small. (2016) 12:4633–48. doi: 10.1002/smll.201600877
- Kargar M, Fayazmanesh K, Alavi M, Spyropoulos F, Norton IT. Investigation into the potential ability of pickering emulsions (food-grade particles) to enhance the oxidative stability of oil-in-water emulsions. J Colloid Interface Sci. (2012) 366:209–15. doi: 10.1016/j.jcis.2011.09.073
- Lu X, Zhang H, Li Y, Huang Q. Fabrication of milled cellulose particles-stabilized pickering emulsions. *Food Hydrocoll.* (2018) 77:427– 35. doi: 10.1016/j.foodhyd.2017.10.019
- Tan H, Sun G, Fau-Lin W, Lin W, Fau-Mu C, Mu C, et al. Gelatin Particle-Stabilized High Internal Phase Emulsions as Nutraceutical Containers. ACS Appl Mater Interfaces. (2014) 6:13977–84. doi: 10.1021/am503341j
- Gonzalez Ortiz D, Pochat-Bohatier C, Cambedouzou J, Bechelany M, Miele P. Current trends in pickering emulsions: particle morphology and applications. *Engineering*. (2020) 6:468–82. doi: 10.1016/j.eng.2019.08.017
- Cossu A, Wang MS, Chaudhari A, Nitin N. Antifungal activity against candida albicans of starch pickering emulsion with thymol or amphotericin B in suspension and calcium alginate films. *Int J Pharm.* (2015) 493:233– 42. doi: 10.1016/j.ijpharm.2015.07.065
- Gao Z, Zhao J, Huang Y, Yao X, Zhang K, Fang Y, et al. Edible Pickering Emulsion Stabilized by Protein Fibrils. Part 1: Effects of Ph and Fibrils Concentration. *LWT-Food Science and Technology*. (2017) 76:1– 8. doi: 10.1016/j.lwt.2016.10.038
- Zhang T, Xu J, Chen J, Wang Z, Wang X, Zhong J. Protein nanoparticles for pickering emulsions: a comprehensive review on their shapes, preparation methods, and modification methods. *Trends Food Sci Technol.* (2021) 113:26–41. doi: 10.1016/j.tifs.2021.04.054
- Chevalier Y, Bolzinger M-A. Emulsions Stabilized with Solid Nanoparticles: Pickering Emulsions. Colloids and Surfaces A: Physicochemical and Engineering Aspects. (2013) 439:23–34. doi: 10.1016/j.colsurfa.2013.02.054
- 24. Binks BP, Horozov TS. Colloidal Particles at Liquid Interfaces. Cambridge University Press (2006).
- 25. Low LE, Siva SP, Ho YK, Chan ES, Tey BT. Recent advances of characterization techniques for the formation, physical properties and stability of pickering emulsion. Adv Colloid Interface Sci. (2020) 277:102117. doi: 10.1016/j.cis.2020.1 02117
- Tavernier I, Wijaya W, Van der Meeren P, Dewettinck K, Patel AR. Food-Grade Particles for Emulsion Stabilization. *Trends Food Sci Technol.* (2016) 50:159–74. doi: 10.1016/j.tifs.2016. 01.023
- Dickinson E. Food emulsions and foams: stabilization by particles. Curr Opin Colloid Interface Sci. (2010) 15:40–9. doi: 10.1016/j.cocis.2009.11.001
- Burgess DJ, Sahin NO. Interfacial rheological and tension properties of protein films. J Colloid Interface Sci. (1997) 189:74–82. doi: 10.1006/jcis.1997.4803
- Dickinson E. Structuring of colloidal particles at interfaces and the relationship to food emulsion and foam stability. J Colloid Interface Sci. (2015) 449:38–45. doi: 10.1016/j.jcis.2014.09.080

- Wasan DT, Nikolov AD, Aimetti F. Texture and Stability of Emulsions and Suspensions: Role of Oscillatory Structural Forces. Advances in Colloid and Interface Science. (2004) 108–9:187–95. doi: 10.1016/j.cis.2003.10.021
- Ribeiro EF, Morell P, Nicoletti VR, Quiles A, Hernando I. Protein- and polysaccharide-based particles used for pickering emulsion stabilisation. *Food Hydrocoll.* (2021) 119:106839. doi: 10.1016/j.foodhyd.2021.106839
- Sarkar A, Dickinson E. Sustainable food-grade pickering emulsions stabilized by plant-based particles. *Curr Opin Colloid Interface Sci.* (2020) 49:69– 81. doi: 10.1016/j.cocis.2020.04.004
- 33. Xu YT, Tang CH, Binks BP. Ultraefficient stabilization of high internal phase emulsions by globular proteins in the presence of polyols: importance of a core-shell nanostructure. *Food Hydrocoll.* (2020) 107:105968. doi: 10.1016/j.foodhyd.2020.105968
