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The Role of Alpha-MSH as a Modulator of Ocular Immunobiology Exemplifies Mechanistic Differences between Melanocortins and Steroids

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Abstract

Melanocortins are a highly conserved family of peptides and receptors that includes multiple proopiomelanocortin-derived peptides and five defined melanocortin receptors. The melanocortins have an important role in maintaining immune homeostasis and in suppressing inflammation. Within the healthy eye, the melanocortins have a central role in preventing inflammation and maintaining immune privilege. A central mediator of the anti-inflammatory activity is the nonsteroidogenic melanocortin peptide alpha-melanocyte stimulating hormone. In this review we summarize the major findings of melanocortin regulation of ocular immunobiology with particular interest in the ability of melanocortin to induce immune tolerance and cytoprotection. The melanocortins have therapeutic potential because their mechanisms of action in regulating immunity are distinctly different from the actions of steroids.

Keywords

alpha-MSH; eye; glucocorticoids; melanocortins

INTRODUCTION

The melanocortin (MC) system encompasses multiple peptides including α -, β -, γ melanocyte stimulating hormone (MSH), adrenocorticotropic hormone (ACTH), and five MC receptors (MCR1-5) that are expressed in a multitude of cells and tissues.^{1,2} The MC peptides and receptors are highly conserved across multiple species³ from lower vertebrates to vertebrates and appeared early in evolution² but remain largely unchanged,⁴ suggesting a critical and ubiquitous role. The role of the MC system in the neuroendocrine control of inflammation is understood and known to be affected by the release of corticotropin-

DECLARATION OF INTEREST

CMC and JY are employees of Mallinckrodt Pharmaceuticals, and AWT has been a consultant for Mallinckrodt Pharmaceuticals.

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releasing hormone in response to infection or stress; this stimulates the production of proopiomelanocortin (POMC), which is processed to generate the MC peptides. The myriad functions of the MCs are carried out through their binding to the five known MC receptors, each of which is expressed in specific cell types.^{5,6} ACTH stimulates melanocortin receptor 2 (MC2R) on the adrenal cortex to produce glucocorticoids (GCs). Although the antiinflammatory effects of GCs are well known and are one way in which MCs control inflammation indirectly, there is a growing understanding of the distinct and direct effects of GCs and MCs as immune modulators (Table 1).

It has long been established that GC production is controlled selectively by ACTH, and that ACTH induces steroidogenesis via binding to MC2R. MC2R is abundantly expressed from the adrenal cortex, specifically the zona fasciculata, where the GCs are produced, and the zona glomerulosa, where the mineralocorticoid aldosterone is produced.⁷ The essential role of MC2R in steroidogenesis is clear as mutations in MC2R result in familial GC deficiencies⁸ and *Mc2r* null mice lack adrenal gland development and steroid production.⁹ The MC receptors have varying affinities for the different MCs, but all recognize a highly conserved sequence (HFRW) that is common to all MC peptides.¹⁰ Unlike the other MC receptors, MC2R exclusively binds ACTH^{7,11} and this specificity is conferred by peptide sequences that are unique to ACTH.¹² Animal studies confirm that α -MSH is not capable of stimulating GC secretion^{13,14} and only ACTH was able to restore wild-type GC production in a *Pomc* null model.¹⁵ Thus, the anti-inflammatory effects of α -MSH are not due to the induction of GC production.

To dissect the direct, non-steroid-mediated effects of MCs in whole organisms multiple approaches have been used. Adrenalectomization,^{16–18} MCR knockouts,^{19–23} and administration of specific MCs such as α -MSH^{24–27} have all been applied to demonstrate the direct pleiotropic effects of the MC system in controlling inflammation. MCs control inflammation through specific downstream modulation of immune cells, processes, cytokines, and chemokines.^{2,4,28} These studies have helped to uncover how MCs utilize different MCRs in specific cells and tissues to regulate acute, systemic, and chronic inflammation and provide a strong foundation for their use to treat a variety of inflammatory conditions.

ADVERSE EFFECTS OF GLUCOCORTICOIDS ON THE EYE

Although GC therapy has been a mainstay in the treatment of inflammation for decades, it affects many different cells beyond the immune system. Acting via the GC receptor, GCs utilize both transactivation of target genes and transrepression mechanisms to regulate a number of transcription factors including NFkB, AP-1, and p38 MAP kinase^{29,30} and influence a number of different pathways including stress responses, metabolism, and development. As such, the profile of undesirable side effects from prolonged systemic GC therapy is significant and includes bone loss, diabetes, muscle wasting, impaired wound healing, gastric ulcers, weight gain, cardiovascular disease, and neuropsychiatric issues.^{31,32}

In the eye, the adverse effects of GCs include increased intraocular pressure (IOP), which is directly associated with the development of glaucoma.³³⁻³⁵ IOP is regulated in part by

outflow of the aqueous humor via the trabecular meshwork (TM), and increased resistance in this outflow pathway is one of the primary causes of elevated intraocular pressure that can lead to glaucoma.-³⁶ GCs are known to increase outflow resistance of the TM in some individuals.^{37,38} This GC sensitivity could be due to variation in the relative levels of alternatively spliced GC receptor isoforms in the TM.^{39,40} GCs have numerous effects on the TM, including changes in TM morphology, gene expression, extracellular matrix, and cvtoskeleton as well as inhibition of phagocytosis, proliferation, and migration of TM cells.⁴¹ Also, GC treatment causes apoptosis in bovine TM cells,³⁸ and in human TM cells at high concentrations.⁴² While the exact mechanism by which GCs increase IOP is unclear, it has been reported that 40-60% of patients undergoing chronic GC therapy develop ocular hypertension.⁴³ These adverse effects are also dependent on the type of GC therapy. A recent study on the treatment of uveitis with steroid implants^{44,45} found that 65% of patients treated with these implants developed increased IOP and 23% developed glaucoma. In comparison, increased IOP was noted in 49% of patients undergoing periocular GC injections,⁴⁶ and 24% of those receiving systemic therapy, composed of GCs and immunosuppressive medications, developed increased IOP.45

Beyond influencing intraocular pressure, GC therapy has been linked to cataract formation, primarily posterior subcapsular cataracts.^{47–51} Lens epithelial cells are known to express GC receptors⁵² and GC treatment induces significant gene expression changes in these cells.^{53,54} The exact mechanism by which GC treatment induces cataract formation is probably complex. GCs have been shown to induce apoptosis in lens epithelial cells,^{55,56} but GC treatment also disrupts the differentiation and proliferation of lens cells and causes their aberrant migration to the posterior pole of the lens.⁵⁷ These factors, as well as changes to the lens environment, via GC effects on other ocular tissues, may all contribute to cataract formation. One meta-analysis of several studies reported that approximately 22% of patients undergoing systemic treatment with GCs developed cataracts, which was correlated with the dosage and duration of treatment.⁴⁹ Similarly, 20% of patients receiving periocular GC injections developed cataracts within 12 months.⁴⁶ However, a recent study found that 91% of patients receiving steroid implants for uveitis developed cataracts within 24 months.⁴⁴

α -MSH AND THE EYE

The microenvironment of the eye is immune privileged in that it has several mechanisms to suppress the action of inflammation. Over the past couple of decades it has become clear that melanocortin pathways have an important role in maintaining immune privilege.^{58–60} The melanocortin pathways are involved in suppressing proinflammatory signals and, unlike steroids, promote the immune system to regulate itself within the ocular microenvironment. Aqueous humor, the fluid filling the anterior chamber of the eye, suppresses the *in vitro* activation of immune cells that mediate hypersensitivity;⁶¹ moreover, these aqueous humor-treated immune cells are able to suppress other hypersensitivity-mediating T cells.⁶² At the molecular level this immunoregulatory activity of aqueous humor is mediated by α -MSH. Although TGF- β 2 is the most abundant anti-inflammatory factor in aqueous humor, $^{63-65}$ it exists in a latent form and must be activated before it can affect immune cells whereas the effects of α -MSH are more immediate. *In vitro* studies have demonstrated that TGF- β 2 is an enhancer of the immune regulating activity induced by α -MSH.

