

Communication

# Antibody Responses after a Third Dose of COVID-19 Vaccine in Kidney Transplant Recipients and Patients Treated for Chronic Lymphocytic Leukemia

Julien Marlet <sup>1,2,\*</sup> , Philippe Gatault <sup>3</sup>, Zoha Maakaroun <sup>4,5</sup> , H el ene Longuet <sup>3</sup>, Karl Stefic <sup>1,2</sup> ,  
Lynda Handala <sup>1,2</sup>, S ebastien Eymieux <sup>1,2,6</sup> , Emmanuel Gyan <sup>7</sup>, Caroline Dartigeas <sup>7</sup>  
and Catherine Gaudy-Graffin <sup>1,2</sup>

- <sup>1</sup> INSERM U1259, Universit e de Tours et CHRU de Tours, 37000 Tours, France; karl.stefic@univ-tours.fr (K.S.); L.HANDALA@chu-tours.fr (L.H.); sebastien.eymieux@univ-tours.fr (S.E.); catherine.gaudy-graffin@univ-tours.fr (C.G.-G.)
- <sup>2</sup> Service de Bact riologie-Virologie-Hygi ne, CHRU de Tours, 37000 Tours, France
- <sup>3</sup> Transplantation r enale-Immunologie clinique, CHRU de Tours, 37000 Tours, France; philippe.gatault@univ-tours.fr (P.G.); H.LONGUET@chu-tours.fr (H.L.)
- <sup>4</sup> Centre de vaccination, CHRU de Tours, 37000 Tours, France; Z.MAAKAROUN-VERMESSE@chu-tours.fr
- <sup>5</sup> Service de m decine p diatrique, CHRU de Tours, 37000 Tours, France
- <sup>6</sup> Plate-Forme IBiSA de Microscopie Electronique, Universit e de Tours and CHRU de Tours, 37000 Tours, France
- <sup>7</sup> H matologie et Th rapie Cellulaire, CHRU de Tours, 37000 Tours, France; E.GYAN@chu-tours.fr (E.G.); c.dartigeas@chu-tours.fr (C.D.)
- \* Correspondence: Julien.marlet@univ-tours.fr



**Citation:** Marlet, J.; Gatault, P.; Maakaroun, Z.; Longuet, H.; Stefic, K.; Handala, L.; Eymieux, S.; Gyan, E.; Dartigeas, C.; Gaudy-Graffin, C. Antibody Responses after a Third Dose of COVID-19 Vaccine in Kidney Transplant Recipients and Patients Treated for Chronic Lymphocytic Leukemia. *Vaccines* **2021**, *9*, 1055. <https://doi.org/10.3390/vaccines9101055>

Academic Editors: Ger Rijkers and Jean-Luc Murk

Received: 31 August 2021  
Accepted: 16 September 2021  
Published: 23 September 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:**   2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

**Abstract:** The impact of a third dose of COVID-19 vaccine on antibody responses is unclear in immunocompromised patients. The objective of this retrospective study was to characterize antibody responses induced by a third dose of mRNA COVID-19 vaccine in 160 kidney transplant recipients and 20 patients treated for chronic lymphocytic leukemia (CLL). Prevalence of anti-spike IgG  $\geq 7.1$  and  $\geq 30$  BAU/mL after the third dose were 47% (75/160) and 39% (63/160) in kidney transplant recipients, and 57% (29/51) and 50% (10/20) in patients treated for CLL. Longitudinal follow-up identified a moderate increase in SARS-CoV-2 anti-spike IgG levels after a third dose of vaccine in kidney transplant recipients (0.19 vs. 5.28 BAU/mL,  $p = 0.03$ ) and in patients treated for CLL (0.63 vs. 10.7 BAU/mL,  $p = 0.0002$ ). This increase in IgG levels had a limited impact on prevalence of anti-spike IgG  $\geq 30$  BAU/mL in kidney transplant recipients (17%, 2/12 vs. 33%, 4/12,  $p = 0.64$ ) and in patients treated for CLL (5%, 1/20 vs. 45%, 9/20,  $p = 0.008$ ). These results highlight the need for vaccination of the general population and the importance of non-medical preventive measures to protect immunocompromised patients.