- Rayner M, Marku D, Eriksson M, Sjöö M, Dejmek P, Wahlgren M. Biomassbased particles for the formulation of pickering type emulsions in food and topical applications. *Colloids and Surfaces A: Physicochemical and Engineering Aspects.* (2014) 458:48–62. doi: 10.1016/j.colsurfa.2014.03.053
- 35. Shi A, Feng X, Wang Q, Adhikari B. Pickering and high internal phase pickering emulsions stabilized by protein-based particles: a review of synthesis, application and prospective. *Food Hydrocoll.* (2020) 109:106117. doi: 10.1016/j.foodhyd.2020.106117
- 36. Zhang T, Ding M, Tao L, Liu L, Tao N, Wang X, et al. Octenyl succinic anhydride modification of bovine bone and fish skin gelatins and their application for fish oil-loaded emulsions. *Food Hydrocoll.* (2020) 108:106041. doi: 10.1016/j.foodhyd.2020.106041
- Tong Q, Yi Z, Ran Y, Chen X, Chen G, Li X. Green tea polyphenol-stabilized gel-like high internal phase pickering emulsions. ACS Sustain Chem Eng. (2021) 9:4076–90. doi: 10.1021/acssuschemeng.0c08633
- Zhang X, Zuo Z, Yu P, Li T, Guang M, Chen Z, et al. Rice peptide nanoparticle as a bifunctional food-grade pickering stabilizer prepared by ultrasonication: structural characteristics, antioxidant activity, and emulsifying properties. *Food Chem.* (2021) 343:128545. doi: 10.1016/j.foodchem.2020.128545
- Shah BR, Xu W, Mráz J. Formulation and characterization of zein/chitosan complex particles stabilized pickering emulsion with the encapsulation and delivery of vitamin D3. J Sci Food Agric. (2021). doi: 10.1002/jsfa.11190
- Li F, Li X, Huang K, Luo Y, Mei X. Preparation and characterization of pickering emulsion stabilized by hordein-chitosan complex particles. *J Food Eng.* (2021) 292:110275. doi: 10.1016/j.jfoodeng.2020.110275
- 41. Liu Q, Chang X, Shan Y, Fu F, Ding S. Fabrication and characterization of pickering emulsion gels stabilized by zein/pullulan complex colloidal particles. *J Sci Food Agric.* (2021) 101:3630–43. doi: 10.1002/jsfa.10992
- 42. Wang L, Lu S, Deng Y, Wu W, Wang L, Liu Y, et al. Pickering emulsions stabilized by luteolin micro-nano particles to improve the oxidative stability of pine nut oil. *J Sci Food Agric*. (2021) 101:1314–22. doi: 10.1002/jsfa.10739
- Hazt B, Bassani HP, Elias-Machado JP, Aldinucci Buzzo JL, Silveira JLM, de Freitas RA. Effect of ph and protein particle shape on the stability of amylopectin–xyloglucan water-in-water emulsions. *Food Hydrocoll.* (2020) 104:105769. doi: 10.1016/j.foodhyd.2020.105769
- 44. Spasojević L, Bučko S, Kovačević D, Bohinc K, Jukić J, Abram A, et al. Interactions of zein and zein/rosin nanoparticles with natural polyanion gum arabic. *Colloids Surf B Biointerfaces*. (2020) 196:111289. doi: 10.1016/j.colsurfb.2020.111289
- Ngwabebhoh FA, Erdagi SI, Yildiz U. Pickering emulsions stabilized nanocellulosic-based nanoparticles for coumarin and curcumin nanoencapsulations: *in vitro* release, anticancer and antimicrobial activities. *Carbohydr Polym.* (2018) 201:317–28. doi: 10.1016/j.carbpol.2018.08.079
- Kasiri N, Fathi M. Production of cellulose nanocrystals from pistachio shells and their application for stabilizing pickering emulsions. *Int J Biol Macromol.* (2018) 106:1023–31. doi: 10.1016/j.ijbiomac.2017.08.112
- Liu F, Zheng J, Huang C-H, Tang C-H, Ou S-Y. Pickering high internal phase emulsions stabilized by protein-covered cellulose nanocrystals. *Food Hydrocoll.* (2018) 82:96–105. doi: 10.1016/j.foodhyd.2018.03.047
- Kalashnikova I, Bizot H, Bertoncini P, Cathala B, Capron I. Cellulosic nanorods of various aspect ratios for oil in water pickering emulsions. *Soft Matter.* (2013) 9:952–9. doi: 10.1039/C2SM26472B
- Lehle H, Noruzifar E, Fau-Oettel M, Oettel M. Ellipsoidal particles at fluid interfaces. *Eur Phys J E Soft Matter*. (2008) 26:151–60. doi: 10.1140/epje/i2007-10314-1