In the healthy eye there is constitutive expression of α -MSH. In aqueous humor the physiological concentration is around 30 pg/mL and the level of a-MSH is relatively constant among mammals. The sources of a-MSH production in the eye are not fully known; however, retinal pigment epithelial (RPE) cells have been shown to be a source of a-MSH.⁶⁰ In culture, RPE cells produce about 2 ng of a-MSH in 24 hours^{58,60} and express the necessary endopeptidases and post-translational enzymes to process a functional α -MSH peptide from POMC.⁶⁷ Other cells in the eve, such as the cells of the iris and ciliary body. also express these enzymes, suggesting that they are also potential sources of a-MSH.⁶⁸ At ocular physiological concentrations under serum-free conditions in vitro, as found within the ocular microenvironment, a-MSH is highly potent and suppresses most signals of inflammation while stimulating regulatory activity.^{26,58,60,62,66,69} Evidence suggests that there is a loss of a-MSH expression in eyes with autoimmune disease or damaged retinas.⁶⁰ Treatment of mice suffering from ocular inflammation (uveitis) with α -MSH suppresses the inflammation, and may re-establish some features of immune privilege and immune regulation.^{22,70–72} The finding of α -MSH activity within the eye has not only promoted examination of the role of neuropeptides in regulating immunity but, more importantly, has also provided molecular mechanisms for the anti-inflammatory activity seen within the immune privileged eye.

Activated effector T cells treated with aqueous humor or α -MSH produce their own TGF- β , are FoxP3 positive, and function as CD4⁺CD25⁺ inducible Treg (iTreg) cells.^{69,73} In addition, these α -MSH-induced iTreg cells require reactivation to their cognate antigen to mediate suppression, although the mechanism of suppression mediated by the α -MSH-induced iTreg cells is non-specific and will suppress all inflammatory activity in their immediate microenvironment. The induction of regulatory immunity instead of complete suppression of inflammation provides the eye with a directed mechanism of protection against autoimmune attack. Induction of regulatory immunity by α -MSH is mediated through its effects on both the antigen presenting cells (APC) and on effector T cells through their expression of melanocortin receptors 1, 3, and 5.^{26,72,74–77} The induction of regulatory immunity and signaling through melanocortin receptors other than MC2R provides a strong indication that α -MSH might be an alternative non-steroidogenic therapeutic approach to regulate immunity.

The RPE, through α -MSH together with another neuropeptide NPY, promotes the differentiation of macrophages into suppressor cells that suppress inflammation and immunity.⁶⁰ This regulation also appears to convert the retinal microglial cells into suppressor cells. The benefit of such cells is that, although they occupy a tissue microenvironment that is hypoxic and full of oxidized byproducts, they are not activated to mediate inflammation;^{60,78,79} on the contrary, they suppress the activation of inflammation and effector T cells. RPE cells from wounded retinas are diminished in α -MSH production and support the activation of macrophages that mediate inflammation.⁶⁰ Therefore, in ocular disease, changes in RPE production of α -MSH may contribute to the disease pathology. The RPE-induced suppressor cells are not themselves suppressed in immune activity. While they continue to phagocytize opsonized materials, α -MSH suppresses the activation of phagolysosomes.^{80,81} The implication is that although α -MSH-affected cells can clear materials through phagocytosis, they cannot process or degrade the materials through

conventional acidic-lysosome pathways. This may be a mechanism of immune privilege by which processing of recognizable autoantigen peptides for MHC presentation is prevented. One interesting phenomenon is that, without α -MSH, healthy RPE cells induce apoptosis in macrophages.^{60,81} While the mechanism by which RPE induces apoptosis in the macrophages is unknown, this unique finding suggests that a macrophage responding to α -MSH is protected from apoptosis;⁸² however, such a rescued macrophage will now be a suppressor cell. This could be part of a mechanism of immune privilege to select for immune cells that can be manipulated to be suppressor cells within the healthy retina. This mechanism is highly dependent on the production of α -MSH by the RPE.

There is increasing evidence that specific melanocortin receptors may be associated with different actions of a-MSH (Figure 1). While the suppression of pro-inflammatory signals on innate immune cells by a-MSH is thought to occur most often through MC1R and MC3R,^{74,75} the activation of regulatory activity appears to be through MC5R.^{26,77} The role of MC5R in regulation is seen in the difference between wild-type mice and Mc5r knockout mice that have recovered from experimental autoimmune uveitis (EAU).⁸³ Mice, unlike humans, recover from EAU without medical intervention. Post-EAU spleens of wild-type mice exhibit regulatory immunity that provides resistance to reactivation of the autoimmune uveitis.^{77,84} Mice without MC5R recover from EAU on their own, but they lack this regulatory immunity in the spleen.⁸⁴ If the post-EAU Mc5r knockout mice are reimmunized to reactivate EAU, they display an immediate reactivation of uveitis that becomes extremely severe. If the spleen cells from post-EAU wild-type mice are adoptively transferred to Mc5rknockout mice before the reimmunization, the second episode is suppressed. This finding demonstrates that mice possess a melanocortin-dependent induction of regulatory immunity that prevents a memory immune response to autoantigen of the retina and a systemic resistance to retinal autoantigens.

The MC5R-dependent regulatory immunity can be induced during uveitis by injecting α -MSH peptide into the EAU mice.^{22,72,77} This has also been demonstrated in the mouse model of multiple sclerosis, experimental autoimmune encephalomyelitis.^{85,86} Both antigen presenting cells (APC) and effector T cells express MC5R, and can be the target of α -MSH induction of regulatory activity. As described above, treating activated effector T cells with α -MSH induces the expansion and activation of inducible Treg cells.^{-26,69} Blocking MC5R or using T cells from immunized MC5R knock-out mice prevents α -MSH induction of regulatory activity. α -MSH upregulates expression of FoxP3 in T cells, indicating that α -MSH promotes T cell differentiation into regulatory cells.⁶⁹ There is very little cytokine production by these α -MSH-treated T cells, other than TGF- β . The cells function as regulatory T cells, and through adoptive transfer, suppress inflammation within the tissue microenvironment where their antigen specificity is presented.^{62,77,87} Therefore, by stimulating MC5r with α -MSH peptide or a melanocortin analogue that targets only MC5r, it is possible to create a unique population of Treg cells from antigen-specific T cells that can be used to target tissues of inflammation, graft rejection, and autoimmune disease.