**Keywords:** COVID-19; vaccine; kidney transplant; chronic lymphocytic leukemia; antibody

## 1. Introduction

Vaccine effectiveness against symptomatic COVID-19 was estimated at between 70 and 97% after two doses of COVID-19 vaccine in immunocompetent patients [1–3]. In contrast, few immunocompromised patients have been enrolled in COVID-19 phase 3 clinical trials and vaccine effectiveness in this population remains unclear. In a recent observational study, solid organ transplant recipients had an 82-fold higher risk of breakthrough infection and a 485-fold higher risk of breakthrough infection with associated hospitalization and death [4]. Antibody responses after two doses of COVID-19 vaccine, defined as anti-spike IgG seroconversion, are impaired in these patients [5]. They were estimated between 38 to 42% for kidney transplant patients [5,6] and between 23 to 52% for patients with chronic lymphocytic leukemia, despite the fact that most of these CLL patients were not

undergoing cancer therapy [7–9]. The impact of a third dose of COVID-19 vaccine on antibody responses is unclear in these patients. Response rates to a third dose ranged between 47 and 68% in kidney transplant recipients [10–12] and limited data are available in patients treated for chronic lymphocytic leukemia (CLL). The objective of this retrospective study was to characterize antibody responses induced by a third dose of mRNA COVID-19 vaccine in 160 kidney transplant recipients and 20 patients treated for CLL.

## 2. Materials and Methods

SARS-CoV-2 anti-spike IgG was tested for all patients at least 21 days after the second and/or third dose of mRNA COVID-19 vaccine, using a SARS-CoV-2 IgG II Quant assay on an Alinity i system (Abbott). Assay results in AU/mL were converted into BAU/mL (international standard units [13]) using a conversion factor of 0.142, according to the manufacturer's recommendations. We defined responders as patients with positive SARS-CoV-2 IgG, corresponding to levels  $\geq 7.1$  binding antibody units (BAU)/mL, according to the manufacturer's recommendations and previous studies [14–18]. This interpretation is limited by its lack of association with vaccine effectiveness and its lack of comparability with other assays, using different cut-offs. As such, results were also compared with a cut-off of 30 BAU/mL, recently associated with 50% vaccine effectiveness against symptomatic COVID-19 in immunocompetent patients [19]. All calculations were performed with GraphPad Prism 9.0 using Wilcoxon test (paired values), Mann-Whitney (unpaired values) and Fisher's exact test (binomial variables).  $p$ -values  $< 0.05$  (2-sided) were considered significant.

## 3. Results

### 3.1. Kidney Transplant Recipients

Prevalence of positive SARS-CoV-2 anti-spike IgG ( $\geq 7.1$  BAU/mL) in kidney transplant recipients tested either after the second ( $n = 97$ ) or third dose ( $n = 160$ ) of COVID-19 vaccine was not significantly different (43%, 42/97 vs. 47%, 75/160,  $p = 0.61$ , Table 1). Prevalence of SARS-CoV-2 anti-spike IgG  $\geq 30$  BAU/mL was also not significantly different between these two groups (30%, 29/97 vs. 39%, 63/160,  $p = 0.14$ ). SARS-CoV-2 anti-spike median IgG levels were not significantly different between these two groups (2.7 vs. 5.5 BAU/mL,  $p = 0.42$ , Figure 1).

In contrast, when considering only the 12 patients with longitudinal follow-up after both the second and third dose, an increase in anti-spike IgG levels was observed (0.19 vs. 5.28 BAU/mL,  $p = 0.03$ , Figure 1). In this subgroup, prevalence of positive SARS-CoV-2 anti-spike IgG and anti-spike IgG  $\geq 30$  BAU/mL did not increase after the third dose (17%, 2/12 vs. 42%, 5/12,  $p = 0.37$  and 17%, 2/12 vs. 33%, 4/12,  $p = 0.64$ ).