- Loudet J-C, Yodh AG, Pouligny B. Wetting and contact lines of micrometer-sized ellipsoids. *Physical Review Letters*. (2006) 97:18304. doi: 10.1103/PhysRevLett.97.018304
- Madivala B, Vandebril S, Fransaer J, Vermant J. Exploiting particle shape in solid stabilized emulsions. *Soft Matter*. (2009) 5:1717–27. doi: 10.1039/b816680c
- Winuprasith T, Suphantharika M. Properties and stability of oil-in-water emulsions stabilized by microfibrillated cellulose from mangosteen rind. *Food Hydrocoll.* (2015) 43:690–9. doi: 10.1016/j.foodhyd.2014.07.027
- Chen Q-H, Zheng J, Xu Y-T, Yin S-W, Liu F, Tang C-H. Surface modification improves fabrication of pickering high internal phase emulsions stabilized by cellulose nanocrystals. *Food Hydrocoll.* (2018) 75:125–30. doi: 10.1016/j.foodhyd.2017.09.005
- Sarker M, Tomczak N, Lim SA-O. Protein nanocage as a phswitchable pickering emulsifier. ACS Appl Mater Interfaces. (2017) 9:11193–201. doi: 10.1021/acsami.6b14349
- 55. Hikima T, Nonomura Y. Pickering emulsions and capsules stabilized by wool powder particles. *J Oleo Sci.* (2011) 60:351–4. doi: 10.5650/jos.60.351
- Hikima T, Nonomura Y. Powderization of wool keratin by alkali hydrolysis in higher alcohol/water binary systems. *Chem Lett.* (2008) 37:338– 9. doi: 10.1246/cl.2008.338
- 57. Guan T, Liu B, Wang R, Huang Y, Luo J, Li Y. The enhanced fatty acids flavor release for low-fat cheeses by carrier immobilized lipases on O/W pickering emulsions. *Food Hydrocoll.* (2021) 116:106651. doi: 10.1016/j.foodhyd.2021.106651
- Paunov VN, Cayre OJ, Noble PF, Stoyanov SD, Velikov KP, Golding M. Emulsions stabilised by food colloid particles: role of particle adsorption and wettability at the liquid interface. J Colloid Interface Sci. (2007) 312:381– 9. doi: 10.1016/j.jcis.2007.03.031
- Binks B, Clint JH, Mackenzie G, Simcock C, Whitby CP. Naturally occurring spore particles at planar fluid interfaces and in emulsions. *Langmuir*. (2005) 18:8161–7. doi: 10.1021/la0513858
- Liu F, Tang C-H. Phytosterol colloidal particles as pickering stabilizers for emulsions. J Agric Food Chem. (2014) 62:5133–41. doi: 10.1021/jf404930c
- Davidov-Pardo G, Joye IJ, McClements DJ. Chapter nine food-grade protein-based nanoparticles and microparticles for bioactive delivery: fabrication, characterization, and utilization. *Adv Protein Chem Struct Biol.* (2015) 98:293–325. doi: 10.1016/bs.apcsb.2014.11.004
- Liu F, Ma C, McClements DJ, Gao Y. A comparative study of covalent and non-covalent interactions between zein and polyphenols in ethanol-water solution. *Food Hydrocoll.* (2017) 63:625–34. doi: 10.1016/j.foodhyd.2016.09.041
- Vu Huu T, Shibasaki K. Major Proteins of Soybean Seeds. Reconstitution of beta-conglycinin from its subunits. J Agric Food Chem. (1978) 26:695– 8. doi: 10.1021/jf60217a027
- Lakemond CMM, de Jongh HHJ, Hessing M, Gruppen H, Voragen AGJ. Soy glycinin: influence of ph and ionic strength on solubility and molecular structure at ambient temperatures. J Agric Food Chem. (2000) 48:1985– 90. doi: 10.1021/jf9908695
- Nicolai T, Chassenieux C. Heat-induced gelation of plant globulins. Current Opinion in Food Science. (2019) 27:18–22. doi: 10.1016/j.cofs.2019.04.005
- Nishinari K, Fang Y, Guo S, Phillips GO. Soy proteins: a review on composition, aggregation and emulsification. *Food Hydrocoll.* (2014) 39:301– 18. doi: 10.1016/j.foodhyd.2014.01.013
- 67. Nugmanova AG, Kalinina MA. Self-assembly of metal-organic frameworks in pickering emulsions stabilized with graphene oxide. *Colloid J.* (2021) 83:614–26. doi: 10.1134/S1061933X21050094
- Graveland-Bikker JF, de Kruif CG. Unique Milk Protein Based Nanotubes: Food and Nanotechnology Meet. *Trends Food Sci Technol.* (2006) 17:196– 203. doi: 10.1016/j.tifs.2005.12.009
- Zou Q, Habermann-Rottinghaus Sm Fau-Murphy KP, Murphy KP. Urea effects on protein stability: hydrogen bonding and the hydrophobic effect. *Proteins: Structure, Function, and Bioinformatics.* (1998) 31:107–15.