When the spleen cells from the post-EAU mice were examined, it was not the T cells but rather the APC that were MC5R dependent for regulatory activity.⁷⁷ These APC are unique in that they express markers of both myeloid and granulocytic cells. They are found in the

spleens of mice, but mediate regulatory immunity only after EAU and only if MC5R is expressed. Resting APC stimulated by α -MSH express the ectoenzymes that convert ATP into adenosine.⁷² There is also an increase in their expression of PD-L1, a surface adhesion protein that regulates T cell activation.⁷³ They process and present antigen and, when mixed with antigen specific effector T cells, the APC promote regulatory activity by the T cells. These Treg cells have the characteristics of inducible Treg cells, suggesting that α -MSH, through MC5R, upregulates APC generation of adenosine and expression of PD-L1 to promote the counterconversion of effector T cells into inducible Treg cells.^{72,73} Therefore, as part of the resolution of EAU, there is a melanocortin-dependent pathway that localizes in the spleen a unique population of APC that promote the activation of protective autoreactive T cells (Figure 2). This pathway can be induced by α -MSH therapy and, since expression of the melanocortins is conserved, has the potential to be a new therapeutic approach to suppression of autoimmune disease in humans. It will be of interest to see whether this is a pathway that, while spontaneously activated in mice, needs to be awakened in humans with a melanocortin-based therapy.

These studies have shown that the MCs have an extremely important role in preventing inflammation within the eye. In addition, there is strong potential for use of the MC system in therapy for uveitis. Also, therapeutic benefits of using the MCs may go beyond immune regulation within the eye. Through MC1R, α-MSH protects the RPE from oxidative stressinduced apoptosis.⁸⁸ In addition, a-MSH may have neurotrophic activity that promotes photoreceptor survival.⁸⁹ This suggests that, besides the anti-inflammatory and immune regulating activity of a-MSH therapy, there will be an additional benefit of maintaining and promoting the health of other cells of the retina. Therefore, the MCs hold an important role in maintaining an anti-inflammatory microenvironment and in the general health of the retina. Moreover, the anti-inflammatory and immune-regulating activity of the melanocortins has the potential to provide a novel therapeutic approach to many autoimmune and inflammatory diseases. This approach contrasts with the general suppression of immunity by GCs and with the biologic therapies that target one specific proinflammatory cytokine at a time. MC therapy manipulates the immune cells to function in a manner that suppresses inflammation and promotes immune tolerance and cell survival to potentially regenerate a healthy ocular microenvironment (Figure 3).

MELANOCORTINS AS ALTERNATIVES TO GLUCOCORTICOIDS

GCs have been the primary therapeutic option for the treatment of ocular inflammatory disorders for over 60 years.⁹⁰ Despite the popularity of GCs for treatment, they have a number of side effects that cause serious complications. As mentioned above, a significant portion of the population is susceptible to increased IOP from treatment with GCs. In addition to the well-documented relationship of GCs with glaucoma and cataracts, GC therapy can also lead to a number of other adverse effects in the eye including viral retinitis-⁹¹ and central serous chorioretinopathy,^{92,93} which is characterized by serous fluid accumulation in the retina resulting in localized detachment of the neurosensory retina.⁹⁴ GC therapy is also associated with an increased risk of infection, impaired wound healing, and thinning of the cornea.^{95,96}

In contrast, a-MSH is present in the aqueous humor of the eye and acts to prevent inflammation that could be damaging to the eye.^{58,97} a-MSH has the ability to influence T cells in a positive immune modulating capacity by suppressing immune response through the release of inhibitory cytokines, metabolic inhibition, or cytolysis of effector T-cells and by suppressing the maturation of dendritic cells.^{58,59,66,69,98,99} a-MSH suppresses production of proinflammatory cytokines, including IFN- γ^{58} and TNFa,¹⁰⁰ while increasing production of anti-inflammatory cytokines such as IL-10.101 In addition to its anti-inflammatory actions, a-MSH has been shown to suppress apoptosis in a number of cell types.^{82,102–104} The immune modulation exhibited by a-MSH, coupled with its distinctive cytoprotective effect, makes α-MSH a potentially advantageous alternative treatment and has generated a number of mechanistic studies to examine a-MSH as a therapeutic tool in the treatment of ocular inflammation. Use of α -MSH and its analogs to treat uveitis in a variety of model systems has shown them to be effective in reducing the severity and promoting resolution of inflammation.^{22,70,105} In addition to the treatment of inflammatory disorders, topical a-MSH has been shown to transiently lower intraocular pressure,¹⁰⁶ which contrasts with the increase in pressure often seen with GC treatment. While the anti-inflammatory actions of GCs suppress the immune system, MCs appear to have multipathway immunomodulatory properties to handle the acute inflammation with pro-resolving properties that alter the phenotype of immune cells, allowing them to be more modulatory.²⁰

MCs have potent anti-inflammatory and cytoprotective functions in reducing inflammation and inducing resolution in other tissues and conditions. α-MSH normalizes oxidative stress, diminishes apoptosis, and reduces retinal damage in diabetic retinopathy.^{104,107} In rodent model systems, MCs promote restoration of nerve damage from spinal cord injury,^{108,109} modulate inflammation, and attenuate apoptosis and damage after traumatic brain injury.¹¹⁰ Recently, α-MSH has been demonstrated to protect against brain damage and cognitive decline in murine models of Alzheimer's disease.^{111–113}

The neuroprotective functions of α-MSH can also reduce the severity of autoimmune encephalomyelitis (EAE) in rodent models.^{85,114} This contrasts with GCs, which have been found to increase apoptosis of retinal nerve ganglion cells in EAE models.¹¹⁵ EAE is an inflammatory demyelinating disease of the central nervous system which, importantly, is a model system for multiple sclerosis (MS). ACTH has been known to be effective in the treatment of MS for many years.¹¹⁶ Historically, the effects of ACTH therapy in MS were thought to be due to the promotion of steroidogenesis through binding of MC2R. However, several studies now indicate that ACTH functions in alleviating MS exacerbations via binding to other MC receptors rather than by the induction of steroid production.¹¹⁷ This, along with results from model systems such as EAE, suggests that MS may be successfully treated with MC peptides.

MCs also exhibit cytoprotective properties that oppose the damaging effects of GCs. A significant side effect of systemic GC therapy is osteoporosis.¹¹⁸ Although ACTH induces steroidogenesis, this results in a lower plasma level of GCs than typical steroid administration^{119,120} and, moreover, has a cytoprotective effect on bone and can protect against osteonecrosis induced by GCs.¹²¹ In addition, while GC therapy can increase the

The differences noted above suggest that MCs may offer a less toxic alternative to GCs for the treatment of inflammation. Toxicity studies found that massive doses of α -MSH, 5000-fold greater than the dose necessary to affect thermoregulation in rabbits, were required before toxicity became lethal in some animals.¹²⁴ Furthermore, α -MSH and analogues have been shown to have no significant adverse effects in clinical studies, especially in comparison to other anti-inflammatory therapies.^{125–131}

CONCLUSION

Melanocortins have shown promise in the treatment of a number of autoimmune disorders. α -MSH administration reduces disease activity in model systems of rheumatoid arthritis,¹³² systemic lupus erythematosus,¹³³ and inflammatory bowel disease.¹³⁴ Recent evidence has shown that ACTH is effective in lupus,¹³⁵ dermatomyositis or polymyositis,¹³⁶ infantile spasms, MS, and nephrotic syndrome.¹³⁷ These are all indications in which steroids are generally considered the mainstay of treatment, and where their toxic side effects often require an alternative therapy. Importantly, in several of these indications, MCs have shown to be efficacious where steroids alone have been ineffective,^{137–139} further suggesting their mechanism of action is distinct from steroids (Table 1).