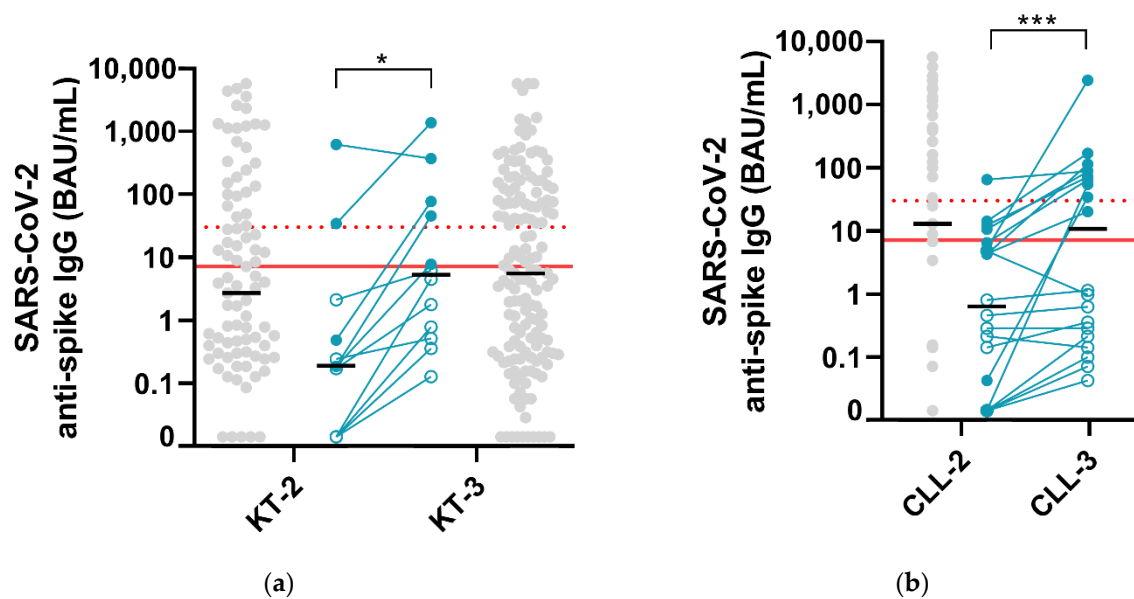
Age was associated with poor response to COVID-19 vaccines ( $>65$  year,  $p = 0.02$ ) but a history of COVID-19 RT-PCR was associated with better responses ( $p = 0.04$ , Table 1). Nine patients had a history of COVID-19, which occurred at a median of 206 days (IQR: 168–248) before the measurement of SARS-CoV-2 anti-spike IgG.

The third dose of vaccine was injected at a median of 43 days (IQR: 33–63) after the second dose. SARS-CoV-2 anti-spike IgG was measured at a median of 95 days (IQR: 48–124) and 52 days (IQR: 34–76) after the second and third dose of vaccine, respectively.

**Table 1.** Factors associated with response to COVID-19 vaccines in 245 kidney transplant recipients and 51 patients treated for chronic lymphocytic leukemia.

Response to COVID vaccine	Kidney Transplant						Chronic Lymphocytic Leukemia					
	After 2nd Dose (n = 97)			After 3rd Dose (n = 160)			After 2nd Dose (n = 51)			After 3rd Dose (n = 20)		
	No (n = 55)	Yes (n = 42)	<i>p</i>	No (n = 85)	Yes (n = 75)	<i>p</i>	No (n = 22)	Yes (n = 29)	<i>p</i>	No (n = 10)	Yes (n = 10)	<i>p</i>
Age > 65 year	31 (56)	13 (31)	<b>0.02</b>	41 (48)	26 (35)	0.08	17 (77)	20 (69)	0.55	8 (80)	6 (60)	0.63
Age < 50 year	5 (9)	8 (19)	0.23	15 (18)	24 (32)	0.06	0	1 (4)	1.0	0	0	-
Sex (female)	18 (33)	21 (5)	0.10	36 (42)	21 (28)	0.07	7 (32)	11 (38)	0.77	2 (20)	4 (40)	0.64
History of positive SARS-CoV-2 RT-PCR	1 (1.8)	6 (14)	<b>0.04</b>	0	2 (2.7)	0.22	-	-	-	-	-	-
COVID-19 vaccine												
Pfizer (BNT162b2)	43 (78)	32 (76)	0.81	53 (62)	45 (60)	0.88	-	-	-	-	-	-
Moderna (mRNA-1273)	3 (5.5)	4 (9.5)	0.46	11 (13)	16 (21)	0.20	-	-	-	-	-	-
Tyrosine kinase inhibitors or venetoclax	-	-	-	-	-	-	19 (86)	14 (48)	<b>0.007</b>	9 (90)	9 (90)	1.0
Prior-CLL directed therapy	-	-	-	-	-	-	15 (68)	12 (41)	0.54	9 (90)	7 (70)	0.58
Anti-CD20 monoclonal antibody within 2 years	-	-	-	-	-	-	2 (9)	0	0.18	1 (10)	0	1.0

