

AMERICAN SOCIETY FOR MICROBIOLOGY

Draft Genome Sequence of the Crude Oil-Degrading and Biosurfactant-Producing Strain *Cobetia* sp. QF-1

Ping Guo,^a Binxia Cao,^a Xue Qiu,^a Jianguo Lin^a

^aCollege of Environmental Science and Engineering, Dalian Maritime University, Dalian, China

ABSTRACT We report here the draft genome of *Cobetia* sp. QF-1, a cold-adapted bacterium isolated from crude oil-contaminated seawater of the Yellow Sea, China. This genome is approximately 4.1 Mb (G+C content, 57.44%) with 3,513 protein-coding sequences. *Cobetia* sp. QF-1 shows crude oil degradation and biosurfactant production activity at low temperature.

The *Cobetia* genus is 1 of 11 genera of the family *Halomonadaceae* and is most closely related to members of the genus *Halomonas* (1). The type strain has been described as aerobic, Gram-negative, motile, rod-shaped, and slightly halophilic (2). *Cobetia* sp. QF-1 can degrade crude oil and produce biosurfactants at 0°C. Although some isolated marine bacteria can degrade oil efficiently around 0°C, most of them have been isolated from polar environments (3, 4); a few have been isolated from China. Several genomes of *Cobetia* strains have been sequenced before (5–7), but a marine *Cobetia* strain from China with the capacity to degrade hydrocarbons and produce biosurfactants at 0°C is lacking. Here, we report the draft genome sequence of *Cobetia* sp. QF-1, isolated from the seawater of Yellow Sea, China.

The genome sequencing of *Cobetia* sp. QF-1 was performed using the Illumina HiSeq platform at NovogenBio, Beijing, China. Reads were assembled using SOAP-denovo software version 2.21 (8). Protein-coding sequences were functionally annotated with the Clusters of Orthologous Groups of Proteins (COG) (9), Gene Ontology (GO) (10), and KEGG databases (11). tRNAs and rRNAs were detected using tRNAscan-SE (12) and rRNAmmer software (13), respectively.

The genome of *Cobetia* sp. QF-1 was 4,084,184 bp with a 57.44% GC content. The genome contained 3,513 predicted protein-coding sequences, and the total length of the genes was 3,470,121 bp, which accounted for 84.96% of the genome. The genome also encoded 71 tRNAs and 5 rRNAs.

According to the annotation results, many genes involved in petroleum hydrocarbon degradation, biosurfactant production, and cold adaption were found in the genome of *Cobetia* sp. QF-1. This draft genome sequence of *Cobetia* sp. QF-1 will help in understanding the genetic basis of crude oil degradation and biosurfactant production by *Cobetia* sp.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number NBVS00000000. The version described in this paper is the first version, NBVS01000000.

ACKNOWLEDGMENT

This work was supported by Fundamental Research Funds for Central Universities, China (grant number 3132015079).

Received 21 November 2017 Accepted 28 November 2017 Published 18 January 2018 Citation Guo P, Cao B, Qiu X, Lin J. 2018. Draft genome sequence of the crude oil-degrading and biosurfactant-producing strain *Cobetia* sp. QF-1. Genome Announc 6:e01456-17. https:// doi.org/10.1128/genomeA.01456-17.

Copyright © 2018 Guo et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Jianguo Lin, lin7017@126.com.

REFERENCES

- Arahal DR, Castillo AM, Ludwig W, Schleifer KH, Ventosa A. 2002. Proposal of *Cobetia marina* gen. nov., comb. nov., within the family *Halomonadaceae*, to include the species *Halomonas marina*. Syst Appl Microbiol 25:207–211. https://doi.org/10.1078/0723-2020-00113.
- Ugbenyen AM, Vine N, Simonis JJ, Basson AK, Okoh AI. 2015. Characterization of a bioflocculant produced from the consortium of three marine bacteria of the genera *Cobetia* and *Bacillus* and its application for wastewater treatment. J Water Sanit Hyg Dev 5:81–88. https://doi.org/ 10.2166/washdev.2014.181.
- Whyte LG, Hawari J, Zhou E, Bourbonnière L, Inniss WE, Greer CW. 1998. Biodegradation of variable-chain-length alkanes at low temperatures by a psychrotrophic *Rhodococcus* sp. Appl Environ Microbiol 64:2578–2584.
- Yakimov MM, Giuliano L, Gentile G, Crisafi E, Chernikova TN, Abraham WR, Lünsdorf H, Timmis KN, Golyshin PN. 2003. *Oleispira antarctica* gen. nov., sp. Number v., a novel hydrocarbonoclastic marine bacterium isolated from Antarctic coastal sea water. Int J Syst Evol Microbiol 53:779–785. https://doi .org/10.1099/ijs.0.02366-0.
- Balabanova LA, Golotin VA, Kovalchuk SN, Babii AV, Shevchenko LS, Son OM, Kosovsky GY, Rasskazov VA. 2016. The genome of the marine bacterium *Cobetia marina* KMM 296 isolated from the mussel *Crenomytilus grayanus* (Dunker, 1853). Russ J Mar Biol 42:106–109. https://doi.org/ 10.1134/S106307401601003X.
- Ibacache-Quiroga C, Canales C, Charifeh M, Dinamarca MA. 2017. Genome sequence of *Cobetia* sp. strain MM1IDA2H-1, a hydrocarbondegrading and biosurfactant-producing marine bacterium. Genome Announc 5(15):e00132-17. https://doi.org/10.1128/genomeA.00132-17.
- 7. Alexiev A, Krusor ML, Jospin G, Lang JM, Eisen JA, Coil DA. 2016. Draft

genome sequence of *Cobetia* sp. UCD-24C, isolated from roots and leaves of the seagrass *Zostera marina*. Genome Announc 4(2):e00116-16. https://doi.org/10.1128/genomeA.00116-16.

- Li RQ, Yu C, Li Y, Lam TW, Yiu SM, Kristiansen K, Wang J. 2009. SOAP2: an improved ultrafast tool for short read alignment. Bioinformatics 25:1966–1967. https://doi.org/10.1093/bioinformatics/btp336.
- Tatusov RL, Fedorova ND, Jackson JD, Jacobs AR, Kiryutin B, Koonin EV, Krylov DM, Mazumder R, Mekhedov SL, Nikolskaya AN, Rao BS, Smirnov S, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA. 2003. The COG database: an updated version includes eukaryotes. BMC Bioinformatics 4:41. https://doi.org/10.1186/1471-2105-4-41.
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G. 2000. Gene Ontology: tool for the unification of biology. Nat Genet 25:25–29. https://doi.org/10.1038/75556.
- Kanehisa M, Goto S, Kawashima S, Okuno Y, Hattori M. 2004. The KEGG resource for deciphering the genome. Nucleic Acids Res 32:D277–D280. https://doi.org/10.1093/nar/gkh063.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25:955–964.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. https://doi.org/10.1093/nar/ gkm160.