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References

- 1. Catania A, Gatti S, Colombo G, et al. Targeting melanocortin receptors as a novel strategy to control inflammation. Pharmacol Rev. 2004; 56(1):1–29. [PubMed: 15001661]
- Catania A, Lonati C, Sordi A, et al. The melanocortin system in control of inflammation. ScientificWorldJournal. 2010; 10:1840–1853. [PubMed: 20852827]
- Schioth HB, Haitina T, Ling MK, et al. Evolutionary conservation of the structural, pharmacological, and genomic characteristics of the melanocortin receptor subtypes. Peptides. 2005; 26(10):1886–1900. [PubMed: 15985310]
- Ottaviani E, Franchini A, Genedani S. ACTH and its role in immune-neuroendocrine functions. A comparative study. Curr Pharm Des. 1999; 5(9):673–681. [PubMed: 10495359]
- Cooray SN, Clark AJ. Melanocortin receptors and their accessory proteins. Mol Cell Endocrinol. 2011; 331(2):215–221. [PubMed: 20654690]
- Cone RD, Mountjoy KG, Robbins LS, et al. Cloning and functional characterization of a family of receptors for the melanotropic peptides. Ann N Y Acad Sci. 1993; 680:342–363. [PubMed: 8390157]
- Mountjoy KG, Robbins LS, Mortrud MT, et al. The cloning of a family of genes that encode the melanocortin receptors. Science. 1992; 257(5074):1248–1251. [PubMed: 1325670]
- Clark AJ, McLoughlin L, Grossman A. Familial glucocorticoid deficiency associated with point mutation in the adrenocorticotropin receptor. Lancet. 1993; 341(8843):461–462. [PubMed: 8094489]
- Chida D, Nakagawa S, Nagai S, et al. Melanocortin 2 receptor is required for adrenal gland development, steroidogenesis, and neonatal gluconeogenesis. Proc Natl Acad Sci U S A. 2007; 104(46):18205–18210. [PubMed: 17989225]

- Schioth HB, Muceniece R, Larsson M, et al. Binding of cyclic and linear MSH core peptides to the melanocortin receptor subtypes. Eur J Pharmacol. 1997; 319(2–3):369–373. [PubMed: 9042613]
- 11. Schioth HB, Chhajlani V, Muceniece R, et al. Major pharmacological distinction of the ACTH receptor from other melanocortin receptors. Life Sci. 1996; 59(10):797–801. [PubMed: 8761313]
- Costa JL, Bui S, Reed P, et al. Mutational analysis of evolutionarily conserved ACTH residues. Gen Comp Endocrinol. 2004; 136(1):12–16. [PubMed: 14980791]
- Nussdorfer GG, Mazzocchi G, Malendowicz LK. Acute effects of alpha-MSH on the rat zona glomerulosa in vivo. Biochem Biophys Res Commun. 1986; 141(3):1279–1284. [PubMed: 3028391]
- Shenker Y, Villareal JZ, Sider RS, et al. Alpha-melanocyte-stimulating hormone stimulation of aldosterone secretion in hypophysectomized rats. Endocrinology. 1985; 116(1):138–141. [PubMed: 2981062]
- 15. Costa JL, Forbes S, Brennan MB, et al. Genetic modifications of mouse proopiomelanocortin peptide processing. Mol Cell Endocrinol. 2011; 336(1–2):14–22. [PubMed: 21195130]
- Getting SJ, Christian HC, Flower RJ, et al. Activation of melanocortin type 3 receptor as a molecular mechanism for adrenocorticotropic hormone efficacy in gouty arthritis. Arthritis Rheum. 2002; 46(10):2765–2775. [PubMed: 12384937]
- Gubner R, Cote L, Hughes J, et al. Comparative effects of aminopterin, cortisone and ACTH in experimental formaldehyde arthritis and psoriatic arthritis. J Invest Dermatol. 1952; 19(4):297– 305. [PubMed: 12990838]
- Yeh JK, Evans JF, Niu QT, et al. A possible role for melanocortin peptides in longitudinal growth. J Endocrinol. 2006; 191(3):677–686. [PubMed: 17170224]
- Auriemma M, Brzoska T, Klenner L, et al. alpha-MSH-stimulated tolerogenic dendritic cells induce functional regulatory T cells and ameliorate ongoing skin inflammation. J Invest Dermatol. 2012; 132(7):1814–1824. [PubMed: 22418871]
- Montero-Melendez T, Patel HB, Seed M, et al. The melanocortin agonist AP214 exerts antiinflammatory and pro-resolving properties. Am J Pathol. 2011; 179(1):259–269. [PubMed: 21703408]
- Patel HB, Bombardieri M, Sampaio AL, et al. Anti-inflammatory and antiosteoclastogenesis properties of endogenous melanocortin receptor type 3 in experimental arthritis. FASEB J. 2010; 24(12):4835–4843. [PubMed: 20702773]
- Lee DJ, Biros DJ, Taylor AW. Injection of an alpha-melanocyte stimulating hormone expression plasmid is effective in suppressing experimental autoimmune uveitis. Int Immunopharmacol. 2009; 9(9):1079–1086. [PubMed: 19426838]
- Lee SN, Ryu JH, Joo JH, et al. Alpha-melanocyte-stimulating hormone inhibits tumor necrosis factor alpha-stimulated MUC5AC expression in human nasal epithelial cells. Am J Respir Cell Mol Biol. 2011; 44(5):716–724. [PubMed: 20639461]
- Cooper A, Robinson SJ, Pickard C, et al. Alpha-melanocyte-stimulating hormone suppresses antigen-induced lymphocyte proliferation in humans independently of melanocortin 1 receptor gene status. J Immunol. 2005; 175(7):4806–4813. [PubMed: 16177130]
- Buggy JJ. Binding of alpha-melanocyte-stimulating hormone to its G-protein-coupled receptor on B-lymphocytes activates the Jak/STAT pathway. Biochem J. 1998; 331(Pt 1):211–216. [PubMed: 9512481]
- 26. Taylor A, Namba K. In vitro induction of CD25+ CD4+ regulatory T cells by the neuropeptide alpha-melanocyte stimulating hormone (alpha-MSH). Immunol Cell Biol. 2001; 79(4):358–367. [PubMed: 11488983]
- Loser K, Brzoska T, Oji V, et al. The neuropeptide alpha-melanocyte-stimulating hormone is critically involved in the development of cytotoxic CD8+ T cells in mice and humans. PLoS One. 2010; 5(2):e8958. [PubMed: 20126537]
- Brzoska T, Luger TA, Maaser C, et al. Alpha-melanocyte-stimulating hormone and related tripeptides: biochemistry, antiinflammatory and protective effects in vitro and in vivo, and future perspectives for the treatment of immune-mediated inflammatory diseases. Endocr Rev. 2008; 29(5):581–602. [PubMed: 18612139]