- Zheng HG. Preparation of soluble soybean protein aggregates (Sspa) from insoluble soybean protein concentrates (Spc) and its functional properties. *Food Res Int.* (2008) 41:154–63. doi: 10.1016/j.foodres.2007.10.013
- 71. Queirós RA-O, Saraiva JA-O, da Silva JA-O. Tailoring structure and technological properties of plant proteins using high

hydrostatic pressure. Crit Rev Food Sci Nutr. (2018) 58:1538-56. doi: 10.1080/10408398.2016.1271770

- Zhang H, Chen Y, Wang S, Ma L, Yu Y, Dai H, et al. Extraction and comparison of cellulose nanocrystals from lemon (citrus limon) seeds using sulfuric acid hydrolysis and oxidation methods. *Carbohydr Polym.* (2020) 238:116180. doi: 10.1016/j.carbpol.2020.116180
- Zoppe JO, Venditti RA, Rojas OJ. Pickering emulsions stabilized by cellulose nanocrystals grafted with thermo-responsive polymer brushes. J Colloid Interface Sci. (2012) 369:202–9. doi: 10.1016/j.jcis.2011.12.011
- Dutta S, Lim K-T. Micro and Nano Technologies. Abd-Elsalam KA, editor. Elsevier (2020). p. 461–83.
- Loudet JC, Alsayed AM, Zhang J, Yodh AG. Capillary interactions between anisotropic colloidal particles. *Phys Rev Lett.* (2005) 94:018301. doi: 10.1103/PhysRevLett.94.018301
- Vella D, Mahadevan L. The "Cheerios Effect". Am J Phys. (2005) 73:817– 25. doi: 10.1119/1.1898523
- Dinsmore AD, Hsu MF, Nikolaides MG, Marquez M, Bausch AR, Weitz DA. Colloidosomes: selectively permeable capsules composed of colloidal particles. *Science*. (2002) 298:1006–9. doi: 10.1126/science.1074868
- Danov KD, Kralchevska SD, Kralchevsky PA, Ananthapadmanabhan KP, Lips A. Mixed solutions of anionic and zwitterionic surfactant (betaine): surface-tension isotherms, adsorption, and relaxation kinetics. *Langmuir*. (2004) 20:5445–53. doi: 10.1021/la049576i
- Tordeux C, Fournier JB, Galatola P. Analytical characterization of adhering vesicles. *Physical Review*. (2002) 65:041912–21. doi: 10.1103/PhysRevE.65.041912
- Kralchevsky PA, Denkov ND. Capillary Forces and Structuring in Layers of Colloid Particles. *Current Opinion in Colloid and Interface Science*. (2001) 6:383–401. doi: 10.1016/S1359-0294(01)00105-4
- Stamou D, Duschl C, Johannsmann D. Long-range attraction between colloidal spheres at the air-water interface: the consequence of an irregular meniscus. *Physical Review*. (2000) 62:5263–72. doi: 10.1103/PhysRevE.62.5263
- Kralchevsky PA, Nagayama K. Capillary interactions between particles bound to interfaces, liquid films and biomembranes. *Adv Colloid Interface Sci.* (2000) 85:145–92. doi: 10.1016/S0001-8686(99)00016-0
- Pääkkö M, Ankerfors M, Kosonen H, Nykänen A, Ahola S, Österberg M, et al. Enzymatic hydrolysis combined with mechanical shearing and highpressure homogenization for nanoscale cellulose fibrils and strong gels. *Biomacromolecules*. (2007) 8:1934–41. doi: 10.1021/bm061215p
- Xhanari K, Syverud K, Stenius P. Emulsions stabilized by microfibrillated cellulose: the effect of hydrophobization, concentration and O/W ratio. J Dispers Sci Technol. (2011) 32:447–52. doi: 10.1080/01932691003658942
- Souza AG, Ferreira RR, Paula LC, Setz LFG, Rosa DS. The effect of essential oil chemical structures on pickering emulsion stabilized with cellulose nanofibrils. J Mol Liq. (2020) 320:114458. doi: 10.1016/j.molliq.2020.114458
- Gestranius M, Stenius P, Kontturi E, Sjöblom J, Tammelin T. Phase Behaviour and Droplet size of oil-in-water pickering emulsions stabilised with plantderived nanocellulosic materials. *Colloids and Surfaces a Physicochemical and Engineering aspects.* (2017) 519:60–70. doi: 10.1016/j.colsurfa.2016. 04.025
- Sarker M, Lee H, Gonçalves RA, Lam YM, Su H, Lim S. Supramolecular protein assembly retains its structural integrity at liquid-liquid interface. *Appl Mater Interfaces.* (2020) 7:1901674. doi: 10.1002/admi.201901674