Results are represented as median (range) or absolute values (percentages); *p* < 0.05 are in bold.



**Figure 1.** (a) SARS-CoV-2 anti-spike IgG levels measured in 245 kidney transplant (KT) recipients after the second (KT-2,  $n = 97$ ) and third dose of COVID-19 vaccine (KT-3,  $n = 160$ ). (b) SARS-CoV-2 anti-spike IgG levels measured in patients treated for chronic lymphocytic leukemia (CLL) after the second (CLL-2,  $n = 51$ ) and the third dose of vaccine (CLL-3,  $n = 20$ ). Longitudinal follow-up was performed for 12 KT patients and 20 CLL patients (responders and non-responders to the 3rd dose of vaccine in plain and empty blue dots, respectively). Light grey dots, patients without longitudinal follow-up; plain red bar, SARS-CoV-2 IgG II Quant assay cut-off ( $\geq 7.1$  BAU/mL); dashed red bar, anti-spike IgG levels previously associated with 50 % vaccine effectiveness against symptomatic COVID-19 ( $\geq 30$  BAU/mL) [19]; black bars, medians. \*  $p < 0.05$ ; \*\*\*  $p < 0.001$ .

### 3.2. Patients Treated for Chronic Lymphocytic Leukemia

Prevalence of positive SARS-CoV-2 anti-spike IgG and prevalence of SARS-CoV-2 anti-spike IgG  $\geq 30$  BAU/mL in 51 patients treated for CLL did not increase between the second and third dose of vaccine (57%, 29/51 vs. 50%, 10/20,  $p = 0.61$  and 45%, 23/51 vs. 45%, 9/20,  $p = 1.0$ , Table 1). SARS-CoV-2 anti-spike median IgG levels were comparable between these two groups (12.9 vs. 10.7 BAU/mL,  $p = 0.32$ , Figure 1).

In contrast, when considering only the 20 patients with longitudinal follow-up after both the second and third dose, we observed an increase in anti-spike IgG levels (0.63 vs. 10.7 BAU/mL,  $p = 0.0002$ , Figure 1) and an increase in the prevalence of anti-spike IgG  $\geq 30$  BAU/mL (5%, 1/20 vs. 45%, 9/20,  $p = 0.008$ ). In this subgroup, prevalence of positive SARS-CoV-2 anti-spike IgG did not increase after the third dose (20%, 4/20 vs. 50%, 10/20,  $p = 0.10$ ).

Treatment with tyrosine kinase inhibitors or venetoclax was associated with poor response to the second dose of COVID-19 vaccine ( $p = 0.007$ , Table 1). The third dose of vaccine was injected at a median of 63 days (IQR: 48–81) after the second dose. SARS-CoV-2 anti-spike IgG was measured at a median of 43 days (IQR: 36–57) and 42 days (IQR: 31–45) after the second and third dose of vaccine, respectively.