- Ratman D, Vanden Berghe W, Dejager L, et al. How glucocorticoid receptors modulate the activity of other transcription factors: a scope beyond tethering. Mol Cell Endocrinol. 2013; 380(1–2):41– 54. [PubMed: 23267834]
- 30. Cruz-Topete D, Cidlowski JA. One hormone, two actions: anti- and pro-inflammatory effects of glucocorticoids. Neuroimmunomodulation. 2015; 22(1–2):20–32. [PubMed: 25227506]
- Schacke H, Docke WD, Asadullah K. Mechanisms involved in the side effects of glucocorticoids. Pharmacol Ther. 2002; 96(1):23–43. [PubMed: 12441176]
- Harris E, Tiganescu A, Tubeuf S, et al. The prediction and monitoring of toxicity associated with long-term systemic glucocorticoid therapy. Curr Rheumatol Rep. 2015; 17(6):513. [PubMed: 25903665]
- Sommer A, Tielsch JM, Katz J, et al. Relationship between intraocular pressure and primary open angle glaucoma among white and black Americans. The Baltimore Eye Survey. Arch Ophthalmol. 1991; 109(8):1090–1095. [PubMed: 1867550]
- AGIS-Investigators. The Advanced Glaucoma Intervention Study (AGIS): 7. The relationship between control of intraocular pressure and visual field deterioration. The AGIS Investigators. Am J Ophthalmol. 2000; 130(4):429–440. [PubMed: 11024415]
- Gordon MO, Beiser JA, Brandt JD, et al. The Ocular Hypertension Treatment Study: baseline factors that predict the onset of primary open-angle glaucoma. Arch Ophthalmol. 2002; 120(6): 714–720. discussion 829–730. [PubMed: 12049575]
- Tan JC, Peters DM, Kaufman PL. Recent developments in understanding the pathophysiology of elevated intraocular pressure. Curr Opin Ophthalmol. 2006; 17(2):168–174. [PubMed: 16552252]
- 37. Wordinger RJ, Clark AF. Effects of glucocorticoids on the trabecular meshwork: towards a better understanding of glaucoma. Prog Retin Eye Res. 1999; 18(5):629–667. [PubMed: 10438153]
- Gu Y, Zeng S, Qiu P, et al. Apoptosis of bovine trabecular meshwork cells induced by dexamethasone. Zhonghua Yan Ke Za Zhi. 2002; 38(5):302–304. [PubMed: 12133380]
- Zhang X, Clark AF, Yorio T. Regulation of glucocorticoid responsiveness in glaucomatous trabecular meshwork cells by glucocorticoid receptor-beta. Invest Ophthalmol Vis Sci. 2005; 46(12):4607–4616. [PubMed: 16303956]
- 40. Jain A, Wordinger RJ, Yorio T, et al. Role of the alternatively spliced glucocorticoid receptor isoform GRbeta in steroid responsiveness and glaucoma. J Ocul Pharmacol Ther. 2014; 30(2–3): 121–127. [PubMed: 24506296]
- Clark AF, Wordinger RJ. The role of steroids in outflow resistance. Exp Eye Res. 2009; 88(4):752– 759. [PubMed: 18977348]
- 42. Sharma A, Patil AJ, Mansoor S, et al. Effects of dexamethasone on human trabecular meshwork cells in vitro. Graefes Arch Clin Exp Ophthalmol. 2013; 251(7):1741–1746. [PubMed: 23640538]
- Lewis JM, Priddy T, Judd J, et al. Intraocular pressure response to topical dexamethasone as a predictor for the development of primary open-angle glaucoma. Am J Ophthalmol. 1988; 106(5): 607–612. [PubMed: 3189477]
- 44. Kempen JH, Altaweel MM, Holbrook JT, et al. Randomized comparison of systemic antiinflammatory therapy versus fluocinolone acetonide implant for intermediate, posterior, and panuveitis: the multicenter uveitis steroid treatment trial. Ophthalmology. 2011; 118(10):1916– 1926. [PubMed: 21840602]
- Friedman DS, Holbrook JT, Ansari H, et al. Risk of elevated intraocular pressure and glaucoma in patients with uveitis: results of the multicenter uveitis steroid treatment trial. Ophthalmology. 2013; 120(8):1571–1579. [PubMed: 23601801]
- 46. Sen HN, Vitale S, Gangaputra SS, et al. Periocular corticosteroid injections in uveitis: effects and complications. Ophthalmology. 2014; 121(11):2275–2286. [PubMed: 25017415]
- Black RL, Oglesby RB, Von Sallmann L, et al. Posterior subcapsular cataracts induced by corticosteroids in patients with rheumatoid arthritis. JAMA. 1960; 174:166–171. [PubMed: 13801171]
- 48. Butcher JM, Austin M, McGalliard J, et al. Bilateral cataracts and glaucoma induced by long term use of steroid eye drops. BMJ. 1994; 309(6946):43. [PubMed: 8044069]
- Urban RC Jr, Cotlier E. Corticosteroid-induced cataracts. Surv Ophthalmol. 1986; 31(2):102–110. [PubMed: 3541262]

- Costagliola C, Cati-Giovannelli B, Piccirillo A, et al. Cataracts associated with long-term topical steroids. Br J Dermatol. 1989; 120(3):472–473. [PubMed: 2713268]
- 51. Klein BE, Klein R, Lee KE, et al. Drug use and five-year incidence of age-related cataracts: The Beaver Dam Eye Study. Ophthalmology. 2001; 108(9):1670–1674. [PubMed: 11535471]
- James ER, Robertson L, Ehlert E, et al. Presence of a transcriptionally active glucocorticoid receptor alpha in lens epithelial cells. Invest Ophthalmol Vis Sci. 2003; 44(12):5269–5276. [PubMed: 14638726]
- James ER, Fresco VM, Robertson LL. Glucocorticoid-induced changes in the global gene expression of lens epithelial cells. J Ocul Pharmacol Ther. 2005; 21(1):11–27. [PubMed: 15718824]
- 54. Gupta V, Galante A, Soteropoulos P, et al. Global gene profiling reveals novel glucocorticoid induced changes in gene expression of human lens epithelial cells. Mol Vis. 2005; 11:1018–1040. [PubMed: 16319822]
- Petersen A, Carlsson T, Karlsson JO, et al. Effects of dexamethasone on human lens epithelial cells in culture. Mol Vis. 2008; 14:1344–1352. [PubMed: 18648526]
- 56. Sharma A, Pirouzmanesh A, Patil J, et al. Evaluation of the toxicity of triamcinolone acetonide and dexamethasone sodium phosphate on human lens epithelial cells (HLE B-3). J Ocul Pharmacol Ther. 2011; 27(3):265–271. [PubMed: 21574867]
- 57. James ER. The etiology of steroid cataract. J Ocul Pharmacol Ther. 2007; 23(5):403–420. [PubMed: 17900234]
- Taylor AW, Streilein JW, Cousins SW. Identification of alpha-melanocyte stimulating hormone as a potential immunosuppressive factor in aqueous humor. Curr Eye Res. 1992; 11(12):1199–1206. [PubMed: 1490338]
- Taylor AW, Alard P, Yee DG, et al. Aqueous humor induces transforming growth factor-beta (TGFbeta)-producing regulatory T-cells. Curr Eye Res. 1997; 16(9):900–908. [PubMed: 9288451]
- Kawanaka N, Taylor AW. Localized retinal neuropeptide regulation of macrophage and microglial cell functionality. Journal of Neuroimmunology. 2011; 232(1–2):17–25. [PubMed: 20965575]
- 61. Kaiser CJ, Ksander BR, Streilein JW. Inhibition of lymphocyte proliferation by aqueous humor. Regional Immunology. 1989; 2(1):42–49. [PubMed: 2534948]
- Nishida T, Taylor A. Specific aqueous humor factors induce activation of regulatory T cells. Investigative Ophthalmology & Visual Science. 1999; 40(10):2268–2274. [PubMed: 10476792]
- 63. Cousins SW, McCabe MM, Danielpour D, et al. Identification of transforming growth factor-beta as an immunosuppressive factor in aqueous humor. Invest Ophthalmol Vis Sci. 1991; 32:33–43.
- 64. Granstein R, Staszewski R, Knisely T, et al. Aqueous humor contains transforming growth factor-b and a small (<3500 daltons) inhibitor of thymocyte proliferation. Journal of Immunology. 1990; 144:3021–3027.
- 65. Jampel HD, Roche N, Stark WJ, et al. Transforming growth factor-beta in human aqueous humor. Curr Eye Res. 1990; 9(10):963–969. [PubMed: 2276273]
- 66. Namba K, Kitaichi N, Nishida T, et al. Induction of regulatory T cells by the immunomodulating cytokines alpha-melanocyte-stimulating hormone and transforming growth factor-beta2. J Leukoc Biol. 2002; 72(5):946–952. [PubMed: 12429716]
- Zmijewski MA, Sharma RK, Slominski AT. Expression of molecular equivalent of hypothalamicpituitary-adrenal axis in adult retinal pigment epithelium. The Journal of endocrinology. 2007; 193(1):157–169. [PubMed: 17400813]
- Ortego J, Coca-Prados M. Molecular characterization and differential gene induction of the neuroendocrine-specific genes neurotensin, neurotensin receptor, PC1, PC2, and 7B2 in the human ocular ciliary epithelium. J Neurochem. 1997; 69(5):1829–1839. [PubMed: 9349525]
- 69. Taylor AW, Lee DJ. The alpha-melanocyte stimulating hormone induces conversion of effector T cells into treg cells. J Transplant. 2011; 2011:246856. [PubMed: 21941624]
- 70. Nishida T, Miyata S, Itoh Y, et al. Anti-inflammatory effects of alpha-melanocyte-stimulating hormone against rat endotoxin-induced uveitis and the time course of inflammatory agents in aqueous humor. Int Immunopharmacol. 2004; 4(8):1059–1066. [PubMed: 15222980]