- Xiao M, Xu A, Zhang T, Hong L. Tailoring the wettability of colloidal particles for pickering emulsions *via* surface modification and roughness. *Front Chem.* (2018) 6:225. doi: 10.3389/fchem.2018.00225
- Tsabet È, Fradette L. Effect of the Properties of oil, particles, and water on the production of pickering emulsions. *Chem Eng Res Des.* (2015) 97:9– 17. doi: 10.1016/j.cherd.2015.02.016
- Kim I, Worthen AJ, Lotfollahi M, Johnston KP, DiCarlo DA, Huh C. Nanoparticle-stabilized emulsions for improved mobility control for adverse-mobility waterflooding. (2017): 2214–4609. doi: 10.3997/2214-4609.201700314
- Hosseini RS, Rajaei A. Potential pickering emulsion stabilized with chitosan-stearic acid nanogels incorporating clove essential oil to produce fish-oil-enriched mayonnaise. *Carbohydr Polym.* (2020) 241:116340. doi: 10.1016/j.carbpol.2020.116340

- 92. Li S, Jiao B, Meng S, Fu W, Faisal S, Li X, et al. Edible mayonnaiselike pickering emulsion stabilized by pea protein isolate microgels: effect of food ingredients in commercial mayonnaise recipe. *Food Chem.* (2021):131866. doi: 10.1016/j.foodchem.2021.131866
- 93. Jiao B, Shi A, Wang Q, Binks BP. High-internal-phase pickering emulsions stabilized solely by peanut-protein-isolate microgel particles with multiple potential applications. *Angew Chem Int Ed.* (2018) 57:9274– 8. doi: 10.1002/anie.201801350
- Liu H, Wang C, Zou S, Wei Z, Tong Z. Simple, reversible emulsion system switched by ph on the basis of chitosan without any hydrophobic modification. *Langmuir.* (2012) 28:11017–24. doi: 10.1021/la30 21113
- Li M-F, He Z-Y, Li G-Y, Zeng Q-Z, Su D-X, Zhang J-L, et al. The formation and characterization of antioxidant pickering emulsions: effect of the interactions between gliadin and chitosan. *Food Hydrocoll.* (2019) 90:482–9. doi: 10.1016/j.foodhyd.2018.12.052
- Tcholakova S, Denkov Nd, Fau Lips A, Lips A. Comparison of Solid Particles, Globular Proteins and Surfactants as Emulsifiers. *Phys Chem Chem Phys.* (2008) 10:1608–27. doi: 10.1039/b715933c
- Destribats M, Rouvet M, Gehin-Delval C, Schmitt C, Binks BP. Emulsions stabilised by whey protein microgel particles: towards food-grade pickering emulsions. *Soft Matter*. (2014) 10:6941–54. doi: 10.1039/C4SM00179F
- Arditty S, Schmitt V, Giermanska-Kahn J, Leal-Calderon F. Materials based on solid-stabilized emulsions. J Colloid Interface Sci. (2004) 275:659– 64. doi: 10.1016/j.jcis.2004.03.001
- Arditty S, Whitby CP, Binks BP, Schmitt V, Leal-Calderon F. Some General Features of Limited Coalescence in Solid-Stabilized Emulsions. *The European Physical Journal E.* (2003) 11:273–81. doi: 10.1140/epje/i2003-10018-6
- 100. Mwangi WW, Ho K-W, Tey B-T, Chan E-S. Effects of environmental factors on the physical stability of pickeringemulsions stabilized by chitosan particles. *Food Hydrocoll.* (2016) 60:543–50. doi: 10.1016/j.foodhyd.2016.04.023
- Xiao J, Wang Xa, Perez Gonzalez AJ, Huang Q. Kafirin nanoparticlesstabilized pickering emulsions: microstructure and rheological behavior. *Food Hydrocolloids*. (2016) 54:30–9. doi: 10.1016/j.foodhyd.2015.09.008
- 102. de Folter JWJ, van Ruijven MWM, Velikov KP. Oil-in-water pickering emulsions stabilized by colloidal particles from the water-insoluble protein zein. Soft Matter. (2012) 8:6807–15. doi: 10.1039/c2sm07417f
- Liu F, Tang C-H. Soy protein nanoparticle aggregates as pickering stabilizers for oil-in-water emulsions. J Agric Food Chem. (2013) 61:8888– 98. doi: 10.1021/jf401859y
- 104. Nan F, Wu J, Qi F, Fan Q, Ma G, Ngai T. Preparation of uniformsized colloidosomes based on chitosan-coated alginate particles and its application for oral insulin delivery. J Mater Chem B. (2014) 2:7403– 9. doi: 10.1039/C4TB01259C
- 105. Ge S, Xiong L, Li M, Liu J, Yang J, Chang R, et al. Characterizations of pickering emulsions stabilized by starch nanoparticles: influence of starch variety and particle size. *Food Chem.* (2017) 234:339–47. doi: 10.1016/j.foodchem.2017.04.150