## 4. Discussion

In 12 kidney transplant recipients with longitudinal follow-up, a moderate increase in SARS-CoV-2 anti-spike IgG levels was observed after the third dose of COVID-19 vaccine (0.19 vs. 5.28 BAU/mL,  $p = 0.03$ ). This increase in IgG levels had no significant impact on the prevalence of anti-spike IgG  $\geq 30$  BAU/mL (17%, 2/12 vs. 33%, 4/12,  $p = 0.64$ ). In 20 patients treated for CLL with longitudinal follow-up, a moderate increase in SARS-CoV-2 anti-spike IgG levels was observed after the third dose of COVID-19 vaccine (0.63

vs. 10.7 BAU/mL,  $p = 0.0002$ ). This resulted in an increase in the prevalence of anti-spike IgG  $\geq 30$  BAU/mL (5%, 1/20 vs. 45%, 9/20,  $p = 0.008$ ). When considering all patients, with or without longitudinal follow-up, prevalence of anti-spike IgG  $\geq 30$  BAU/mL after the third dose was 39% (63/160) in kidney transplant recipients and 50% (10/20) in patients treated for CLL. Overall, this suggests that a third dose of COVID-19 vaccine in immunocompromised patients can increase antibody levels. Still, this increase might not be clinically relevant because few patients reached anti-spike IgG levels  $\geq 30$  BAU/mL, previously associated with 50% vaccine effectiveness [19]. These results are in line with other studies in kidney transplant recipients [10–12] and provide new data regarding patients with CLL.

In kidney transplant recipients, our results confirm the association between young age (<65 years) and response to COVID-19 vaccines ( $p = 0.02$ ) [6,20] and suggest a better response in patients with a history of COVID-19 ( $p = 0.04$ ). In patients treated for CLL, our results confirm the association between treatment with tyrosine kinase inhibitors or venetoclax and non-response to COVID-19 vaccines ( $p = 0.007$ ) [8].

One limitation of our study is the small number of patients with longitudinal follow-up. Indeed, antibody responses in immunocompromised patients are probably too heterogeneous to compare groups of unpaired patients. Other limitations of our study are the lack of antibody measurements prior to vaccination, the lack of in vitro investigation of neutralizing Ab titers, especially against circulating variants, and cellular responses. Other parameters such as disease stage and other treatments, not included in this study, could also have an impact on response to COVID-19 vaccines.

Overall, we believe these results highlight the need for vaccination of the general population and the importance of upholding non-medical preventive measures to protect immunocompromised patients, especially those at higher risk of non-response to COVID-19 vaccines.

**Author Contributions:** Conceptualization, J.M., P.G., Z.M., H.L., K.S., L.H., S.E., C.D., E.G. and C.G.-G.; methodology, J.M., P.G., Z.M., H.L. and C.D.; software, J.M.; validation, J.M., P.G., Z.M., C.D. and C.G.-G.; formal analysis, J.M., P.G., Z.M., H.L. and C.D.; investigation J.M., P.G., Z.M., H.L., E.G. and C.D.; resources, J.M., K.S., L.H., S.E. and C.G.-G.; data curation, J.M., P.G., Z.M., H.L. and C.D.; writing—original draft preparation, J.M.; writing—review and editing, J.M., P.G., Z.M., C.D. and C.G.-G.; visualization, J.M.; supervision, J.M., P.G., Z.M., C.D., E.G. and C.G.-G.; project administration, J.M., P.G., Z.M., C.D. and C.G.-G.; All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** This study was performed after patient information and anonymization of data according to French Reference Methodology MR-004 and to the guidelines of the Declaration of Helsinki. Data collection was approved by the French Commission Nationale de l'Informatique et des Libertés (n° 2020\_097) and sample collection was approved by the Ministère de l'Enseignement Supérieur et de la Recherche (authorization n° DC-2020-3961).

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** Data supporting reported results can be provided by contacting the corresponding author.

**Acknowledgments:** The authors thank Vincent Roux for its participation to data collection and all the medical and paramedical staff, especially Léa Bouijoux, involved in data collection regarding this study.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Haas, E.J.; Angulo, F.J.; McLaughlin, J.M.; Anis, E.; Singer, S.R.; Khan, F.; Brooks, N.; Smaja, M.; Mircus, G.; Pan, K.; et al. Impact and Effectiveness of mRNA BNT162b2 Vaccine against SARS-CoV-2 Infections and COVID-19 Cases, Hospitalisations, and Deaths Following a Nationwide Vaccination Campaign in Israel: An Observational Study Using National Surveillance Data. *Lancet* **2021**, *397*, 1819–1829. [[CrossRef](#)]