- 71. Shiratori K, Ohgami K, Ilieva IB, et al. Inhibition of endotoxin-induced uveitis and potentiation of cyclooxygenase-2 protein expression by alpha-melanocyte-stimulating hormone. Invest Ophthalmol Vis Sci. 2004; 45(1):159–164. [PubMed: 14691168]
- 72. Lee DJ, Taylor AW. Both MC5r and A2Ar are required for protective regulatory immunity in the spleen of post-experimental autoimmune uveitis in mice. J Immunol. 2013; 191(8):4103–4111. [PubMed: 24043903]
- 73. Lee DJ, Taylor AW. Recovery from experimental autoimmune uveitis promotes induction of antiuveitic inducible Tregs. J Leukoc Biol. 2015; 97(6):1101–1109. [PubMed: 25877928]
- 74. Li D, Taylor AW. Diminishment of alpha-MSH anti-inflammatory activity in MC1r siRNAtransfected RAW264.7 macrophages. J Leukoc Biol. 2008; 84(1):191–198. [PubMed: 18388300]
- 75. Getting SJ, Christian HC, Lam CW, et al. Redundancy of a functional melanocortin 1 receptor in the anti-inflammatory actions of melanocortin peptides: studies in the recessive yellow (e/e) mouse suggest an important role for melanocortin 3 receptor. J Immunol. 2003; 170(6):3323–3330. [PubMed: 12626592]
- Neumann Andersen G, Nagaeva O, Mandrika I, et al. MC (1) receptors are constitutively expressed on leucocyte subpopulations with antigen presenting and cytotoxic functions. Clin Exp Immunol. 2001; 126(3):441–446. [PubMed: 11737060]
- Lee DJ, Taylor AW. Following EAU recovery there is an associated MC5r-dependent APC induction of regulatory immunity in the spleen. Invest Ophthalmol Vis Sci. 2011; 52(12):8862– 8867. [PubMed: 21989727]
- Robertson MJ, Erwig LP, Liversidge J, et al. Retinal microenvironment controls resident and infiltrating macrophage function during uveoretinitis. Invest Ophthalmol Vis Sci. 2002; 43(7): 2250–2257. [PubMed: 12091424]
- 79. Dick AD, Carter D, Robertson M, et al. Control of myeloid activity during retinal inflammation. J Leukoc Biol. 2003; 74(2):161–166. [PubMed: 12885931]
- Phan TA, Taylor AW. The neuropeptides alpha-MSH and NPY modulate phagocytosis and phagolysosome activation in RAW 264.7 cells. J Neuroimmunol. 2013; 260(1–2):9–16. [PubMed: 23689030]
- 81. Taylor AW, Dixit S, Yu J. Retinal Pigment Epithelial Cell Line Suppression of Phagolysosome Activation. Int J Ophthalmol Eye Sci. 2015; (Suppl 2(1)):1–6. [PubMed: 25905107]
- 82. Taylor AW. Alpha-melanocyte stimulating hormone (alpha-MSH) is a post-caspase suppressor of apoptosis in RAW 264.7 macrophages. PLoS One. 2013; 8(8):e74488. [PubMed: 24009773]
- Caspi RR, Roberge FG, Chan CC, et al. A new model of autoimmune disease. Experimental autoimmune uveoretinitis induced in mice with two different retinal antigens. J Immunol. 1988; 140(5):1490–1495. [PubMed: 3346541]
- Kitaichi N, Namba K, Taylor A. Inducible immune regulation following autoimmune disease in the immune-privileged eye. Journal of Leukocyte Biology. 2005; 77(4):496–502. [PubMed: 15647326]
- Taylor AW, Kitaichi N. The diminishment of experimental autoimmune encephalomyelitis (EAE) by neuropeptide alpha-melanocyte stimulating hormone (alpha-MSH) therapy. Brain Behav Immun. 2008; 22(5):639–646. [PubMed: 18171609]
- 86. Yin P, Luby TM, Chen H, et al. Generation of expression constructs that secrete bioactive alphaMSH and their use in the treatment of experimental autoimmune encephalomyelitis. Gene Ther. 2003; 10(4):348–355. [PubMed: 12595893]
- Ng TF, Kitaichi N, Taylor AW. In vitro-generated autoimmune regulatory T cells enhance intravitreous allogeneic retinal graft survival. Investigative Ophthalmology & Visual Science. 2007; 48(11):5112–5117. [PubMed: 17962463]
- 88. Cheng LB, Cheng L, Bi HE, et al. Alpha-melanocyte stimulating hormone protects retinal pigment epithelium cells from oxidative stress through activation of melanocortin 1 receptor-Akt-mTOR signaling. Biochem Biophys Res Commun. 2014; 443(2):447–452. [PubMed: 24316214]
- Naveh N. Melanocortins applied intravitreally delay retinal dystrophy in Royal College of Surgeons rats. Graefes Arch Clin Exp Ophthalmol. 2003; 241(12):1044–1050. [PubMed: 14586589]