- Tsuji S, Kawaguchi H. thermosensitive pickering emulsion stabilized by poly(N-isopropylacrylamide)-carrying particles. *Langmuir*. (2008) 24:3300– 5. doi: 10.1021/la701780g
- 107. Zhang Y, Chen K, Cao L, Li K, Wang Q, Fu E, et al. Stabilization of pickering emulsions by hairy nanoparticles bearing polyanions. *Polymers*. (2019) 11:816. doi: 10.3390/polym11050816
- Sharma T, Kumar GS, Chon BH, Sangwai JS. Thermal stability of oilin-water pickering emulsion in the presence of nanoparticle, surfactant, and polymer. J Ind Eng Chem. (2015) 22:324–34. doi: 10.1016/j.jiec.2014. 07.026
- 109. Marto J, Gouveia LF, Gonçalves L, Chiari-Andréo BG, Isaac V, Pinto P, et al. Design of novel starch-based pickering emulsions as platforms for skin photoprotection. J Photochem Photobiol. (2016) 162:56–64. doi: 10.1016/j.jphotobiol.2016.06.026
- 110. Shen P, Zhao M, Zhou F. Design of soy protein/peptidebased colloidal particles and their role in controlling the lipid digestion of emulsions. *Current Opinion in Food Science*. (2022) 43:61–70. doi: 10.1016/j.cofs.2021.10.003

- 111. Zhou L, Luo X, Gao J, Liu G, Ma L, He Y, et al. Facile synthesis of covalent organic framework derived Fe-Cofs composites as a peroxidase-mimicking artificial enzyme. *Nanoscale Advances*. (2020) 2:1036–9. doi: 10.1039/D0NA00025F
- McClements DJ. Advances in fabrication of emulsions with enhanced functionality using structural design principles. *Curr Opin Colloid Interface Sci.* (2012) 17:235–45. doi: 10.1016/j.cocis.2012.06.002
- 113. Zhang T, Liu F, Wu J, Ngai T. Pickering emulsions stabilized by biocompatible particles: a review of preparation, bioapplication, and perspective. *Particuology*. (2022) 64:110– 20. doi: 10.1016/j.partic.2021.07.003
- 114. Fan L, Lu Y, Ouyang X-k, Ling J. Development and characterization of soybean protein isolate and fucoidan nanoparticles for curcumin encapsulation. *Int J Biol Macromol.* (2021) 169:194– 205. doi: 10.1016/j.ijbiomac.2020.12.086
- 115. Mor N, Raghav N. Alginate hydrogels: sustained release system to analyze the effect of traditional excipients on curcumin availability. *Bioorg Chem.* (2021) 107:104513. doi: 10.1016/j.bioorg.2020.104513
- 116. Adahoun MA, Al-Akhras MH, Jaafar MS, Bououdina M. Enhanced anti-cancer and antimicrobial activities of curcumin nanoparticles. Artif Cells Nanomed Biotechnol. (2017) 45:98– 107. doi: 10.3109/21691401.2015.1129628
- Rauf A, Imran M, Orhan IE, Bawazeer S. Health perspectives of a bioactive compound curcumin: a review. *Trends Food Sci Technol.* (2018) 74:33– 45. doi: 10.1016/j.tifs.2018.01.016
- 118. Huang XM, Luo ZJ, Guo JA-O, Ruan QJ, Wang JA-O, Yang XQ. Enzyme-adsorbed chitosan nanogel particles as edible pickering interfacial biocatalysts and lipase-responsive phase inversion of emulsions. *J Agric Food Chem.* (2020) 68:8890–9. doi: 10.1021/acs.jafc.0c00116
- 119. Xue J, Wang T, Hu Q, Zhou M, Luo Y. Insight into natural biopolymer-emulsified solid lipid nanoparticles for encapsulation of curcumin: effect of loading methods. *Food Hydrocoll.* (2018) 79:110–6. doi: 10.1016/j.foodhyd.2017.12.018
- 120. Shah BR, Li Y, Jin W, An Y, He L, Li Z, et al. Preparation and optimization of pickering emulsion stabilized by chitosan-tripolyphosphate nanoparticles for curcumin encapsulation. *Food Hydrocoll.* (2016) 52:369– 77. doi: 10.1016/j.foodhyd.2015.07.015
- Leung MH, Colangelo H, Fau-Kee TW, Kee TW. Encapsulation of curcumin in cationic micelles suppresses alkaline hydrolysis. *Langmuir*. (2008) 24:5672–5. doi: 10.1021/la800780w
- 122. Lv P, Wang D, Dai L, Wu X, Gao Y, Yuan F. Pickering emulsion gels stabilized by high hydrostatic pressure-induced whey protein isolate gel particles: characterization and encapsulation of curcumin. *Food Res Int.* (2020) 132:109032. doi: 10.1016/j.foodres.2020.109032