2. Tenforde, M.W.; Olson, S.M.; Self, W.H.; Talbot, H.K.; Lindsell, C.J.; Steingrub, J.S.; Shapiro, N.I.; Ginde, A.A.; Douin, D.J.; Prekker, M.E.; et al. Effectiveness of Pfizer-BioNTech and Moderna Vaccines Against COVID-19 Among Hospitalized Adults Aged  $\geq 65$  Years—United States, January–March 2021. *MMWR Morb. Mortal Wkly. Rep.* **2021**, *70*, 674–679. [[CrossRef](#)] [[PubMed](#)]
3. Lopez Bernal, J.; Andrews, N.; Gower, C.; Robertson, C.; Stowe, J.; Tessier, E.; Simmons, R.; Cottrell, S.; Roberts, R.; O’Doherty, M.; et al. Effectiveness of the Pfizer-BioNTech and Oxford-AstraZeneca Vaccines on Covid-19 Related Symptoms, Hospital Admissions, and Mortality in Older Adults in England: Test Negative Case-Control Study. *BMJ* **2021**, *373*, n1088. [[CrossRef](#)] [[PubMed](#)]
4. Qin, C.X.; Moore, L.W.; Anjan, S.; Rahamimov, R.; Sifri, C.D.; Ali, N.M.; Morales, M.K.; Tsapepas, D.S.; Basic-Jukic, N.; Miller, R.A.; et al. Risk of Breakthrough SARS-CoV-2 Infections in Adult Transplant Recipients. *Transplantation* **2021**. [[CrossRef](#)]
5. Stumpf, J.; Siepmann, T.; Lindner, T.; Karger, C.; Schwöbel, J.; Anders, L.; Faulhaber-Walter, R.; Schewe, J.; Martin, H.; Schirutschke, H.; et al. Humoral and Cellular Immunity to SARS-CoV-2 Vaccination in Renal Transplant versus Dialysis Patients: A Prospective, Multicenter Observational Study Using mRNA-1273 or BNT162b2 mRNA Vaccine. *Lancet Reg. Health Eur.* **2021**, 100178. [[CrossRef](#)] [[PubMed](#)]
6. Grupper, A.; Rabinowich, L.; Schwartz, D.; Schwartz, I.F.; Ben-Yehoyada, M.; Shashar, M.; Katchman, E.; Halperin, T.; Turner, D.; Goykhman, Y.; et al. Reduced Humoral Response to mRNA SARS-Cov-2 BNT162b2 Vaccine in Kidney Transplant Recipients without Prior Exposure to the Virus. *Am. J. Transplant.* **2021**. [[CrossRef](#)]
7. Roeker, L.E.; Knorr, D.A.; Thompson, M.C.; Nivar, M.; Lebowitz, S.; Peters, N.; Deonaraine, I.; Momotaj, S.; Sharan, S.; Chanlatte, V.; et al. COVID-19 Vaccine Efficacy in Patients with Chronic Lymphocytic Leukemia. *Leukemia* **2021**. [[CrossRef](#)] [[PubMed](#)]
8. Herishanu, Y.; Avivi, I.; Aharon, A.; Shefer, G.; Levi, S.; Bronstein, Y.; Morales, M.; Ziv, T.; Shorer Arbel, Y.; Scarfò, L.; et al. Efficacy of the BNT162b2 mRNA COVID-19 Vaccine in Patients with Chronic Lymphocytic Leukemia. *Blood* **2021**, *137*, 3165–3173. [[CrossRef](#)] [[PubMed](#)]
9. Agha, M.; Blake, M.; Chilleo, C.; Wells, A.; Haidar, G. Suboptimal Response to COVID-19 mRNA Vaccines in Hematologic Malignancies Patients. *medRxiv* **2021**. [[CrossRef](#)]
10. Benotmane, I.; Gautier, G.; Perrin, P.; Olagne, J.; Cognard, N.; Fafi-Kremer, S.; Caillard, S. Antibody Response After a Third Dose of the mRNA-1273 SARS-CoV-2 Vaccine in Kidney Transplant Recipients With Minimal Serologic Response to 2 Doses. *JAMA* **2021**. [[CrossRef](#)] [[PubMed](#)]