- Steffensen EH. Corticotropin, cortisone, and hydrocortisone in treatment of ocular disease. J Am Med Assoc. 1952; 150(17):1660–1664. [PubMed: 12999486]
- Takakura A, Tessler HH, Goldstein DA, et al. Viral retinitis following intraocular or periocular corticosteroid administration: a case series and comprehensive review of the literature. Ocul Immunol Inflamm. 2014; 22(3):175–182. [PubMed: 24655372]
- Bouzas EA, Karadimas P, Pournaras CJ. Central serous chorioretinopathy and glucocorticoids. Surv Ophthalmol. 2002; 47(5):431–448. [PubMed: 12431693]
- Karadimas P, Bouzas EA. Glucocorticoid use represents a risk factor for central serous chorioretinopathy: a prospective, case-control study. Graefes Arch Clin Exp Ophthalmol. 2004; 242(9):800–802. [PubMed: 14986014]
- Wang M, Munch IC, Hasler PW, et al. Central serous chorioretinopathy. Acta Ophthalmol. 2008; 86(2):126–145. [PubMed: 17662099]
- 95. Cunningham ET Jr, Wender JD. Practical approach to the use of corticosteroids in patients with uveitis. Can J Ophthalmol. 2010; 45(4):352–358. [PubMed: 20648092]
- McGhee CN, Dean S, Danesh-Meyer H. Locally administered ocular corticosteroids: benefits and risks. Drug Saf. 2002; 25(1):33–55. [PubMed: 11820911]
- Taylor AW. Ocular immunosuppressive microenvironment. Chem Immunol Allergy. 2007; 92:71– 85. [PubMed: 17264484]
- Vignali DA, Collison LW, Workman CJ. How regulatory T cells work. Nat Rev Immunol. 2008; 8(7):523–532. [PubMed: 18566595]
- 99. Taylor AW, Streilein JW. Inhibition of antigen-stimulated effector T cells by human cerebrospinal fluid. Neuroimmunomodulation. 1996; 3(2–3):112–118. [PubMed: 8945726]
- 100. Wong KY, Rajora N, Boccoli G, et al. A potential mechanism of local anti-inflammatory action of alpha-melanocyte-stimulating hormone within the brain: modulation of tumor necrosis factoralpha production by human astrocytic cells. Neuroimmunomodulation. 1997; 4(1):37–41. [PubMed: 9326743]
- 101. Bhardwaj RS, Schwarz A, Becher E, et al. Pro-opiomelanocortin-derived peptides induce IL-10 production in human monocytes. J Immunol. 1996; 156(7):2517–2521. [PubMed: 8786313]
- 102. Bohm M, Wolff I, Scholzen TE, et al. alpha-Melanocyte-stimulating hormone protects from ultraviolet radiation-induced apoptosis and DNA damage. J Biol Chem. 2005; 280(7):5795–5802. [PubMed: 15569680]
- 103. Chai B, Li JY, Zhang W, et al. Melanocortin-4 receptor-mediated inhibition of apoptosis in immortalized hypothalamic neurons via mitogen-activated protein kinase. Peptides. 2006; 27(11): 2846–2857. [PubMed: 16806584]
- 104. Zhang L, Dong L, Liu X, et al. alpha-Melanocyte-stimulating hormone protects retinal vascular endothelial cells from oxidative stress and apoptosis in a rat model of diabetes. PLoS One. 2014; 9(4):e93433. [PubMed: 24695675]
- 105. Edling AE, Gomes D, Weeden T, et al. Immunosuppressive activity of a novel peptide analog of alpha-melanocyte stimulating hormone (alpha-MSH) in experimental autoimmune uveitis. J Neuroimmunol. 2011; 236(1–2):1–9. [PubMed: 21640392]
- 106. Naveh N, Kaplan-Messas A, Marshall J. Mechanism related to reduction of intraocular pressure by melanocortins in rabbits. Br J Ophthalmol. 2000; 84(12):1411–1414. [PubMed: 11090484]
- Catania A. Neuroprotective actions of melanocortins: a therapeutic opportunity. Trends Neurosci. 2008; 31(7):353–360. [PubMed: 18550183]
- 108. Joosten EA, Majewska B, Houweling DA, et al. Alpha-melanocyte stimulating hormone promotes regrowth of injured axons in the adult rat spinal cord. J Neurotrauma. 1999; 16(6):543–553. [PubMed: 10391370]
- 109. Bharne AP, Upadhya MA, Kokare DM, et al. Effect of alpha-melanocyte stimulating hormone on locomotor recovery following spinal cord injury in mice: Role of serotonergic system. Neuropeptides. 2011; 45(1):25–31. [PubMed: 21036396]
- 110. Schaible EV, Steinstrasser A, Jahn-Eimermacher A, et al. Single administration of tripeptide alpha-MSH(11–13) attenuates brain damage by reduced inflammation and apoptosis after experimental traumatic brain injury in mice. PLoS One. 2013; 8(8):e71056. [PubMed: 23940690]

- 111. Giuliani D, Galantucci M, Neri L, et al. Melanocortins protect against brain damage and counteract cognitive decline in a transgenic mouse model of moderate Alzheimers disease. Eur J Pharmacol. 2014; 740:144–150. [PubMed: 25034807]
- 112. Giuliani D, Bitto A, Galantucci M, et al. Melanocortins protect against progression of Alzheimer's disease in triple-transgenic mice by targeting multiple pathophysiological pathways. Neurobiol Aging. 2014; 35(3):537–547. [PubMed: 24094579]
- 113. Ma K, McLaurin J. alpha-Melanocyte stimulating hormone prevents GABAergic neuronal loss and improves cognitive function in Alzheimer's disease. J Neurosci. 2014; 34(20):6736–6745. [PubMed: 24828629]
- 114. Fang J, Han D, Hong J, et al. SValpha-MSH, a novel alpha-melanocyte stimulating hormone analog, ameliorates autoimmune encephalomyelitis through inhibiting auto-reactive CD4(+) T cells activation. J Neuroimmunol. 2014; 269(1–2):9–19. [PubMed: 24518673]
- 115. Diem R, Hobom M, Maier K, et al. Methylprednisolone increases neuronal apoptosis during autoimmune CNS inflammation by inhibition of an endogenous neuroprotective pathway. J Neurosci. 2003; 23(18):6993–7000. [PubMed: 12904460]
- Miller H, Newell DJ, Ridley A. Multiple sclerosis. Treatment of acute exacerbations with corticotrophin (A. C.T.H.). Lancet. 1961; 2(7212):1120–1122. [PubMed: 14474011]
- 117. Arnason BG, Berkovich R, Catania A, et al. Mechanisms of action of adrenocorticotropic hormone and other melanocortins relevant to the clinical management of patients with multiple sclerosis. Mult Scler. 2013; 19(2):130–136. [PubMed: 23034287]
- Weinstein RS. Clinical practice. Glucocorticoid-induced bone disease. N Engl J Med. 2011; 365(1):62–70. [PubMed: 21732837]
- 119. Besser GM, Butler PW, Plumpton FS. Adrenocorticotrophic action of long-acting tetracosactrin compared with corticotrophin-gel. Br Med J. 1967; 4(5576):391–394. [PubMed: 4293374]
- 120. Coburg AJ, Gray SH, Katz FH, et al. Disappearance rates and immunosuppression of intermittent intravenously administered prednisolone in rabbits and human beings. Surg Gynecol Obstet. 1970; 131(5):933–942. [PubMed: 4919384]
- 121. Zaidi M, Sun L, Robinson LJ, et al. ACTH protects against glucocorticoid-induced osteonecrosis of bone. Proc Natl Acad Sci U S A. 2010; 107(19):8782–8787. [PubMed: 20421485]
- 122. Grieco P, Carotenuto A, Auriemma L, et al. Novel alpha-MSH peptide analogues with broad spectrum antimicrobial activity. PLoS One. 2013; 8(4):e61614. [PubMed: 23626703]
- 123. Singh M, Mukhopadhyay K. Alpha-melanocyte stimulating hormone: an emerging antiinflammatory antimicrobial peptide. Biomed Res Int. 2014; 2014:874610. [PubMed: 25140322]
- 124. Holdeman M, Lipton JM. Effects of massive doses of alpha-MSH on thermoregulation in the rabbit. Brain Res Bull. 1985; 14(4):327–330. [PubMed: 3873978]
- 125. Ugwu SO, Blanchard J, Dorr RT, et al. Skin pigmentation and pharmacokinetics of melanotan-I in humans. Biopharm Drug Dispos. 1997; 18(3):259–269. [PubMed: 9113347]
- 126. Dorr RT, Lines R, Levine N, et al. Evaluation of melanotan-II, a superpotent cyclic melanotropic peptide in a pilot phase-I clinical study. Life Sci. 1996; 58(20):1777–1784. [PubMed: 8637402]
- 127. Dorr RT, Dvorakova K, Brooks C, et al. Increased eumelanin expression and tanning is induced by a superpotent melanotropin [Nle4-D-Phe7]-alpha-MSH in humans. Photochem Photobiol. 2000; 72(4):526–532. [PubMed: 11045725]
- 128. Dorr RT, Ertl G, Levine N, et al. Effects of a superpotent melanotropic peptide in combination with solar UV radiation on tanning of the skin in human volunteers. Arch Dermatol. 2004; 140(7):827–835. [PubMed: 15262693]
- 129. Royalty JE, Konradsen G, Eskerod O, et al. Investigation of safety, tolerability, pharmacokinetics, and pharmacodynamics of single and multiple doses of a long-acting alpha-MSH analog in healthy overweight and obese subjects. J Clin Pharmacol. 2014; 54(4):394–404. [PubMed: 24166760]
- 130. Safarinejad MR. Evaluation of the safety and efficacy of bremelanotide, a melanocortin receptor agonist, in female subjects with arousal disorder: a double-blind placebo-controlled, fixed dose, randomized study. J Sex Med. 2008; 5(4):887–897. [PubMed: 18179455]
- 131. Rosen RC, Diamond LE, Earle DC, et al. Evaluation of the safety, pharmacokinetics and pharmacodynamic effects of subcutaneously administered PT-141, a melanocortin receptor