- 123. Li S, Zhang B, Li C, Fu X, Huang Q. Pickering emulsion gel stabilized by octenylsuccinate quinoa starch granule as lutein carrier: role of the gel network. *Food Chem.* (2020) 305:125476. doi: 10.1016/j.foodchem.2019.125476
- 124. Zhao H, Yang Y, Chen Y, Li J, Wang L, Li C, et al. Review of multiple pickering emulsions: solid stabilization, preparation, particle effect, and application. *Chem Eng Sci.* (2022) 248:117085. doi: 10.1016/j.ces.2021.117085
- 125. Beicht J, Zeeb B, Gibis M, Fischer L, Weiss J. Influence of layer thickness and composition of cross-linked multilayered oil-in-water emulsions on the release behavior of lutein. *Food Funct.* (2013) 4:1457– 67. doi: 10.1039/c3fo60220f
- 126. Wang J, Shi A, Agyei D, Wang Q. Formulation of water-in-oil-in-water (W/O/W) emulsions containing trans-resveratrol. RSC Adv. (2017) 7:35917– 27. doi: 10.1039/C7RA05945K
- 127. Mat Dian NLH, Sundram K, Idris NA. Dsc study on the melting properties of palm oil, sunflower oil, and palm kernel olein blends before and after chemical interesterification. J Am Oil Chem Soc. (2006) 83:739– 45. doi: 10.1007/s11746-006-5032-z
- 128. Micha R, Mozaffarian D. Trans fatty acids: effects on cardiometabolic health and implications for policy. *Prostaglandins, Leukotrienes and Essential Fatty Acids.* (2008) 79:147–52. doi: 10.1016/j.plefa.2008. 09.008
- 129. Chivero P, Gohtani S, Yoshii H, Nakamura A. Assessment of soy soluble polysaccharide, gum arabic and osa-starch as emulsifiers for

mayonnaise-like emulsions. LWT-Food Science and Technology. (2016) 69:59-66. doi: 10.1016/j.lwt.2015.12.064

- Firoozmand H, Rousseau D. Microbial cells as colloidal particles: pickering oil-in-water emulsions stabilized by bacteria and yeast. *Food Res Int.* (2016) 81:66–73. doi: 10.1016/j.foodres.2015.10.018
- 131. Lu Z, Zhou S, Ye F, Zhou G, Gao R, Qin D, et al. A novel cholesterolfree mayonnaise made from pickering emulsion stabilized by apple pomace particles. *Food Chem.* 353:129418. doi: 10.1016/j.foodchem.2021.129418
- Liu X, Guo J, Wan Z-L, Liu Y-Y, Ruan Q-J, Yang X-Q. Wheat gluten-stabilized high internal phase emulsions as mayonnaise replacers. *Food Hydrocoll.* (2018) 77:168–75. doi: 10.1016/j.foodhyd.2017.09.032
- Romoscanu A, Mezzenga R. Emulsion-templated fully reversible protein-inoil gels. *Langmuir*. (2006) 22:7812–8. doi: 10.1021/la060878p
- Nechyporchuk O, Belgacem MN, Pignon F. Current progress in rheology of cellulose nanofibril suspensions. *Biomacromolecules*. (2016) 17:2311– 20. doi: 10.1021/acs.biomac.6b00668
- Aaen R, Simon S, Wernersson Brodin F, Syverud K. The potential of tempo-oxidized cellulose nanofibrils as rheology modifiers in food systems. *Cellulose.* (2019) 26:5483–96. doi: 10.1007/s10570-019-02448-3
- Quennouz N, Hashmi SM, Choi HS, Kim JW, Osuji CO. Rheology of cellulose nanofibrils in the presence of surfactants. Soft Matter. (2016) 12:157–64. doi: 10.1039/C5SM01803J
- Velásquez-Cock J, Serpa A, Vélez L, Gañán P, Gómez Hoyos C, Castro C, et al. Influence of cellulose nanofibrils on the structural elements of ice cream. *Food Hydrocoll.* (2019) 87:204–13. doi: 10.1016/j.foodhyd.2018.07.035
- 138. Sun L, Chen W, Liu Y, Li J, Yu H. Soy protein isolate/cellulose nanofiber complex gels as fat substitutes: rheological and textural properties and extent of cream imitation. *Cellulose*. (2015) 22:2619– 27. doi: 10.1007/s10570-015-0681-4
- 139. Wang L-J, Yin S-W, Wu L-Y, Qi J-R, Guo J, Yang X-Q. Fabrication and characterization of pickering emulsions and oil gels stabilized by highly charged zein/chitosan complex particles (Zccps). *Food Chem.* (2016) 213:462–9. doi: 10.1016/j.foodchem.2016.06.119
- 140. Chu Y, Xu T, Gao C, Liu X, Zhang N, Feng X, et al. Evaluations of physicochemical and biological properties of pullulan-based films incorporated with cinnamon essential oil and tween 80. *Int J Biol Macromol.* (2019) 122:388–94. doi: 10.1016/j.ijbiomac.2018.10.194