11. Werbel, W.A.; Boyarsky, B.J.; Ou, M.T.; Massie, A.B.; Tobian, A.A.R.; Garonzik-Wang, J.M.; Segev, D.L. Safety and Immunogenicity of a Third Dose of SARS-CoV-2 Vaccine in Solid Organ Transplant Recipients: A Case Series. *Ann. Intern. Med.* **2021**. [[CrossRef](#)] [[PubMed](#)]
12. Kamar, N.; Abravanel, F.; Marion, O.; Couat, C.; Izopet, J.; Del Bello, A. Three Doses of an mRNA Covid-19 Vaccine in Solid-Organ Transplant Recipients. *N. Engl. J. Med.* **2021**, *385*, 661–662. [[CrossRef](#)] [[PubMed](#)]
13. Kristiansen, P.A.; Page, M.; Bernasconi, V.; Mattiuzzo, G.; Dull, P.; Makar, K.; Plotkin, S.; Knezevic, I. WHO International Standard for Anti-SARS-CoV-2 Immunoglobulin. *Lancet* **2021**, *397*, 1347–1348. [[CrossRef](#)]
14. Ebinger, J.E.; Fert-Bober, J.; Printsev, I.; Wu, M.; Sun, N.; Figueiredo, J.C.; Eyk, J.E.V.; Braun, J.G.; Cheng, S.; Sobhani, K. Prior COVID-19 Infection and Antibody Response to Single Versus Double Dose mRNA SARS-CoV-2 Vaccination. *medRxiv* **2021**. [[CrossRef](#)]
15. Narasimhan, M.; Mahimainathan, L.; Araj, E.; Clark, A.E.; Markantonis, J.; Green, A.; Xu, J.; SoRelle, J.A.; Alexis, C.; Fankhauser, K.; et al. Clinical Evaluation of the Abbott Alinity SARS-CoV-2 Spike-Specific Quantitative IgG and IgM Assays in Infected, Recovered, and Vaccinated Groups. *medRxiv* **2021**. [[CrossRef](#)]
16. Prendecki, M.; Clarke, C.; Brown, J.; Cox, A.; Gleeson, S.; Guckian, M.; Randell, P.; Pria, A.D.; Lightstone, L.; Xu, X.-N.; et al. Effect of Previous SARS-CoV-2 Infection on Humoral and T-Cell Responses to Single-Dose BNT162b2 Vaccine. *Lancet* **2021**, *397*, 1178–1181. [[CrossRef](#)]
17. Gobbi, F.; Buonfrate, D.; Moro, L.; Rodari, P.; Piubelli, C.; Caldrea, S.; Riccetti, S.; Sinigaglia, A.; Barzon, L. Antibody Response to the BNT162b2 mRNA COVID-19 Vaccine in Subjects with Prior SARS-CoV-2 Infection. *Viruses* **2021**, *13*, 422. [[CrossRef](#)] [[PubMed](#)]
18. Perkmann, T.; Perkmann-Nagele, N.; Koller, T.; Mucher, P.; Radakovics, A.; Marculescu, R.; Wolzt, M.; Wagner, O.F.; Binder, C.J.; Haslacher, H. Anti-Spike Protein Assays to Determine SARS-CoV-2 Antibody Levels: A Head-to-Head Comparison of Five Quantitative Assays. *Microbiol. Spectr.* **2021**, *9*. [[CrossRef](#)] [[PubMed](#)]
19. Feng, S.; Phillips, D.J.; White, T.; Sayal, H.; Aley, P.K.; Bibi, S.; Dold, C.; Fuskova, M.; Gilbert, S.C.; Hirsch, I.; et al. Correlates of Protection against Symptomatic and Asymptomatic SARS-CoV-2 Infection. *medRxiv* **2021**. [[CrossRef](#)]
20. Danthu, C.; Hantz, S.; Dahlem, A.; Duval, M.; Ba, B.; Guibbert, M.; El Ouafi, Z.; Ponsard, S.; Berrahal, I.; Achard, J.M.; et al. Humoral Response after SARS-Cov-2 mRNA Vaccine in a Cohort of Hemodialysis Patients and Kidney Transplant Recipients. *JASN* **2021**. [[CrossRef](#)] [[PubMed](#)]