agonist, in healthy male subjects and in patients with an inadequate response to Viagra. Int J Impot Res. 2004; 16(2):135–142. [PubMed: 14999221]

- Ceriani G, Diaz J, Murphree S, et al. The neuropeptide alpha-melanocyte-stimulating hormone inhibits experimental arthritis in rats. Neuroimmunomodulation. 1994; 1(1):28–32. [PubMed: 8528882]
- Botte DA, Noronha IL, Malheiros DM, et al. Alpha-melanocyte stimulating hormone ameliorates disease activity in an induced murine lupus-like model. Clin Exp Immunol. 2014; 177(2):381– 390. [PubMed: 24666423]
- 134. Rajora N, Boccoli G, Catania A, et al. alpha-MSH modulates experimental inflammatory bowel disease. Peptides. 1997; 18(3):381–385. [PubMed: 9145424]
- 135. Fiechtner J, Montroy T. Treatment of moderately to severely active systemic lupus erythematosus with adrenocorticotropic hormone: a single-site, open-label trial. Lupus. 2014; 23(9):905–912. [PubMed: 24795067]
- 136. Levine T. Treating refractory dermatomyositis or polymyositis with adrenocorticotropic hormone gel: a retrospective case series. Drug Des Devel Ther. 2012; 6:133–139.
- 137. Bomback AS, Tumlin JA, Baranski J, et al. Treatment of nephrotic syndrome with adrenocorticotropic hormone (ACTH) gel. Drug Des Devel Ther. 2011; 5:147–153.
- 138. Bomback AS, Canetta PA, Beck LH Jr, et al. Treatment of resistant glomerular diseases with adrenocorticotropic hormone gel: a prospective trial. Am J Nephrol. 2012; 36(1):58–67. [PubMed: 22722778]
- Baram TZ, Mitchell WG, Tournay A, et al. High-dose corticotropin (ACTH) versus prednisone for infantile spasms: a prospective, randomized, blinded study. Pediatrics. 1996; 97(3):375–379. [PubMed: 8604274]



FIGURE 1.

Effects of different melanocortin receptors on immune activity. Through MC1r, MC3r, and MC5r, α-MSH suppresses inflammation and promotes the activation of anti-inflammatory activity and regulatory immunity. In macrophages, through MC1r and MC3r, α-MSH mediates alternative activation by which the macrophages suppress inflammation.^{74,75} Through MC5r, α-MSH mediates the activation of suppressor cell activity in macrophages that suppresses effector T cell activation and viability.^{58,60} Through MC5r on T cells, α-MSH promotes the activation of regulatory T cell activity and the suppression of effector T cells.^{26,69} In addition, through MC5r, α-MSH induces antigen presenting cells to promote activation inducible Treg cells^{72, 77} (see Figure 2). Ag, antigen; LPS, lipopolysaccharide; MØ, macrophage.





FIGURE 2.

Melanocortin-driven induction of autoantigen-specific protective immunity. Self-resolution or α -MSH treatment of experimental autoimmune uveitis in mice is followed by the induction of autoantigen-specific Treg cells that provide resistance to the recurrence of uveitis.^{72,84} This is driven by the action of α -MSH, through MC5r on macrophages, to induce antigen presenting activity that counter-converts autoantigen-specific effector T cells into inducible Treg cells.^{2,77} α -MSH upregulates APC generation of adenosine and expression of PD-L1.⁷² This process requires autoantigen-specific effector T cells expressing the adenosine 2A receptor.⁷² Ag, antigen, CD39, ecto-nucleoside triphosphate diphosphohydrolase 1; CD73, ecto-5'-nucleotidase; MHC II, major histocompatibility class II antigen; TCR, T-cell receptor.



FIGURE 3.

Effects of α -MSH on uveitis and retinal cell health. Therapeutic application of α -MSH has the potential to suppress inflammation, induce immune tolerance, and promote retinal cell survival (see text). α -MSH can directly influence immune cells through their expression of melanocortin receptors to suppress activation of effector T cells (Th1, Th17 cells), while promoting iTreg cell activation. In macrophages, α -MSH suppresses production of proinflammatory cytokines and promotes antigen presenting activity that converts effector T cells into functional iTreg cells. In addition, RPE and photoreceptors express melanocortin receptors through which α -MSH promotes cell survival. EC, endothelial cell; IL-1 β , -6, -12; interleukin-1beta, -6, -12; iTreg: inducible Treg cells; RPE, retinal pigment epithelial cells; PR, photoreceptor cell; ROS, reactive oxygen species; Th: T helper cells; TNF α ; tumor necrosis factor-alpha; VEGF, vascular endothelial growth factor.

TABLE 1

Effects of glucocorticoids and $\alpha\text{-MSH}$ on the eye.

	Glucocorticoid	a–MSH
Immune effects	 Suppresses proinflammatory signals Suppresses immune cell activity 	 Suppresses proinflammatory signals Promotes production of anti-inflammatory cytokines Induces suppressor antigen presenting cells Induces the activation of regulatory T cells Natural regulator of immunity in healthy eyes
Optic nerve effects	 Indirect damage of optic nerve due to increased intraocular pressure May sensitize nerve cells to disease-induced apoptosis 	Transiently lowers intraocular pressureCan protect neuronal cells from damage
Adverse effects	 Causes cataracts Induces increased intraocular pressure and glaucoma Many other adverse effects 	No known adverse effects