- 141. Bonilla J, Atarés L, Vargas M, Chiralt A. Effect of essential oils and homogenization conditions on properties of chitosan-based films. *Food Hydrocoll.* (2012) 26:9–16. doi: 10.1016/j.foodhyd.2011. 03.015
- 142. Hassan B, Chatha SAS, Hussain AI, Zia KM, Akhtar N. Recent advances on polysaccharides, lipids and protein based edible films and coatings: a review. *Int J Biol Macromol.* (2018) 109:1095–107. doi: 10.1016/j.ijbiomac.2017.11.097
- 143. Liu Z, Shen R, Yang X, Lin D. Characterization of a novel konjac glucomannan film incorporated with pickering emulsions: effect of the emulsion particle sizes. *Int J Biol Macromol.* (2021) 179:377– 87. doi: 10.1016/j.ijbiomac.2021.02.188
- 144. Fortunati E, Peltzer M, Armentano I, Torre L, Jiménez A, Kenny JM. Effects of Modified cellulose nanocrystals on the barrier and migration properties of Pla nano-biocomposites. *Carbohydr Polym.* (2012) 90:948–56. doi: 10.1016/j.carbpol.2012. 06.025
- 145. Liu Q-R, Wang W, Qi J, Huang Q, Xiao J. Oregano essential oil loaded soybean polysaccharide films: effect of pickering type immobilization on physical and antimicrobial properties. *Food Hydrocoll.* (2019) 87:165–72. doi: 10.1016/j.foodhyd.2018. 08.011

- 146. Atef M, Rezaei M, Behrooz R. Characterization of physical, mechanical, and antibacterial properties of agar-cellulose bionanocomposite films incorporated with savory essential oil. *Food Hydrocoll.* (2015) 45:150– 7. doi: 10.1016/j.foodhyd.2014.09.037
- 147. Pera-Titus M, Leclercq L, Fau-Clacens J-M, Clacens Jm, Fau-De Campo F, De Campo F, et al. Pickering Interfacial Catalysis for Biphasic Systems: From Emulsion Design to Green Reactions. *Angew Chem Int.* (2015) 54:2006– 21. doi: 10.1002/anie.201402069
- 148. Dong Z, Liu Z, Shi J, Tang H, Xiang X, Huang F, et al. Carbon nanoparticle-stabilized pickering emulsion as a sustainable and high-performance interfacial catalysis platform for enzymatic esterification/transesterification. ACS Sustain Chem Eng. (2019) 7:7619–29. doi: 10.1021/acssuschemeng.8b05908
- 149. Wang T, Chen K, Li Z, Jiang L, Yu D, Cheng J, et al. Construction of an enzyme-pickering emulsion catalytic system and its application in the interfacial catalytic reaction of rice bran oil deacidification. *LWT-Food Science and Technology.* (2021) 150:111921–31. doi: 10.1016/j.lwt.2021.111921
- 150. Yang T, Zhang Y, Wang J, Huang F, Zheng M. Magnetic switchable pickering interfacial biocatalysis: one-pot cascade synthesis of phytosterol esters from high-acid value oil. ACS Sustain Chem Eng. (2021) 9:12070– 8. doi: 10.1021/acssuschemeng.1c02448
- 151. Yang X, Wang Y, Bai R, Ma H, Wang W, Sun H, et al. Pickering emulsionenhanced interfacial biocatalysis: tailored alginate microparticles act as particulate emulsifier and enzyme carrier. *Green Chemistry*. (2019) 21:2229– 33. doi: 10.1039/C8GC03573C
- 152. Wang M, Wang M, Zhang S, Chen J. Pickering gel emulsion stabilized by enzyme immobilized polymeric nanoparticles: a robust and recyclable biocatalyst system for biphasic catalysis. *React Chem Eng.* (2019) 4:1459– 65. doi: 10.1039/C9RE00158A
- 153. Sun T, Dong Z, Wang J, Huang F-H, Zheng M-M. Ultrasound-assisted interfacial immobilization of lipase on hollow mesoporous silica spheres in a pickering emulsion system: a hyperactive and sustainable biocatalyst. ACS Sustain Chem Eng. (2020) 8:17280–90. doi: 10.1021/acssuschemeng.0c06271
- Cui F, Zhao S, Guan X, McClements DJ, Liu X, Liu F, et al. Polysaccharidebased pickering emulsions: formation, stabilization and applications. *Food Hydrocoll.* (2021) 119:106812. doi: 10.1016/j.foodhyd.2021.106812
- 155. Abdullah WJ, Ahmad T, Zhang C, Zhang H. A review of recent progress on high internal-phase pickering emulsions in food science. *Trends Food Sci Technol.* (2020) 106:91–103. doi: 10.1016/j.tifs.2020.10.016

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Li, Jiao, Li, Faisal, Shi, Fu, Chen and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

17